MIBEFRADIL IS A P-GLYCOPROTEIN SUBSTRATE AND A POTENT INHIBITOR OF BOTH P-GLYCOPROTEIN AND CYP3A IN VITRO

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ABSTRACT:

Mibefradil, a calcium T- and L-channel blocker developed for use in hypertension, was recently removed from the market after reports of severe drug-drug interactions. Mibefradil is known to inhibit various cytochrome P450 enzymes involved in drug metabolism, particularly CYP3A. However, the extent and the severity of the observed drug interactions in humans suggest that inhibition of additional systems important to drug disposition, such as the drug transporter P-glycoprotein (P-gp), may also have contributed to the severity of the mibefradil interactions. A polarized epithelial cell line, LLC-PK1, which does not express P-gp, and the derived L-MDR1 cell line, which overexpresses human P-gp, were used to study the effects of mibefradil on drug transport. A markedly greater basal-to-apical versus apical-to-basal transport of [3H]mibefradil was seen in the L-MDR1, but not in the LLC-PK1 cells, suggesting that the drug is a substrate of P-gp. Using a human intestinal cancer-derived cell line Caco-2, which constitutively expresses P-gp, mibefradil was shown to be a potent inhibitor of P-gp-mediated digoxin transport, with an IC50 of 1.6 μM. Additionally, the effect of mibefradil on CYP3A was assessed using human liver microsomes. Mibefradil inhibited CYP3A-mediated nefedipine oxidase activity with an IC50 of 0.8 μM, and a K, of 0.6 μM. Thus, mibefradil is not only a P-gp substrate, but also a potent inhibitor of both P-gp and CYP3A. These data suggest that the severity of drug interactions seen with mibefradil use is due to the dual inhibition of both P-gp and CYP3A.

Experimental Procedures

Materials. [3H]Digoxin (15 Ci/mmol) was supplied by DuPont-New England Nuclear (Boston, MA), and [3H]mibefradil (specific activity, 15 Ci/mmol, >98% purity by HPLC) was a gift from Hoffmann-La Roche (Basel, Switzerland). L-MDR1 and LLC-PK1 cells were provided by Dr. A.H. Schinkel (Netherlands Cancer Institute, Amsterdam, the Netherlands) and Dr. E.G. Schuetz (St. Jude’s Children’s Research Hospital, Memphis, TN), and Caco-2 cells were obtained from Dr. R.J. Coffey (Vanderbilt University, Nashville, TN). All other chemicals and reagents, unless stated otherwise, were obtained from Sigma-Aldrich (St. Louis, MO) and were of the highest quality available.

Transport in Cultured LLC-PK1, L-MDR1, and Caco-2 Cells. LLC-PK1 and L-MDR1 cells were cultured in M199 medium (Life Technologies, Gaithersburg, MD) supplemented with 50 μg streptomycin/ml and 10% v/v fetal calf serum at 37°C in the presence of 5% CO2; cells were subcultured after trypsinization every 6 to 7 days (Schinkel et al., 1995). Cells were grown as monolayers on polycarbonate membrane filters (Transwell; Costar Corporation, Cambridge, MA) as outlined previously (Kim et al., 1998). Transepithelial resistance was measured in each well using a Millicell ERS ohmmeter (Millipore, Bedford, MA); wells registering a resistance of 200 ohms or greater, after correcting for the resistance obtained in control blank wells, were used for transport studies.

When a drug enters widespread use after its approval, previously unexpected severe adverse drug interactions can occur. Severe interactions have been reported with the recently developed calcium channel blocker mibefradil (Posicor, Roche Laboratories), an antihypertensive agent that inhibits both T (transient) channel as well as the L (long) channel (Mullins et al., 1998). In the limited population studied before its approval, this drug had appeared to be as efficacious and as well tolerated as comparable drugs currently on the market. However, mibefradil was withdrawn from the market by its manufacturer after drug interactions were reported (Bursztyn et al., 1997; Kobrin et al., 1997). However, mibefradil was withdrawn from the market by its manufacturer after drug interactions were reported (Bursztyn et al., 1997; Kobrin et al., 1997). Nevertheless, this work was supported in part by U.S. Public Health Service Grants GM 31304, HL 56251, OA 44353, ES 00267, and the Deutsche Forschungsgemeinschaft (C.W.).
used in the transport experiments. About 1 to 2 h before the start of the transport experiments, the medium in each compartment was replaced with a serum-free medium (OptiMEM; Life Technologies). At the beginning of each experiment, the medium in each compartment was replaced with 700 μl of serum-free medium (OptiMEM), with or without drug (radiolabeled or unlabeled). The amount of the drug appearing in the opposite compartment after 1, 2, 3, and 4 h was measured in 25-μl aliquots taken from each compartment.

