**Short Communication**

**Amlodipine Metabolism in Human Liver Microsomes and Roles of CYP3A4/5 in the Dihydropyridine Dehydrogenation**

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**ABSTRACT**

Amlodipine is a commonly prescribed calcium channel blocker for the treatment of hypertension and ischemic heart disease. The drug is slowly cleared in humans primarily via dehydrogenation of its dihydropyridine moiety to a pyridine derivative (M9). Results from clinical drug-drug interaction studies suggest that CYP3A4/5 mediate metabolism of amlodipine. However, attempts to identify a role of CYP3A5 in amlodipine metabolism in humans based on its pharmacokinetic differences between CYP3A5 expressers and nonexpressers failed. Objectives of this study were to determine the metabolite profile of amlodipine (a racemic mixture and S-isomer) in human liver microsomes (HLM), and to identify the cytochrome P450 (P450) enzyme(s) involved in the M9 formation. Liquid chromatography/mass spectrometry analysis showed that amlodipine was mainly converted to M9 in HLM incubation. M9 underwent further O-demethylation, O-dealkylation, and oxidative deamination to various pyridine derivatives. This observation is consistent with amlodipine metabolism in humans. Incubations of amlodipine with HLM in the presence of selective P450 inhibitors showed that both ketoconazole (an inhibitor of CYP3A4/5) and CYP3cide (an inhibitor of CYP3A4) completely blocked the M9 formation, whereas chemical inhibitors of other P450 enzymes had little effect. Furthermore, metabolism of amlodipine in expressed human P450 enzymes showed that only CYP3A4 had significant activity in amlodipine dehydrogenation. Metabolite profiles and P450 reaction phenotyping data of a racemic mixture and S-isomer of amlodipine were very similar. The results from this study suggest that CYP3A4, rather than CYP3A5, plays a key role in metabolic clearance of amlodipine in humans.

**Introduction**

Amlodipine, a dihydropyridine calcium channel blocker, is one of the most commonly prescribed drugs for the treatment of hypertension and ischemic heart disease. In a clinical study, amlodipine showed a long elimination of half-life (35 hours) after a single 10-mg intravenous dose (Abernethy, 1991), likely due to its high volume of distribution and a low rate of plasma clearance. The drug also exhibits a good oral bioavailability in the range of 52%–88% following a single 10-mg oral dose to human subjects (Faulkner et al., 1986). After an oral dose of radiolabeled amlodipine to humans, the total radioactivity recovery is 59.3% in urine and 23.4% in feces (Beresford et al., 1988a; Stopher et al., 1988). Similarly, 62% and 22.7% of a radiolabeled intravenous dose is recovered in human urine and faces, respectively. S-amlodipine is the active component of racemic amlodipine. There are no significant differences in pharmacodynamic and pharmacokinetic parameters after a single dose of 5 mg of S-amlodipine and 10 mg of amlodipine racemate (Kim et al., 2010). The amlodipine pyridine metabolite (Fig. 1C, M9), raised from dehydrogenation of the dihydropyridine moiety, and its derivatives are major drug-related components in human urine. The mass balance and metabolite profiling data suggest that amlodipine dehydrogenation to M9 followed by multiple oxidative reactions of M9 is the major clearance pathway of amlodipine in humans. Similar metabolic pathways have been observed in rats and dogs (Beresford et al., 1988b). It has been reported that many dihydropyridine analogs undergo CYP3A4-mediated dehydrogenation to form the corresponding pyridine metabolites in vitro (Guengerich et al., 1991). However, information on in vitro metabolism of amlodipine in humans, including P450 reaction phenotyping data, is not available in the literature.

