Metabolism and Pharmacokinetics of Allitinib in Cancer Patients: The Roles of Cytochrome P450s and Epoxide Hydrolase in its Biotransformation

Lishan Lin, Cen Xie, Zhiwei Gao, Xiaoyan Chen, and Dafang Zhong
Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai, China
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ABSTRACT
Allitinib, a novel irreversible selective inhibitor of the epidermal growth factor receptor (EGFR) 1 and human epidermal receptor 2 (ErbB2), is currently in clinical trials in China for the treatment of solid tumors. It is a structural analog of lapatinib but has an acrylamide side chain. Sixteen metabolites of allitinib were detected by ultra-high-performance liquid chromatography/quadrupole time-of-flight mass spectrometry. The pharmacologically active \(\alpha,\beta\)-unsaturated carbonyl group was the major metabolic site. The metabolic pathways included \(\alpha\)-dealkylation, amide hydrolysis, dihydrodiol formation, hydroxylation, and secondary phase 2 conjugation. The metabolite of amide hydrolysis (M6) and 27,28-dihydrodiol allitinib (M10) were the major pharmacologically active metabolites in the circulation. The steady-state exposures to M6 and M10 were 11% and 70% of that of allitinib, respectively. The biotransformation of allitinib was determined using microsomes and recombinant metabolic enzymes. In vitro phenotyping studies demonstrated that multiple cytochrome P450 (P450) isoforms, mainly CYP3A4/5 and CYP1A2, were involved in the metabolism of allitinib. ThiO conjugates (M14 and M16) and dihydrodiol metabolites (M5 and M10) were detected in humans, implying the formation of reactive intermediates. The formation of a glutathione conjugate of allitinib was independent of NADPH and P450 isoforms, but was catalyzed by glutathione-S-transferase. P450 enzymes and epoxide hydrolase were involved in M10 formation. Overall, our study showed that allitinib was metabolized by the \(\alpha\)-dealkylation pathway similar to lapatinib, but that amide hydrolysis and the formation of dihydrodiol were the dominant metabolic pathways. The absorbed allitinib was extensively metabolized by multiple enzymes.

Introduction
The ErbB family of receptor tyrosine kinases consists of four members: epidermal growth factor receptor (EGFR), ErbB2, ErbB3, and ErbB4 (Klapper et al., 2000; Olajioye et al., 2000). Activation of these receptor tyrosine kinases plays important roles in the proliferation, survival, adhesion, migration, and apoptosis of cells. ErbB2 predominantly exists in an active conformation for lack of any known endogenous ligands, and it prevents the degradation of EGFR. Because of these features, ErbB2 has attracted increasing attention in recent years (Garrett et al., 2003; Xie et al., 2011). Abnormal expression of these receptors is related to the development and migration of tumors (Gonzales et al., 2008). Thus, the ErbB family of receptor tyrosine kinases is an important therapeutic target for the design of anticancer drugs.

Lapatinib (Tykerb, GlaxoSmithKline, Brentford, UK), a 4-anilinoquinazolone derivative, is an orally administered small-molecule reversible inhibitor of EGFR and ErbB2. Lapatinib blocks the phosphorylation and activation of these receptors to prevent downstream signaling events by binding to the ATP-binding site of protein kinases and competing with the ATP substrate (Shewchuk et al., 2000). Although the safety profile of lapatinib is acceptable when treating breast cancer (Bence et al., 2005; Burris et al., 2005; Geyer et al., 2006), hepatotoxicity has been reported in some patients treated with lapatinib (Peroukides et al., 2011).

Allitinib (AST1306; N-(4-(4-(3-fluorobenzyloxy)-3-chlorophenylamino)quinazolin-6-yl)-acrylamide) is a novel analog of lapatinib. In contrast to lapatinib, it is an irreversible inhibitor of EGFR and ErbB2. Allitinib is currently being evaluated in clinical trials in China for the treatment of solid tumors. Unlike other known irreversible EGFR and ErbB2 inhibitors, such as HKI-272 (Rabindran et al., 2004), BIBW2992 (Eskens et al., 2008; Takezawa et al., 2010; Yap et al., 2010), and PF-00299804 (Gonzales et al., 2008), allitinib is more effective in ErbB2-dependent models than in EGFR-dependent models. Compared with lapatinib, allitinib is associated with greater inhibition of EGFR and ErbB2. After incubation of allitinib at different concentrations with kinases at 37°C for 1 hour, allitinib exhibited IC50 values of 0.5 nM and 3.0 nM toward EGFR and ErbB2, respectively. Furthermore, allitinib shows inhibitory activity against...
The pharmacokinetics, metabolism, and disposition of allitinib to their pharmacological or toxicity profiles in vivo (Baillie et al., 2002). However, because this $\alpha,\beta$-unsaturated carbonyl group is electrophilic, allitinib might covalently bind to proteins and other nucleophilic biomolecules, with potentially adverse effects. To evaluate the pharmacology and safety of new drugs, it is necessary to determine their disposition and metabolism during the clinical development program. In particular, the metabolites of new drugs might contribute to their pharmacological or toxicity profiles in vivo (Baillie et al., 2002). The pharmacokinetics, metabolism, and disposition of allitinib have been investigated in rats (data not shown). Allitinib was absorbed quickly, and the maximum concentration was achieved at 1.0–3.0 hours after administration. Its concentration decreased quickly, with $t_{1/2}$ of 3.4–4.4 hours. Systemic available allitinib was extensively metabolized to thiol conjugates and dihydrodiol metabolites, which indicated that allitinib tended to form reactive intermediates. The bioavailability of allitinib was only 5.7% in rats, which might be related to 1) poor solubility, 2) first-pass metabolism, 3) poor permeability, and 4) efflux transport.

The objectives of the current study were as follow: 1) to characterize the metabolic pathways of allitinib in cancer patients at the steady state following its repeated oral administration, 2) to determine the pharmacokinetic profile and routes of excretion of allitinib, and 3) to explore the mechanisms involved in the formation of the major metabolites in vitro to understand the metabolism of the acrylamide group.

