Two Nonsynonymous Single Nucleotide Polymorphisms of Human Carbonyl Reductase 1 Demonstrate Reduced In Vitro Metabolism of Daunorubicin and Doxorubicin

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ABSTRACT:

Carbonyl reductases (CBRs) are a group of metabolic enzymes belonging to the short-chain dehydrogenase family with NADPH-dependent oxidoreductase activity. These enzymes are known to metabolize the anthracyclines doxorubicin (DOX) and daunorubicin (DAUN). Both DOX and DAUN are highly effective in cancer therapy; however, there is considerable interpatient variability in adverse effects seen in patients undergoing treatment with these drugs. This may be attributed to altered metabolism associated with nonsynonymous single nucleotide polymorphisms (ns-SNPs) in the genes encoding for CBRs. In this study, we examine the effect of the V88I and P131S mutations in the human CBR1 gene on the metabolism of anthracyclines to their respective major metabolites, doxorubicinol and daunorubicinol. Kinetic studies using purified, histidine-tagged, recombinant enzymes in a high-performance liquid chromatography-fluorescence assay demonstrated that the V88I mutation leads to a significantly reduced maximal rate of activity ($V_{\text{max}}$) ($2090 \pm 112$ and $257 \pm 11 \text{ nmol/min \cdot mg}$ of purified protein for DAUN and DOX, respectively) compared with that for the wild-type ($3430 \pm 241$ and $364 \pm 37 \text{ nmol/min \cdot mg}$ of purified protein for DAUN and DOX, respectively). In the case of the P131S mutation, a significant increase in substrate affinity ($K_{\text{m}}$) was observed for DAUN only ($89 \pm 13 \mu M$) compared with that for the wild-type ($51 \pm 13 \mu M$). In the presence of either anthracycline, both variants exhibited a 20 to 40% decrease in catalytic efficiency ($k_{\text{cat}}/K_{\text{m}}$) compared with that for the wild-type enzyme. Therefore, the ns-SNPs generating both these mutations may alter bioavailability of these anthracyclines in cancer patients and should be examined in clinical studies as potential biomarkers for DAUN- and DOX-induced adverse effects.

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ABBR EVIATIONS: CBR, carbonyl reductase; DOX, doxorubicin; DAUN, daunorubicin; SNP, single nucleotide polymorphism; ns, nonsynonymous; DAUNol, daunorubicinol; DOXol, doxorubicinol; IPTG, isopropyl β-D-thiogalactopyranoside; 6x-His, six histidine; FXa, factor Xa; Ni-NTA, nickel-nitrioltriacetic acid; HPLC, high-performance liquid chromatography; ROS, reactive oxygen species.
cyclohexane and 26% in adults treated with \(\geq 550\) mg/m\(^2\) DOX (Deng and Wojnowski, 2007). Both the frequency of chronic cardiotoxicity and the mortality rate are associated with increased dosage of anthracyclines. In addition, the risk of chronic cardiotoxicity increases with the concurrent use of other anticancer drugs, such as Herceptin, paclitaxel, and cyclophosphamide (Minotti et al., 1999; Wojtacki et al., 2000; Danesi et al., 2002; Gianni et al., 2007).

The underlying cause(s) of interpatient variations in DAUN- or DOX-induced adverse drug events may be altered enzyme activity associated with nonsynonymous nucleotide polymorphisms (nsSNPs) of the CBR genes, particularly human CBR1. There are two documented nsSNPs, one producing a change from a valine to isoleucine at amino acid position 88 (V88I; SNP ID: rs1143663 from National Center for Biotechnology Information Database; \(q = 2.1\%\)) and the other from a proline to serine at amino acid position 131 (P131S; SNP ID: rs41557318; \(q = 2.2\%\)). Although there is evidence for the catalytic properties of the V88I enzyme with DAUN (Gonzalez-Covarrubias et al., 2007), no information has been reported for DOX. In addition, there is no information on the catalytic properties of the P131S variant in the presence of either anthracycline. Therefore, our goal for this study was to compare the wild-type and variant enzymes for their ability to metabolize DAUN and DOX to their corresponding carbon-13 alcohol metabolites, daunorubicinol (DAU-Nol) and doxorubicinol (DOXol). Using purified, bacterially expressed, recombinant, human histidine-tagged enzymes, we demonstrated that the V88I variant resulted in significantly reduced metabolic activity for both DOX and DAUN, whereas the P131S gene product exhibited a significant increase in affinity with DAUN compared with that for the wild-type enzyme. We also found that DAUN is a better substrate than DOX for both the wild-type and variant enzymes. The metabolic/affinity differences seen with the mutant V88I and P131S enzymes compared with that of the wild-type enzyme could result in increased concentrations of DOX or DAUN in cancer patients and the increased occurrence of adverse side effects.