Telaprevir, a potent inhibitor of both CYP3A4 and CYP3A5, increases the amlodipine mean area under the curve (AUC) by 2.79-fold and the mean half-life from 41.3 to 95.1 hours when the two drugs are coadministered (Lee et al., 2011). Similarly, combined dosing of indinavir and ritonavir, both of which are CYP3A inhibitors, increases the median amlodipine AUC0-24 by 90% (Glesby et al., 2005). These clinical drug-drug interaction observations suggest that CYP3A4/5 enzymes play significant roles in metabolic clearance of amlodipine in humans. CYP3A4 and CYP3A5 are two major enzymes of the CYP3A family and have similar catalytic specificities. However, unlike CYP3A4, CYP3A5 is a polymorphic enzyme. To determine if CYP3A5 contributes to the metabolism of amlodipine, the pharmacokinetics of amlodipine in CYP3A5 nonexpressers (CYP3A5*3/*3 carriers) and CYP3A5 expressers (CYP3A5*1/*3 and CYP3A5*1/*1 carriers) have been recently determined and compared with each other. After a single dose of amlodipine, its exposure in CYP3A5 expressers is slightly higher than that in CYP3A5 nonexpressers (Kim et al., 2006). However, the AUC and the maximal concentrations of amlodipine in CYP3A5 expressers are about 2-fold higher than those in CYP3A5 expressers after 5-mg once-daily oral doses for 7 days (Zuo et al., 2013). Although roles of CYP3A5 in the disposition and elimination of ABRIDATIONS: AUC, area under the curve; AUC0-24, area under the curve during 24 hours; HLM, human liver microsomes; LC, liquid chromatography; MH+, protonated molecular ions; MS, mass spectrometry; MSn, multiple stage MS/MS fragmentation; 1'-OH-midazolam, 1'-hydroxy-midazolam; Thio-TEPA, N,N',N'-triethylenethiophosphoramid.
amlodipine in humans have been suggested by investigators of these clinical studies, it is very challenging to draw meaningful conclusions without in vitro P450 phenotyping data of amlodipine. Furthermore, several studies have investigated the role of CYP3A5 polymorphism in blood pressure response to amlodipine, but results are not consistent (Zhang et al., 2013). The major objectives of this study were to determine the amlodipine metabolite profile in human liver microsomes (HLM) and P450 enzyme(s) responsible for amlodipine dehydrogenation to M9. In particular, the roles of CYP3A4 and CYP3A5 in amlodipine metabolism were investigated. In addition, the in vitro metabolism of racemic amlodipine was compared with that of S-amlodipine. Results from this study provide a biochemical basis for better understanding of the clinically observed amlodipine drug-drug interactions with CYP3A inhibitors and pharmacokinetics of amlodipine in both CYP3A5 non-expressers and expressers.

Materials and Methods

Chemicals and Reagents. A racemic amlodipine (2-[(2-aminoethoxy)methyl]-4-(2-chlorophenyl)-1,4-dihydro-6-methyl-3,5-pyridinedicarboxylic acid 3-ethyl 5-methyl ester benzenesulfonate norvasc), quinidine, ketoconazole, N,N,N’-triethylenethiophosphoramide (Thio-TEPA), montelukast, furafylline, sulfaphenazole, CYP3cide, and benzylnirvanol were purchased from Sigma (St. Louis, MO). S-amlodipine was purchased from Energy Chemical (Shanghai, China). All solvents (acetonitrile, methanol, and water) were of high-performance liquid chromatography (LC) grade. Pooled HLM was obtained from BD Biosciences (Woburn, MA). Expressed P450 enzymes, CYP1A2, 2B6, 2C8, 2C9, 2C19, 2D6, 3A4, and 3A5 were purchased from BD Biosciences (San Jose, CA).

HLM Incubation for Metabolite Profiling. Racemic amlodipine and S-amlodipine (30 μM) were separately incubated with HLM (2 mg protein/ml) for 60 minutes. The reaction mixtures (1 ml) also contained potassium phosphate buffer (69 mM, pH 7.4) and NADPH (4 mM). Metabolic reactions were initiated by addition of NADPH after a 3-minute preincubation and stopped by the addition of 2 ml of acetonitrile. Precipitate was removed by centrifugation. Supernatant was dried under nitrogen and then reconstituted in 200 μl of 5% acetonitrile/water (v/v) for liquid chromatography/mass spectrometry (LC/MS) analysis.

Time Course of Amlodipine Metabolism in HLM. Racemic amlodipine and S-amlodipine (1 μM) were separately incubated in duplicates in HLM (1 mg protein/ml) for 0, 5, 10, 15, 30, and 60 minutes. The reaction mixtures (0.25 ml) also contained potassium phosphate buffer (69 mM, pH 7.4) and NADPH (2 mM). Metabolic reactions were initiated by the addition of NADPH after a 3-minute preincubation and stopped at designated time points by the addition of 0.5 ml of acetonitrile. Precipitate was removed by centrifugation. Supernatant was dried under nitrogen and then reconstituted in 50 μl of 5% acetonitrile/water (v/v) for determination of disappearance of the parent drug and the formation of the pyridine metabolite (M9).

