SEX DIFFERENCE IN INHIBITION OF IN VITRO MEXAZOLAM METABOLISM BY VARIOUS 3-HYDROXY-3-METHYLGLUTARYL-COENZYME A REDUCTASE INHIBITORS IN RAT LIVER MICROSOMES

MICHIGISHAM, WATABAKASAKI, TOSHIHIKOKEEDA, TORUKOMAI, KIYOMIITO, ANDYUCHISUGIYAMA

Drug Metabolism and Pharmacokinetics Research Laboratories, New Drug Development Division and Product Strategy Department, Sankyo Co., Ltd., Shinagawa-ku, Tokyo, Japan (M.I., W.T., T.I., T.K.); School of Pharmaceutical Sciences, Kitasato University, Minato-ku, Tokyo, Japan (K.I.); and Graduate School of Pharmaceutical Sciences, University of Tokyo, Bunkyo-ku, Tokyo, Japan (Y.S.)

(Received August 23, 2001; accepted April 19, 2002)

This article is available online at http://dmd.aspetjournals.org

ABSTRACT:
To identify an appropriate animal model for the study of drug interaction via CYP3A4 inhibition, the inhibition of in vitro mexazolam metabolism by various 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) reductase inhibitors [simvastatin (lactone), simvastatin acid, fluvastatin, atorvastatin, cerivastatin, pravastatin lactone, and pravastatin (acid)] in male and female rat liver microsomes was investigated and compared with that by HMG-CoA reductase inhibitors in human liver microsomes reported previously. The metabolism of mexazolam in male rat and female rat liver microsomes was inhibited by all the HMG-CoA reductase inhibitors examined except pravastatin (acid). The K_i values in female rats were lower than those in male rats, demonstrating the presence of a sex difference in the inhibition potency of HMG-CoA reductase inhibitors toward mexazolam. Using anti-cytochrome P450 P450 (P450) antisera, the main P450 isoform responsible for the metabolism of mexazolam was identified as CYP3A in female rats and CYP2C11 in male rats. Based on these results, we speculate that the sex difference in the inhibition potency of HMG-CoA reductase inhibitors for mexazolam observed in rats is caused by their different inhibition potencies against CYP2C11 and CYP3A isoforms. For mexazolam metabolism, the results obtained in female rats, rather than those in male rats, seem to be a much better reflection of the results in humans. Since species and sex differences were observed in P450 isoforms in the present study, our results show that establishing appropriate experimental conditions, in particular with respect to the P450 isoforms responsible for the drug metabolism in question, is indispensable for the investigation of drug interactions using rats as a model animal for humans.

Among the human liver cytochrome P450 (P450) isoforms, CYP3A4 is present in the highest concentration and responsible for the metabolism of various kinds of drugs. For this reason, it is considered an important isoform for the investigation of drug interactions via P450. In a previous study, we reported that mexazolam, a benzodiazepine antianxiety agent, was metabolized by CYP3A4 (Ono et al., 1993). Using mexazolam as a probe, the inhibition of CYP3A4 activity by lactone or acid forms of HMG-CoA reductase inhibitors was investigated in a series of in vitro studies using human liver microsomes. Each of the HMG-CoA reductase inhibitors, except for pravastatin, (simvastatin, simvastatin acid, lovastatin, fluvastatin, atorvastatin, and cerivastatin) was found to inhibit mexazolam metabolism in human liver competitively (Ishigami et al., 2001a). Although the inhibition of CYP3A4/5 by HMG-CoA reductase inhibitors can now be tested directly in man, if the question had been raised preclinically, the choice of a suitable model would be of primary importance, since interspecies differences in drug disposition are well known (Nelson et al., 1996). Sex differences have also been observed in liver content and activity of P450 isoforms (Kato and Kamataki, 1982; Kamataki et al., 1983), providing another important reason why results obtained in experimental animals do not always reflect drug interactions observed in humans. Therefore, we carried out an investigation to see whether the inhibition of mexazolam metabolism by HMG-CoA reductase inhibitors observed in human liver microsomes could occur in both male and female rats, the most commonly used animal species in pharmacokinetic studies. We previously found that the in vitro metabolism of simvastatin was inhibited by itraconazole in female rat liver microsomes but not in male rat liver microsomes (Ishigami et al., 2001b). Although female rat liver contains CYP3A at an extremely low level (Cooper et al., 1993), its activity is considered to be strongly inhibited by itraconazole. On the other hand, male rat liver contains much higher levels of CYP2C11 than CYP3A (Cooper et al., 1993), and CYP2C11 seems to be mainly responsible for the metabolism of simvastatin, which is not inhibited by itraconazole. Thus, there has been speculation that the sex difference in the inhibition of simvastatin metabolism by itraconazole may be attributable to both the sex difference in P450 isoforms responsible for simvastatin metabolism and the difference in the inhibition potency of itraconazole against CYP3A and CYP2C11.
In the present study, taking into account the sex difference in the drug interactions observed between simvastatin and itraconazole, the inhibition of mexazolam metabolism by HMG-CoA reductase inhibitors (Fig. 1) was investigated in an in vitro system using female and male rat liver microsomes, and the results were compared with those obtained in humans to identify an appropriate animal model for the study of drug interaction via CYP3A4 inhibition. In addition, the inhibition of mexazolam metabolism by HMG-CoA reductase inhibitors and itraconazole was also investigated under the condition in which the content of P450 isozymes was changed, using liver microsomes of rats treated with dexamethasone, which is known to induce CYP3A in rats (Ghosal et al., 1996a).

