One of the fastest growing populations suffering from HIV infection is women in their reproductive years (1). The number of HIV infections in the pediatric population is also increasing at an alarming rate (2). HIV infection in children can occur in several ways, but the dominant route is by vertical transmission from an HIV-infected mother to her infant (3). This vertical viral transmission may occur early or late in pregnancy, during birth, or postnatally through breast feeding (4). Although the fetus can be infected with the HIV virus during gestation, only ~30% of children born to HIV-infected mothers are documented HIV-positive in the first several years of life (5). Treatment of HIV-infected pregnant women with anti-HIV therapeutics may, therefore, inadvertently expose many uninfected and healthy fetuses to these maternally administered and potentially toxic drugs.

The majority of agents approved for the treatment of AIDS and infection with HIV are nucleoside analog reverse-transcriptase inhibitors, which include AZT. AZT has recently been shown to reduce the vertical transmission of HIV from mother to fetus (6), but little is known about its fetal tissue distribution and metabolism. AZT is a deoxynucleoside with structural similarities to endogenous nucleosides. The mechanism of anti-HIV activity of AZT is believed to involve the inhibition of HIV reverse transcriptase and DNA chain elongation by incorporation of AZT-TP into the viral DNA (7). The toxicity of AZT may also be due to the interaction of AZT-TP with mammalian DNA polymerases β and γ (mitochondrial) (8, 9). These triphosphates are potent inhibitors of reverse transcriptase, and differences in potency are primarily due to differences in metabolism to the active form (10). Although these compounds are efficacious in both adult and pediatric HIV-infected populations (11, 12), and reduce the risk of vertical transmission during pregnancy (6), they are associated with serious dose-limiting side effects. The major dose-limiting toxicity of AZT is the inhibition of the development of bone marrow cells resulting in anemia and neutropenia (13).

Our laboratory has previously demonstrated that the anti-HIV nucleosides ddC and 2’,3’-dideoxyinosine reach the fetal circulation...
after maternal intravenous administration, but the active triphosphates of these agents were not detectable in fetal tissues 3 hr after maternal dosing (14). The rhesus monkey’s placental structure and function, as well as pharmacokinetic similarities to humans, make this an appropriate model for examining AZT distribution during pregnancy (15, 16). The purpose of the present study was to examine fetal exposure to AZT by determining the placental transfer and fetal distribution of the parent compound and its glucuronidated and phosphorylated metabolites in the late-term rhesus monkey. This knowledge will provide a foundation for subsequent estimations concerning both the efficacy and the potential toxicity of these agents.

Methods

Animals. Four pregnant rhesus macaques (Macaca mulatta) were obtained from the NCTR nonhuman primate colony. Animals were maintained in accordance with the National Institute of Health Guide for the Care and Use of Laboratory Animals, and experimental procedures were approved by the NCTR’s Institutional Animal Care and Use Committee. Food (Purina Hi Protein Monkey Chow,Ralston Purina, St. Louis, MO) supplemented with fresh fruit and chewable multiple vitamins with iron (Arkansas Cooperative Assoc., Inc., North Little Rock, AR) was provided daily, and water was available ad libitum. The gestational age at the time of surgery was 150 ± 5 days (term = 165 days).

Chemicals. AZT, AZTG, and AZddU were purchased from Sigma Chemical Co. (St. Louis, MO). AZT- DP, AZT-TP, [3H]-AZT, [methyl-3H]-3’-azido-3’-deoxythymidine 5’-monophosphate (diammonium salt) and [methyl-3H]-3’-azido-3’-deoxythymidine 5’-triphosphate (tetraammonium salt) were purchased from Moravek Biochemicals (Brea, CA).

Pharmacokinetics. Two weeks before surgery, a pharmacokinetic study was performed to determine the dose of AZT needed to reach steady-state plasma concentrations in the mother. Approximately 2 weeks before this kinetic study (1 month before surgery), animals were acclimated to sitting in a restraint chair. On the day of the kinetic study, an intravenous bolus of 8 mg/kg AZT was administered to the awake animal while chair-restrained. Venous blood samples were collected beginning 2 min after the bolus dose for up to 3.5 hr, and immediately processed and analyzed for AZT and AZTG using reverse-phase HPLC.