Materials and Methods

Chemicals

Reference standards of allitinib (99.7% purity), NB-2 ((E)-N-(4-((3-chloro-4-(3-fluorobenzyl)oxy)phenyl)amino)quinazolin-6-yl)but-2-enamide, 96% purity), O-dealkyl-27,28-dihydrogenated allitinib (M2), and the metabolite of amide hydrolysis (M6) were supplied by Allist Pharmaceuticals Inc. (Shanghai, China). The metabolites O-dealkylallitinib (M1), O-dealky-27,28-dihydrodiol allitinib (M5), 27,28-dihydrodiol allitinib (M10), and allitinib epoxide (M23) were synthesized and purified in our laboratory. Pooled mixed-gender human liver microsomes, pooled mixed-gender human liver cytosol, pooled mixed-gender human intestinal microsomes, pooled mixed-gender human pulmonary microsomes, and recombinant cytochrome P450 (P450) enzymes (CYP1A2, CYP1B1, CYP2A6, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP2E1, CYP3A4, CYP3A5, and CYP4A11) were purchased from BD Gentest (Woburn, MA). Caco-2 cells at passage 17 were purchased from the American Type Culture Collection (Rockville, MD). $\beta$-Glucuronidase, sulfatase, 1-amino-2-naphthoate (ABT), $\alpha$-naphthoflavone ($\alpha$-NF), sulfaphenazole, ticlopidine, quinidine, clomethiazole, ketoconazole (KET), quercetin, and 1-aminobenzotriazole (ABT), were acquired from Sigma-Aldrich (St. Louis, MO). Caco-2 cells at passage 17 were purchased from the American Type Culture Collection (Rockville, MD). $\beta$-Glucuronidase, sulfatase, 1-amino-2-naphthoate (ABT), $\alpha$-naphthoflavone ($\alpha$-NF), sulfaphenazole, ticlopidine, quinidine, clomethiazole, ketoconazole (KET), quercetin, and 1-aminobenzotriazole (ABT), were acquired from Sigma-Aldrich (St. Louis, MO). All solvents were of analytical or high-performance liquid chromatography (HPLC) grade. Purified water was generated with a Milli-Q Gradient Water Purification System (Millipore, Molsheim, France).

Clinical Study Design

The study was performed at Fudan University Shanghai Cancer Center (Shanghai, China) and was approved by the Ethics Committee of Fudan University Shanghai Cancer Center. All subjects gave their written informed consent before enrollment. Three Chinese cancer patients (two males and one female) were given 1,000 mg of allitinib tosylate three times a day orally for 21 consecutive days. Blood samples were collected in heparinized tubes at the following times on days 1 and 24: before and at 0.5, 1, 1.5, 2, 3, 4, and 6 hours after the first dose; before and at 1, 2, 3, 4, 5, and 6 hours after the second dose; and before and at 1, 2, 3, 4, 8, 12, 24, and 36 hours after the third dose. Additional samples were obtained before and 2 hours after the dose on days 10 and 23. Plasma samples were separated and stored at −20°C until analysis. Fecal samples were collected on days 10–12. The total fecal weight was recorded at the end of each collection interval. Five volumes of acetonitrile-water (2:1, v/v) were added to each fecal sample. The mixture was blended by a motor-driven homogenizer and then vibrated by ultrasound waves for 30 minutes. A 50-ml aliquot of the mixture was removed and centrifuged at 3,500 $\times$ g for 5 minutes to produce fecal homogenate.

Another 10 subjects received a single oral dose of 1,000 mg of allitinib tosylate. Urine samples were collected before and at intervals of 0–6, 6–12, 12–24, 24–48, and 48–72 hours after the start of oral administration. The total urine volume was recorded at each collection interval. The fecal homogenate and urine samples were stored at −20°C until analysis.

Metabolite Profiling of Human Plasma, Urine, and Fecal Samples

Sample Preparation. Representative pooled plasma, urine, and fecal samples were prepared for metabolite profiling and identification experiments. Plasma samples taken at 0 hour on day 1 and at 2, 4, and 6 hours after the third dose on day 24 were pooled using equal volumes (100 µl) from each individual sample. Urine samples collected at 0 hour and from 0–24 hours were pooled across all subjects by combining volumes proportional to the total volume excreted by each subject in each collection interval. Three volumes of acetonitrile were added to each plasma and urine sample. After vortex mixing and centrifugation at 11,000 $\times$ g for 5 minutes, the supernatant was transferred into a clean plastic tube and evaporated to dryness under a stream of air at 40°C. The residues of the plasma and urine samples were reconstituted in 100 µl of 90% aqueous acetonitrile (v/v). Then, 10 µl of the reconstituted solution was injected into an ultra-high-performance liquid chromatograph coupled to a quadrupole time-of-flight mass spectrometer (UHPLC/Q-TOF MS) system for analysis. Fecal samples were pooled by combining volumes proportional to the total volume of each fecal homogenate from all subjects. The pooled fecal homogenates were directly injected into the UHPLC/Q-TOF MS system for analysis.

Enzyme Hydrolysis

Pooled human urine samples were incubated with sulfatase and $\beta$-glucuronidase in 1 M citrate buffer (pH 5.0) in a water bath at 37°C for 16 hours. The mixtures were treated as described above and analyzed.

Synthesis of M1

Allitinib was stirred with aluminum chloride in acetone at room temperature for 10 hours. The mixture was then separated on a YMC-Pack ODS column (250 × 10 mm I.D., 5 µM; YMC Company Ltd., Kyoto, Japan) to obtain pure M1.