Materials and Methods

Chemicals and Enzymes. Agarose, chloramphenicol, daunorubicin, doxorubicin, iabadicin, kanamycin sulfate, lysozyme, menadione, methanol, potassium phosphate (KH\(_2\)PO\(_4\)), RNase I, sodium phosphate (NaH\(_2\)PO\(_4\)), N,N,N′,N′-tetramethylethylenediamine and NADPH were supplied by Sigma-Aldrich (St. Louis, MO). HPLC-grade acetonitrile, agar, ammonium persulfate, formic acid, ethanolic, glycine, glycerol, glacial acetic acid, imidazole, and tris were purchased from Thermo Fisher Scientific (Waltham, MA). NaCl and yeast extract were ordered from EMD Chemicals Inc. (Darmstadt, Germany). Bacto tryptone and isopropyl \(\beta\)-D-thiogalactopyranoside (IPTG) were obtained from BD Biosciences (Franklin Lakes, NJ) and MBI Fermentas (Hanover, MD), respectively. Tween 20 was purchased from EMD Biosciences (La Jolla, CA), and DNase I was provided by Boehringer Mannheim GmbH (Mannheim, Germany). Klunow fragment and T4 ligase were obtained from MBI Fermentas (Burlington, ON, Canada). Factor Xa and restriction enzymes were purchased from New England Biolabs (Ipswich, MA). Doxorubicinol was obtained from Qvents Inc. (Branford, CT).

Molecular Cloning of Human CBR1 Gene and Creation of the Genetic Variants. The CBR1 coding region, which was excised from a pOTB7 recombinant plasmid (MGC-1920; American Type Culture Collection, Manassas, VA) using BssHI (blunt end with Klenow fragment) and XhoI, was subcloned into Nhel (blunt end)-Xhol sites of the pET2a prokaryotic expression vector (Novagen, San Diego, CA) with T4 ligase. These constructs gave rise to a human CBR1 enzyme with a 6x-His tag separated by an 18-amino acid residue linker on the amino terminus. A factor Xa (FXa) site was inserted at the amino terminus between the linker and CBR1 gene using the QuikChange Site-Directed Mutagenesis Kit (Stratagene, La Jolla, CA) with the 5′-CCCGTTCACGCCATAGAAGGAAGATGTCGCCGCGCCG3′ (forward) and 5′-GCCGAGACGACATCTCTCTCTCTATGGCTGAACCGGGG3′ (reverse) primers. Afterward, the pET2a-V88I and pET2a-P131S constructs were created by site-directed mutagenesis with the following primers: 1) V88I: 5′-CTGAGCTGTTGTCAATCAGCCTACGCA-3′ (forward) and 5′-GGCGCTTGGTGATCAGCACGTCCAG-3′ (reverse); and 2) P131S: 5′-CTGTTGTCAATCAGCCTACGCA-3′ (reverse) and 5′-CCA-CCTCCCTCTGTTGGATTTATAGGAGGAG-3′ (reverse). A modified site-directed mutagenesis procedure was used in which two separate 50-µl reactions (one for each primer) were subjected to 10 cycles of denaturation at 95°C for 1 min, annealing at 60°C for 1.5 min, and extension at 68°C for 6.5 min. Afterward, 25 µl of each reaction were combined and 0.75 µl of PfuTurbo DNA polymerase (Stratagene) was added. This reaction was subjected to 18 cycles using the same PCR protocol. All constructs were verified by dideoxy sequencing at the University of British Columbia Nucleic Acid Protein Service unit.