Inhibition of P-gp-mediated transport by Caco-2 cells was determined in a similar manner after the addition of the putative inhibitor to both the apical and basal compartments, and using radiolabeled digoxin as a P-gp substrate (DeAnnoy and Silverman, 1992) as described previously (Wandel et al., 1999). Complete inhibition of P-gp-mediated transport would be expected to result in the loss of the basal-to-apical (B→A) versus apical-to-basal (A→B) transport difference for digoxin. Accordingly, percent inhibition was estimated by eq. 1:

\[
\text{Degree of inhibition} = \left(1 - \frac{i_{B\rightarrow A} - a_{B\rightarrow A}}{a_{B\rightarrow A}}\right) \times 100\% \quad (1)
\]

where \(i\) and \(a\) are the percentages of digoxin transport in the presence and absence of the putative inhibitor, according to the direction of transport. Data shown represent results obtained from studies carried out on at least three preparations on different days. For statistical analysis, unpaired Student’s t test or Mann-Whitney U test was used (Prism; Graph Pad, San Diego, CA) with \(P < .05\) as the limit of significance.

**CYP3A Activity Measured Using Human Liver Microsomes.** Human liver microsomes were prepared as previously described (Guengerich, 1994), and the total cytochrome P-450 content was measured using a standard method (Omura and Sato, 1964). CYP3A catalytic activity was determined by incubating the microsomes in a total volume of 500 μl of 0.1 M phosphate buffer (pH 7.4) containing 100 pmol of cytochrome P450 and 1.5 mM NADPH, with varying concentrations of nifedipine (15, 20, 30, 60, and 200 μM), in the presence and absence of varying concentrations of inhibitors. All samples including controls contained 2% dimethyl sulfoxide because 2% dimethyl sulfoxide itself inhibited CYP3A activity by 25% in the microsomes used. Incubations were performed at 37°C and stopped after 10 min by the addition of 1 ml of CHCl₃. Nifedipine metabolite concentrations were determined using an HPLC-based procedure (Guengerich and Bocker, 1988; Guengerich, 1994). The Michaelis-Menten kinetics was calculated using a nonlinear regression program (kcat; BioMetallics, Princeton, NJ). Inhibition type and constant (\(K_i\)) were derived using a Lineweaver-Burk plot, and IC₅₀ values using the Hill equation.

**Results**

**Identification of Mibefradil as a P-gp Substrate Using L-MDR1 and LLC-PK1 Cells.** Studies with LLC-PK1 cells, a cell line lacking P-gp, and the derived cell line L-MDR1, which overexpresses human MDR1-encoded P-gp, revealed that mibefradil cellular translocation was markedly polarized (Fig. 1); transport is more extensive and significantly greater (\(P < .01\)) in the basolateral to apical direction.
Inhibition Studies Using Human Liver Microsomes. Nifedipine oxidation in human liver microsomes was characterized by an apparent $K_m$ of 30 $\mu$M, and a $V_{max}$ of 7 nmol metabolite/nmol of cytochrome P450/min. Mibebradil competitively inhibited oxidation of nifedipine (CYP3A) with an apparent $K_i$ of 0.6 $\mu$M. The IC$_{50}$ value was 1 $\mu$M at a nifedipine concentration of 20 $\mu$M. The comparative effects of ketoconazole and PSC833 on microsomal nifedipine oxidation are shown in Table 1 and Fig. 3.

