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Kinetic characterization of the 1A subfamily of recombinant human UDP-glucuronosyltransferases

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ABSTRACT

The initial glucuronidation rates were determined for eight recombinant human UDP-glucuronosyltransferases (UGTs) of the 1A subfamily, and the bisubstrate kinetics and inhibition patterns were analyzed. At low substrate concentrations the reactions followed general ternary complex kinetics, whereas at higher concentrations of both substrates the reactions were mostly characterized by ternary complex kinetics with substrate inhibition. The glucuronidation of entacapone by UGT1A9 was inhibited by 1-naphthol in a competitive fashion with respect to entacapone and uncompetitive fashion with respect to UDP-glucuronic acid (UDPGA). Its inhibition by UDP, on the other hand, was noncompetitive with respect to entacapone and competitive with respect to UDPGA. These inhibition patterns are compatible with a compulsory ordered bi bi mechanism in which UDPGA is the first-binding substrate. Despite the identical primary structure of the C-terminal halves of the UGT1A isoforms, there were marked differences in the respective K_m values for UDPGA, ranging from 52 μM for UGT1A6 to 1256 μM for UGT1A8. Relative specificity constants were calculated for the eight UGT1A isoforms with 1-hydroxypyrene, 4-nitrophenol, scopoletin, 4-methylumbelliferone, and entacapone as aglycone substrates. The results demonstrated that seven of the UGT1A isoforms are capable of conjugating phenolic substrates with similar highest k_{cat} values, and UGT1A4 has lower relative turnover rate. The highest specificity constants were obtained for 1-hydroxypyrene, even with UGT1A6, which has been regarded as a specific isoform for small planar phenols. A k_{cat} value of 1.9 s^{-1} was calculated for the glucuronidation of scopoletin by purified UGT1A9.

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UDP-glucuronosyltransferases (UGTs, EC 2.4.1.17) catalyze the transfer of glucuronic acid from the co-substrate uridine 5'-diphospho- α -D-glucuronic acid (UDPGA) to various aglycone compounds, such as steroid hormones, bile acids, and bilirubin, as well as to a large number of xenobiotics including drugs and drug metabolites.

Expressed in the liver and several extrahepatic tissues (Tukey and Strassburg, 2000), human UGTs are integral membrane proteins that reside within the endoplasmic reticulum, with their catalytic sites, as well as most of their mass, on the luminal side of the membrane. They have a single trans-membrane segment close to their C-terminus and their last 19-26 residues are located on the cytoplasmic side of the endoplasmic reticulum membrane. UGTs appear to be built of two large domains of almost equal size, namely the N- and C-terminal domains. At least three subfamilies, UGT1A, UGT2A, and UGT2B are distinguished on the basis of sequence homology and gene structure (Mackenzie et al., 1997). The 1A subfamily comprises nine functional isoforms. The N-terminal domains of UGT1A isoforms are encoded by different first exons and their primary structures are more variable. The C-terminal domains are encoded by the same exons 2-5, so that the primary structure of the C-terminal domains, roughly corresponding to the C-terminal halves are identical in all UGT1A isoforms (Gong et al., 2001).

This very high degree of homology of the C-terminal halves of all the UGTs, as well as interactions of periodate-oxidized UDPGA with UGT1A6 (Battaglia et al., 1998), strongly suggest that the C-terminal domain is directly involved in UDPGA binding, whereas the N-terminal halves contain the aglycone binding site(s) (Mackenzie, 1990; Senay et al., 1999; Gong et al., 2001). It has been suggested, however, that the N-terminal part of the mature protein may also participate in UDPGA binding (Pillot et al., 1993). In addition, recent findings suggest that protein-protein interactions between the N-terminal and C-terminal halves of the UGTs may have functional importance, perhaps providing a mechanism for one of these two major domains to affect substrate or co-substrate binding in the other (Kurkela et al., 2004).

