Sulfinpyrazone \([1,2\text{-diphenyl-4-(phenylsulfinylethyl)-3,5-pyrazolidinedione}\) (SFZ) is a uricosuric agent used primarily to treat hyperuricemia associated with gout (Pittman and Bross, 1999). SFZ also possesses antiplatelet and antithrombotic actions (Margulies et al., 1980). It has been demonstrated that C-glucuronidation is a major biotransformation pathway of SFZ in humans (Dieterle et al., 1975, 1979), resulting in the formation of a conjugate (sulfinpyrazone \(\beta\)-d-glucuronide; SFZG). SFZ C-glucuronidation by HLMs exhibited Michaelis-Menten kinetics, with mean \((\pm S.D.)\) \(K_m\) and \(V_{\max}\) values of 51 \(\pm\) 21 \(\mu\)M and 2.6 \(\pm\) 0.6 pmol/min \(\cdot\) mg, respectively. Fifteen recombinant human UDP-glucuronosyltransferases (UGTs), expressed in HEK293 cells, were screened for their capacity to catalyze SFZ C-glucuronidation. Of the hepatically expressed enzymes, only UGT1A9 formed SFZG. UGTs 1A7 and 1A10, which are expressed in the gastrointestinal tract, also metabolized SFZ, but rates of metabolism were low compared with UGT1A9. SFZ glucuronidation by UGT1A9 exhibited “weak” negative cooperative kinetics, which was modeled by the Hill equation \((S_{50} 16 \ \mu\text{M})\). The data indicate that UGT1A9 is the enzyme responsible for hepatic SFZ C-glucuronidation and that SFZ may be used as a substrate “probe” for UGT1A9 activity in HLMs.

**Materials.** Alamethicin (from *Trichoderma viride*), \(\beta\)-glucuronidase (from *Escherichia coli*), SFZ, and UDPGA (trisodium salt) were purchased from Sigma-Aldrich (Sydney, Australia). All other reagents and solvents were of analytical reagent grade.

**Human Liver Microsomes and Recombinant UGT Proteins.** Human livers (HL10, 12, 13, and 40) were obtained from the human liver “bank” of the Department of Clinical Pharmacology, Flinders Medical Centre. Micro-
somes were prepared by differential centrifugation, as described by Bower et al. (2005), and stored at −80°C until use. Human liver microsomes (HLMs) were activated by preincubation with alamethicin (50 μg/ml protein) on ice for 30 min before use in incubations (Boase and Miners, 2002). UGTs 1A1, 1A3, 1A4, 1A5, 1A7, 1A8, 1A9, 1A10, 2B4, 2B7, 2B10, 2B15, 2B17, and 2B28 cDNAs were stably expressed in a human embryonic kidney cell line (HEK293), as described previously (Stone et al., 2003; Uchaipichat et al., 2004). Expression of each UGT protein was demonstrated by immunoblotting with a commercial UGT1A antibody and a nonelective UGT antibody (raised against purified mouse Ugt1A) (Uchaipichat et al., 2004). Activities of recombinant UGTs (except UGTs 1A4, 1A5, and 2B10) were confirmed using the nonselective substrate 4-methylumbelliflorone, as described by Rowland et al. (2006). The activity of UGT1A4 was confirmed using trifluoroacetic acid as substrate (Uchaipichat et al., 2006).

**SFZ Glucuronidation Assay.** Incubation mixtures, in a total volume of 200 μl, contained HLMs or HEK293 cell lysate protein (2 mg/ml), UDPGA (5 mM), MgCl2 (4 mM), and SFZ (11–12 substrate concentrations in the range of 2.5–400 μM for kinetic studies) in 0.1 M phosphate buffer, pH 7.4. SFZ stock solutions were prepared in methanol, such that incubations contained 1% solvent (v/v). This concentration of methanol has been shown to have a negligible effect on UGT activities (Uchaipichat et al., 2004). After a 5-min preincubation at 37°C, reactions were initiated by the addition of UDPGA. Incubations were carried out for 3 h at 37°C in a shaking water bath. “Blank” incubations, which contained methanol (1% v/v), were performed in the absence of UDPGA. Reactions were terminated by the addition of ice-cold 4% acetic acid/96% methanol (200 μl). After centrifugation (12,000 g for 10 min), a 40-μl aliquot of the supernatant fraction was injected onto an Agilent 1100 HPLC system (Agilent Technologies, Palo Alto, CA) fitted with a Waters Nova-Pak C18 column (3.9 mm, 4 μm; Waters, Milford, MA) with a Phenomenex Security Guard Cartridge (Phenomenex, Torrance, CA). Peaks were separated with 90% 10 mM triethylamine, pH 2.5 (adjusted with HClO4)/10% acetonitrile (mobile phase A) and 100% acetonitrile (mobile phase B) as follows: initial conditions of 80% A/20% B for 2 min, followed by a linear gradient over 10 min to 60% A/40% B, which was held for 1 min before returning to the starting conditions. The mobile phase flow rate was 1.0 ml/min and peaks were monitored by UV detection at 240 nm (the absorbance maximum for SFZ in the mobile phase). Under these conditions, retention times for SFZG and SFZ were 6.0 and 12.1 min, respectively. SFZG was quantified by comparison of peak areas to those of an SFZ standard curve prepared over the concentration range 0.25 to 2 μM. SFZ standard curves were linear over this concentration range, with r2 values >0.99. The lower limit of quantification, defined as 5 times background, was 0.01 μM (equivalent to a rate of product formation of 0.93 pmol/min mg). Overall within-day assay reproducibility was assessed by measuring SFZG formation in 10 separate incubations of the same batch of HLMs. Within-day coefficients of variation were 1.8% and 2.7% for substrate concentrations of 10 and 250 μM, respectively. The formation of SFZG was linear with incubation times to at least 4 h and microsomal protein concentrations to at least 4 mg/ml (data not shown).