Discussion

It has been estimated that nearly 100,000 patients each year die of adverse drug reactions in the United States (Lazarou et al., 1998). Some of these are due to drug-drug interactions. The voluntary withdrawal of mibebradil by its manufacturer demonstrated that interactions were detected only after the drug was released for general use. There are a number of potent CYP3A inhibitors on the market that are used safely. Therefore why did mibebradil appear to be so toxic? We proposed that this was due to the multiplicity of pathways inhibited by mibebradil, including metabolism by such key cytochrome P450 enzymes as CYP2D6 and CYP3A4 (Abernethy, 1997), the enzymes responsible for the metabolism of a variety of drugs (Guengerich, 1995; Thummel and Wilkinson, 1998); in addition, drug transport by the drug efflux transporter, P-gp was inhibited. This membrane-bound efflux transporter is expressed not only in cancer cells, but also in normal tissues, including the intestine, liver, and kidney (Thiebaut et al., 1989; Pastan and Gottesman, 1991), where P-gp contributes to the elimination of drug substrates into the gut, bile, and urine, respectively. In addition, P-gp also determines drug entry into the central nervous system (Kim et al., 1998). Known drug substrates of P-gp include various calcium channel blockers, some beta blockers, immunosuppressive agents such as cyclosporine and tacrolimus, HIV-1 protease inhibitors, and cardiac glycosides including digoxin (Mayer et al., 1996; Schinkel et al., 1996; Terao et al., 1996; Kim et al., 1998). There is considerable overlap in substrates of P-gp and CYP3A, resulting in many drugs being both transported by P-gp and metabolized by CYP3A (Wacher et al., 1995; Wandel et al., 1999; Shirato et al., 1999). Therefore, the simultaneous inhibition of both systems will result in a substantial increase in plasma drug concentrations with resultant toxicity.

It is important to emphasize the potential for drug interactions to occur due to both CYP3A and P-gp inhibition in the gut. CYP3A and P-gp in enterocytes contribute to reducing systemic availability of drugs administered orally so that inhibition of both systems may result in a substantial increase in plasma drug concentrations and patient exposure to drug.

We have previously shown that the predominant mechanism to account for the clinically significant digoxin-quinidine interaction is the inhibition of P-gp by quinidine, resulting in reduced P-gp-mediated digoxin elimination (Fromm et al., 1998). We therefore hypothesized that the previously described drug-drug interactions involving

**TABLE 1**

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<thead>
<tr>
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<th>IC$_{50}$ ($\mu$M)</th>
<th>$K_i$ ($\mu$M)</th>
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<tr>
<td>P-gp</td>
<td></td>
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<tr>
<td>CYP3A</td>
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<tr>
<td>Mibebradil</td>
<td>1.6</td>
<td>0.80</td>
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<tr>
<td>Ketoconazole</td>
<td>1.4</td>
<td>0.15</td>
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<tr>
<td>PSC 833</td>
<td>0.11</td>
<td>10</td>
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Each experiment was performed on at least two different experimental days.

**Fig. 2.** Inhibition of P-gp by PSC833 (●), mibebradil (▲), and ketoconazole (■) measured using 5 $\mu$M digoxin transport in Caco-2 cells.

**Fig. 3.** Inhibition of CYP3A by PSC833 (●), mibebradil (▲), and ketoconazole (■) measured using human liver microsomal nifedipine oxidase activity.
mibefradil and digoxin (Siepmann et al., 1995) might be explained by similar P-gp inhibition by mibefradil. This study has confirmed that hypothesis, and demonstrated that mibefradil is both a substrate and an inhibitor of P-gp. In addition, we confirmed the potent inhibitory effect of mibefradil on CYP3A. However, it is the likely dual inhibition of both systems that contributed to its notable toxicity in practice. As shown in Table 1, mibefradil inhibits nifedipine oxidation activity with an IC50 and Ki values similar to the potent CYP3A inhibitor, ketoconazole. The potency of mibefradil as a P-gp inhibitor, although less than that of drugs specifically developed to inhibit P-gp, such as PSC833, is remarkably high so that inhibition of both P-gp and CYP3A will occur at clinically relevant concentrations. After chronic mibefradil dosing of 100 mg/day in human subjects, the trough levels are approximately 1 μM (Abernethy, 1997). Accordingly, patients receiving mibefradil will have plasma concentrations throughout the dosing interval that may significantly inhibit both P-gp and CYP3A, resulting in both distributional and metabolic drug interactions.

In conclusion, our data demonstrate that mibefradil is both a substrate with similar basal to apical translocation rates to digoxin and a potent inhibitor of the P-gp transporter while also being a potent inhibitor of human CYP3A and is therefore another example of the overlapping substrate specificity of CYP3A and P-gp. Accordingly, the severity of drug-drug interactions observed during clinical use of mibefradil is explained by its ability to inhibit multiple systems important to drug disposition. Studies aimed at determining the role of drug transport systems, and not just drug metabolism at an early stage in drug development, have the potential to better predict drug-drug interactions before widespread general use of a drug and prevent subsequent toxicity.

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References