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The most common glucuronidation sites are nucleophilic O or N atoms of the sugar acceptor substrate. Glucuronidation occurs as an S_N2 substitution by the attack of a nucleophilic heteroatom of the aglycone on the C_1 atom of glucuronic acid, while both UDPGA and the aglycone substrate are bound on the active site of the enzyme. The S_N2 mechanism is supported by inversion of the α -configuration of the C_1 atom to β -configuration in the glucuronide (Johnson and Fenselau, 1978), and by the substituent effects on the glucuronidation rate (Yin et al., 1994). The 3D structure of the active site is unknown. A unanimous conclusion of all previous mechanistic studies is that the reaction involves the formation of a ternary complex (Potrepka and Spratt, 1972; Vessey and Zakim, 1972; Sanchez and Tephly, 1975; Rao et al., 1976; Koster and Noordhoek, 1983; Falany et al., 1987; Matern et al., 1991; Yin et al., 1994). Product and dead-end inhibition studies conducted to determine the order of substrate binding have given ambiguous results, however. Two reports based on partially purified rat UGTs (Falany et al., 1987; Yin et al., 1994), and one report based on beef liver microsomes (Vessey and Zakim, 1972) have proposed a random ordered bi bi mechanism, while other studies with animal tissue fractions have proposed a compulsory ordered bi bi (Potrepka and Spratt, 1972; Sanchez and Tephly, 1975; Koster and Noordhoek, 1983) or a Theorell-Chance mechanism (Rao et al., 1976; Koster and Noordhoek, 1983).

The availability of recombinant UGTs has aided in understanding the contribution of glucuronidation to xenobiotic metabolism. Substrate specificities of human UGTs have been characterized by determining specific activities and kinetic parameters for a large number of substrates. Nevertheless, the role and significance of individual UGT isoforms are still poorly understood, and the bisubstrate kinetics of UGTs has not previously been studied with individual isoenzymes. Meaningful substrate specificity assessment can only be made after the catalytic constants (k_{cat}), and especially the specificity constants (k_{cat}/K_m), have been determined, or at least normalized values have been assigned for individual isoenzymes. The other parameters of the

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bisubstrate kinetics should also be determined. Detailed study of the kinetic mechanism of UGTs could also assist the design of potent and specific inhibitors.

To better understand the kinetics of glucuronidation, we studied the bisubstrate kinetics of eight human recombinant UGTs of the 1A subfamily. The kinetic mechanism and substrate inhibition were characterized and the substrate specificities were evaluated on the basis of relative specificity constants.

Materials and methods

Materials. Insect cell membranes containing one of the following human UGTs—1A1, 1A3, 1A4 and 1A6-1A10—together with purified UG1A9, were prepared in our laboratory as previously described (Kurkela et al., 2003; Kuuranne et al., 2003). Phosphatidylcholine X-E from dried egg yolk, ethinylestradiol, scopoletin, 4-aminobiphenyl, 1-naphthol, 4-hydroxyestrone, 4-nitrophenol, umbelliferone, uridine 5'-diphosphate sodium salt (UDP), D-saccharic acid 1,4-lactone (saccharolactone), and uridine 5'-diphosphoglucuronic acid trisodium salt (UDPGA) were purchased from Sigma-Aldrich (St Louis, MO), and 1-hydroxypyrene was from Acros Organics (New Jersey, USA). Entacapone was kindly provided by Orion Pharma (Espoo, Finland), and entacapone glucuronide (Luukkanen et al., 1999), 1-hydroxypyrene glucuronide (Luukkanen et al., 2001), and scopoletin glucuronide were synthesized in our laboratory. α -Naphthyl- β -D-glucuronide, 4-nitrophenyl- β -D-glucuronide, and umbelliferyl- β -D-glucuronide were purchased from Sigma-Aldrich (St Louis, MO). Radiolabelled [14 C]UDPGA was from PerkinElmer Life Sciences (Boston, MA).