Synthesis of M5 and M10

M1 and citric acid (0.2 molar equivalent) were dissolved in acetonewater (5:1, v/v) in a 100-ml round-bottom flask. Potassium osmate (0.2 molar equivalent) was then added, followed by 4-methylmorpholin N-oxide (1.1 molar equivalent). The mixture was stirred at room temperature for 48 hours and quenched by the addition of sodium hyposulfite. Acetone was removed by placing the flask on a rotary evaporator. The aqueous residue was extracted with ethyl acetate and separated on a YMC-Pack ODS column (250 × 10 mm I.D., 5 µM; YMC Company Ltd.) to yield pure M5. The M10 synthetic standard was acquired with the same chemical reaction after replacing M1 with allitinib.

Synthesis of Allitinib Epoxide

Allitinib tosylate (0.18 mmol) was dissolved in 3.75 ml formic acid, and 2 ml H$_2$O$_2$ was added. The contents were heated at 45°C for 3 hours. After adding two volumes of acetonitrile-water (2:1, v/v) and adjusting the pH value to 3 with 1% ammonia, the mixture was then separated on a Shim-Pack PREP-ODS (H) KIT column (250 × 20 mm I.D., 5 µM; Shimadzu, Kyoto, Japan) to yield pure allitinib epoxide.
Pharmacokinetic Analysis

The pharmacokinetic characteristics of allitinib, M6, and M10 were analyzed using the noncompartmental model with Winnonlin 5.3 software (Pharsight, Mountain View, CA). The $C_{max}$ and the time to reach the maximum concentration were directly determined from the experimental data. The linear-log trapezoidal method was used to calculate the area under the plasma concentration-time curve for $0-24$ hours ($AUC_{0-24}$). The accumulation ratio (day 24/day 1) for $AUC_{0-24}$ was also calculated. The elimination rate constant ($k_e$) was determined by least squares regression of the terminal log-linear phase of the concentration-time curve. The terminal half-life was estimated as $\ln 2/k_e$.

In Vitro Pharmacological Activity of the Metabolites of Allitinib

The inhibitory activities of M1, M6, and M10 against EGFR and ErbB2 were determined as previously described (Xie et al., 2011).

Microsomal Incubation

A stock solution of allitinib was prepared in dimethyl sulfoxide (DMSO). A mixture containing 1-$\mu$M allitinib was mixed with human liver microsomes (HLM) (1 mg/ml), human intestinal microsomes (HIM) (1 mg/ml), and human pulmonary microsomes (HPM) (1 mg/ml), in addition to phosphate-buffered...
saline (PBS) (100 mM), at pH 7.4 without or with 1 mM glutathione (GSH) at a final volume of 200 μl. The final DMSO concentration was 0.1%. After preincubation at 37°C for 3 minutes, 1 mM NADPH was added to initiate the reaction. After incubation for 60 minutes, two volumes of ice-cold acetonitrile were added to terminate the reaction. Control samples without NADPH or microsomes were also prepared. To evaluate the contribution of glutathione transferase (GST) to the formation of the reactive metabolites, separate samples were prepared with 1 mg/ml human liver cytosol (HLC), 1 mM allitinib, and 1 mM GSH. Each incubation was performed in duplicate. After the samples were centrifuged at 11,000 g, the supernatants were evaporated to dryness under an air stream at 40°C. The residues were reconstituted in 100 μl water/acetonitrile (90:10, v/v). Then, a 10-μl aliquot of the reconstituted solution was injected into the UHPLC/Q-TOF MS system for analysis.

Identification of CYP Enzymes Involved in the Metabolism of Allitinib

A stock solution of allitinib was prepared in DMSO. Allitinib (1 μM) was incubated in the presence of individually expressed recombinant P450 enzymes (50 pmol/ml 1A2, 1B1, 2A6, 2B6, 2C8, 2C9, 2C19, 2D6, 2E1, 3A4, 3A5, or 4A11 suspended in 100-mM PBS, pH 7.4) and 2-nM NADPH at 37°C for 60 minutes. The final DMSO concentration during incubation was 0.1%. Two volumes of ice-cold acetonitrile containing an internal standard (500 ng/ml NB-2) were added to terminate the reaction. A sample incubated without NADPH was used as the negative control.

Inhibition of the Oxidative Metabolism of Allitinib by Selective P450 Inhibitors

To evaluate the relative contributions of different microsomal enzyme systems to the metabolism of allitinib in HLM, 1 μM allitinib was incubated at 37°C for 60 minutes with 1 mg/ml HLM alone or in the presence of different chemical inhibitors of P450, including the nonspecific P450 inhibitor 1-aminobenzotriazole (ABT) (1 mM), the specific CYP1A1/2 inhibitor α-NF (2 μM), the CYP2C8 inhibitor quercetin (1 μM), the CYP2C9 inhibitor sulfaphenazole (6 μM), the CYP2B6/2C19 inhibitor ticlopidine (0.4 μM), the CYP2D6 inhibitor quinidine (2 μM), the CYP2E1 inhibitor clomethiazole (0.1 μM), or the CYP3A4/5 inhibitor KET (1 μM). After incubation, two volumes of ice-cold acetonitrile containing NB-2 were added to terminate the reaction. Each incubation was performed in duplicate. The MS peak area ratios of allitinib and the detected metabolites to the internal standard in each incubation system were recorded to determine the contributions of the P450 enzymes to the metabolism of allitinib. The results were compared with those for control samples lacking the inhibitors.

Mechanism of M10 Formation

Valpromide (10, 100, or 1,000 μM), an inhibitor of epoxide hydrolase, was preincubated with HLM and NADPH for 3 minutes. Allitinib was added to the mixture to a final volume of 200 μl. A control sample without valpromide was...

Fig. 2. Metabolic profiles of pooled plasma samples at the peak (A) (2 hours after the third dose on day 24) and trough (B) (4 hours after the third dose on day 24) steady-state levels of allitinib after the oral administration of 1,000 mg of allitinib tosylate three times a day for 21 days to cancer patients. Upper trace, UHPLC-UV chromatogram. Lower trace, mass defect filter–processed chromatogram.

