GLUCURONIDATION OF ETOPOSIDE IN HUMAN LIVER MICROSONES IS SPECIFICALLY CATALYZED BY UDP-GLUCURONOSYLTRANSFERASE 1A1

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ABSTRACT:
A metabolite formed by incubation of human liver microsomes, etoposide, and UDP-glucuronic acid was identified as etoposide glucuronide by liquid chromatography-tandem mass spectrometry analysis. According to the derivatization with trimethylsilylimidazole (Tri-Sil-Z), it was confirmed that the glucuronic acid is linked to an alcoholic hydroxyl group of etoposide and not to a phenolic group. Among nine recombinant human UGT isoforms (UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A8, UGT1A9, UGT1A10, UGT2B7, and UGT2B15), only UGT1A1 exhibited the catalytic activity of etoposide glucuronidation. The enzyme kinetics in pooled human liver microsomes and recombinant UGT1A1 microsomes showed a typical Michaelis-Menten plot. The kinetic parameters of etoposide glucuronidation were \( K_m = 439.6 \pm 70.7 \mu M \) and \( V_{max} = 255.6 \pm 19.2 \) pmol/min/mg of protein in human liver microsomes and \( K_m = 503.2 \pm 110.2 \mu M \) and \( V_{max} = 266.5 \pm 28.6 \) pmol/min/mg of protein in recombinant UGT1A1. The etoposide glucuronidation in pooled human liver microsomes was inhibited by bilirubin (IC \(_{50} = 31.7 \mu M \)) and estradiol (IC \(_{50} = 34 \mu M \)) as typical substrates for UGT1A1. The inhibitory effects of 4-nitrophenol (IC \(_{50} = 121.0 \mu M \)) as a typical substrate for UGT1A6 and UGT1A9, imipramine (IC \(_{50} = 393.8 \mu M \)) as a typical substrate for UGT1A3 and UGT1A4, and morphine (IC \(_{50} = 109.3 \mu M \)) as a typical substrate for UGT2B7 were relatively weak. The interindividual difference in etoposide glucuronidation in 13 human liver microsomes was 78.5-fold (1.4–109.9 pmol/min/mg of protein). The etoposide glucuronidation in 10 to 13 human liver microsomes was significantly correlated with \( \beta \)-estradiol-3-glucuronidation (\( r = 0.841, p < 0.01 \)), bilirubin glucuronidation (\( r = 0.935, p < 0.01 \)), and the immunonquantified UGT1A1 protein content (\( r = 0.800, p < 0.01 \)). These results demonstrate that etoposide glucuronidation in human liver microsomes is specifically catalyzed by UGT1A1.

Etoposide [4’-demethyllepipodophyllotoxin-9-(4,6-O-ethylidene)-\( \beta \)-d-glucopyranoside] is one of the clinically important antitumor agents derived from 4’-deoxylepothophyllotoxin, which is an extract from the plants Podophyllum peltatum and Podophyllum emodi (Clark and Slevin, 1987; Stühlen and von Wartburg, 1991). It is widely used in the treatment of testicular cancer, small cell lung cancer, and certain lymphomas (O’Dwyer et al., 1985). Etoposide causes tumor cell killing through DNA strand breakage resulting from the interaction of etoposide with the enzyme topoisomerase II and DNA (Ross et al., 1984).

In humans, the disposition of etoposide is described as a biphasic process with a distribution half-life of about 1.5 h and terminal elimination half-life ranging from 4 to 11 h (PDR, 2000). Clearance of etoposide occurs by direct renal excretion and metabolism. Roughly 35% of the administered drug is excreted into urine as a parent drug (Hande et al., 1984; Sinkule et al., 1984), but less than 3% is excreted into bile (Joel et al., 1996). Several metabolites were identified in human plasma and urine such as hydroxy acid derivatives, cis-(picro) lactone, 3’-demethylleptoside, and etoposide glucuronide (Clark and Slevin, 1987; Stewart, 1994). It has been reported that 58 and 19% of the administered drug is excreted as hydroxy acid into urine and bile, respectively (Clark and Slevin, 1987); less than 5 and 1% of the administered drug is found as cis-(picro) lactone in plasma and urine, respectively (Holthuis et al., 1986). It has been reported that 3’-demethylation of etoposide, a minor metabolite, is catalyzed by CYP3A4 (Relling et al., 1994). Etoposide glucuronides account for the disposition of 15 to 35% of administered etoposide (D’Incalci et al., 1986; Hande et al., 1988).