Materials and Methods

**Chemicals and Reagents.** Pravastatin (acid), pravastatin lactone, simvastatin (lactone), simvastatin acid (Na salt), fluvastatin (acid), atorvastatin (acid), cerivastatin (acid), itraconazole, mexazolam, and a metabolite of mexazolam (M-1; Fig. 2) used in the present study were synthesized at Sankyo Co., Ltd. (Tokyo, Japan). Anti-rat P450 antisera preparations (anti-rat CYP2C11 prepared from goat serum and anti-rat CYP3A2 prepared from rabbit serum) were purchased from Daiichi Pure Chemicals Co., Ltd. (Tokyo, Japan). All other chemicals and reagents used were commercially available and of reagent grade.

**Preparation of Rat Liver Microsomes.** Rat liver microsomes were prepared from male and female Sprague-Dawley rats (Japan SLC, Hamamatsu, Japan).
concentrations of 5 to 100 mM ethanol solution of mexazolam was added to each reaction mixture to make 10 mM glucose-6-phosphate dehydrogenase, and 10 mM MgCl₂ was added to rat liver microsomes. Liver microsomes were also prepared in the same manner as described above from livers of female and male rats, 24 h after a single intraperitoneal administration of dexamethasone (100 mg/kg, corn oil, Japan) according to standard methods. To each isolated liver sample was added 10 mM phosphate buffer containing potassium chloride at a concentration of 1.15%, and the resulting mixture was homogenized and then centrifuged at 9,000g for 20 min. The supernatant was centrifuged again at 105,000g for 1 h, and the phosphate buffer containing glycerol was added to the obtained pellet to prepare a microsomal suspension. Liver microsomes in the same manner as described above from livers of female and male rats, 24 h after a single intraperitoneal administration of dexamethasone (100 mg/kg, corn oil solution) (Dex-treated rats).

**Metabolism of Mexazolam in Rat Liver Microsomes.** An NADPH-generating system containing 2.5 mM NADP, 25 mM glucose 6-phosphate, 2 units glucose-6-phosphate dehydrogenase, and 10 mM MgCl₂, was added to rat liver microsomes (0.2 mg of protein/ml) in a total volume of 0.2 ml, and the resulting mixtures were preincubated at 37°C for 3 min. Then 2 μl of an ethanol solution of mexazolam was added to each reaction mixture to make concentrations of 5 to 100 μM. After incubation at 37°C for 2 min, 0.4 ml of methanol was added followed by vortex mixing to stop the obtained reaction and centrifugation at 19,000g for 3 min. The amount of mexazolam metabolite (M-1; Fig. 2) in the supernatants was analyzed by high-performance liquid chromatography (HPLC). For the HPLC operating conditions, a Deverosil ODS-UG-5 column (250 × 4.6 mm; Nomura Chemical Co., Ltd., Aichi, Japan), a mobile phase of acetonitrile/50 mM phosphate buffer (pH 8.0) (55:45, v/v), a flow rate of 1 ml/min, and a wavelength at 235 nm for UV measurement were used.

