INHIBITION OF HUMAN DRUG METABOLIZING CYTOCHROMES P450 BY ANASTROZOLE, A POTENT AND SELECTIVE INHIBITOR OF AROMATASE

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Abstract:
Anastrozole (2,2'[5-(1H-1,2,4-triazol-1-ylmethyl)-1,3-phenylene]-bis[2-methylpropionitrile]) is a potent third-generation inhibitor of aromatase, currently marketed as a treatment for postmenopausal women with advanced breast cancer. While its potency and selectivity for inhibition of estrogen synthesis has been established in both preclinical and clinical studies, this study used in vitro methods to examine the effects of anastrozole on several drug metabolizing CYP enzymes in human liver. Human liver microsomes were co-incubated with anastrozole and probe substrates for CYP1A2 (phenacetin), CYP2A6 (coumarin), CYP2C9 (tolbutamide), CYP2D6 (dextromethorphan), and CYP3A (nifedipine). The formation of the CYP-specific metabolites following co-incubation with various anastrozole concentrations was determined to establish IC50 and Ki values for these enzymes. While anastrozole did not inhibit CYP2A6 and CYP2D6 activities at concentrations below 500 µM, this compound inhibited CYP1A2, CYP2C9, and CYP3A activities with Ki values of 8, 10, and 10 µM, respectively. Dixon plots used to determine the Ki values for the inhibition of CYP1A2 and CYP3A activities by anastrozole were biphasic, indicating additional lower affinity Ki values. Major metabolites of anastrozole did not retain the ability to inhibit the metabolism of nifedipine (CYP3A). The results of this study indicate that, although anastrozole can inhibit CYP1A2, 2C9, and 3A-mediated catalytic activities, this compound would not be expected to cause clinically significant interactions with other CYP-metabolized drugs at physiologically relevant concentrations achieved during therapy with Arimidex (Zeneca, Ltd., Macclesfield, UK) 1-mg.

Anastrozole (ZD1033), an achiral triazole derivative known as 2,2'[5-(1H-1,2,4-triazol-1-ylmethyl)-1,3-phenylene]bis[2-methylpropionitrile] (fig. 1), is a potent inhibitor of aromatase (CYP19), which converts androgens to estrogens. This compound is currently marketed as a treatment for postmenopausal women with advanced breast cancer. Anastrozole exhibits high intrinsic potency demonstrated by the in vitro inhibition of human placental aromatase with an IC50 of 15 nM. Preclinical studies with anastrozole demonstrated its selectivity for aromatase in vivo as compared with inhibition of other enzyme activities responsible for steroid biosynthetic pathways. Cholesterol biosynthesis was minimally inhibited in vitro and plasma cholesterol concentrations were unchanged in preclinical species at doses that were at least 30 times higher than its maximally effective dose (0.1 mg/kg) required for aromatase inhibition (1). Anastrozole, at doses 100–200 times its maximally effective aromatase inhibitory dose, did not interfere with cholesterol side-chain cleavage nor did it affect plasma aldosterone levels, sodium and potassium excretion, or adrenal and prostate gland weights. Anastrozole was a comparatively weak inhibitor of bovine adrenal 11β-hydroxylase in vitro (IC50 ~ 12 µM), and in vivo changes in circulating 11-deoxycorticosterone or hypokalemia were not observed at 3 and 10 mg/kg doses in monkeys and dogs, respectively. Androgen synthesis was not affected in rats and monkeys at 10 times or, in dogs, at 100 times the effective aromatase inhibitory dose. The selectivity of anastrozole was also established in early clinical studies where doses of 1 to 10 mg in postmenopausal female volunteers produced maximal suppression of estradiol without affecting cortisol or aldosterone secretion (1, 2).

