Role of UDP-Glucuronosyltransferase Isoforms in 13-cis Retinoic Acid Metabolism in Humans

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ABSTRACT:

13-cis Retinoic acid (13cisRA, isotretinoin) is an important drug in both dermatology, and the treatment of high-risk neuroblastoma. 13cisRA is known to undergo cytochrome P450-mediated oxidation, mainly by CYP2C8, but phase II metabolic pathways have not been characterized. In the present study, the glucuronidation activities of human liver (HLM) and intestinal microsomes (HIM), as well as a panel of human UDP-glucuronosyltransferases (UGTs) toward both 13cisRA and the 4-oxo metabolite, 4-oxo 13cisRA, were compared using high-performance liquid chromatography. Both HLM and, to a greater extent, HIM catalyzed the glucuronidation of 13cisRA and 4-oxo 13cisRA. Based on the structures of 13cisRA and 4-oxo 13cisRA, the glucuronides formed are conjugated at the terminal carboxylic acid. Further analysis revealed that UGT1A1, UGT1A3, UGT1A7, UGT1A8, and UGT1A9 were the major isoforms responsible for the glucuronidation of both substrates. For 13cisRA, a pronounced substrate inhibition was observed with individual UGTs and with HIM. UGT1A3 exhibited the highest rate of activity toward both substrates, and a high rate of activity toward 13cisRA glucuronidation was also observed with UGT1A7. However, for both substrates, Km values were above concentrations reported in clinical studies. Therefore, UGT1A9 is likely to be the most important enzyme in the glucuronidation of both substrates as this enzyme had the lowest Km and is expressed in both the intestine and at high levels in the liver.

Retinoids are derivatives of retinol (vitamin A) that regulate numerous biological processes, including embryogenesis, growth, differentiation, vision, and reproduction (Evans and Kaye, 1999). Retinoid is the main biologically active derivative of retinol and exists in several stereoisomeric forms, including all-trans retinoic acid (ATRA), 13-cis retinoic acid (13cisRA), and 9-cis retinoic acid. Retinoids exhibit several pharmacological properties useful in dermatology (Peck and DiGiovanna, 1994), as well as in cancer prevention and chemotherapy (Hong and Itri, 1994). The most dramatic clinical benefits with retinoids are observed with ATRA for the treatment of acute promyelocytic leukemia and 13cisRA in the treatment of high-risk neuroblastoma (Reynolds and Lemons, 2001; Matthey et al., 2009).

The oxidative metabolism of 13cisRA has been extensively examined and involves the cytochrome P450 (P450) enzyme system, a superfamily of heme-containing monooxygenases that have a central role in the oxidative metabolism of a variety of endogenous and exogenous compounds. As a result of P450-mediated metabolism, intermediates that may exert toxicity or carcinogenicity are formed. Such intermediates are also targets for phase II drug metabolism, which renders them inactive, with greater polarity and water solubility. The human UGTs 1A8 and 2B7 have been identified as being responsible for the glucuronidation of ATRA, 4-oxo ATRA, 4-hydroxy ATRA, and 5,6-epoxy ATRA (Cheng et al., 1999; Czernik et al., 2000; Samokyszyn et al., 2000). Analysis of the glucuronidation products revealed that ATRA, 4-oxo ATRA, and 5,6-epoxy ATRA formed carboxyl-linked glucuronides, whereas 4-hydroxy ATRA was glucuronidated at its hydroxyl group (Samokyszyn et al., 2000). In view of its strong homology with ATRA, it is probable that 13cisRA also undergoes glucuronidation.

To date, 21 functional UGT isoforms have been identified in humans (http://www.ugtalleles.ulaval.ca), many of which catalyze the glucuronidation of a wide variety of environmental carcinogens, dietary chemopreventatives, and anticancer agents (Nagar and Remmel, 2006). However, no studies examining the glucuronidation of 13cisRA have as yet been reported. In the present study the activities and kinetics of human liver and intestinal microsomes as well as a panel of recombinant human UGTs were determined to identify those likely to be responsible for 13cisRA and 4-oxo 13cisRA glucuronidation in humans.