Expression and Purification of Recombinant Human CBR1 Wild-Type and Variant Enzymes. The pET2a constructs of the CBR1 wild-type and variant enzymes were heat-shock transformed into Escherichia coli BL21 (DE3) pLYsS-competent cells and expressed under the control of an IPTG-inducible T7 polymerase. Cells were plated on Luria-Bertani (1% Bacto tryptone, 0.5% yeast extract, and 0.5% NaCl) broth agar supplemented with 50 µg/ml kanamycin sulfate and 25 µg/ml chloramphenicol for selection. Colonies were randomly picked and cultured overnight at 37°C in 3 ml of Luria-Bertani broth with kanamycin and chloramphenicol at the same concentrations stated previously. Cultures were expanded to 800 ml and grown at 37°C until an OD\(_{600}\) of 0.5 was reached. IPTG was added to a final concentration of 1 mM, and cells were allowed to grow for an additional 3 h. The cultures remaining after the induction were harvested by centrifugation (4000g for 20 min at 4°C), and the recombinant 6x-His-tagged proteins were purified using nickel-nitrilotriacetic acid (Ni-NTA) affinity chromatography and elution with imidazole according to the procedure described in Bains et al. (2008).

Protein purity was assessed by running elution fractions on 18% SDS-polyacrylamide gels, which were stained with SYPRO Ruby (Invitrogen Canada, Inc., Burlington, ON, Canada) overnight (16 h). After staining, the protein was detected using a Storm 840 Molecular Dynamics Imager (GE Healthcare, Chalfont St. Giles, UK) at excitation and emission wavelengths of 450 and 520 nm, respectively.

Western blot analyses of the purified fractions were conducted according to the procedure described for the Odyssey Infrared Imaging system (LI-COR Biosciences, Lincoln, NE). After 18% SDS-polyacrylamide gel electrophoresis, proteins were transferred at 20 V in Towbin’s buffer (25 mM Tris, 192 mM glycine, and 20% v/v methanol) overnight (at 4°C) to a Hybond-C Extra nitrocellulose membrane (GE Healthcare). The membranes were blocked in Odyssey blocking buffer, and the enzymelocalized antibody was detected using a monoclonal mouse anti-human CBR1 (Abnova Corporation, Taipei City, Taiwan) antibody (diluted 1:3000) as the primary antibody and IRDye 800CW goat anti-mouse IgG as the secondary antibody (diluted 1:5000) (LI-COR Biosciences). Both primary and secondary antibodies were in blocking buffer containing 0.1% Tween 20. The bound secondary antibodies were detected using the Odyssey Infrared Imaging system.

Kinetische Analysis of CBR1 and Variant Enzymes. The enzyme activities of the purified 6x-His-tagged CBR1 wild-type and variant enzymes were measured at 37°C using a Fluoroskan Ascent FL (Thermo Fisher Scientific) by following the initial rate of NADPH oxidation at excitation and emission wavelengths of 355 and 460 nm, respectively. The assays were conducted as described previously using menadione as the test substrate (Gonzalez-Covarrubias et al., 2007). In brief, purified protein was incubated with 1 mM NADPH and menadione (20–150 µM) in a reaction mixture of 150 µl of 100 mM potassium phosphate (pH 7.4). The protein amount and incubation times were selected for each enzyme and substrate concentration to ensure that rates were in the linear range of the enzyme kinetic curve. In these assays, the concentration of 95% ethanol, which was required to dissolve the substrate, was kept below 4% (v/v) in the final reaction mixture. Reactions were collected at 20-s intervals for 1.5 h with shaking between each reading. Maximal rates were calculated from the Ascent program (version 2.6) using a 5-min interval (15 total readings) with the steepest slope. Enzymatic activity (nanomoles of NADPH consumed per minute per milligram of purified protein) was calculated from these rates using a standard curve constructed from the fluorescence measurements of solutions of known NADPH concentrations.
Activity measurements for the reduction of the anthracyclines were performed by incubating either DOX or DAUN (1–400 μM) with purified enzyme in a total volume of 150 μl containing 100 mM potassium phosphate (pH 7.4) and 1 mM NADPH at 37°C. Protein concentrations were based on the Bradford protein assay using bovine serum albumin as a standard. The reaction was stopped by adding 300 μl of ice-cold acetonitrile, which contained idarubicin as an internal standard, followed by vortex mixing and centrifuging at 10,000g for 10 min at 4°C to remove protein. The supernatant was removed for high-performance liquid chromatography (HPLC) analysis. The procedures for HPLC separation and fluorescence detection of DOX and DAUN and their carbon-13 hydroxy metabolites, DOXol and DAUNol, were consistent with the protocol in Bains et al. (2008).