Infusion and Surgery. On the day of the surgery, the awake mother was administered an intravenous loading dose of AZT (1.2–2.3 mg/kg), including 40–60 µCi of [3H]-AZT, followed by at least a 3-hr steady-state infusion (via maternal radial vein) of AZT (30–50 µg/min/kg, 15 ml/hr; Sage Instruments syringe pump model 351) and an additional 200 µCi of [3H]-AZT. Maternal venous blood samples were collected every 30 min after the start of the infusion. Approximately 2.5 hr after infusion, anesthesia was induced with 10 mg/kg in ketamine hydrochloride (Ketaset, Fort Dodge Laboratories, Fort Dodge, IA), and at 3 hr a mixture of 1% halothane, 20–30% N2O, and oxygen was delivered via an endotracheal tube to maintain general anesthesia. When stable surgical anesthesia was achieved, the fetus was delivered via hysterotomy and exsanguinated. Amniotic fluid and maternal blood (via uterine vein) were also collected. The mother was allowed to recover and returned to the colony. Plasma, amniotic fluid, and fetal tissues were frozen at −70°C until analysis by HPLC.

Sample Analysis. Plasma and amniotic fluid were analyzed for AZT and AZTG by solid-phase extraction, followed by reverse-phase HPLC according to a modification of the method of Qian et al. (17). AZddU (750 ng) and water (200 µl) were added to 100 µl plasma and the mixture applied to a BondElut column (1 ml column/50 mg solid phase; Varian, Harbor City, CA). The column was rinsed with 1 ml of 0.01 M phosphate buffer (pH 2.0) and the compounds eluted with 300 µl methanol. Methanol was evaporated to dryness and the compounds reconstituted in 200 µl of mobile phase. An aliquot of the samples were injected onto a Hypersil ODS column (5 µm, 150 × 4.6 mm; Alltech Associates, Inc., Deerfield, IL) using a Waters 178 plus Autosampler (Waters Corp., Milford, MA). Elution of the compounds was achieved by using a mobile phase that consisted of a mixture of acetonitrile:water (7:93) adjusted to pH 2.5 with phosphoric acid and a flow rate of 2 ml/min. Compounds were analyzed and quantitated at 267 nm using a Waters 486 Tunable Absorbance Detector linked to a computer workstation with Millen-
The majority of the radioactivity in the kidney (62%) and plasma (54%) was due to the glucuronidated metabolite AZTG, and yielded estimates of 14.34 nmol/g and 6.78 nmol/ml, respectively. In all of the other tissues, the majority of radioactivity (48–100%) was associated with the parent compound (yielding estimates 2.35–7.53 nmol/g). AZT-MP was the only phosphorylated metabolite detected in any of the tissues examined (0.613–5.913 nmol/g) and was the major metabolite (88% of total metabolites) in the spleen (5.913 nmol/g). No significant radioactivity eluted near AZT-DP or AZT-TP (fig. 5). The plasma did not contain any measurable amounts of AZT-MP. Brain regions examined contained only parent compound, except for the cerebellum, which consistently contained a small amount (0.613 nmol/g) of AZT-MP and one frontal cortex sample that also contained AZT-MP (0.3 nmol/g).

Discussion
Pharmacokinetic parameters determined from either the single dose intravenous administration or the steady-state infusion of AZT were similar to data for humans and other nonhuman primates reported by others. The values for clearance (21 ml/min/kg) and volume of distribution (0.8 liters/kg) in the present study compare favorably with those for the pregnant pigtailed macaque (Macaca nemestrina) (CL: 455

![Structure of AZT and its major metabolites.](image)

Fig. 1. Structure of AZT and its major metabolites.

![Plasma concentration-time profiles for AZT and AZTG decay in parallel in pregnant rhesus macaques after an intravenous bolus dose of 8 mg/kg AZT.](image)

Fig. 2. Plasma concentration-time profiles for AZT and AZTG decay in parallel in pregnant rhesus macaques after an intravenous bolus dose of 8 mg/kg AZT.

Venous blood samples were collected beginning 2 min after the dose for up to 3.5 hr (see table 1). Values are expressed as μg AZT or AZTG/ml plasma (means ± SD) for four animals.
Late-term maternal rhesus monkeys were administered 8 mg/kg AZT iv, and venous blood samples were collected beginning 2 min after administration for up to 3.5 hr.