Materials and Methods

Materials. 13-cis Retinoic acid, all-trans retinoic acid, and β-glucuronidase (from Helix pomatia) were purchased from Sigma-Aldrich (Poole, UK), 4-Oxo...
13-cis retinoic acid (Ro 22-6595) and 4-oxo all-trans retinoic acid (Ro 12-4824) were generously provided by F. Hoffmann-La Roche (Basel, Switzerland). UGT Reaction Mix Solution A (25 mM UDP) and UGT Reaction Mix Solution B (250 mM Tris·HCl, 40 mM MgCl₂, and 0.125 mg/ml alamethicin) were purchased from BD Gentest (Oxford, UK). Pooled human liver and intestinal microsomes as well as Supersomes expressing the recombinant human UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A7, UGT1A8, UGT1A9, UGT1A10, UGT2B4, UGT2B7, UGT2B15, and UGT2B17 enzymes were also purchased from BD Gentest. Control microsomes with wild-type baculovirus were obtained. High-performance liquid chromatography (HPLC)-grade acetonitrile and glacial acetic acid were obtained from Thermo Fisher Scientific (Loughborough, UK).

**Characterization of 13cisRA and 4-Oxo 13cisRA Glucuronidation.** Glucuronidation activity was determined in pooled human liver microsomes.

**Fig. 1.** Representative HPLC chromatograms illustrating the in vitro glucuronidation of 13cisRA and 4-oxo 13cisRA by UGT1A3. Absorbance units are measured on the y-axis, and time in minutes is measured on the x-axis. A, 13cisRA glucuronidation. 13cisRA (i) eluted at 23.1 min and 13cisRA glucuronide (ii) at 6.2 min. B, HPLC chromatogram of the same incubation shown in A, after treatment with β-glucuronidase. C, 4-oxo 13cisRA glucuronidation. 4-Oxo 13cisRA (i) eluted at 7.9 min, 4-oxo ATRA (ii) at 7.2 min, and 4-oxo 13cisRA glucuronide (iii) at 2.4 min. D, HPLC chromatogram of the same incubation shown in C, after treatment with β-glucuronidase.
(HLM), human intestinal microsomes (HIM), and Supersomes containing different cDNA-expressed UGT isoforms (UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A7, UGT1A9, UGT1A10, UGT2B4, UGT2B7, UGT2B15, and UGT2B17). Conditions for linearity with respect to time and protein concentration were optimized in preliminary studies. The incubation mixture consisted of enzyme fractions (0.5 mg/ml total protein), 13cisRA (100 μM), or 4-oxo 13cisRA (100 μM), alanethicin (25 μg/ml), MgCl₂ (8 mM), UDPGA (2 mM), and methanol (0.25%) made up to a final volume (200 μl) with Tris-HCl buffer (50 mM, pH 7.5). Incubations were performed in a shaking water bath at 37°C for 60 min. Reactions were initiated by the addition of enzyme and terminated with ice-cold acetonitrile (400 μl). Incubations conducted with control Supersomes served as a negative control. Samples were vortex mixed and centrifuged at 17,900g for 5 min. Supernatant (200 μl) was removed and evaporated to dryness, and the residue was reconstituted in 200 μl of 0.1% glacial acetic acid, pH 5.0, before HPLC analysis. All incubations were performed in triplicate.

Hydrolysis of 13cisRA and 4-oxo 13cisRA Glucuronides. To samples prepared from the incubations as described above, β-glucuronidase (80 units) in potassium phosphate buffer (75 mM, pH 6.8) was added. The mixture was incubated in a shaking water bath at 37°C for 30 min. Reactions were terminated by the addition of ice-cold acetonitrile (400 μl). Incubations conducted were prepared under the same conditions, but with the addition of β-glucuronidase after the acetonitrile. Samples were vortex mixed and centrifuged at 17,900g for 5 min. Supernatant (200 μl) was removed and evaporated to dryness. Residue was reconstituted in 200 μl of 0.1% glacial acetic acid, pH 5.0, before HPLC analysis. All reactions were performed in triplicate.