The kinetic constants of maximal rate of reaction (V_{max}) and substrate affinity (K_m) were determined by fitting rate measurement data using nonlinear least-squares fitting of a Michaelis-Menten hyperbola (version 4.0, GraphPad Prism; GraphPad Software Inc., San Diego, CA). The turnover values (kcat) were calculated from V_{max} values using the apparent molecular weight for the 6x-His-tagged CBRs and variant proteins of 34,000. Catalytic efficiency (kcat/K_m) was also calculated. After Michaelis-Menten data analysis, Eadie-Hofstee plots were created to check for deviation from linearity with changing substrate concentrations.

**Statistical Analysis.** Statistical analyses were performed using GraphPad Instat (version 3.6; GraphPad Software Inc.). Results were expressed as means ± S.D. Enzyme activities were compared using a one-way analysis of variance followed by Tukey-Kramer multiple comparisons tests. Differences were considered significant at p < 0.05.

**Results**

**Expression and Purification.** The expression of the recombinant 6x-His-tagged human CBR1 wild-type along with the V88I and P131S enzymes was confirmed by Western blot analysis showing bands with mobility corresponding to the calculated molecular mass of the tagged CBRs (~34 kDa) (Fig. 1). Total protein staining of the SDS-polyacrylamide electrophoresis gels demonstrated that these fractions were successfully purified from their respective transformed bacterial lysates because no other proteins were detected (Fig. 1). A majority of the pure enzyme was recovered in the 50 mM imidazole elution fractions; no further protein was eluted with 100 or 250 mM imidazole (data not shown).

**Kinetic Characterization of Wild-Type and Variant Enzymatic Activities with Menadione.** Our initial goal was to determine whether the purified human CBR1 protein was active and, if so, to determine the kinetic properties of the protein and the effect of the ns-SNP mutations on these properties. To accomplish this, Michaelis-Menten kinetic studies were conducted using menadione as a standard test substrate because it is specific for the CBR1 enzyme (Gonzalez-Covarrubias et al., 2007). In the presence of menadione, the 6x-His-tagged CBR1 wild-type enzyme was found to have a maximal rate of reaction (V_{max}) of 537 ± 61 nmol NADPH consumed/min · mg of purified protein (Fig. 2; Table 1). In comparison, the V88I variant with the same substrate had a V_{max} that was significantly lower (334 ± 9 nmol/min · mg of protein); however, no significant difference in V_{max} was seen with the P131S variant (581 ± 68 nmol/min · mg of protein). These values for the CBR1 wild-type and V88I variant enzymes were approximately 2-fold higher than the V_{max} values previously reported in the literature for bacterially expressed recombinant human 6x-His-tagged CBR1 and the V88I variant form (220 ± 27 and 168 ± 16 nmol/min · mg, respectively) (Gonzalez-Covarrubias et al., 2007). Greater protein stability after purification as well as our use of fluorescence detection as well as our use of fluorescence detection, a more sensitive method of measuring the rate of NADPH oxidation, over UV-visible spectrophotometry may account for the differences in value of the kinetic parameters between these studies. Both studies illustrate that the V88I variant enzyme has significantly decreased metabolic activity using menadione as substrate (p < 0.05).

No significant difference was detected with the K_m values between the wild-type and both variant isoforms. kcat/K_m values for the V88I and P131S variants were found to be 20 and 40% lower than those for the wild type, respectively. Eadie-Hofstee plots for the CBR1 wild-type, V88I, and P131S enzymes verified linearity at the same concentrations of menadione used to conduct the assays (r^2 = 0.91 for CBR1 wild-type; r^2 = 0.99 for V88I; and r^2 = 0.97 for P131S).

After the kinetic studies with menadione, the CBR1 wild-type and variant enzymes were exposed to FXa for 6 h at 23°C to remove the tag and linker. The native enzyme was separated from the tag and linker by mixing the cleaved preparation with Ni-NTA resin and filtering the mixture. The tag and linker were retained by the resin,
and the purified enzyme was collected in the filtrate. Western blot analysis demonstrated that this treatment completely removed the tag and linker from the wild-type and variant enzymes (data not shown). The metabolic activities of tagged and native (untagged) enzymes were compared. With the use of 150 μM menadione, there were no differences in enzymatic activity between the tagged and native enzyme (CBR1: 504 ± 48 versus 479 ± 36 nmol/min · mg of protein for tagged and untagged, respectively; V88I: 365 ± 24 versus 343 ± 30 nmol/min · mg of protein; P131S: 484 ± 51 versus 463 ± 42 nmol/min · mg of protein), suggesting that the amino acid linker and 6x-His tag engineered on the amino terminus of the wild-type and mutant gene products has no effect on enzyme activity. Based on these findings, the uncleaved wild-type and variant enzymes were used for subsequent activity assays involving the anthracyclines.