Inhibition of Amlodipine Dehydrogenation HLM by P450 Inhibitors. Racemic amlodipine and S-amlodipine (1 μM) were separately incubated in triplicates in HLM (0.5 mg/ml) in triplicates for 13 minutes. Reaction mixtures (0.25 ml) also contained potassium phosphate buffer (69 mM, pH 7.4), a chemical inhibitor of P450 enzyme, and NADPH (2 mM). Final concentrations of the P450 inhibitors were 1 μM for benzylnirvanol, quinidine, ketoconazole, montelukast, and CYP3cide; 10 μM for furafylline and sulfaphenazole; and 50 μM for Thio-TEPA. For the incubations with time-dependent inhibitors (Thio-TEPA, furafylline, and CYP3cide), the reaction was initiated by the addition of amlodipine or S-amlodipine after 10-minute preincubations of HLM and NADPH in the presence of the individual P450 inhibitors. All other reactions were initiated by the addition of NADPH after 3-minute preincubations of HLM with amlodipine or S-amlodipine in the presence of the individual P450 inhibitors. All of the reactions were stopped by the addition of 0.5 ml of acetonitrile, and the precipitate was removed by centrifugation. Supernatants were
dried under nitrogen and then reconstituted in 50 μl of 5% acetonitrile/water (v/v) for LC/MS analysis.

Amlodipine Dehydrogenation by Expressed P450 Enzymes. Racemic amlodipine and S-amlodipine (1 μM) were separately incubated with individual expressed human P450 enzymes (100 nM) (CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP3A4, and CYP3A5) in triplicates for 13 minutes. Reaction mixtures (0.25 ml) also contained potassium phosphate buffer (69 mM, pH 7.4) and NADPH (2 mM). After 3-minute preincubation, metabolic reactions were initiated by the addition of NADPH and stopped by the addition of 0.5 ml of acetonitrile. Precipitate was removed by centrifugation. The supernatant was dried under nitrogen and then reconstituted in 50 μl of 5% acetonitrile/water (v/v) for LC/MS analysis. In addition, racemic amlodipine was incubated with expressed CYP3A4 and CYP3A5 with or without CYP3cide under the conditions described earlier. In the same experiment, midazolam (10 μM) was incubated with expressed CYP3A4 or CYP3A5 (100 nM) in triplicates for 5 minutes. Reaction mixtures (0.04 ml) also contained potassium phosphate buffer (69 mM, pH 7.4) and NADPH (2 mM). After 3-minute preincubation, the metabolic reaction converting midazolam to 1’-hydroxy-midazolam (1’-OH-midazolam) was started by the addition of NADPH and stopped by the addition of 0.08 ml of acetonitrile. Precipitate was removed by centrifugation. Concentrations of 1’-OH-midazolam in the supernatant were quantitatively determined using LC/MS and 1’-OH-midazolam standard.

Liquid Chromatography/ Mass Spectrometry Analysis. An LC/MS system consisted of an LC instrument (Thermo Accela HPLC with a photodiode array detector; Thermo Fisher Scientific, Waltham, MA) equipped with an Xbridge C18 column (5 μm, 2.1 × 150 mm; Waters, Milford, MA) and an ion trap mass spectrometer (LTQ XL; Thermo Fisher Scientific) was used for profiling and identification of metabolites formed in HLM. LC separation was carried out using a mobile phase composed of 0.1% formic acid (v/v) in water (solvent A) and 0.1% formic acid in acetonitrile (v/v) (solvent B). A linear gradient of solvent A starting at 95% to 10% from 0 to 23 minutes was applied followed by remaining at 10% for 3 minutes before returning to 95%. The total run time was 30 minutes, and the flow rate was 0.3 ml/min. The mass spectrometer was operating in the positive electrospray ionization mode in the m/z range of 100–1000. Capillary temperature was at 275°C. Capillary voltage was at 46 V. The UV profile of racemic amlodipine and its metabolites in HLM incubations was acquired at 245 nm for semiquantitative estimation of the relative abundance of metabolites to the parent drug. Their full-scan MS and MSn data were acquired using a data-dependent method for metabolite structure characterization. The same LC/MS instrument and method, except for a short LC run time (13 minutes), were used for the determination of relative amounts of the dehydrogenation metabolite (M9) in the time course, HLM inhibition, and metabolism in P450 experiments. In the analysis, full-scan MS data were acquired, and then extracted ion chromatogram at m/z 409 was generated. The peak area corresponding to M9 was calculated as the measurement of the relative abundance of M9.