Fig. 3. Metabolic profiles of pooled urine samples collected at 0–24 hours after the oral administration of 1,000 mg of allitinib tosylate daily in cancer patients (A) and pooled feces samples collected at days 10 to 12 after the oral administration of 1,000 mg of allitinib tosylate three times a day for 21 days in cancer patients (B). Upper trace, UHPLC-UV chromatogram. Lower trace, mass defect filter–processed chromatogram.
Caco-2 Cell Permeability Study

For the cell permeability experiments, Caco-2 cells were seeded in 24-well inserts at a density of 10^5 cells/cm^2 to generate Caco-2 monolayers. The cells were cultured for 21 days, during which time the medium was replaced every 3 days. Cell layers with a transepithelial electrical resistance value > 300 Ω·cm^2 were used in the permeability experiment. Before the assay, the monolayers were gently washed twice with warm HBSS (pH 7.4, 37°C). HBSS containing 2 μM allitinib was added to the apical side (150 μl) or the basolateral side (750 μl) of the inserts to initiate the experiment. The cells were then incubated at 37°C for 2 hours in a humidified atmosphere containing 5% CO₂. Next, 200 μl of transport buffer was collected from the apical or basolateral side and mixed with the same volume of methanol. HBSS containing allitinib at the donor side was collected before and after the assay to calculate assay recovery. After washing three times, cell monolayers were lysed with 200-μl acetonitrile. All incubations were performed in triplicate. The concentration of allitinib was analyzed by LC-MS/MS.

The apparent permeability coefficients (P_{app}, cm/s) were calculated with the following equation:

$$P_{app} = \frac{\Delta Q/\Delta t}{A \times C_{donor}}$$

where \(\Delta Q/\Delta t\) is the rate of permeability (nmol/s), \(A\) is the surface area of the insert (cm^2), and \(C_{donor}\) is the initial concentration on the donor side (nmol/ml). The efflux ratio (ER) was determined as follows:

$$ER = \frac{P_{app}(B-A)}{P_{app}(A-B)}$$

where \(P_{app}(A-B)\) and \(P_{app}(B-A)\) represent the apparent permeability coefficients of the substrate from the apical to the basolateral side and from the basolateral to the apical side, respectively.

Analytical Conditions

Metabolite Profiling by UHPLC-UQ-TOF-MS. Metabolite profiling of allitinib in biologic samples was performed on a Waters Acquity UHPLC system (Waters, Milford, MA) equipped with a binary solvent delivery pump, column oven, UV detector, and autosampler. Chromatographic separation was performed on an Acquity UPLC HSS T3 column (100 × 2.1 mm I.D., 1.8 μm; Waters) at 40°C. The mobile phase was a mixture of 5-mM ammonium formate in water containing 0.05% formic acid (A) and acetonitrile (B) at a flow rate of
0.4 ml/min. Elution started with a 1-minute isocratic run with 10% solvent B, followed by a linear gradient of 10% to 55% solvent B in 14 minutes, 55% to 99% solvent B in 1 minute, maintained for 1 minute, and then reduced to 10% B to equilibrate the column. The eluate was monitored by UV detection at 300 nm.

For metabolite profiling, MS detection was conducted with a Synapt Q-TOF high-resolution mass spectrometer (Waters) in the positive electrospray ionization mode. Nitrogen and argon were used as the desolvation and collision gases, respectively. The capillary and cone voltages were set at 3,000 and 40 V, respectively. Data from 80 to 1,000 Da were acquired using a source temperature of 120°C and a desolvation temperature of 350°C. Data were corrected during acquisition using a reference (LockSpray) sample consisting of 400 ng/ml leucine encephalin ([m/z] 556.277) infused at 20 µl/min. An MS² with two separate scan functions programmed with independent collision energies (CEs) was used for data acquisition. At low CE, the transfer and trap CEs were 2 and 3 eV, respectively. At high CE, the transfer CE was 10 eV, and the trap CE ranged from 10 to 20 eV. This mode of data collection allowed us to detect intact precursor ions and fragments.

Data processing was performed using a MetaboLynx subroutine of the MassLynx software (Waters). Mass defect filtering was used to screen metabolites using a 40-mDa filter between the filter and the target metabolites. The fragment ion spectra were compared between the parent compound and the metabolites to help identify the metabolites, including the structure and site(s) of modification in the parent molecule.

To determine M23 in HLC incubation, MS detection was conducted on a triple TOF 5600+ MS/MS system (AB Sciex, Concord, Ontario, Canada) in the positive ESI mode. Mass range was set at [m/z] 100–1,000. The following parameter settings were used: ion spray voltage: 5,500 V, declustering...
TABLE 2

<table>
<thead>
<tr>
<th>Epoxide</th>
<th>Position</th>
<th>Allitinib</th>
<th>M6</th>
<th>M10</th>
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<tbody>
<tr>
<td>M1</td>
<td>a</td>
<td>8.89, s</td>
<td>8.67, s</td>
<td>8.50, s</td>
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<tr>
<td>M1</td>
<td>b</td>
<td>8.89, d, J = 9.2 Hz</td>
<td>8.67, d, J = 9.2 Hz</td>
<td>8.50, d, J = 9.2 Hz</td>
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<tr>
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<td>a</td>
<td>8.40, s</td>
<td>8.42, s</td>
<td>8.40, s</td>
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<td>M3</td>
<td>b</td>
<td>8.40, d, J = 9.2 Hz</td>
<td>8.42, d, J = 9.2 Hz</td>
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<tr>
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<td>a</td>
<td>8.31, s</td>
<td>8.33, s</td>
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<td>b</td>
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<td>a</td>
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<td>8.04, s</td>
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<td>b</td>
<td>8.02, d, J = 9.2 Hz</td>
<td>8.04, d, J = 9.2 Hz</td>
<td>8.02, d, J = 9.2 Hz</td>
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<td>a</td>
<td>7.93, s</td>
<td>7.95, s</td>
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<td>b</td>
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<td>a</td>
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<td>b</td>
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<td>7.77, d, J = 9.2 Hz</td>
<td>7.75, d, J = 9.2 Hz</td>
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Note: a = NMR, nuclear magnetic resonance.