Various azole-containing compounds have been shown to be potent inhibitors and inducers of cytochrome P450-mediated drug metabolism both in vitro and in vivo. Lipophilic compounds containing nitrogen heterocycles such as phenylimidazoles and phenylpyridines inhibit rat liver monoxygenase activities with varying microsomal binding affinities depending on the position of heterocycle substitution (3). Many investigators have demonstrated that N- and C4-substituted imidazole drugs such as ketoconazole and cimetidine inhibit CYP3A activity and

1 Abbreviations used are: CYP, cytochrome P450; BSA, bovine serum albumin; BMPN-benzoic acid, 3,5-bis-(2-methylpropionitrile)-benzoic acid; PMSF, phenylmethyl-sulfonfluoride; HEPES, 4-(2-hydroxyethyl)-1-piperazineethane-sulfonic acid; TFA, trifluoroacetic acid; TCA, trichloroacetic acid; HCl, hydrochloric acid; MTBE, methyl-tert-butyl ether; Cmax, maximum concentration.
drug clearance in both animals and man (4–8). The potency of inhibition by these agents varies greatly and is determined by their hydrophobic nature and the effect of steric hindrance on the strength of the bond between their heteroatomic lone pair of electrons and the CYP heme iron. Although many azole-containing compounds are generally thought to be nonselective inhibitors of monoxygenase activity, the selectivity of CYP inhibition by compounds such as ketoconazole has been shown to be concentration-dependent in human in vitro systems (9).

Information on potential drug:drug interactions is necessary during the development of all compounds intended for therapeutic use. In vitro methodologies are now routinely used to study the potential of a drug candidate to inhibit specific CYP enzymes and classify the drugs most likely to interact during clinical use. This study examined the effects of anastrozole on several of the major drug metabolizing CYP enzymes in human liver using in vitro approaches. Comparisons of the inhibitory effects were made to other known inhibitors.

Materials and Methods

Chemicals. Anastrozole, BMPN-benzoic acid, dehydronifedipine, and dextromethorphan were synthesized by Xeneca Pharmaceuticals (Macclesfield, UK or Wilmington, DE). Phenacetin and acetaminophen were obtained from US Pharmacopoeia Convention, Inc. (Rockville, MD). Cimetidine, coumarin, dextromethorphan, ketoconazole, nifedipine, tolbutamide, and umbelliferone were obtained from Sigma Chemical Co. (St. Louis, MO). 4-Hydroxytolbutamide was obtained from Ultratine Chemicals, Ltd. (Manchester, UK). All other reagents and supplies were obtained from standard commercial sources.

Microsome preparations. Transplant quality samples of human liver were obtained from the International Institute for the Advancement of Medicine (Exton, PA). Microsomes were prepared from liver tissue immediately upon receipt by differential centrifugation using previously described methods (10). Microsomal suspensions were stored for subsequent use at ~80°C in 10 mM Tris-acetate (pH 7.4) containing 0.1 mM EDTA, 0.1 mM PMSF, and 20% glycerol (v/v). The total protein content of each microsomal sample was determined using the bicinchoninic acid reagent (Pierce Chemical Co., Rockford, IL) using BSA as the standard.

Determination of Human CYP Activities. Phenacetin deethylation to acetaminophen was used to assess CYP1A2 activity in human liver microsomes. Incubation mixtures of 0.5 ml contained 0.1 mg microsomal protein, 50 mM HEPES (pH 7.5), 15 mM MgCl2, 1 mM NADPH, and phenacetin (50, 100, and 200 μM). Reactions were stopped after 15 min by the addition of 2 ml isopropanol:ethyl acetate (1:9 v/v). The samples were vortexed and then centrifuged to separate aqueous and organic layers. The organic layer was transferred to a clean tube and evaporated under nitrogen to dryness. Samples were reconstituted in HPLC mobile phase (10:90 acetonitrile: 0.1% TFA in water). The concentration of acetaminophen produced in the microsomal incubation was determined by HPLC with UV detection at 248 nm. Separation of acetaminophen from phenacetin and other reaction components was accomplished using a Zorbax SB-C8 column, 4.6×3 mm, with a precolumn 0.5 μm filter (Uptisphere Scientific, Oak Harbor, WA). Acetaminophen (rt = 1.6 min) was eluted from the column using acetonitrile: 0.1% TFA in HPLC grade water (10:90 v/v) at a flow of 1.5 ml min⁻¹. Phenacetin (rt = 4.8 min) was washed from the column following a step gradient to 25% acetonitrile.