**Kinetic Analysis of Mexazolam Metabolism.** The formation of a metabolite from mexazolam by rat liver microsomes (pooled liver microsomes from five male or five female rats) was fitted to eq. 1 using a nonlinear least-squares regression program (WinNonlin; Scientific Consulting, Inc., Apex, NC), and the Km and Vmax values were calculated:

\[ V_o = V_{\text{max}} \times S/(K_m + S) \]

where V₀, Vmax, S, and Km represent the initial rate of metabolism, maximum rate of metabolism, substrate concentration, and Michaelis constant, respectively. Intrinsic metabolic clearance (CLint) was calculated by the following equation:

\[ \text{CL_{int}} = \frac{V_{\text{max}}}{K_m} \]

**Inhibition of Mexazolam Metabolism by Anti-P450 Antisera.** To identify the enzymes responsible for the metabolism of mexazolam, inhibition experiments using anti-rat P450 antisera were performed. Anti-rat P450 antisera or corresponding control sera (0–0.25 mg of IgG, 25 μl) were added to rat liver microsomes (pooled from five male rats or five female rats, 10 mg of protein/ml, 10 μl) separately, and the resulting mixtures were preincubated at room temperature for 30 min. Then, as described above for the metabolism of mexazolam in rat liver microsomes, an NADPH-generating system was added to each mixture, and the resulting solutions were preincubated at 37°C for 3 min. Subsequently, an ethanol solution of mexazolam was added (final concentration, 20 μM mexazolam) and incubated at 37°C for 2 min (0.2 mg of protein/ml; volume of reaction mixture, 0.5 ml). The reaction was stopped by the addition of 1 ml of methanol and the amount of metabolite (M-1) formed in the supernatants obtained after centrifugation was determined by HPLC.

**Inhibition of Mexazolam Metabolism by HMG-CoA Reductase Inhibitors and Itraconazole.** To 0.2 ml of each of male and female rat liver microsomal preparations (0.2 mg of protein/ml), 2 μl of an ethanol solution of each of the HMG-CoA reductase inhibitors [pravastatin (acid), pravastatin lactone, simvastatin acid, fluvastatin (acid), atorvastatin (acid), and cerivastatin (acid), 20 to 200 μM; and simvastatin (lactone), 2 to 20 μM] was added separately. The resulting mixtures were preincubated at 37°C for 3 min, and an ethanol solution of mexazolam (2 μl) was added to each microsomal prepa-

---

**TABLE 1**

<table>
<thead>
<tr>
<th></th>
<th>Control Rat</th>
<th>Dex-Treated Rat</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Km (μM)</td>
<td>Vmax (nmol/min/mg protein)</td>
</tr>
<tr>
<td>Male</td>
<td>10.7 ± 0.4</td>
<td>3.23 ± 0.04</td>
</tr>
<tr>
<td>Female</td>
<td>10.3 ± 0.8</td>
<td>0.29 ± 0.01</td>
</tr>
</tbody>
</table>
Fig. 6. Dixon plots for the inhibition of M-1 formation from mexazolam in male (a) and female (b) rat liver microsomes by HMG-CoA reductase inhibitors.

Mexazolam (5–50 μM) was incubated for 2 min at 37°C with rat liver microsomes (one male rat or one female rat, 0.2 mg of protein/ml) in the presence or absence of HMG-CoA reductase inhibitors [pravastatin (acid), 50–400 μM; simvastatin (lactone), 5–20 μM; pravastatin lactone, simvastatin acid, fluvastatin (acid), atorvastatin (acid), and cerivastatin (acid), 20–200 μM]. ○, 5 μM mexazolam; ◆, 10 μM mexazolam; □, 20 μM mexazolam; ▲, 50 μM mexazolam. V₀, M-1 formation rate (nanomoles per minute per milligram of protein).
ration to give a final concentration of 10 to 50 μM. The amount of metabolite, M-1, formed in each reaction mixture was determined in the same manner as described above for the metabolism of mexazolam in rat liver microsomes. Inhibition by itraconazole was also examined in the same manner as mentioned above by adding 2 μL of a dimethylacetamide solution of itraconazole (0.1–1 μM) to each of the male and female rat liver microsomal preparations, separately.