Glucuronidation is catalyzed by UDP-glucuronosyltransferase (UGT1) enzymes (Miners and Mackenzie, 1991). It is well known that there are many isoforms of mammalian UGT enzymes (Tukey and Strassburg, 2000). UGT1 and UGT2 have been shown to catalyze the glucuronidation of xenobiotics. The UGT1 and UGT2 genes seem to be structurally different in that UGT1 proteins result from alternate splicing of different first exons with five shared exons encoded by the UGT1 gene complex, whereas UGT2 proteins seem to be encoded by unique genes. In the human genome, at least 13 different first exons have been identified for the UGT1 gene (Gong et al., 2001). Currently, several common genetic polymorphisms in UGTs have been described, although their effects on the potency of glucuronidations have not been well studied. The purpose of the present study was to identify UGT isoform(s) involved in etoposide glucuronidation and to

1 Abbreviations used are: UGT, UDP-glucuronosyltransferase; LC-MS/MS, liquid chromatography-tandem mass spectrometry; ESI, electrospray ionization; APCI, atmospheric pressure chemical ionization.
characterize the interindividual variability in etoposide glucuronida-
tion using human liver microsomes. UGT isoforms relevant to this
variability were identified by screening a panel of recombinant UGTs
microsomes and by chemical inhibition of specific UGT substrates.
Furthermore, correlation analyses of etoposide glucuronosyltransfer-
ase activity with glucuronosyltransferase activities of other sub-
strates and immunoquantified UGT1A1 content were determined.

Materials and Methods

Materials. Etoposide, bilirubin, 4-nitrophenol, and imipramine hydrochlor-
ide were purchased from Wako Pure Chemicals (Osaka, Japan). UDP-glucu-
ronic acid, alamethicin, β-estradiol, p-nitrophenyl β-glucuronide, and α-naph-
thyl β-glucuronide were from Sigma-Aldrich (St. Louis, MO). Morphine
hydrochloride was purchased from Takeda Chemical Industries (Osaka, Ja-
pan). Morphine-3-glucuronide was kindly provided by Dr. Kazuta Oguri
(Kyusyu University, Fukuoka, Japan). Tri-Sil-Z, a mixture of trimethylsilyl-
limazolode in dry pyridine (1.5 mM/Pml = 1 part trimethylsilylimidazole:4
parts pyridine), was purchased from Pierce Chemical (Rockford, IL). Pooled
human liver microsomes (H161) and microsomes from 13 individual human
livers (H003, H023, H042, H043, H056, H064, H066, H089, H093, H095,
H112, HK23, and HK34) were prepared from BD Gentest (Woburn, MA).
The glucuronosyltransferase activities of β-estradiol (UGT1A1), trifluoroe-
prazine (UGT1A4), and propofol (UGT1A9) in these human liver microsomes
were provided by the manufacturer. The glucuronosyltransferase activities
of bilirubin (UGT1A1) and immunoquantified UGT1A1 protein in these human
liver microsomes except for H064, H095, and HK34 were also provided by the
manufacturer. Recombinant human UGT1A1, UGT1A3, UGT1A4, UGT1A6,
UGT1A8, UGT1A9, UGT1A10, UGT2B7, and UGT2B15 expressed in bac-
ulovirus-infected insect cells (Supersomes) were also from BD Gentest. All
other chemicals and solvents were of the highest grade commercially available.

Preparation of Rat Liver Microsomes. Male Wistar and Gunn rats (6
weeks old, n = 3) were purchased from Japan SLC (Hamamatsu, Japan). Liver
microsomes from untreated rats were prepared as described previously
(Guengerich, 1994). Liver microsomes were suspended in 10 mM Tris-HCl
buffer (pH 7.4) containing 10 mM EDTA and 20% (v/v) glycerol. After
determination of the protein concentration (Lowry et al., 1951), the suspended
microsomal samples were frozen and stored at −80°C until use.

Etoposide Glucuronidation Assay. A typical incubation mixture (200-μl
total volume) contained 25 mM Tris-HCl buffer (pH 7.4), 5 mM MgCl2, 1 mM
UDP-glucuronic acid, 50 μg/ml alamethicin, 1.0 mg/ml liver microsomes (1
mg/ml for recombinant UGT), and 400 μM etoposide. The reactions were
initiated by the addition of UDP-glucuronic acid and were then incubated at
37°C for 60 min. The reactions were terminated by boiling at 100°C for 10
min. After removal of the protein by centrifugation at 10,000 rpm for 5 min,
the glucuronosyltransferase activity with glucuronosyltransferase activities of other sub-
strates and immunoquantified UGT1A1 content were determined.