Coumarin hydroxylation to umbelliferone (hydroxycoumarin) was used to assess CYP2A6 activity in human liver microsomes. Incubation mixtures of 0.5 ml contained 50 μg microsomal protein, 25 mM phosphate buffer (pH 7.4), 2.5 mM MgCl2, 1 mM NADPH, and 50 μM coumarin. Reactions were stopped after 15 min by the addition of 0.5 ml 6% TCA (w/v). The samples were vortexed and then centrifuged to remove precipitated protein. A 0.5 ml aliquot of the supernatant was transferred to a tube containing 3 ml 0.8M Tris(0.8M glycine (pH 9.0). Umbelliferone concentrations in samples and standards were analyzed by fluorescence detection at excitation and emission wavelengths set at 380 nm and 460 nm, respectively.

Tolbutamide methyl hydroxylation was used to assess CYP2C9 activity in human liver microsomes. Incubation mixtures of 0.5 ml contained 0.5 mg microsomal protein, 50 mM HEPES (pH 7.5), 15 mM MgCl2, 1 mM NADP⁺, 10 mM glucose-6-phosphate, 0.33 U glucose-6-phosphate dehydrogenase, and tolbutamide (100, 200, and 1000 μM). Reactions were stopped after 30 min by the addition of 25 μl 1N HCl. The samples were extracted by vortexing with 2 ml MTBE. After centrifugation, the organic layer was transferred to a clean tube and evaporated under nitrogen to dryness. Samples were reconstituted in HPLC mobile phase (47:53 methanol: 0.1% TFA in water, pH 2.6 with NH₄OH). The concentration of hydroxytolbutamide in each sample was determined by HPLC using a Zorbax SB-C8 column, 4.6×150 mm, and UV detection at 230 nm. At a flow rate of 1.5 ml min⁻¹, hydroxytolbutamide, chlorpropanide (internal standard), and tolbutamide eluted at approximately 3, 7, and 11 min.

CYP2D6 activity in human liver microsomes was assessed by determining dextrophan formed from dextromethorphan using methods modified from published procedures (11). Incubation mixtures of 0.15 ml contained 50 μg microsomal protein, 50 mM HEPES (pH 7.5), 15 mM MgCl2, 2 mM NADPH, and 100 μM dextromethorphan. Reactions were stopped after 15 min by the addition of a drop of 10% NH₄OH. The samples were extracted by vortexing with 1 ml MTBE. The MTBE layer was transferred after centrifugation to a clean tube and evaporated under nitrogen to dryness. Samples were reconstituted in HPLC mobile phase (25:75 acetonitrile: 30 mM ammonium acetate, pH 4.0). The concentration of dextrophan in each sample was determined by HPLC using a Zorbax SB-C8 column, 4.6×150 mm, and UV detection at 230 nm. At a flow rate of 1.5 ml min⁻¹, dextrophan and dextromethorphan eluted at approximately 3 and 10 min.