HPLC Analysis. Analysis of 13cisRA and 13cisRA glucuronide was performed using a 2695 Separations Module and a 996 Photodiode Array UV Detector with Empower 2 software for data acquisition and analysis (Waters, Milford, MA). The UV detector was set to monitor 200 to 400 nm, isolating at 374 nm. Separation was performed on a Luna C18 (2) column (50 × 2.0 mm, 3 μm) (Phenomenex, Torrance, CA) by gradient reverse-phase chromatography. The mobile phase consisted of 0.1% glacial acetic acid, pH 5.0 (A) and 100% acetonitrile (B). Analysis was performed at a constant flow rate of 0.2 ml/min and an injection volume of 20 μl, and the gradient ran from 60% A to 60% B between 0 and 2 min. It remained at 60% B for 2 min and then switched back to 60% A between 4 and 5 min, remaining at 60% A for 10 min before switching to 100% B between 15 and 16 min. It remained at 100% B for 10 min and finally switched back to 60% A between 26 and 30 min, remaining at 60% A until 40 min. Analysis of 4-oxo 13cisRA and 4-oxo 13cisRA glucuronide was performed as described above, but with an alternative gradient, running from 60% A to 60% B between 0 and 2 min and remaining at 60% B for 2 min before switching back to 60% A between 4 and 5 min, remaining until 20 min. The 13cisRA and 4-oxo 13cisRA glucuronide peaks were identified by treating the metabolites with β-glucuronidase, which hydrolyzes the glucuronide back to the aglycone. Analysis of 13cisRA and 4-oxo 13cisRA glucuronides was considered to be semiquantitative, because authentic standards were unavailable for the 13cisRA and 4-oxo 13cisRA glucuronides.

Results

Identification of 13cisRA and 4-Oxo 13cisRA Glucuronides. Incubation of 13cisRA with HLM, HIM, recombinant UGT isoforms, and UDPGA resulted in the appearance of a novel chromatographic peak that was visible using a HPLC UV method previously optimized for the detection of 13cisRA (Fig. 1A). The glucuronide peak had a shorter retention time (6.2 min) compared with that for 13cisRA (23.1 min), and the rate of generation of this peak was dependent on microsomal protein concentration and incubation time. After incubation of 4-oxo 13cisRA with HLM, HIM, recombinant UGT isoforms, and UDPGA, another novel chromatographic peak was visible using a HPLC UV method that had previously been optimized for the detection of 4-oxo 13cisRA (Fig. 1C). An additional substrate peak was also observed. This was identified as 4-oxo ATRA and was formed as a result of minor isomerization of 4-oxo 13cisRA. The glucuronide peak retention time was 2.4 min, compared with 7.9 min for 4-oxo 13cisRA and 7.2 min for 4-oxo ATRA; again, the peak area increased with microsomal protein and incubation time. Treatment of...
both incubates with β-glucuronidase resulted in complete disappearance of the metabolite peaks, indicating that the metabolites are glucuronide conjugates of 13cisRA and 4-oxo 13cisRA, respectively (Fig. 1, B and D).

**Characterization of 13cisRA and 4-Oxo 13cisRA Glucuronidation.** Screening of 13cisRA and 4-oxo 13cisRA glucuronosyltransferase activity was performed using HLM and HIM, as well as 12 recombinant UGT isoforms (UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A7, UGT1A8, UGT1A9, UGT1A10, UGT2B4, UGT2B7, UGT2B15, and UGT2B17). HLM, HIM, UGT1A1, UGT1A3, UGT1A7, UGT1A8, and UGT1A9 were found to catalyze both 13cisRA and 4-oxo 13cisRA glucuronide formation (Fig. 2). UGT1A4, UGT1A6, UGT1A10, UGT2B4, UGT2B7, UGT2B15, and UGT2B17 exhibited no detectable catalytic activity toward either 13cisRA or 4-oxo 13cisRA under the incubation conditions used.

![Graphs showing kinetic profiles for glucuronide formation](image-url)
Kinetic parameter estimates for the formation of 13cisRA and 4-oxo 13cisRA glucuronides in human liver and intestinal microsomes and recombinant UGT isoforms

Data are expressed as means ± S.E.M.; n = 3.