Activity assays. All enzyme assays (total volume 250 μ l) contained 50 mM phosphate buffer pH 7.4, 5 mM MgCl₂, 5 mM saccharolactone, 25-5000 μ M UDPGA, and 10-50 μ g membrane protein

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of the appropriate recombinant UGT or 50 ng of purified UGT1A9. Phosphatidylcholine type X-E (1 mg/ml) was added to the assay mixtures containing the purified UGT1A9. The aglycone substrate concentrations ranged from ca. 0.2 to 10 times the respective K_m values (determined in preliminary assays) unless method sensitivity or substrate solubility demanded otherwise. The aglycone substrates were added as DMSO solutions to a final DMSO concentration of 2% in all the enzyme assays. All samples were prepared in duplicate.

The reactions were performed at 37°C and the reaction time varied, within the linear range, from 10 to 60 min. The reactions were terminated by the addition of 25 μ l of 4 M ice-cold perchloric acid or 250 μ l of ice-cold methanol (4-aminobiphenyl). The precipitated proteins were removed by centrifugation (5 min at 14 000 rpm). Blank incubations were prepared in a similar way at the highest aglycone substrate concentrations without UDPGA.

Aliquots of the supernatants were analyzed using an Agilent model 1100 or a Hewlett Packard model 1090 HPLC with UV or fluorescence detection. Details of the analytical conditions are given in Table 1. The quantitation of entacapone glucuronide, 1-naphthylglucuronide, 1-hydroxypyrene glucuronide, scopoletin glucuronide, and 4-nitrophenylglucuronide was based on authentic reference standards. When pure glucuronide was not available the quantitation of the glucuronidated products was achieved with the HPLC connected to a flow scintillation analyzer (150TR, Packard, Meriden, CT) that was fitted with a 500- μ l flow cell into which 3 ml/min of scintillation liquid (Monoflow 3, National Diagnostics, Atlanta, GA) was pumped. The radiochemical detection was coupled to UV or fluorescence detection as described previously (Kaivosari et al., 2001).

Product inhibition studies with UDP (UGT1A9) were conducted at sub-saturating (100 μ M) concentrations of the fixed substrate. Inhibition studies with a competing substrate, 1-naphthol, were conducted at 100 μ M entacapone when UDPGA was the varied substrate, and at 1000 μ M UDPGA when entacapone was the varied substrate. UDP was added to the reaction mixture as an

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aqueous solution to a final concentration of 0, 25, 50, 100, 250, or 500 μM , and 1-naphthol in DMSO was added to a final concentration of 0, 0.5, 2.5, or 10 μM .

Protein concentrations were measured using the bicinchoninic acid (BCA) protein assay kit (Pierce, Rockford, IL) with bovine serum albumin as standard, and the relative enzyme concentrations in the membranes were estimated by dot-blot analyses as described recently (Kurkela et al., 2004).

Kinetic analysis. Kinetic parameters were estimated by fitting the initial rate data to appropriate bisubstrate and inhibition rate equations (Cornish-Bowden, 1995; Copeland, 2000) by nonlinear least-squares regression using a weighting factor $1/y$. Data analysis was performed by SigmaPlot Enzyme kinetics module 1.1S and the goodness of fit was evaluated on the basis of standard deviations of the parameter estimates, R^2 values, and residual plots. The individual rate constants for the glucuronidation of scopoletin by purified UGT1A9 were calculated according to Cornish-Bowden (Cornish-Bowden, 1995).

Results

Kinetic mechanism. The initial rates for the reactions catalyzed by the recombinant human UGTs 1A1, 1A3, 1A4, 1A6, 1A7, 1A8, 1A9, and 1A10 were determined by varying the concentrations of both substrates: the sugar donor UDP-glucuronic acid (AX) and the sugar acceptor (B). The measured initial rates were in good agreement with a ternary complex mechanism, for which the reaction velocity can be given by equation 1 or equation 2 (Copeland, 2000), where K_{AX} is the dissociation constant for the enzyme–AX complex, K_B is the dissociation constant for the enzyme–B complex, and K_{mAX} and K_{mB} are the Michaelis constants for AX and B, respectively.

$$v = \frac{V_{\max} [AX][B]}{(\alpha K_{AX} K_B + \alpha K_{AX} [B] + \alpha K_B [AX] + [AX][B])} \quad (\text{Eq. 1})$$

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$$v = \frac{V_{\max} [AX][B]}{(K_{AX} K_{mB} + K_{mAX} [B] + K_{mB} [AX] + [AX][B])} \quad (\text{Eq. 2})$$

Equation 1 is derived for random ordered bi bi mechanism (Fig. 2A) assuming rapid equilibrium and equation 2 is derived for compulsory ordered bi bi mechanism (Fig. 2B) using steady state assumptions.