CD3OD, 400 MHz.

DMSO-d6, 500 MHz.

Potential: 80 V, ion source heater: 550°C, curtain gas: 40 psi, ion source gas 1: 55 psi, ion source gas 2: 55 psi. For TOF MS scans, collision energy was 10 eV; for product ion scans, CE was 55 eV, and collision energy spread was 10 in the MSMS experiment. Information-dependent acquisition (IDA) was used to trigger acquisition of MSMS spectra for ions matching the IDA criteria. The real-time Multiple Mass Defect Filter was used in IDA criteria.

Quantification of Allitinib and its Metabolites in Plasma with LC-MS/MS.
The concentrations of allitinib, M6, and M10 were quantified simultaneously using a previously described LC-MS/MS method (Lin et al., 2013).

Quantification of M1 in Urine with LC-MS/MS.
The LC system consisted of two LC-20ADXR pumps and a SIL-20ACXR autosampler (Shimadzu, Kyoto, Japan). A TSQ Quantum Vantage triple-quadrupole MS (Thermo Fisher Scientific, Waltham, MA) equipped with a positive electrospray ionization source was used for MS detection. Selected reaction monitoring was used to detect the analyte and internal standard. The multiple reaction monitoring transitions m/z 341.0 → m/z 251.0 + 277.0 + 285.0 + 305.0 and m/z 387.0 → 167.0 were used to detect M1 and irinotecan, respectively.

The concentration of M1 in urine was determined using a validated LC-MS/MS method. After incubating equal volumes of β-glucuronidase (2,000 units of type HA-4, Sigma-Aldrich) and sulfatase (15 units of type VIII, Sigma-Aldrich) in 1 M citrate buffer (pH 5.0) at 37°C for 16 hours, M1 and the metabolites of allitinib were identified by comparing their chromatographic and MS fragmentation behaviors with those of the parent drug and the synthesized reference standards. We first examined the chromatographic and MS fragmentation behaviors of the reference allitinib. Allitinib was eluted at 14.1 minutes and a protonated molecule [M+H]+ was detected at m/z 449.120 in the positive ion mode. The

Results

MS Fragmentation Behavior of Allitinib. The metabolites of allitinib were identified by comparing their chromatographic and MS fragmentation behaviors with those of the parent drug and the synthesized reference standards. We first examined the chromatographic and MS fragmentation behaviors of the reference allitinib. Allitinib was eluted at 14.1 minutes and a protonated molecule [M+H]+ was detected at m/z 449.120 in the positive ion mode. The
oral administration of 1,000 mg of allitinib tosylate three times a day of allitinib, M6, and M10 in human plasma were quantified after the Table 2.

mass spectra with those of reference standards. The 1H-nuclear confirmed by comparing their chromatographic retention times and the dihydrodiol metabolites in plasma suggested the formation of reactive Supp.

Results. The detection of cysteine conjugates and proposed metabolic pathways of allitinib in humans are shown in Fig. 4.

elemental compositions, and characteristic fragment ions. The pro-

metabolites of allitinib, including their retention times, proposed M0 was the predominant compound in feces, whereas M2 and M6 whereas those in urine were M1 and its glucuronide conjugate, M13. were relatively minor. Table 1 lists the characteristics of the possible M16 based on their

human plasma, urine, and feces, respectively (Fig. 2 and Fig. 3). The results of in vitro pharmacological studies of allitinib and its metabolites

Analyte  
EGFR  
ErbB2  

<table>
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<th>IC50 (nM)</th>
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<td>Allitinib</td>
<td>0.263 ± 0.044</td>
<td>1.10 ± 0.40</td>
</tr>
<tr>
<td>M1</td>
<td>0.773 ± 0.023</td>
<td>25.0 ± 5.0</td>
</tr>
<tr>
<td>M6</td>
<td>1.600 ± 0.960</td>
<td>282 ± 84</td>
</tr>
<tr>
<td>M10</td>
<td>0.755 ± 0.573</td>
<td>72.0 ± 13.0</td>
</tr>
</tbody>
</table>

TABLE 4
Results of in vitro pharmacological studies of allitinib and its metabolites

All values are mean ± S.D.

high-CE MS (Fig. 1A) yielded fragment ions at m/z 340.074, 339.069, 313.054 (100% abundance), 311.074, 305.109, 287.071, 277.107, and 213.081. The predominant ion at m/z 313.054 was formed via the loss of terminal olefin and cleavage of the alkyl C-O bond. The radical ion at m/z 340.074 formed from the hemolysis of the alkyl C-O bond, which was followed by the loss of a specific 109 Da moiety (1-fluoro-3-toluene free radical), while m/z 339.069 was proposed to form from the cleavage of alkyl C-O bond. The ions at m/z 311.074, 305.109, 287.071, 277.107, and 213.081 formed from the cleavage of N-(N4-((3-chloro-4-hydroxyphenyl)amino)quinazolin-6-yl)acrylamide. According to the fragmentation pattern, the structure of allitinib was divided into segments A, B, and C (Fig. 1B). The structures of the metabolites were characterized by determining the changes in the m/z values of these three segments.

A total of 11, 12, and 10 metabolites of allitinib were detected in human plasma, urine, and feces, respectively (Fig. 2 and Fig. 3). The metabolites were designated as M0 (parent compound, allitinib) to M16 based on their m/z values. According to the UV chromatograms, the major components in plasma were M0 followed by M6 and M10, whereas those in urine were M1 and its glucuronide conjugate, M13. M0 was the predominant compound in feces, whereas M2 and M6 were relatively minor. Table 1 lists the characteristics of the possible metabolites of allitinib, including their retention times, proposed elemental compositions, and characteristic fragment ions. The proposed metabolic pathways of allitinib in humans are shown in Fig. 4. The information used to assign each metabolite is summarized in the Supplemental Results. The detection of cysteine conjugates and dihydrodiol metabolites in plasma suggested the formation of reactive intermediates. The structures of M1, M2, M5, M6, and M10 were confirmed by comparing their chromatographic retention times and the mass spectra with those of reference standards. The 1H-nuclear magnetic resonance data for the reference standards are listed in Table 2.