<table>
<thead>
<tr>
<th>Animal ID No.</th>
<th>Weight</th>
<th>CL</th>
<th>Vdss</th>
<th>t1/2</th>
</tr>
</thead>
<tbody>
<tr>
<td>R4707</td>
<td>9.8</td>
<td>22.42</td>
<td>0.73</td>
<td>31.00</td>
</tr>
<tr>
<td>E480</td>
<td>9.0</td>
<td>15.21</td>
<td>0.64</td>
<td>34.86</td>
</tr>
<tr>
<td>R21</td>
<td>12.8</td>
<td>25.02</td>
<td>1.18</td>
<td>43.66</td>
</tr>
<tr>
<td>R4912</td>
<td>9.0</td>
<td>20.51</td>
<td>0.74</td>
<td>33.10</td>
</tr>
<tr>
<td>Mean</td>
<td></td>
<td>20.79</td>
<td>0.82</td>
<td>35.07</td>
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<tr>
<td>± SD</td>
<td></td>
<td>±4.15</td>
<td>±0.24</td>
<td>±4.82*</td>
</tr>
</tbody>
</table>

*Harmonic mean ± pseudo-SD.

23 ml/min/kg; Vdss: 0.8 liters/kg) (19) and in pregnant women (CL: 26 ml/min/kg; Vdss: 1.2 liters/kg) (20). The current recommended intrapartum AZT regimen for HIV-positive pregnant women is a 2 mg/kg/hr (33 µg/min/kg) loading dose during the first hour of labor, followed by 1 mg/kg/hr (17 µg/min/kg) for the duration of labor and delivery (6). In the present study, steady-state infusions of 30–50 µg/min/kg in pregnant rhesus monkeys resulted in maternal steady-state concentrations approaching 1.7 µg/ml. Plasma concentrations achieved herein are, therefore, at or slightly above those predicted by the pharmacokinetics and dosing for pregnant women during labor and delivery (1–1.5 µg/ml).

Very few studies to date have examined fetal-to-maternal plasma concentration ratios for AZT in humans at steady-state. A single case study in which AZT was administered via continuous intravenous infusion (0.12 mg/kg/hr) for 24 hr before delivery reported a cord blood-to-maternal blood ratio of 1.68 (21). Clinical fetal-to-maternal blood ratios of 0.76–2.50 have, however, been reported after single intravenous doses of AZT with single time point collections at delivery (20). In the present study, fetal-to-maternal AZT plasma concentration ratio averaged 0.85, which is nearly identical to the ratio reported for the pigtailed macaque at steady-state (mean of 0.83) (22). It has been suggested that AZT does not accumulate in the fetus and that placental transfer of AZT occurs rapidly by passive diffusion (22, 23). The fetal plasma-to-amniotic fluid concentration ratio of AZT was near unity. This ratio was expected because AZT concentrations in the amniotic fluid and fetal plasma should achieve equilibrium relatively quickly because the fetus swallows amniotic fluid; AZT is readily absorbed through the gastrointestinal tract (24), after which it is excreted in the urine.

Concentrations of AZT and AZTG in both the fetal plasma and amniotic fluid at the time of the hysterotomy were also included. Ketamine was administered at ~2.5 hr, and gaseous anesthesia began at ~3 hr. Values are expressed as µg AZT or AZTG/ml plasma.

It should be noted that an increase in the maternal concentration of AZT and AZTG was observed toward the end of the infusion. This was likely due to the induction of the anesthesia. A nearly 2-fold increase in the maternal levels of AZT and AZTG was observed in the pigtailed macaque after continuous AZT infusion, within the first hour after the initiation of anesthesia (22). Halothane and other volatile anesthetics have been shown to inhibit metabolism of drugs (25, 26), which may have affected the clearance of AZT and AZTG. In the present study, the hysterotomy was typically performed in <1 hr after the initiation of anesthesia, so the increase in maternal plasma AZT and AZTG levels was minimal. Also, the magnitude of the effect of the anesthesia may be smaller in the rhesus than in the pigtailed macaque.