To determine their inhibitory effects on CYP3A activity, various concentrations of anastrozole, BMPN-benzoic acid, triazole, ketoconazole, or cimetidine were co-incubated in vitro as potential inhibitors with nifedipine using modified procedures from those described by Guengerich et al. (13). Incubation mixtures of 0.5 ml contained 0.1 mg microsomal protein, 50 mM HEPES (pH 7.5), 15 mM MgCl₂, 1 mM NADPH+, 10 mM glucose-6-phosphate, 0.33 U glucose-6-phosphate dehydrogenase, and nifedipine (10, 25, and 50 μM). Reactions were stopped after 20 min by the addition of 0.1 ml Na₂CO₃. The samples were extracted by vortexing with 2 ml MTBE. The MTBE layer was transferred after centrifugation to a clean tube and evaporated under nitrogen to dryness. Samples were reconstituted in HPLC mobile phase (60:40 methanol:water). The concentration of dehydronifedipine in each sample was determined by HPLC using a Zorbax SB-C8 column, 4.6×250 mm, and UV detection at 260 nm. At a flow rate of 1.5 ml min⁻¹, dehydronifedipine and nifedipine eluted at approximately 6 and 9 min. Incubations, extractions, and HPLC analyses of samples and standards containing nifedipine were conducted while minimizing the exposure to light.

Data Analysis. Incubation of the CYP-marker substrates with microsomes from 1 to 3 individual donors in the presence or absence of various concentrations of anastrozole was used to determine IC₅₀ values. IC₅₀ values for the inhibition of the CYP activities were determined by nonlinear regression analysis (Origin, version 3.0, Microcal Software, Inc., Northampton, MA) using the following equation:

\[ y = 100 - (100 - E₀) \times \left(\frac{I}{1 + IC_{50}}\right) \]

where IC₅₀ is the initial concentration of inhibitor in a microsomal incubation, E₀ is the per cent enzyme activity that is not inhibited at I = 0, and IC₅₀ is the inhibitor concentration that inhibits enzyme activity by 50%.

Rates of metabolite formation from pooled human liver microsomes following co-incubation of three concentrations of CYP-marker substrate with multiple inhibitor concentrations were used to determine Kᵢ values using the method of Dixon (14). The type of inhibition was assessed by transforming the data to give S/metabolite formation rate vs. inhibitor concentration (15).

Results

The potential inhibitory effects of the aromatase inhibitor, anastrozole, on drug metabolism were evaluated by determining IC₅₀ and Kᵢ values for metabolic reactions that are selectively catalyzed by five different cytochrome P450 forms in human hepatic microsomes (table 1). Anastrozole inhibited less than 10% of the control coumarin.
TABLE 1

<table>
<thead>
<tr>
<th>CYP</th>
<th>Marker Substrate</th>
<th>Test Substance</th>
<th>IC50 (µM)</th>
<th>K_i (µM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1A2</td>
<td>Phenacetin</td>
<td>Anastrozole</td>
<td>30</td>
<td>80/80</td>
</tr>
<tr>
<td>2A6</td>
<td>Coumarin</td>
<td>Anastrozole</td>
<td>&gt;500</td>
<td>—</td>
</tr>
<tr>
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<td>Tolbutamide</td>
<td>Anastrozole</td>
<td>48</td>
<td>10</td>
</tr>
<tr>
<td>2D6</td>
<td>Dextromethorphan</td>
<td>Anastrozole</td>
<td>&gt;500</td>
<td>—</td>
</tr>
<tr>
<td>3A</td>
<td>Nifedipine</td>
<td>Anastrozole</td>
<td>27</td>
<td>10/55</td>
</tr>
<tr>
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<td>Nifedipine</td>
<td>Triazolide</td>
<td>&gt;250</td>
<td>—</td>
</tr>
<tr>
<td>3A</td>
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<td>BMPN-benzoic acid</td>
<td>&gt;250</td>
<td>—</td>
</tr>
<tr>
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<td>—</td>
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<tr>
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<td>Nifedipine</td>
<td>Cimetidine</td>
<td>650</td>
<td>—</td>
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</tbody>
</table>

*IC50 values were determined using the following concentrations of the specific CYP-marker substrates: phenacetin, 100 µM; coumarin, 50 µM; tolbutamide, 1 mM; dextromethorphan, 100 µM; nifedipine, 25 µM.

a Anastrozole and its metabolites (products formed by N-dealkylation) were not tested for CYP inhibition above 250–500 µM. Because little inhibition was observed at high concentrations, K_i values were not determined.