[M - H]⁻ ion at m/z 587 and [M + CH₃COOH-H]⁻ ion at m/z 647 corresponds to etoposide (A). [M – H]⁻ ion at m/z 763 corresponds to etoposide glucuronide (B). Product ion spectrum showed peaks at m/z 381 corresponding to the podophyllotoxin moiety (C and D). The product ion at m/z 455 corresponded to the trimethylsilylated podophyllotoxin moiety (E and F).
381 (due to the loss of glycopyranosyl moiety of etoposide), 337, 322, and 307 (Fig. 1D) were the same as those of the product ions of etoposide (Fig. 1C). From these observations, it was confirmed that the peak formed by the incubation of etoposide with human liver microsomes and UDP-glucuronic acid was etoposide glucuronide. After the treatment of etoposide with Tri-Sil-Z, the APCI mass spectrum of the peak showed [M+H]/H ion at m/z 805 corresponding to tri-trimethylsilylated etoposide (Fig. 1E). The product ion at m/z 455 corresponded to the trimethylsilylated podophyllotoxin moiety (Fig. 1E). After the treatment of the metabolite with Tri-Sil-Z, the APCI mass spectrum of the peak did not show the parent ion. However, the product ion at m/z 455 (trimethylsilylated podophyllotoxin moiety), which was the same as the product ion obtained from the treatment of etoposide was obtained (Fig. 1, E and F). These results suggested that glucuronic acid is linked to the alcoholic hydroxyl group and not to the phenolic group.

Etoposide Glucuronidation in Recombinant UGT Isoforms. Ten recombinant UGT isoforms expressed in baculovirus-infected insect cells were used to determine their etoposide glucuronosyltransferase activities. As shown in Fig. 2, only UGT1A1 exhibited etoposide glucuronosyltransferase activities (103.2 pmol/min/mg of protein).

Kinetics of Etoposide Glucuronidation in Human Liver Microsomes and Recombinant Human UGT1A1. Kinetic analyses of etoposide glucuronidation in pooled human liver microsomes (H161) and recombinant human UGT1A1 were performed. The kinetics fitted the Michaelis-Menten kinetics (Fig. 3, A and B) and the Eadie-Hofstee plots were monophasic (Fig. 3, C and D). When the apparent enzyme kinetic parameters were estimated, the $K_m$ value was 439.6 ± 70.7 μM and the $V_{max}$ value was 255.6 ± 19.2 pmol/min/mg of protein for human liver microsomes; the $K_m$ value was 503.2 ± 110.2 μM and the $V_{max}$ value was 266.5 ± 28.6 pmol/min/mg of protein for recombinant human UGT1A1.

Inhibition Analyses of Etoposide Glucuronidation in Human Liver Microsomes. The effects of bilirubin (UGT1A1), β-estradiol (UGT1A1 and UGT1A9), 4-nitrophenol (UGT1A6 and UGT1A9), imipramine (UGT1A3 and UGT1A4), and morphine (UGT2B7) on the etoposide glucuronosyltransferase activity in the pooled human liver microsomes were determined. As shown in Fig. 4, the etoposide glucuronosyltransferase activity in the pooled human liver microsomes was inhibited by bilirubin (IC₅₀ = 31.7 μM) and β-estradiol (IC₅₀ = 34 μM). The inhibitory effects of 4-nitrophenol (IC₅₀ = 121.0 μM), imipramine (IC₅₀ = 393.8 μM), and morphine (IC₅₀ = 109.3 μM) were relatively weak.

Interindividual Variability of Etoposide Glucuronosyltransferase Activity in Microsomes from 13 Human Livers and Cor...
Etoposide glucuronosyltransferase activities in microsomes from 13 human livers were determined (A). Human liver microsomes (1 mg/ml microsomal protein) were incubated with 400 μM etoposide and 3 mM UDP-glucuronic acid at 37°C for 60 min. Each data point represents the mean of duplicate determinations. Correlation analysis between the etoposide glucuronosyltransferase activity and bilirubin glucuronosyltransferase activity (B, n = 10), β-estradiol-3-glucuronosyltransferase activity (C, n = 13), and the content of UGT1A1 protein (D, n = 10) in human liver microsomes.

**Fig. 5.** Interindividual variability in etoposide glucuronosyltransferase activity in microsomes from 13 human livers (A) and correlation analyses with enzymatic activities (B and C) and content of UGT1A1 (D).

Etoposide glucuronosyltransferase activities in microsomes from 13 human livers were determined at 400 μM etoposide and 3 mM UDP-glucuronic acid at 37°C for 60 min. Each data point represents the mean of duplicate determinations. Correlation analysis between the etoposide glucuronosyltransferase activity and bilirubin glucuronosyltransferase activity (B, n = 10), β-estradiol-3-glucuronosyltransferase activity (C, n = 13), and the content of UGT1A1 protein (D, n = 10) in human liver microsomes.