### Table 1

<table>
<thead>
<tr>
<th>UGT Protein Source</th>
<th>4-Oxo 13cisRA</th>
<th>13cisRA</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$V_{max}$</td>
<td>$K_m$</td>
</tr>
<tr>
<td>HLM</td>
<td>59 ± 17</td>
<td>974 ± 342</td>
</tr>
<tr>
<td>HIM</td>
<td>1120 ± 800</td>
<td>200 ± 98</td>
</tr>
<tr>
<td>UGT1A1</td>
<td>22$^a$</td>
<td>78$^a$</td>
</tr>
<tr>
<td>UGT1A3</td>
<td>134$^b$</td>
<td>161$^b$</td>
</tr>
<tr>
<td>UGT1A7</td>
<td>114$^c$</td>
<td>186$^c$</td>
</tr>
<tr>
<td>UGT1A8</td>
<td>17$^d$</td>
<td>49$^d$</td>
</tr>
<tr>
<td>UGT1A9</td>
<td>29$^a$</td>
<td>14$^a$</td>
</tr>
</tbody>
</table>

N.A., not applicable.

$^a$Precise estimates of the parameters could not be determined.

### Determination of Kinetic Parameters for 13cisRA and 4-Oxo 13cisRA Glucuronide Formation

Kinetic analyses of 13cisRA and 4-oxo 13cisRA glucuronide formation were performed using HLM, HIM, and recombinant UGT1A1, UGT1A3, UGT1A7, UGT1A8, and UGT1A9 isoforms. Both 13cisRA and 4-oxo 13cisRA glucuronides were formed in varying amounts and rates across all UGT sources studied. HIM, which comprised a pool of more than one UGT enzyme, had the highest rate of glucuronidation toward both substrates. The formation of the 13cisRA glucuronide exhibited atypical kinetics across all UGT sources (Fig. 3; Table 1), except HLM for which data were fitted by the hyperbolic Michaelis-Menten model. The $V_{max}$ and $K_m$ for HLM were 59 ± 17 $\mu$M/s and 974 ± 342 $\mu$M, respectively. For HIM and recombinant UGT isoforms, data were fitted by the hyperbolic Michaelis-Menten model. The formation of the 13cisRA glucuronide showed evidence of substrate inhibition. Although it was not possible to determine the parameters of this phenomenon precisely, inspection of the kinetic plots and the estimated parameters suggests that for the majority of enzyme sources, atypical kinetics of glucuronidation is not clinically important. The exception to this is UGT1A9, which showed signs of substrate inhibition at 50 $\mu$M. Substrate inhibition kinetics have been reported for other UGT substrates, including resveratrol (Iwuchukwu and Nagar, 2010).

The kinetic profiles for 4-oxo 13cisRA glucuronidation by HLM, HIM, and all of the UGT isoforms studied followed Michaelis-Menten kinetics, allowing determination of apparent $K_m$ and $V_{max}$ values. The 4-oxo metabolite appeared to be a better substrate for glucuronidation in HLM, in that the $V_{max}$ value was higher than that for the parent compound. However, it should be noted that the $K_m$ values for both substrates were higher than the maximal plasma concentrations observed in patients (Veal et al., 2007). In addition, glucuronidation in the intestine may be more relevant as the rate of reaction was highest for HIM, particularly at concentrations typically achieved in plasma. The intestine may be exposed to higher concentrations during drug absorption, but the degree of substrate inhibition was not marked at concentrations less than 100 $\mu$M in enzyme sources other than UGT1A9.

Caution in the interpretation of the estimated parameters is necessary because, unlike P450s, UGTs do not have a measurable chromophore representative of active protein; therefore, the amount of active UGT in any given recombinant preparation is unknown, and activities are normalized to milligrams of total protein rather than picomoles of UGT. The inability to quantify UGT expression levels in recombinant systems makes it difficult to verify the relative importance of individual isoforms in a particular glucuronidation pathway in which multiple UGTs may be involved. One approach to overcome this problem is through implementation of the relative activity factor method. This method was first used to investigate P450-dependent activity (Crespi and Miller, 1999) and measures the activities of an enzyme-selective probe substrate in recombinant UGT enzymes and pooled HLM. However, because of the broad overlapping substrate specificity between the UGTs, one caveat with the rela-

### Discussion

An in vitro HPLC-based assay has been developed to characterize the generation of 13cisRA and 4-oxo 13cisRA glucuronides using HLM, HIM, and recombinant UGT isoforms. Positive identification of these metabolites as glucuronides was based on sensitivity to β-glucuronidase treatment, resulting in complete disappearance of the metabolite peaks generated. Both HIM and HLM were found to catalyze the glucuronidation of 13cisRA and 4-oxo 13cisRA, with the highest rate of activity toward both substrates seen for HIM. This result could be particularly significant for an orally administered drug such as 13cisRA. Further investigations revealed that UGT1A1, UGT1A3, UGT1A7, UGT1A8, and UGT1A9 were the major isoforms responsible for the glucuronidation of both substrates.