Calculation of inhibition constant (K_i) values from Dixon plots was performed by the simultaneous fitting of the data to the following equation using the WinNonlin program.

\[ \frac{1}{V_0} = \frac{[1 + K_a/S]V_{max} + [I \times K_i]/(K_i \times S \times V_{max})}{1 + I/K_i} \]

where S and I represent the concentration of the substrate (mexazolam) and each inhibitor (HMG-CoA reductase inhibitors or itraconazole), respectively.

**Results**

**Effects of Anti-P450 Antisera on Mexazolam Metabolism in Rat Liver Microsomes.** The mexazolam metabolism in male rat liver microsomes was inhibited by about 80% after the addition of anti-CYP2C11 antiserum (0.25 mg as IgG), whereas it was scarcely affected by the addition of anti-CYP3A2 antiserum (Fig. 3a). On the other hand, the mexazolam metabolism in female rat liver microsomes was inhibited by about 80% after the addition of anti-CYP3A2 antiserum (0.25 mg as IgG) but was scarcely affected by the addition of anti-CYP2C11 antiserum (Fig. 3b). In the Dex-treated rats, however, the mexazolam metabolism in liver microsomes was inhibited by about 80% in male rats and by about 90% in female rats, after the addition of anti-CYP2C11 antiserum (0.25 mg as IgG). In addition, following the addition of anti-CYP2C11 antiserum (0.25 mg as IgG), the metabolism in male and female rats was also inhibited by about 40% and by about 50%, respectively (Fig. 4, a and b).

**Evaluation of Kinetic Parameters for Mexazolam Metabolism in Rat Liver Microsomes.** The concentration dependence of mexazolam metabolism in control and Dex-treated rat liver microsomes was examined by Eadie-Hofstee plots in Fig. 5. As can be seen, saturation was reached in the formation of mexazolam metabolite, M-1. The calculated kinetic parameters of K_m and V_max are summarized in Table 1. In the control rats, the calculated K_m values in female and male rats were almost identical, but the V_max value and the CL_int in male rats were about 11 times higher than those in female rats. In the Dex-treated rats as well, the calculated K_m values in the female and male rats were almost identical, and no difference was observed between the control and Dex-treated rats. The V_max value in the Dex-treated female rats was about 1.3 times higher than that in the Dex-treated male rats, and the V_max values in the Dex-treated female and male rats were higher than those in the control female and male rats by about 39- and 2.7-fold, respectively. The intrinsic clearance values in Dex-treated female and male rats were also higher than those obtained in the control female and male rats by about 37- and 4.0-fold, respectively.

**Effects of HMG-CoA Reductase Inhibitors on Mexazolam Metabolism in Rat Liver Microsomes.** The K_i values of various HMG-CoA reductase inhibitors for mexazolam metabolism in female and male rat liver microsomes were obtained from Dixon plots (Fig. 6, a and b) and summarized in Table 2. Mexazolam metabolism in female rat liver microsomes was not inhibited by pravastatin (acid) but was inhibited competitively by other HMG-CoA reductase inhibitors (Fig. 6b). The order of inhibition potency of these HMG-CoA reductase inhibitors observed, in decreasing order, was simvastatin (lactone), pravastatin (lactone), atorvastatin (acid), cerivastatin (acid), fluvas statin (acid), and simvastatin acid. On the other hand, mexazolam metabolism in male rat liver microsomes was inhibited by all the HMG-CoA reductase inhibitors except simvastatin acid and pravastatin (acid) (Fig. 6a). The order of inhibition potency in decreasing order was simvastatin (lactone), atorvastatin (acid), cerivastatin (acid), fluvastatin (acid), and pravastatin (lactone), demonstrating that the order of inhibition potency differed between female and male rats. Furthermore, the HMG-CoA reductase inhibitors were less potent as inhibitors of mexazolam metabolism in male rat liver microsomes than in female rat liver microsomes (Table 2). In the Dex-treated female and male rat liver microsomes, mexazolam metabolism was inhibited competitively by simvastatin (lactone), pravastatin (lactone), and simvastatin acid but was scarcely affected by pravastatin (acid) (Fig. 7, a and b).