**Isolation of SFZG.** SFZ was incubated with pooled HLMs (10 mg), in a total volume of 5 ml, for 15 h. The reaction was terminated with 85% H3PO4 (50 μl) and centrifuged (12,000 g for 10 min). The supernatant fraction was separated and applied to a C18 Sep-Pak solid-phase extraction cartridge (Waters), which was preconditioned with 2 ml of methanol, 5 ml of water, and 2 ml of acidified water (pH 2). After application of the sample, the cartridge was washed with 1 ml of acidified water (pH 2) and 5 ml of water, dried under vacuum, and eluted with methanol (4 ml). The eluate was evaporated to dryness under nitrogen and the residue was reconstituted in 200 μl of methanol/acetic acid/water (50:2:48, v/v). The fraction corresponding to the SFZG peak was separated by HPLC (see above), again extracted on a Sep-Pak C18 cartridge, and eluted with methanol for mass spectral analysis. A portion of this eluate was evaporated to dryness under nitrogen and reconstituted in water for the acid hydrolysis study (see below).

**Identification of SFZG.** The structural identity of the SFZG peak formed by incubations of HLMs was confirmed by mass spectrometry and by enzymatic and acid hydrolysis. Mass spectra were acquired using a Micromass Quattro micro tandem quadrupole mass spectrometer (Waters, Manchester, UK) with electrospray ionization in positive ion mode. The sample was introduced into the electrospray ionization source via an integrated syringe pump at a flow rate of 10 μl/min. Instrument parameters were as follows: capillary voltage, 3.2 kV; cone voltage, 35 V; source temperature, 90°C; desolvation temperature, 300°C; desolvation gas flow, 250 l/h; collision energy, 20 V; and gas cell Pirani pressure 3.78 × 10−3 mbar.

For the enzymatic hydrolysis experiment, an SFZ glucuronidation incubation sample, in a total volume of 200 μl, was terminated with 85% H3PO4 (2 μl) and centrifuged (12,000 g for 10 min). The supernatant fraction was decanted and adjusted to pH 7 with 6 M KOH. Enzymatic hydrolysis was performed by incubation with β-glucuronidase (100 units/ml) at 37°C for 2 h. Control incubations without β-glucuronidase were performed simultaneously. Acid hydrolysis was performed by adding 100 μl of 6 M HCl to 200 μl of the purified aqueous solution of SFZG (see above) and heating at 90°C for 1 h (since acid hydrolysis of incubation mixtures gave an interfering peak close to that of SFZG). Control incubations without HCl were performed simultaneously.

**Data Analysis.** Kinetic constants for SFZ C-glucuronidation by HLMs and recombinant UGTs were obtained by fitting experimental data to the Michaelis-Menten or Hill equations using EnzFitter (Biosoft, Cambridge, UK), to obtain estimates of Km (or Skm), maximal velocity (Vmax), and the Hill coefficient, n (as described by Uchaipichat et al., 2004). Goodness of fit to kinetic models was assessed by comparison of the F statistic, coefficient of determination (r2), parameter standard errors, and 95% confidence intervals.