The bisubstrate initial rate data for all UGT1A enzymes gave a good statistical fit to both equations 1 and 2 up to aglycone substrate concentrations at least 4-5 times the respective K_m values (Table 2) and up to UDPGA concentrations of 1-5 mM. Fit of the kinetic data to equation 2 is shown in Fig. 1. At higher aglycone concentrations, substrate inhibition was observed for all UGT1A isoforms except UGT1A4. At very high concentrations of both substrates (above 2 mM), the measured velocity data showed irregular behavior and poor reproducibility. In the case of UGT1A3, the high K_m for scopoletin limited the rate determination to a concentration of aglycone of only twice the K_m .

Substrate inhibition is expected for a ternary complex mechanism following compulsory ordered kinetics. As depicted in Fig. 2B, binding of the aglycone substrate to the enzyme–UDP complex leads to a non productive dead-end complex that slows the completion of the catalytic cycle. In such a case the reaction velocity is given by equation 3:

$$v = \frac{V_{\max} \cdot [AX] \cdot [B]}{\left(K_{AX} \cdot K_{mB} + K_{mAX} \cdot [B] + K_{mB} \cdot [AX] + [AX] \cdot [B] \cdot \left(1 + \frac{[B]}{K_{siB}} \right) \right)} \quad (\text{Eq. 3})$$

where K_{siB} is a constant that describes the strength of substrate inhibition.

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The bisubstrate kinetic data for 1A1, 1A8, 1A9 and 1A10, including conditions of high aglycone substrate concentration, fitted well to equation 3 (Fig. 3). In the case of UGTs 1A3, 1A6, and 1A7, the substrate inhibition data was measured only at one UDPGA concentration and the data fitted well to the corresponding reduced equation (Eq. 4).

$$v = \frac{V_{\max}}{\left(1 + \frac{K_m}{S} + \frac{S}{K_{siB}}\right)} \quad (\text{Eq. 4})$$

Although formation of the same non-productive complex is also possible in a mechanism that follows random ordered ternary complex kinetics (Fig. 2A), the formation should not occur, and substrate inhibition should not be seen if the mechanism involves rapid equilibrium, because then the concentration of the enzyme-UDP or enzyme-glucuronide complex is zero in the absence of added products. Observation of substrate inhibition may thus be used to exclude rapid equilibrium random ordered bi bi mechanism.

The results of the bisubstrate kinetic experiments suggest that glucuronidation catalyzed by UGTs 1A1, 1A3, and 1A6-1A10 does not involve rapid equilibrium, and the reaction most probably follows the scheme presented in Fig. 2B. Nevertheless, a non-rapid-equilibrium random ordered reaction according to Fig. 2A cannot be excluded on the basis of the bisubstrate kinetic data alone.

Two other possible mechanisms, rapid equilibrium compulsory ordered mechanism (Theorell-Chance) and the double displacement mechanism (Ping-Pong) (Copeland, 2000) can be excluded on the basis of the poor fit of the velocity data to the corresponding equations.