**Relation with the Other Glucuronosyltransferase Activities.** Etoposide glucuronosyltransferase activities in microsomes from 13 human livers were determined at 400 μM etoposide (Fig. 5A). The interindividual difference in etoposide glucuronosyltransferase activity was as large as 78.5-fold (1.4–109.9 pmol/min/mg of protein). Correlation analyses were performed between the etoposide glucuronosyltransferase activity and bilirubin glucuronosyltransferase activity was significantly correlated with the bilirubin (r = 0.935, P < 0.01) and β-estradiol (r = 0.841, P < 0.01) glucuronosyltransferase activities. Furthermore, the etoposide glucuronosyltransferase activity was significantly correlated with the immunoquantified UGT1A1 protein contents (Fig. 5D). In contrast, the etoposide glucuronosyltransferase activity was not correlated with the trifluoperazine (r = 0.204), propofol (r = −0.075), 4-nitrophenol (r = 0.160), 1-naphthol (r = 0.232), or morphine (r = −0.017) glucuronosyltransferase activities.

**Kinetics of Etoposide Glucuronidation in Liver Microsomes from Wistar and Gunn Rats.** Kinetic analyses of etoposide glucuronidation in liver microsomes from Wistar (n = 3) and Gunn (n = 3) rats were performed. The etoposide glucuronidation in liver microsomes from Wistar rats fitted the Michaelis-Menten kinetics (Fig. 6). The Km value was 565.5 μM and Vmax value was 194.7 pmol/min/mg of protein.

**Discussion**

A number of approaches have been developed for the identification of human UGTs involved in the glucuronidation of endogenous and exogenous compounds in vitro (Ritter, 2000; Tukey and Strassburg, 2000). It has been reported that etoposide is metabolized to its glucuronide in rats and humans (D’Incalci et al., 1986; Hande et al.,...
that the etoposide glucuronosyltransferase activity in recombinant UGT1A1. As shown in Fig. 4, inhibition by 4-nitrophenol, imipramine, and morphine, respectively. Therefore, the observed inhibitory effects of these compounds on the etoposide glucuronosyltransferase activity in human liver microsomes could be due to the inhibition of UGT1A1. The etoposide glucuronosyltransferase activities in 13 human liver microsomes were significantly correlated with the bilirubin and β-estradiol glucuronosyltransferase activities and immunoquantified UGT1A1 protein content. These results suggest that UGT1A1 would specifically catalyze the etoposide glucuronidation in human liver microsomes.

The Gunn rat has been reported to lack the UGT1A1 enzymes (Gunn, 1938; Iyanagi, 1991; Sato et al., 1991). In the present study, we demonstrated that the etoposide glucuronosyltransferase activity was not detected in liver microsomes from Gunn rats. In contrast, Wistar rats produced the etoposide glucuronide with a \( K_m \) value that was similar to that in human liver microsomes. It has been reported that human and rat UGT1A1 enzymes have similar substrate specificity (King et al., 1996) with 89% of protein sequence similarity (Mackenzie et al., 1997). Our results suggest that etoposide glucuronidation is also catalyzed by UGT1A1 in rats.

We first demonstrated that there is a large interindividual difference in etoposide glucuronidation in humans (78.5-fold). UGT1A1 has been reported to be induced by phenobarbital, phenytoin, oltipraz, and 3-methylcholanthrene (Fisher et al., 2001). Human liver microsomes of H112 showed the highest etoposide glucuronosyltransferase activity (Fig. 6A). From the manufacturer’s information, the donor had taken phenobarbital. Therefore, UGT1A1 in H112 might be induced by phenobarbital. Furthermore, it is known that there are genetic polymorphisms in UGT1A1 (Tukey and Strassburg, 2000). Several alleles such as UGT1A1*6 (G71R) and UGT1A1*28 (TAa-TAA) of UGT1A1 have been reported to cause a significant reduction of UGT activity toward bilirubin (Yamamoto et al., 1998) and to be associated with Gilbert’s syndrome (Bosma et al., 1995; Monaghan et al., 1996). Therefore, the genetic polymorphisms in UGT1A1 might be a one of factors causing the interindividual differences in etoposide glucuronidation in humans, which will be clarified in the near future.

In conclusion, we found that etoposide glucuronidation in using human liver microsomes is specifically catalyzed by UGT1A1. Large interindividual differences in etoposide glucuronidation in human liver microsomes were observed. These results may have important implications for the clinical variability in efficacy and adverse reactions of etoposide.

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References