Results and Discussions

Identification of Amlodipine Metabolites Formed in HLM. LC/MS analysis showed that a total of nine metabolites were formed in HLM incubations with 30 μM racemic amlodipine or S-amlodipine (Table 1). M9 and M10 exhibited relatively higher abundances and accounted for 5.9% and 3.2% of racemic amlodipine, respectively, based on a UV profile (Fig. 1A). Other metabolites were detectable only by LC/MS (Supplemental Fig. 1). Protonated molecular ions and multiple stage MS/MS fragmentation (MSn) spectra of the parent drug and its metabolites are summarized in Table 1. Based on mass spectral interpretation and comparisons with those reported in the literature (Suchanowa et al., 2006, 2008), metabolite structures and their formation pathways are proposed in Fig. 1C. For example, the structure of M9 was ascertained based on the comparison of its MS/MS spectrum (Fig. 1B) to that of the parent drug (Supplemental Fig. 1) as well as that of the amlodipine pyridine metabolite reported previously. Amlodipine dehydrogenation to M9 was the major metabolic pathway in HLM. M9 underwent further metabolism including oxidative deamination to M10, O-demethylation to M1, and O-dealkylation to M4. In addition, amlodipine underwent O-demethylation to M6 and monohydroxylation to M3, M5, and M8 (Fig. 1C; Table 1), which constituted minor pathways in HLM since the metabolites were not displayed in the UV profile (Fig. 1A). Metabolic profiles of racemic amlodipine and S-amlodipine in HLM were similar except that O-demethylation leading to M1 and M6 was not observed for S-amlodipine (Table 1). The major metabolic pathway observed in HLM is consistent with the major clearance pathway of amlodipine in human subjects: amlodipine dehydrogenation followed by oxidations to multiple metabolites (Beresford et al., 1988a; Stopher et al., 1988).

Time Course of Amlodipine Metabolism in HLM. To optimize incubation conditions for assessing the contribution of P450 enzymes in the metabolism of amlodipine, a time course of the formation of major metabolites (M1, M3, M4, M5, M6, M8, M9, M10, M12) was assessed (Fig. 1C). As shown in Table 1, the total abundance of metabolites increased with incubation time and reached a stable level after 24 minutes. The results suggested that a 24-minute incubation time was sufficient for metabolic analysis of amlodipine in HLM.

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>RT (min)</th>
<th>Identity</th>
<th>MH+ Spectral Data</th>
<th>Racemic Amlodipine</th>
<th>S-amlodipine</th>
</tr>
</thead>
<tbody>
<tr>
<td>M1</td>
<td>10.06</td>
<td>P-2H-CH₂</td>
<td>MS²: 376, 350, 332, 314, 304; MS³: 332: 304, 286</td>
<td>✓</td>
<td>–</td>
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<tr>
<td>M3</td>
<td>11.79</td>
<td>P + O</td>
<td>MS²: 379, 336, 294, 364; MS³: 336: 318, 304, 180</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>M4</td>
<td>12.02</td>
<td>P-2H-C₂H₄NH-C₂H₄</td>
<td>MS³: 318,286; MS⁴: 318</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>M5</td>
<td>12.06</td>
<td>P + O</td>
<td>MS²: 417, 390, 379, 336, 294; MS³: 336: 318,304,286,181</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>M6</td>
<td>12.06</td>
<td>P-CH₂</td>
<td>MS²: 352, 334, 306,180; MS³: 180: 163,144</td>
<td>✓</td>
<td>–</td>
</tr>
<tr>
<td>M9</td>
<td>13.57</td>
<td>P-2H</td>
<td>MS²: 390, 364, 346, 318, 286; MS³: 346: 318, 286</td>
<td>✓</td>
<td>✓</td>
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<tr>
<td>M10</td>
<td>17.88</td>
<td>P-2H-NH+O</td>
<td>MS²: 346,318, 286, 390; MS³: 346: 318</td>
<td>✓</td>
<td>✓</td>
</tr>
<tr>
<td>M12</td>
<td>18.38</td>
<td>P-2H-NH+2O-2H</td>
<td>MS²: 376, 346, 318, 286; MS³: 346: 318, 286</td>
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<td>✓</td>
</tr>
<tr>
<td>P</td>
<td>15.02</td>
<td>Amlodipine</td>
<td>MS²: 392, 346, 348, 320, 294, 238; MS³: 392: 360, 346</td>
<td>✓</td>
<td>✓</td>
</tr>
</tbody>
</table>

MH⁺, protonated molecular ions; MSn, multiple stage MS/MS fragmentation; RT, retention time.
to amlodipine dehydrogenation, racemic amlodipine and S-amlodipine were incubated at a lower concentration (1 μM) with HLM, respectively. Metabolite profiles were determined and time courses of the M9 formation were semiquantified by LC/MS. Under the condition investigated, the formation of M9 in HLM increased linearly with increasing incubation time within 15 minutes. No sequential metabolites of M9 were detected, reinforcing the idea that M9 is the intermediate to other pyridine derivatives during amlodipine metabolism. Disappearance of either racemic amlodipine or S-amlodipine was less than 76% of the initial concentrations following 15-minute incubations. Based on these data, 1 μM substrate concentration and 13-minute incubation time were chosen for the subsequent P450 reaction phenotyping experiments. It was expected that the formation of M9 under this condition would be in a linear range.