Kinetic Characterization of Wild-Type and Variant Enzymatic Activities with DOX and DAUN. To evaluate the impact of the single amino acid substitutions in the human CBR1 enzyme on the reduction of the anthracycline drugs, we measured the formation of

### FIG. 2.
In vitro enzymatic activities for the purified 6x-His-tagged CBR1 wild-type and V88I variant with 20 to 150 μM of the test substrate menadione as measured by following the initial rate of NADPH oxidation. Two independent batches of each enzyme were purified, and assays were performed in triplicate with each batch. Enzymatic activities are reported as mean ± S.D. (n = 6) with the background levels subtracted. The background levels represented the reaction buffer, enzyme, and NADPH cofactor only, without the addition of the test substrate. Eadie-Hofstee plots for the wild-type and variant enzyme are shown to the right of the activity plot (r² > 0.91).

### TABLE 1
Kinetic constants for menadione metabolism by recombinant CBR1 wild-type and variant allele enzymes

<table>
<thead>
<tr>
<th>Kinetic Parameter</th>
<th>CBR1 Wild-Type</th>
<th>V88I Variant</th>
<th>P131S Variant</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vₘₐₓ (nmol/min · mg of protein)</td>
<td>537 ± 61</td>
<td>334 ± 9*</td>
<td>581 ± 68</td>
</tr>
<tr>
<td>Kₘ (μM)</td>
<td>24 ± 10</td>
<td>18 ± 2</td>
<td>45 ± 15</td>
</tr>
<tr>
<td>kₘᵃᵗ /Kₘ (s⁻¹ M⁻¹)</td>
<td>0.30</td>
<td>0.19</td>
<td>0.33</td>
</tr>
<tr>
<td>kₘᵃᵗ (s⁻¹)</td>
<td>12.8 × 10³</td>
<td>10.5 × 10³</td>
<td>7.3 × 10³</td>
</tr>
</tbody>
</table>

*Significantly different from wild type (p < 0.05).

* kₘᵃᵗ calculated from Mₛ 34,000.
the major alcohol metabolites in vitro. Full chromatographic resolution of DAUNol and DOXol from DAUN and DOX, respectively, and of idarubicin (internal standard) was achieved for all chemical standards and in vitro samples. DOXol, DOX, DAUNol, DAUN, and idarubicin were observed to elute at 4.5, 5.5, 6.0, 6.8, and 7.3 min, respectively. Incubation of the 6x-His-tagged CBR1 wild-type and V88I variant enzyme with DOX generated a single new chromatographic peak that was identified as DOXol. Likewise, incubation with DAUN generated a single new chromatographic peak that was identified as DAUNol. The identification of the metabolite peaks was confirmed by incubation of DOX and DAUN with human liver cytosol and the generation of compounds that had identical chromatographic behaviors, as well as correspondence in retention time of the metabolite peak from DOX incubations with that for the chemical standard of DOXol. There were no detectable peaks at the DAUNol or DOXol retention times in the absence of protein.