**Results and Discussion.**

HPLC analysis of incubations containing SFZ and UDPGA revealed a metabolite peak with a retention time of 6.0 min, which was not observed in incubations performed without UDPGA or SFZ. The putative SFZG was resistant to β-glucuronidase and acid hydrolysis, as reported previously for C-glucuronides (Dieterle et al., 1975; Richter et al., 1975; Yasuda et al., 1982). The identity of the SFZG peak was thus confirmed by electrospray mass spectrometry in positive ion mode. Mass spectral analysis showed the presence of a protonated molecular ion [M + H]+ at m/z 581, which is consistent with the molecular weight of protonated SFZG. The major fragment ions, detected at m/z 455, 405, and 279, correspond to the loss of phenylsulfoxide, glucuronic acid, and both groups, respectively. Although phenylbutazone is reported to form both C- and O-glucuronides (the latter presumably via an enol), only SFZ C-glucuronide has been observed in the urine of humans administered SFZ (Dieterle et al., 1975, 1980). The occurrence of a single SFZG peak in microsomal incubations, which was unaffected by β-glucuronidase and acid hydrolysis, is consistent with the in vivo observations. The resistance of C-glucuronides to β-glucuronidase and acid hydrolysis should be...
Microsomal SFZ C-glucuronidation exhibited Michaelis-Menten kinetics in the four livers investigated (Fig. 2A). Mean (± S.D.) derived $K_m$ and $V_{max}$ values for the four livers were 51 ± 21 μM and 2.6 ± 0.6 pmol/min · mg, respectively. As noted previously, standard curves were prepared using SFZ. Thus, rates of SFZ C-glucuronide formation and $V_{max}$ values for this metabolite should be considered "apparent".

UGTs 1A1, 1A3, 1A4, 1A5, 1A6, 1A7, 1A8, 1A9, 1A10, 2B4, 2B7, 2B10, 2B15, 2B17, and 2B28 were screened for SFZ C-glucuronidation activity at substrate concentrations of 10, 50, and 250 μM. Of these enzymes, only UGTs 1A7, 1A9, and 1A10 converted SFZ to its C-glucuronide. UGT1A9 exhibited the highest activity, whereas UGT1A7 and 1A10 exhibited lesser activity (<14% that of UGT1A9) (Fig. 2C). Given the lower activities of UGTs 1A7 and 1A10, and lack of expression of these enzymes in liver (see below), kinetic studies were performed only with UGT1A9. Although kinetic data were adequately fitted to the Michaelis-Menten equation, SFZ C-glucuronidation by UGT1A9 was best described by the Hill equation (with $n = 0.77 ± 0.01$, i.e., negative cooperativity) (Fig. 2B); all goodness of fit parameters (see Data Analysis) were superior using this model.

The $S_{50}$ and $V_{max}$ (parameter ± S.E. of parameter fit) values were 16 ± 0.3 μM and 3.2 ± 0.02 pmol/min · mg, respectively.

UGT1A9 is expressed in liver and a range of other human tissues (Tukey and Strassburg, 2000). In contrast, UGTs 1A7 and 1A10 are expressed only in the gastrointestinal tract and hence cannot contribute to SFZ C-glucuronidation by HLMs (or hepatic SFZ clearance in vivo). Thus, on the basis of the screening data, it is postulated that human hepatic SFZ C-glucuronidation is catalyzed solely by UGT1A9, and SFZ may thus serve as a substrate "probe" for UGT1A9 activity in HLMs. It should be noted that no selective inhibitors of UGT1A9 are currently available, and thus confirmatory inhibition experiments are not possible. Whereas SFZ is a selective substrate of UGT1A9, we have demonstrated recently that both SFZ and phenylbutazone inhibit most UGT1A subfamily enzymes (Uchipingat et al., 2006), although the lowest IC$_{50}$ values were associated with those enzymes shown here to glucuronidate SFZ (UGT1A7, 1A9, and 1A10). This observation provides further evidence for the principle that substrate and inhibitor selectivities may not be identical with drug-metabolizing enzymes. It is interesting to note that differences in the kinetic models for SFZ C-glucuronidation by HLM (Michaelis-Menten) and UGT1A9 expressed in HEK293 cells ("weak" negative cooperativity) were observed here. The reasons for
this are unclear, but non-Michaelis Menten kinetics are not uncommonly observed for xenobiotic glucuronidation reactions by recombinant UGTs (Miners et al., 2004, 2006), including UGT1A9 (Uchaipichat et al., 2004; Bowalgaha et al., 2005).

Apart from SFZ, propofol has been proposed as a selective substrate for hepatic UGT1A9 activity (Court, 2005), but detailed comparative kinetic studies have not been published. Like SFZ, propofol also appears to be a substrate for UGTs expressed exclusively in the gastrointestinal tract (viz. 1A7, 1A8, and 1A10) (Court, 2005). Although the apparent $V_{\text{max}}$ for SFZ C-glucuronidation by HLMs is relatively low, SFZG formation during the course of a standard incubation at a substrate concentration corresponding to the $K_m$ is almost 500 pmol, which is very easily measurable. When considered as a substrate probe for human liver microsomal UGT1A9 activity, SFZ offers advantages compared with propofol, including well characterized hyperbolic kinetics, ease of handling (solid versus liquid), solubility, and negligible nonspecific binding to HLMs (<5% for SFZ (O. Kerdpin and J.O. Miners, unpublished results) versus 95% for propofol (Soars et al., 2002)).

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