The kinetics of phenacetin O-deethylation by the microsomes used in this investigation was best fit to a 2-enzyme model with a K_m of 59 µM for the high affinity site. This result suggests that more than one enzyme contributes to this O-deethylation reaction and may potentially explain the appearance of biphasicity in the Dixon plot. In contrast and although other CYP3A enzymes are known to metabolize nifedipine, the kinetics of dehydro-nifedipine formation was fitted best to a 1-enzyme Michaelis-Menten model (K_m ~ 25 µM).

The inhibition of tolbutamide hydroxylase activity by anastrozole co-incubation on the rate of acetyaminophen formation from phenacetin in human liver microsomes was excluded. The Dixon plot could be divided into two distinct linear phases. Therefore, when the data corresponding to the two phases were subjected to linear regression analysis, two apparent K_i values of 8 and 80 µM were determined for the inhibition of phenacetin metabolism to acetaminophen. The high affinity K_i of 8 µM estimated in this in vitro investigation was used to estimate conservatively the in vivo interaction potential towards CYP1A2 substrates. Parallel plots in the Cornish-Bowden transformation of the data indicate that the high affinity inhibition is competitive in nature.

Tolbutamide methyl-hydroxylase activity, a marker of CYP2C9 activity in human liver microsomes, was inhibited by anastrozole with an IC_50 of 48 µM when tolbutamide was co-incubated at 1000 µM (fig. 2). At this concentration of tolbutamide, CYP2C8 most likely contributes to the methylhydroxylase activity (16, 17) whereas at lower concentrations, the contribution by this CYP2C form would be negligible. An apparent K_i of 10 µM was determined based on plots of the reciprocal hydroxylation rates when 100, 200, and 1000 µM tolbutamide was co-incubated with 0 to 100 µM anastrozole (fig. 4A).

The inhibition of tolbutamide hydroxylase activity by anastrozole exhibited linear behavior on Dixon plots and Cornish-Bowden plots demonstrated parallel best-fit lines indicating competitive-type inhibition (fig. 5B).

Anastrozole inhibited nifedipine oxidation in human liver microsomes with an IC_50 of 27 µM. By comparison, ketoconazole was a
much more potent inhibitor of this CYP3A-catalyzed reaction with an IC\textsubscript{50} of 0.02 μM, while cimetidine was a weaker inhibitor (IC\textsubscript{50} = 650 μM) when co-incubated with nifedipine. Dixon plots of the reciprocal oxidation rates exhibited similar biphasicity to that observed for the inhibition of CYP1A2 by anastrozole (fig. 5A). Two distinct intercepts were obtained by linear regression analysis of the data and thus, two apparent K\textsubscript{i} values of 10 and 55 μM were determined for the inhibition of CYP3A-catalyzed nifedipine oxidation by anastrozole. Cornish-Bowden plots suggested that the CYP3A inhibition was mixed (competitive/noncompetitive) in nature (fig. 5B).

Because CYP3A4 appears to be the most abundant cytochrome P450 enzyme in both human liver and intestine and this enzyme is involved in the metabolism of many drug substrates, triazole and BMPN-benzoic acid were tested for their ability to inhibit nifedipine oxidation. These two compounds are major metabolites of anastrozole that are produced by N-dealkylation (cleavage of the methylene bridge). When co-incubated with nifedipine at concentrations up to 250 μM, these metabolites of anastrozole did not inhibit nifedipine oxidation.