**Evaluation of Kinetic Parameters for Mexazolam Metabolism in Rat Liver Microsomes.** The formation of mexazolam metabolite in control female rat liver microsomes was considered to be inhibited by the dimethylacetamide used to dissolve the itraconazole, and the amount of metabolite formed was less than the detection limit. The mexazolam metabolism in male rat liver microsomes was not inhibited by itraconazole (Fig. 8a). On the other hand, in the Dex-treated female and male rat liver microsomes, mexazolam metabolism was noncompetitively inhibited by itraconazole, with K_i values of 0.693 μM in female rats and 0.877 μM in male rats (Fig. 8, b and c).

**Discussion**

We have previously reported that mexazolam is metabolized mainly by CYP3A4 in humans (Ono et al., 1993) and that this metabolism by human liver microsomes is inhibited by a variety of HMG-CoA reductase inhibitors (Ishigami et al., 2001a). In the present study, we investigated the inhibition of mexazolam metabolism in male and female rat liver microsomes in vitro by HMG-CoA reductase inhibitors and compared the results with those previously re-
ported in humans to identify an appropriate animal model for the study of drug interaction via CYP3A4 inhibition. Experiments using anti-rat P450 antisera demonstrated that the P450 isozymes responsible for mexazolam metabolism in rat liver microsomes differ between female and male rats. In the male rat liver microsomes, CYP2C11 was the major contributor to mexazolam metabolism, whereas in the female rat liver microsomes, the CYP3A isoforms were the main contributors (Fig. 3).

In mature female rat liver, CYP3A2 and CYP3A1 are reported to be expressed at extremely low levels (Cooper et al., 1993). Therefore, the significantly lower $V_{\text{max}}$ value for mexazolam metabolism in female rats than that in male rats may be due to the low expression level of the CYP3A isoforms responsible for mexazolam metabolism in female rats. On the other hand, CYP2C11, the male rat-specific P450 isozyme is expressed in the liver to a greater extent than the CYP3A isoforms, leading to a much higher level of mexazolam metabolism in

Fig. 7. Dixon plots for the inhibition of M-1 formation from mexazolam in Dex-treated male (a) and Dex-treated female (b) rat liver microsomes by simvastatin (lactone), simvastatin acid, pravastatin lactone, and pravastatin (acid).

Mexazolam (5–50 μM) was incubated for 2 min at 37°C with pooled rat liver microsomes (5 Dex-treated male rats or 5 Dex-treated female rats; 0.2 mg of protein/ml) in the presence or absence of simvastatin acid, pravastatin lactone, and pravastatin (acid) (20–200 μM); and simvastatin (lactone) (5–20 μM); □, 5 μM mexazolam; ★, 10 μM mexazolam; □, 20 μM mexazolam; ▲, 50 μM mexazolam. $V_{\text{max}}$, M-1 formation rate (nanomoles per minute per milligram of protein).
Mexazolam (5–50 μM) was incubated for 2 min at 37°C with pooled rat liver microsomes (5 male rats, 5 Dex-treated male rats, or 5 Dex-treated female rats; 0.2 mg of protein/ml) in the presence or absence of itraconazole (0.1–1 μM). □, 5 μM mexazolam; ■, 10 μM mexazolam; ▲, 20 μM mexazolam; ▼, 50 μM mexazolam. Vm, M-1 formation rate (nanomoles per minute per milligram of protein).

In conclusion, since species and sex differences are observed in P450 isoymes, appropriate experimental conditions that take into account P450 isoymes responsible for the drug metabolism in question will be indispensable for the investigation of drug interactions using rats as a model animal for humans.

References