Pharmacokinetics and Excretion of Allitinib. The concentrations of allitinib, M6, and M10 in human plasma were quantified after the oral administration of 1,000 mg of allitinib tosylate three times a day for 21 consecutive days. The pharmacokinetic characteristics of allitinib, M6, and M10 are listed in Table 3. The Cmax values for allitinib, M6, and M10 were achieved at 3.6–5.2 hours after the first dose. The AUC0–24 values for M6 and M10 were 14% and 65% of the corresponding values of the parent drug (based on molar concentrations), respectively. After the last dose, the Cmax values for allitinib, M6, and M10 were achieved at approximately 2.5–3.2 hours.

At steady state, the terminal half-lives of allitinib, M6, and M10 were observed at 4.5–9.9 hours, at 6.3–15.8 hours, and at 4.6–11.6 hours, respectively. The AUC0–24 values for M6 and M10 were 11% and 70% of that for the parent drug, respectively. The accumulation ratios of allitinib, M6, and M10 were 1.39–2.02, 1.06–2.78, and 0.74–2.43, respectively.

At steady state, the daily fecal excretion of allitinib and its major metabolites M2, M5, and M6 accounted for 32.5% ± 17.4% of the dose. In the fecal samples, unchanged allitinib accounted for 30.4% ± 20.3% of the dose, whereas the major metabolites M2, M5, and M6 accounted for 1.41%, 0.067%, and 0.635% of the dose, respectively. After a single oral dose of 1,000 mg of allitinib tosylate, a combination of M1 and its glucuronide conjugate in urine accounted for 0.042% of the dose.

In Vitro Pharmacological Activity of the Major Metabolites of Allitinib. The results of in vitro pharmacological activity studies of the major metabolites are presented in Table 4. The results showed that M1, M6, and M10 were potent inhibitors of EGFR but weak inhibitor of ErbB2.

Microsomal Incubation. After incubation with microsomes for 60 minutes, approximately 58% of the parent compound was consumed by HLM, compared with 10% by HIM. Allitinib was not metabolized by HPM. Therefore, the liver was considered to be the major site of allitinib biotransformation.

Five metabolites were generated by incubating allitinib with HLM in the presence of NADPH. Based on the MS peak areas, the main metabolite in vitro was M5, followed by M1, M10, and M6. In samples lacking NADPH, only M6 was detected, which indicated that the formation of M1, M5, and M10 was NADPH-dependent, and that P450 enzymes played important roles in the formation of these metabolites. The formation of M6 was NADPH-independent, and amidoxygenase was assumed to be responsible for its formation. The properties of the metabolites identified after allitinib was incubated with HLM are listed in Table 5.

Identification of P450 Enzymes Involved in the Oxidative Metabolism of Allitinib. In vitro studies were performed to identify which P450 enzymes were involved in the metabolism of allitinib and the formation of M1 and M10. As shown in Fig. 5, after 1-μM allitinib was incubated with recombinant P450 enzymes in the presence of NADPH, it was metabolized by 11 of the enzymes tested, but not CYP2C9. Based on the mean hepatic expression of the P450 enzymes (Rodrigues, 1999), the predominant enzyme involved in
the metabolism of allitinib was CYP3A4, followed by CYP3A5 and CYP2C8. In the presence of NADPH, M1 was mainly produced by recombinant CYP3A4 and CYP2C8, followed by CYP2C19, CYP2E1, and CYP3A5, whereas M10 was mainly produced by CYP1A2, followed by CYP2D6 and CYP3A5.

To further determine the relative contributions of the different P450s in allitinib biotransformation, ABT (nonspecific P450 inhibitor), α-NF (CYP1A2 inhibitor), sulfaphenazole (CYP2C9 inhibitor), ticlopidine (CYP2B6/CYP2C19 inhibitor), quercetin (CYP2C8 inhibitor), quinidine (CYP2D6 inhibitor), clomethiazole (CYP2E1 inhibitor), and KET (CYP3A4/5 inhibitor) were individually added to HLM preparations. The results are presented in Fig. 6. The metabolism of allitinib was inhibited by all seven P450 inhibitors. The amount of allitinib that remained following incubation with ABT was approximately three times greater than that in the control group, suggesting that P450 enzymes played important roles in the oxidative metabolism of allitinib. The formation of M1 and M10 was markedly reduced in the presence of ABT, indicating that P450 enzymes were important in the production of M1 and M10. The formation of M1 was inhibited by 27.8% and 25.5% in the presence of quinidine and KET, respectively. The formation of M10 was inhibited by 18.6% and 44.0% in the presence of α-NF and KET, respectively. These results indicated that CYP2D6 and CYP3A4/5 were the major enzymes involved in the formation of M1, whereas CYP3A4/5 was the predominant enzyme involved in the production of M10, followed by CYP1A2.

**Epoxide Hydrolase is Responsible for the Formation of M10.** It has been suggested that M10 is formed by P450 enzymes via an epoxide intermediate, and the resulting epoxide is hydrolyzed to M10 by epoxide hydrolase. To evaluate this possibility, HLMs were incubated with valpromide, an epoxide hydrolase inhibitor, in the presence of NADPH. The coincubation of allitinib with valpromide resulted in the production of epoxide, which was eluted at 13.2 minutes. The protonated molecule of epoxide was detected at m/z 465.105, and so was 16 Da larger than that of allitinib, indicating that an oxygen atom was introduced into allitinib. The ion at m/z 356.067 was formed by the specific loss of 109 Da. The product ion at m/z 287.056 was identical to that of the parent drug, whereas the product ions at m/z 339.069, 311.072, 305.109, and 277.103, respectively. These results suggested that an oxygen atom was probably attached to segment C of allitinib to form an epoxide. The identity of allitinib epoxide was further confirmed using synthetic standards.