Enzyme kinetic studies were performed with microsomes from human liver and intestine and with human UGT isoforms UGT1A1, UGT1A3, UGT1A7, UGT1A8, and UGT1A9 for both substrates. Although only a limited range of concentrations of 4-oxo 13cisRA could be explored, the concentrations far exceeded those reported clinically. For the parent drug, all enzyme sources other than HLM showed evidence of substrate inhibition. Although it was not possible to determine the parameters of this phenomenon precisely, inspection of the kinetic plots and the estimated parameters suggests that for the majority of enzyme sources, atypical kinetics of glucuronidation is not clinically important. The exception to this is UGT1A9, which showed signs of substrate inhibition at 50 $\mu$M. Substrate inhibition kinetics have been reported for other UGT substrates, including resveratrol (Iwuchukwu and Nagar, 2010).

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The activity factor method for UGT reaction phenotyping is that probe substrates have not been identified for all of the major UGT isoforms.

Examination of the structures of 13cisRA and 4-oxo 13cisRA (Fig. 5) and comparison with the glucuronidation of ATRA allow tentative assignment of the position of glucuronidation. Because glucuronidation requires either a –COOH, –OH, –NH₂, or –SH functional group for conjugation with ß-D-glucuronic acid, it is likely that the glucuronides formed in this study are conjugated at the terminal carboxylic acid. This prediction is in line with the previous finding by Samokyszyn et al. (2000) that the UGT2B7-mediated glucuronidation of ATRA and 4-oxo ATRA is directed toward the carboxyl function, whereas the 4-hydroxylated metabolite is glucuronidated almost exclusively at the hydroxyl function. It is also likely that 4-hydroxy 13cisRA undergoes glucuronidation. This activity would most likely occur at the hydroxyl function, as has been demonstrated for 4-hydroxy ATRA (Samokyszyn et al., 2000).
However, because this metabolite is not readily available, investigations into the potential glucuronidation of 4-hydroxy 13-cis RA could not be performed.

Despite being unable to determine the exact contribution of each of the recombinant UGT isoforms examined in this study, our data suggest that UGT1A3 has the highest rate of activity toward both substrates, as assessed by the \( V_{\text{max}} \) estimates. UGT1A3 is one of the main isoforms responsible for the glucuronidation of the majority of carboxylic acid compounds (Sakaguchi et al., 2004), supporting the hypothesis that the carboxyl function is the site of 13cisRA and 4-oxo 13cisRA glucuronidation. However, although UGT1A3 is expressed mainly in the liver, the level of expression is significantly lower than that of UGT1A1 or UGT1A9 (Ohno and Nakajin, 2009). Furthermore, because the plasma concentrations of 13cisRA and 4-oxo 13cisRA are reported to be below the \( K_m \) concentrations for the enzymes tested in this study (Veal et al., 2007), it is probable that UGT1A9 is the most important isoform in the glucuronidation of both 13cisRA and 4-oxo 13cisRA, the enzyme with the lowest \( K_m \) value for each substrate, is expressed at high levels in the liver and is also expressed in the intestine (Ohno and Nakajin, 2009). The importance of glucuronidation in the intestines is emphasized by the very high degree of glucuronidation observed with human intestinal microsomes and the high level of expression of the relevant isoforms in this tissue (Ohno and Nakajin, 2009). Identification of UGT1A9 as an important isoform involved in the glucuronidation of both 13cisRA and 4-oxo 13cisRA provides a rationale for investigating the impact of UGT1A9 genetic polymorphisms on the reported variability associated with 13cisRA pharmacokinetics and metabolism in children with neuroblastoma (Veal et al., 2007).

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References


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