Michaelis-Menten kinetic study properties were determined for the wild-type and variant forms of human CBR1, using both DAUN and DOX, separately, as substrates. With DAUN (Fig. 3A), the $V_{max}$ and $K_m$ of the wild-type CBR1 enzyme were $3430 \pm 241 \text{nmol/min} \cdot \text{mg}$ and $51 \pm 13 \mu M$, respectively (Table 2). The V88I variant reduced the $V_{max}$ significantly ($2090 \pm 112 \text{nmol/min} \cdot \text{mg}$) but did not alter the $K_m$ (Table 2). In contrast, the P131S variant exhibited a significant increase in $K_m$ ($89 \pm 13 \mu M$) but did not alter the $V_{max}$ (Table 2). With DOX as a substrate (Fig. 3B), the $V_{max}$ and $K_m$ for the CBR1 wild-type enzyme were $364 \pm 37 \text{nmol/min} \cdot \text{mg}$ and $287 \pm 57 \mu M$, respectively (Table 2). The V88I variant resulted in a $30\%$ reduction in the $V_{max}$ for DOX metabolism ($257 \pm 11 \text{nmol/min} \cdot \text{mg}$), which is a significant difference from that for the wild-type enzyme. The $k_{cat}/K_m$ values obtained from DAUN metabolism with both variant enzymes were $40\%$ lower than those for the wild-type enzyme. With DOX, the $k_{cat}/K_m$ values for the V88I and P131S variants were 20 and $40\%$ lower, respectively, than that for the wild-type enzyme. Eadie-Hofstee plots for the CBR1 wild-type and variant enzymes verified linearity at the same concentrations of DAUN and DOX used to conduct the assays ($r^2 > 0.94$ for all plots). In addition, we observed that DAUN is a better substrate than DOX for the wild-type and the variant enzymes as shown by the significant decrease in $V_{max}$ (V88I) as well a significant increase in $K_m$ (P131S).

**Discussion**

Living organisms are regularly exposed to a broad spectrum of compounds containing carbonyl functional groups such as aldehydes, ketones, carboxylic acids, and quinones, which are endogenous or exogenous in nature. This functional group is of particular concern to human health because it is highly reactive and can induce oxidative stress or interact with nucleophilic centers of proteins and nucleic acids (Matsunaga et al., 2006). To combat the negative effects brought about by carbonyl compounds, organisms have evolved several bio-transformation enzyme systems, one of them being the CBRs (Höffmann and Maser, 2007; Oppermann, 2007).

In humans, CBR1 is one of the most extensively studied carbonyl reductases and has been shown to metabolize drugs containing carbonyl groups, including DOX and DAUN. Treatment with these anthracyclines has played a significant role in improving life expectancy in cancer patients by halting the division of cancer cells. Unfortunately, the mechanism by which they function remains puzzling; a variety of hypotheses have been proposed including induction of apoptosis, RNA synthesis inhibition, DNA intercalation, and interference with DNA unwinding or DNA strand separation (Minotti et al., 2004; Takemura and Fujiwara, 2007).

We conducted in vitro metabolic assays on DOX and DAUN using purified CBR1 wild-type and V88I and P131S mutant enzymes expressed in E. coli. The water-soluble metabolites, DOXol and DAUNol, were quantified because previous studies have acknowledged them to be the major metabolites in cancer patients (Lipp and Boke, 1999; Plebisch et al., 2007). We demonstrated that DOX and DAUN are both readily metabolized to their respective alcohol metabolites with the CBR1 wild-type enzyme having a significantly higher $V_{max}$ for both anticancer drugs compared with the V88I variant and a significantly lower $K_m$ for DAUN compared with the P131S variant. In addition, the CBR1 wild-type enzyme and both variants metabolize DAUN far more efficiently than DOX; there is an 8- to 11-fold increase in $V_{max}$ a 4- to 7-fold decrease in $K_m$, and a 40- to 50-fold increase in $k_{cat}/K_m$. The metabolic preference of the CBR1 wild-type enzyme and the variant forms for DAUN over DOX is interesting and is consistent with what we observed for DAUN metabolism with another metabolic reductase, aldo-keto reductase 1A1 (Bains et al., 2008). It is clear that the metabolic activity of these two reductases can differ substantially despite the minor difference in chemical structures between these two drugs.

Our results with the V88I variant in the presence of DAUN are consistent with those reported by Gonzalez-Covarrubias et al. (2007), who used cofactor (NADPH) consumption to demonstrate that the $V_{max}$ was significantly decreased in the V88I variant compared with that in the wild-type enzyme. Although cofactor usage is an indirect test of substrate conversion, these authors confirmed this kinetic difference by directly measuring DAUNol levels using HPLC-fluorescence at a single DAUN concentration (500 μM), which demonstrated that the variant enzyme produced 47% less metabolite than the wild-type enzyme. These authors suggested that the V88I mutation has an impact on NADPH binding affinity because of its close proximity to the cofactor site (Fig. 4). Their finding demonstrates that the valine to isoleucine substitution leads to a lower affinity for the cofactor. Because NADPH is a vital cofactor in the metabolic conversion of DOX and DAUN to their major alcohol metabolites, this substitution might be the basis for the significant decrease in the levels of DOXol and DAUNol produced with the V88I variant enzyme. The P131S mutation, albeit located further from the active and cofactor binding sites than the V88I mutation (Fig. 4), may alter the conformation of one or both sites, which could explain why there was a significant increase in $K_m$ for DAUN compared with that for the wild-type enzyme, whereas no change was observed for DOX. However, further studies are needed to provide a more definitive explanation.