As shown in Fig. 7, the formation of M10 decreased and the production of allitinib epoxide increased with increasing concentrations...
of valpromide. These results confirmed that epoxide hydrolase displayed an important role in the formation of M10.

Mechanism Involved in Thiol Conjugate Formation. Cysteine conjugates (M9 and M14) and N-acetyl cysteine conjugates (M12 and M16) were detected in humans, indicating the formation of reactive intermediates. To determine the mechanism involved in the formation of these thiol conjugates, in vitro studies were performed using GSH as a trapping agent. The coincubation of allitinib with GSH resulted in the production of a GSH conjugate, M22, which was determined as a major metabolite in rats after intravenous injection. It was eluted at 9.6 minutes. The protonated molecule of M22 was detected at \( m/z \) 756.206, and was 307.090 Da larger than that of allitinib, suggesting the addition of a GSH molecule. Product ions were observed at \( m/z \) 627.163, 449.096, 341.092, and 287.073. The ion at \( m/z \) 627.163 resulted from the loss of the pyroglutamic acid moiety (129 Da). The product ions at \( m/z \) 341.092 and 287.073 were identical to those of the parent drug. These results suggested that the GSH molecule probably attached to segment C of allitinib by Michael addition.

M22 was also observed when allitinib was incubated without NADPH or HLM, implying that the formation of the GSH conjugate was independent of NADPH and P450. The chromatographic peak of M22 increased when allitinib was incubated with human liver cytosol, indicating that the formation of M22 was catalyzed by GST. Cysteine conjugate M14 was also detected when allitinib was incubated with GSH, HLM, and NADPH, suggesting that the cysteine conjugates of allitinib in humans might be partly derived from GSH conjugates via the mercapturate pathway.

In Vitro Metabolism of Allitinib Epoxide by HLC. The coincubation of allitinib epoxide (2 \( \mu \)M) with GSH in HLC allowed the formation of M6, M10, and a GSH conjugate, M23, which was eluted at 9.3 minutes. M23 exhibited \([M+H]^+\) ions at \( m/z \) 772.196, and was 307.083 Da larger than that of allitinib epoxide, suggesting the addition of a GSH molecule. The loss of pyroglutamic acid moiety resulted in the formation of a fragment ion at \( m/z \) 643.154. The product ions at \( m/z \) 355.062, 327.072, and 321.101 were identical to those of allitinib epoxide. These results suggested that the GSH molecule most likely be attached to the epoxy part of allitinib epoxide. After incubation of 20-nM allitinib epoxide with GSH in HLC, M6 and M10, but not M10, were detected (Fig. 8).

Caco-2 Cell Permeability Study. The permeability of Caco-2 cells to allitinib was determined at a concentration of 2 \( \mu \)M. The \( P_{app(A\rightarrow B)} \) of allitinib was 0.77 \( \times 10^{-6} \) cm/s, indicating that the cells showed poor permeability to allitinib. When the transport of allitinib in the apical-to-basolateral direction was compared with that in the basolateral-to-apical direction, no apparent efflux transport was observed based on an ER of 1.40.

Discussion

This study characterized the metabolic profiles of allitinib in cancer patients and established its biotransformation mechanisms. Allitinib was quickly absorbed, with a time to reach maximum concentration of approximately 3.0 hours, consistent with the results of the preclinical study. Significant interpatient variability was observed after the oral administration of allitinib tosylate, indicating that dose modifications may be necessary to meet individual patients’ needs. The unmodified drug was the predominant substance detected in plasma. The amide hydrolysis metabolite (M6) and 27,28-dihydrodiol allitinib (M10) were the major pharmacologically active metabolites, accounting for approximately 11% and 70% of the estimated AUC0–24 for the parent drug at steady state, respectively. The systemic clearance of M6 and M10 was slower than that of allitinib.

A total of 10 and 12 metabolites were identified in feces and urine, respectively. Unchanged allitinib was detected in trace amounts in
urine, suggesting that the systemically available allitinib was metabolized extensively. The proposed structures of the metabolites M1, M2, M5, M6, and M10 were supported by comparisons with synthetic standards. The metabolic scheme of allitinib in humans is presented in Fig. 4. The major metabolites in urine were O-dealkyllallitinib (M1) and its glucuronide conjugate (M13). After the urine was incubated with β-glucuronidase and sulfatase for 16 hours, the recovery of allitinib in terms of M1 was less than 0.1%, indicating that renal elimination makes a negligible contribution to the excretion of allitinib. After the oral administration of allitinib tosylate, the mean fecal excretion ratio was 32.5% ± 17.4% (range from 0.95% to 59.4%). About 93.5% of the total fecal excretion was attributed to the unchanged drug. The major metabolite, M2, accounted for 4.4% of the total fecal extraction, and the other two metabolites (M5 and M6) accounted for <3.0%. The recovery of allitinib in fecal samples seemed to be related to the fecal weight excreted by the subjects. Several factors may contribute to the low recovery of allitinib in humans after its oral administration, including the difficulty in fully sampling the nonhomogeneously processed fecal samples, the loss of samples during collection or processing, or the covalent binding or noncovalent tissue uptake of the drug or its metabolites (Roffey et al., 2007). However, the results suggested that allitinib is predominantly eliminated via fecal excretion. To confirm this possibility, radiolabeled allitinib will be used in future mass balance experiments.

To further investigate the source of unchanged drug in feces, in vitro Caco-2 permeability study was performed. The results showed that the permeability of these cells to allitinib was poor ($P_{app} < 2.0 \times 10^{-6}$ cm/s). Thus, it might be concluded that the parent drug determined in human fecal samples was generated from the unabsorbed parent drug due to poor bioavailability rather than through biliary excretion after absorption.