Identification of P450 Enzyme Responsible for Amlodipine Dehydrogenation. As shown in Fig. 2, the formation of M9 from amlodipine or S-amlodipine was NADPH-dependent and nearly completely inhibited by ketoconazole, a potent inhibitor of both CYP3A4 and CYP3A5, and CYP3cide, a potent and selective inhibitor of CYP3A4 (Walsky et al., 2012). Chemical inhibitors of other major P450 enzymes, including sulfaphenazole (CYP2C9), benzylnirvanol (CYP2C19), quinidine (CYP2D6), montelukast (CYP2C8), furafylline (CYP1A2), and Thio-TEPA (CYP2B6), had no or minimal inhibitory effect on amlodipine dehydrogenation. Similarly, results from metabolism of racemic amlodipine and S-amlodipine by a panel of expressed P450 enzymes showed that CYP3A4 was the single P450 enzyme catalyzing the formation of M9 (Fig. 3).

To further confirm that CYP3A5 has no or minimal catalyzing activity toward amlodipine dehydrogenation, the relative formation rate of M9 from racemic amlodipine in CYP3A4 and CYP3A5 was compared with that of 1'-OH-midazolam from midazolam in CYP3A4 and CYP3A5. Midazolam is a substrate of both CYP3A4 and CYP3A5. Results from the current study showed that the formation rates of 1'-OH-midazolam in expressed CYP3A4 and CYP3A5 were 3.96 and 7.21 (nmol/min/nmol P450), respectively, and the relative formation of 1'-OH-midazolam in CYP3A4 and CYP3A5 was 100:182, which is in agreement with that reported in the literature (Christensen et al., 2011; Li et al., 2012). In contrast, the relative formation of M9 in CYP3A4 and CYP3A5 was 100:11. When CYP3cide, a CYP3A4-specific inhibitor, was coincubated with CYP3A4 and CYP3A5, the M9 formation in CYP3A4 was greatly reduced to the levels in CYP3A5, whereas CYP3cide had no effect on the M9 formation in CYP3A5. These results indicate that CYP3A4 is responsible primarily for the formation of M9 via amlodipine dehydrogenation. CYP3A5 and other P450 enzymes have no or little role in catalyzing amlodipine dehydrogenation. The differences in amlodipine exposures observed between CYP3A5 nonexpressers and expressers (Kim et al., 2006; Zuo et al., 2013) could be due to differences in CYP3A4 expressions between the two groups since CYP3A5 has minimal, if any, effect on the in vitro metabolism of amlodipine (Figs. 2 and 3). The in vitro data also support an early clinical observation that blood pressure response to amlodipine among high-risk African Americans appears to be determined by CYP3A4, but not CYP3A5, genotypes (Bhatnagar et al., 2010).

In summary, racemic amlodipine and S-amlodipine were slowly metabolized in HLM. Amlodipine dehydrogenation to the pyridine metabolite (M9) was the single most important metabolic pathway in HLM; M9 further underwent oxidative deamination, O-demethylation, and O-dealkylation. These in vitro metabolism data are consistent with amlodipine metabolism and disposition observed in vivo in humans. The data derived from amlodipine metabolism in expressed P450 and HLM with P450 inhibitors indicate that CYP3A4, not CYP3A5, is the primary contributor to amlodipine dehydrogenation. Metabolite profiles and P450 reaction phenotyping data of a racemic mixture and S-isomer of amlodipine were very similar, consistent with pharmacokinetics of racemic amlodipine and S-amlodipine in humans. These findings suggest that the observed clinical drug-drug interactions between amlodipine and CYP3A4/5 inhibitors are mediated by CYP3A4 rather than CYP3A5. In addition, polymorphic expressions of CYP3A5 would not affect the pharmacokinetic variability of amlodipine and the blood pressure response to amlodipine in humans.
Metabolism of Amlodipine by HLM and CYP3A4

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References