Reductions in 1) metabolic activity for DOX and DAUN by the V88I variant and 2) substrate affinity for DAUN by the P131S variant may explain the interpatient variability in the serious side effects seen in cancer patients treated with either of these anthracyclines. We propose that these adverse events are the result of excessive buildup of parent drug. An increasing amount of data has shown that these drugs accumulate in mitochondria of cardiomyocytes and interfere with enzyme function of the electron transport chain (Hasinoff et al., 2003; Wallace, 2003; Obara et al., 2007). For example, complex I of the electron transport chain acts on DOX and DAUN such that the quinone moiety within the tetracycline ring undergoes redox cycling between the quinone and semiquinone states. The electrons generated during this cycling are captured by oxidizing agents, including oxygen, leading to the formation of reactive oxygen species (ROS) (Singal et al., 2000; Kim et al., 2005; Wold et al., 2005; Takemura and Fujiwara, 2007). ROS are a major concern because they are capable of mitochondrial respiratory chain disruption, lipid peroxidation, and DNA disintegration, all of which may be the underlying causes of the life-threatening side effects seen in some patients undergoing DOX and DAUN treatment (Wojtacki et al., 2000). The heart seems to be
Fig. 3. In vitro enzymatic activities for the purified 6x-His-tagged CBR1 wild-type and variants with (A) 0 to 400 μM daunorubicin and (B) 0 to 400 μM doxorubicin. Activities are measured by following the rate of daunorubicinol and doxorubicinol production. Two independent batches of each enzyme were purified, and assays were performed in triplicate with each batch. Enzymatic activities are reported as mean ± S.D. (n = 6). Eadie-Hofstee plots for the wild-type and variant enzyme are shown to the right of the activity plot for each anthracycline (r² > 0.94 for each plot).
coadministration of dexrazoxane with DOX has resulted in cardio-
radical damage (Wu and Hasinoff, 2005). Studies have revealed that
iron (Fe$^{2+}$ scavenger, dexrazoxane, an intracellular iron chelator that binds free
radicals) has been demonstrated in studies with the free radical
complexes involving anthracyclines and intracellular free iron. This
process is mediated by enzymes, such as superoxide dismutases and glutathione peroxidases,
because myocardial tissue has low levels of endogenous antioxidant
capacity. The valine and proline are at positions 87 and 130 in the model, respectively,
because amino acid numbering does not include initiator methionine. This molec-
ular graphic image was produced using the Chimera program (Resource for Bio-
computing, Visualization, and Informatics, University of California, San Francisco,
CA).

The most susceptible to the harmful effects associated with ROS
because myocardial tissue has low levels of endogenous antioxidant
enzymes, such as superoxide dismutases and glutathione peroxidases,
to neutralize these free radicals (Takemura and Fujiwara, 2007). ROS
are also generated during Fenton reactions after the formation of
complexes involving anthracyclines and intracellular free iron. This
generation has been demonstrated in studies with the free radical scavenger, dexrazoxane, an intracellular iron chelator that binds free
iron (Fe$^{2+}$ and Fe$^{3+}$), thus preventing site-specific iron-based oxygen
radical damage (Wu and Hasinoff, 2005). Studies have revealed that
coadministration of dexrazoxane with DOX has resulted in cardio-
protection (Wexler et al., 1996; Swain et al., 1997; Wu and Hasinoff,
2005).

In conclusion, this study demonstrates that a 6x-His-tagged CBR1
enzyme, whose activity does not differ from that of the untagged
enzyme, is efficient in metabolizing DOX and DAUN, with higher
specificity for the latter. We have also shown that the V88I variant
significantly reduces reductase activity toward both anthracyclines
and that the P131S variant significantly reduces substrate affinity of
DAUN. Therefore, the ns-SNPs in the human CBR1 gene generating the
V88I and P131S mutations may prove to be clinically useful
 genetic biomarkers for guiding anthracycline therapy in cancer pa-
tients to minimize adverse effects.

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