Similar to studies that examined AZT pharmacokinetics during human pregnancy (27–29), AZTG was also found to be the major metabolite of AZT in the rhesus monkey. A primary difference

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**TABLE 1**

AZT pharmacokinetic parameters in the late-term maternal rhesus monkey

Late-term maternal rhesus monkeys were administered 8 mg/kg AZT iv, and venous blood samples were collected beginning 2 min after administration for up to 3.5 hr.

<table>
<thead>
<tr>
<th>Animal ID No.</th>
<th>Weight</th>
<th>CL</th>
<th>Vdss</th>
<th>t1/2</th>
</tr>
</thead>
<tbody>
<tr>
<td>R4707</td>
<td>9.8</td>
<td>22.42</td>
<td>0.73</td>
<td>31.00</td>
</tr>
<tr>
<td>E480</td>
<td>9.0</td>
<td>15.21</td>
<td>0.64</td>
<td>34.86</td>
</tr>
<tr>
<td>R21</td>
<td>12.8</td>
<td>25.02</td>
<td>1.18</td>
<td>43.66</td>
</tr>
<tr>
<td>R4912</td>
<td>9.0</td>
<td>20.51</td>
<td>0.74</td>
<td>33.10</td>
</tr>
<tr>
<td>Mean</td>
<td></td>
<td>20.79</td>
<td>0.82</td>
<td>35.07</td>
</tr>
<tr>
<td>± SD</td>
<td></td>
<td>±4.15</td>
<td>±0.24</td>
<td>±4.82*</td>
</tr>
</tbody>
</table>

*Harmonic mean ± pseudo-SD.

23 ml/min/kg; Vdss: 0.8 liters/kg) (19) and in pregnant women (CL: 26 ml/min/kg; Vdss: 1.2 liters/kg) (20). The current recommended intrapartum AZT regimen for HIV-positive pregnant women is a 2 mg/kg/hr (33 µg/min/kg) loading dose during the first hour of labor, followed by 1 mg/kg/hr (17 µg/min/kg) for the duration of labor and delivery (6). In the present study, steady-state infusions of 30–50 µg/min/kg in pregnant rhesus monkeys resulted in maternal steady-state concentrations approaching 1.7 µg/ml. Plasma concentrations achieved herein are, therefore, at or slightly above those predicted by the pharmacokinetics and dosing for pregnant women during labor and delivery (1–1.5 µg/ml).

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Similar to studies that examined AZT pharmacokinetics during human pregnancy (27–29), AZTG was also found to be the major metabolite of AZT in the rhesus monkey. A primary difference

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**TABLE 2**

AZT infusion pharmacokinetic parameters

Maternal plasma, fetal plasma, and amniotic fluid were collected and analyzed using reversed-phase HPLC. Cpssm maternal steady-state plasma concentration; Cpool/Cpssm fetal-to-maternal AZT plasma ratios at steady-state; Cpool/AFss fetal plasma-to-amniotic fluid ratios at steady-state.

<table>
<thead>
<tr>
<th>Animal ID No.</th>
<th>Weight</th>
<th>Infusion Ratea</th>
<th>IV Bolusa</th>
<th>Cpssm</th>
<th>CL</th>
<th>Vdss</th>
<th>t1/2</th>
<th>Cpool/Cpssm</th>
<th>Cpool/AFss</th>
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<td>1.466</td>
<td>1.529</td>
<td>29.29</td>
<td>0.96</td>
<td>0.867</td>
<td>1.048</td>
<td></td>
</tr>
<tr>
<td>E480</td>
<td>8.9</td>
<td>30.42</td>
<td>1.281</td>
<td>1.296</td>
<td>23.47</td>
<td>0.99</td>
<td>0.634</td>
<td>1.035</td>
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<tr>
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<td>12.0</td>
<td>50.04</td>
<td>2.256</td>
<td>2.151</td>
<td>23.26</td>
<td>1.10</td>
<td>0.883</td>
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<td>R4912</td>
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<td>41.02</td>
<td>1.476</td>
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<td>0.995</td>
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<tr>
<td>Mean</td>
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<td>1.68</td>
<td>24.90</td>
<td>0.97</td>
<td>0.845</td>
<td>1.187</td>
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<td></td>
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<tr>
<td>± SD</td>
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<td>±0.152</td>
<td>±0.192</td>
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</table>

*Based on the calculations using data from table 1 and 2 µg/ml as the target steady-state plasma concentration.
AZT concentrations in plasma and amniotic fluid

Maternal plasma, fetal plasma, and amniotic fluid were collected and analyzed using reversed-phase HPLC. $C_{\text{pss}}$, maternal plasma concentrations at steady-state; $C_{\text{pss}}$, fetal plasma concentrations at steady-state; $AF_{\text{ss}}$, steady-state concentrations in amniotic fluid.