The primary routes of allitinib biotransformation involved O-dealkylation (M1, M2, M3, M4, M5, M8, M9, M12, and M13), amide hydrolysis (M6), dihydrodiol formation (M5 and M10), and the subsequent phase 2 conjugation of these metabolites. The identified metabolic processes of lapatinib and allitinib are shown in Fig. 9. Allitinib shared several metabolic pathways with its analog lapatinib (Castellino et al., 2012), including the O-dealkylation and hydroxylation of the quinazoline moiety. However, the major metabolic sites of allitinib are located at the α,β-unsaturated carbonyl group, and include amide hydrolysis and dihydrodiol formation. The formation of cysteine conjugate in vivo indicated that allitinib might covalently bind to proteins or biologic macromolecules. The protein covalent binding ratios after incubation of allitinib with human plasma and HLM at 37°C for 1 hour were both less than 10.1%, and protein covalent binding of allitinib was time-dependent (data not shown). Further investigation is needed to understand the impact of protein covalent binding on the potency of this drug. In vitro studies revealed that the reaction between allitinib and GSH was independent of microsomal P450 enzymes and NADPH, but that GST could catalyze the conjugation of allitinib and GSH.

In vitro metabolism studies using recombinant human P450 isozymes and inhibition studies using selective chemical inhibitors of P450 enzymes suggested that the oxidative metabolism of allitinib is mediated by multiple P450 enzymes. Allitinib was primarily metabolized by CYP3A4/5, followed by CYP1A2, CYP1B1, CYP2A6,
CYP2B6, CYP2C8, CYP2C19, CYP2D6, CYP2E1, and CYP4A11. The formation of M1 was primarily mediated by CYP3A4/5 and CYP2D6, whereas CYP1A2 and CYP3A4/5 were mainly involved in the formation of M10. The incubation of allitinib with the CYP1A2 inhibitor α-NF in an HLM preparation markedly increased the production of M1, which was probably attributable to the stimulation of the CYP3A system by α-NF (Ueng et al., 1997; Maenpaa et al., 1998). Because multiple P450 enzymes were involved in the metabolism of allitinib, in vivo exposure to allitinib is unlikely to be markedly affected by P450 enzyme inhibitors. Therefore, the likelihood of drug-drug interactions mediated by the metabolism of allitinib is minimal.

The incubation of allitinib with HLM and valpromide, an inhibitor of epoxide hydrolase, confirmed that allitinib is metabolized to epoxide by P450 enzymes, and the epoxide metabolite is further metabolized to M10 by epoxide hydrolase. Epoxides can react with cellular macromolecules, such as DNA and protein, to exert mutagenic and genotoxic effects. Alternatively, epoxides can be metabolized to GSH conjugates by GST or to dihydrodiol metabolites by epoxide hydrolase with reduced toxicity (Ehrenberg and Hussain, 1981; Hooberman et al., 1993; Faller et al., 2001; Lee et al., 2005; Gonzalez-Perez et al., 2012). The incubation of allitinib with HLM supplemented with NADPH and GSH confirmed that the pharmaco logically active metabolite M10 was the major metabolite, but the GSH conjugate of epoxide M23 was not observed. M10 and M23 were both derived from allitinib epoxide. Our studies showed that allitinib epoxide could form a GSH conjugate at the concentration of 2 μM. However, when a much lower substrate concentration (20 nM) was used, the GSH conjugate of epoxide could not be detected, whereas the formation of M10 was observed, indicating that the epoxide intermediate was more readily metabolized to dihydrodiol than to the GSH conjugate.

An in vitro investigation showed that lapatinib was metabolized to O-dealkylated lapatinib by CYP3A4 and CYP2C8, and GSH and cysteinyl-glycine conjugates were also observed (Teng et al., 2010). Although allitinib was metabolized by O-dealkylation, like lapatinib, we detected no conjugates formed by the reaction between GSH and the quinone imine intermediate. This might be explained by the greater reactivity of the α,β-unsaturated carbonyl group of allitinib. Consequently, GSH reacted with the α,β-unsaturated carbonyl group more readily than with the quinone imine group.

Lapatinib is suggested to be a quasi-irreversible inhibitor of CYP3A4, which is mediated by metabolites derived from N-oxidation (Takakusa et al., 2011). Another in vitro study showed that lapatinib is also a mechanism-based inactivator of CYP3A5, and this inactivation is mediated by quinone imine (Chan et al., 2012). These studies provide good insight into the possible mechanism of hepatotoxicity of lapatinib observed in clinical trials. To determine whether allitinib inhibits P450s because its structure is analogous to that of lapatinib and its introduced acrylamide group, in vitro enzyme induction and inhibition studies were performed. The results demonstrated that allitinib did not inhibit CYP1A2, CYP2C9, CYP2C19, or CYP2D6, but was a weak inhibitor of CYP3A4 (IC₅₀ ~ 100 μM with midazolam as the substrate). This level of inhibition is negligible at clinical doses. Furthermore, we observed no apparent enzyme induction. These studies suggest that, even though the α,β-unsaturated carbonyl group was introduced and a quinone imine intermediate might be formed via O-dealkylation, allitinib does not affect P450 enzyme activity. The probability that allitinib has a pharmacologic effect on coadministered drugs via the induction or inhibition of P450 enzymes is minimal.

**Fig. 9.** Identified metabolic processes of lapatinib (A) and allitinib (B) in humans.
is low, even though the recommended dose of allitinib is 1,000 mg three times a day.

In conclusion, we found that 0-dealkylation is an important metabolic pathway of allitinib, as it is for lapatinib. However, we also found that amid hydrolysis and dihydrodiol formation are major metabolic pathways, because the reactivity of the α,β-unsaturated carbonyl group is high. Multiple enzymes, including P450 and epoxide hydrolase, are involved in the metabolism of allitinib, but allitinib does not induce or inhibit tested P450 enzymes, suggesting a low potential for drug-drug interaction mediated by tested P450 enzymes in cases of coadministration.

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Authorship Contributions
Participated in research design: Lin, Zhong, Chen.
Conducted experiments: Lin, Xie, Gao.
Contributed new reagents or analytic tools: Lin, Zhong, Chen.
Performed data analysis: Lin, Zhong, Xie, Gao, Chen.
Contributed to the writing of the manuscript: Lin, Zhong.

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