**Discussion**

Many, if not most, clinically relevant drug interactions are caused by inhibition of drug metabolizing enzymes leading to a decreased metabolic clearance and increased exposure to the inhibited drug. Cytochrome P450 enzymes mediate the rate limiting step of the primary Phase I metabolic reactions for the vast majority of drug compounds as well as the biosynthesis or catabolism of a number of endogenous substances with potent physiological and pathophysiological functions. Specific cytochrome P450 enzymes in the CYP1A, 2A, 2C, 2D, and 3A subfamilies are responsible for the oxidative metabolism of most drugs in humans (18, 19). Toxicities and other adverse events may therefore develop during clinical therapy with drugs that have narrow therapeutic margins when the concomitant administration of CYP inhibitors causes potent metabolic inhibition of drug clearance. Probably the most prominent example of this type of potentially harmful interaction is the cardiotoxicity that can be produced following co-administration of CYP3A4 inhibitors such as ketoconazole and erythromycin with terfenadine, a prodrug metabolized exclusively by CYP3A enzyme(s) (20).

In vitro studies are now being used routinely to investigate the interaction potential for drugs resulting from inhibition of CYP-mediated metabolism (21, 22). The experiments reported here examined the ability of anastrozole to inhibit the metabolism of substrates for CYP1A2, CYP2A6, CYP2C9, CYP2D6, and CYP3A. The results were used 1) to compare the inhibition of drug metabolizing CYPs by anastrozole to its in vitro ability to inhibit estrogen synthesis mediated by human aromatase and 2) to predict the potential for anastrozole to cause metabolic drug interactions.

Anastrozole inhibited specific reactions catalyzed by CYP1A2, CYP2C9, and CYP3A but did not inhibit CYP2A6 or CYP2D6-mediated pathways. The inhibition of both CYP1A2 and CYP3A
activities were biphasic in nature although the basis of the nonlinear results has not been established. Two apparent \( K_i \) values were exhibited by anastrozole for inhibition of CYP1A2 and CYP3A metabolism. The low \( K_i \) values for these two enzymes was similar to the single apparent \( K_i \) determined for inhibition of CYP2C9 activity by anastrozole (\( K_i = 8 \times 10^{-10} \) M corresponds to 2.3 to 2.9 \( \mu g \) ml \(^{-1} \)). The low \( K_i \) values were used to estimate conservatively the potential for inhibitor drug interactions with anastrozole during clinical use of the drug. Average steady-state \( C_{\text{max}} \) concentrations in patients chronically administered the 1-mg marketed dose of Arimidex were approximately 0.08 \( \mu g \) ml \(^{-1} \) (0.3 \( \mu M \)). These plasma concentrations are 30-fold lower than the apparent \( K_i \) values determined in these in vitro studies, suggesting that anastrozole would cause little or no inhibition in vivo of the metabolism of drugs that are substrates for CYP1A2, CYP2C9, and CYP3A enzymes (per cent inhibition in vivo predicted to be approximately 3%). Although a lack of inhibition potential can be predicted from these in vitro results, any effect of intracellular binding or accumulation of anastrozole in hepatocytes on the in vivo inhibition have not been accounted for in these investigations.\(^3\)

Nifedipine oxidation in human liver microsomes was not decreased by two major metabolites (triazole and BMPN-benzoic acid) of anastrozole at concentrations up to 250 \( \mu M \). The lack of CYP inhibition by these metabolites is consistent with requirement for both hydrophobic character for active site access and the heteratomic lone pair of electrons in triazole for Type II binding to the heme iron (23).

The results herein also show that the ability of anastrozole to inhibit drug metabolizing CYP enzymes in liver microsomes is much weaker than the inhibition of aromatase. When comparing the IC\(_{50}\) for inhibition of human placental aromatase activity (15 nM) with the inhibition of the hepatic microsomal CYP produced by anastrozole in vitro, a margin of selectivity of at least 500-fold is obtained.

In conclusion, the results of the in vitro experiments performed in human liver microsomes indicate that, although anastrozole can inhibit CYP1A2, 2C9, and 3A-mediated catalytic activities, the level of inhibition in vivo during clinical therapy with Arimidex (Zeneca, Ltd., Macclesfield, UK) 1-mg would not be expected to cause clinically significant interactions with other CYP-metabolized drugs.

References