<table>
<thead>
<tr>
<th>Animal ID No.</th>
<th>$C_{\text{pss}}$</th>
<th>$C_{\text{pss}}$</th>
<th>$C_{\text{pss}}$</th>
<th>Concentration Ratios</th>
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<td>$C_{\text{pss}}/C_{\text{pss}}$</td>
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<td>Mean</td>
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<tr>
<td>$\pm$ SD</td>
<td></td>
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<td></td>
<td>$\pm$0.107</td>
</tr>
</tbody>
</table>

Values are expressed as AZT equivalents ng/g or ng/ml (means $\pm$ SD; n = 4). AF, amniotic fluid; Sm Int, small intestine; Sk Musc, skeletal muscle; CB, cerebellum; Thal, thalamus; CN, caudate nucleus; Hypo, hypothalamus; BS, brainstem; Hipo, hippocampus; PaPCG, parietal cortex plus precentral gyrus; PrCPG, premotor cortex plus postcentral gyrus; FC, frontal cortex; ROB, rest of brain; OC, occipital cortex.

between the findings in this study and those from a study in pigtailed macaques (19) was the relative amount of AZT glucuronidation. At steady-state, the AZTG:AZT plasma ratio of 0.24 in children after a continuous intravenous administration of AZT (12). In the present study, the cerebellum,
AZT-MP. However, the specific activity of the radiolabel used in the transcriptase (0.1 mCi) and fetal spleen AZT-MP levels ranged from 1 to 10 nmol/g to generate enough AZT-TP to allow detection. Because in the present study would not permit this low concentration of AZT-TP to be detected.

Because significant toxicity has occurred in adults during the course of AZT treatment (13, 34), the possibility for long-term toxicity in children exposed prenatally to AZT is of concern. There are indications from other animal studies that AZT exposure before or early in pregnancy could negatively impact pregnancy outcome. In mice, AZT exposure during gestation was associated with a reduction in viable offspring (35, 36). Pigtailed macaques, prenatally exposed to AZT, exhibited a lower postnatal weight gain and slower acquisition of a Black-White Learning discrimination task (37), and rhesus macaques treated with AZT during the second and third trimesters had increased numbers of abortions, fetal/neonatal deaths, and premature deliveries (38). These could result from both the direct and indirect effects of AZT during development. Infants in the AIDS Clinical Trials Group (protocol 076) study tolerated acute AZT treatment rather well (6), with a mild, transient anemia being the most frequent adverse effect. No AZT exposures occurred, however, before gestational week 14.

In summary, the intravenous kinetics of AZT in Macaca mulatta are similar to those reported for humans and other nonhuman primates. AZT, when administered intravenously was rapidly converted to its major metabolite, the glucuronide AZTG. When infused intravenously over a 3-hr period, AZT readily crossed the placenta and was found in fetal plasma, and in all fetal tissues examined as either parent compound, the glucuronide, or monophosphate metabolite. The AZT fetal-to-maternal plasma concentration ratio of 0.85 suggests that AZT does not accumulate in the fetus. In placenta and spleen, AZT-MP concentrations equaled or exceeded AZTG concentrations. Although fetal plasma AZT concentrations obtained in the present study were similar to peak plasma AZT concentrations observed clinically, the putative active antiviral metabolite, AZT-TP, was not detected in any monkey fetal tissue. However, based on the amount of the monophosphorylated metabolite observed in some tissues, clinically relevant concentrations of the triphosphate metabolite may still have been present.

Acknowledgments. We thank John R. Johnson for his technical assistance during surgery. We also thank Mrs. Betty White and the primate colony staff at the NCTR for their excellent care and handling of the monkeys, and for their assistance during the study.

References