An inhibitor that binds to the same site as one of the substrates would kinetically distinguish between the two ternary complex mechanisms. In the case of compulsory ordered mechanism an inhibitor that resembles the second-binding substrate would be expected to bind to the enzyme-first-binding-substrate complex rather than to the free enzyme. Hence, a competitive inhibitor with

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respect to the second-binding substrate would behave as an uncompetitive inhibitor with respect to the first-binding substrate. In the case of a random ordered mechanism, on the other hand, no uncompetitive inhibition pattern should be observed. Carrying out of such an inhibition experiment has been hampered by the lack of effective UGT inhibitors that bind to the aglycone-binding site. However, we found that 1-naphthol, although it is a substrate for UGT1A9, shows significantly higher affinity and lower reactivity ($K_m = 0.09 \mu\text{M}$, $V_{max} = 27.8 \text{ pmol/ml/min}$) than entacapone (Table 2) and, hence, could be used as a competitive inhibitor with respect to entacapone in experiments with UGT1A9.

The effects of 1-naphthol on the kinetics of entacapone glucuronidation by UGT1A9 are shown in Fig. 4. As can be seen, 1-naphthol acts as an uncompetitive inhibitor when UDPGA is the varied substrate (Fig 4A), and as a competitive inhibitor when entacapone is the varied substrate (Fig. 4B). Fitting the data to the general equation of reversible inhibition yielded dissociation constants of $0.191 \mu\text{M}$ and $1.46 \mu\text{M}$ for the binding of 1-naphthol to the enzyme–UDPGA complex and to the free enzyme, respectively. These results indicate that 1-naphthol also binds to the free enzyme, but with significantly lower affinity than to the enzyme–UDPGA complex. It is not unexpected to see some inhibitory binding of a hydrophobic substrate to the free enzyme, even though the catalytic binding would require compulsory order.

We also studied UDP inhibition of the entacapone glucuronidation reaction by UGT1A9. The results are presented in Fig. 5. The inhibition by UDP displayed noncompetitive and competitive patterns with respect to entacapone (Fig. 5A) and UDPGA (Fig. 5B), respectively. Such inhibition patterns are possible in both random and compulsory ordered mechanisms. In the case of compulsory ordered reaction, the inhibitory behavior of UDP suggests that UDPGA is the first-binding substrate. The dissociation constant K_i for enzyme–UDP complex was found to be less than one-tenth the K_i value for the enzyme–UDPGA complex (Fig. 5).

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Kinetic parameters. The enzyme kinetic parameters for the glucuronidation of appropriate aglycone substrates by the eight UGT1A isoforms were determined using Equation 2. The results are listed in Table 2. The initial velocities obtained at high substrate concentrations with obvious substrate inhibition were omitted from this kinetic analysis. Considerable variation was observed in UDPGA parameter values for the different UGT1A isoforms. The lowest K_m value, 52 μM , was determined for the glucuronidation of 1-naphthol by UGT1A6, and the highest, 1256 μM , for the glucuronidation of 4-hydroxyestrone by UGT1A8. The K_{AX} values, the dissociation constants of the enzyme–UDPGA complex, in these cases were 163 μM and 1264 μM , respectively. In a compulsory ordered mechanism, the K_{AX} value should be independent of the aglycone substrate, if UDPGA is the first-binding substrate. To test this, the kinetic parameters for UGT1A6 and UGT1A8 were also determined for the glucuronidation of scopoletin (166 μM) and entacapone (875 μM), respectively, and the results are included in Table 2. The K_{AX} values for the latter substrates are similar to those obtained with 1-naphthol (1A6) and 4-hydroxyestrone (1A8), as expected according to the proposed mechanism.

The bisubstrate initial rate data showing obvious substrate inhibition were fitted to Equation 3 and, when only the aglycone substrate was varied, to the corresponding reduced equation. The kinetic parameters K_m , V_{max} , K_{AX} , and K_{siB} are presented in Table 3. The K_m , V_{max} , and K_{AX} values are mostly in reasonable agreement with the values given in Table 2. The K_{siB} values varied from 178 μM for the glucuronidation of ethinylestradiol by UGT1A1 to 2800 μM for the glucuronidation of entacapone by UGT1A10. In all these cases the K_{siB} values were significantly higher than the corresponding K_m values.

Relative k_{cat} and relative specificity constants. The relative specificity constants allow a comparison of the catalytic efficiencies of different enzymes or of the utilization of different substrates by the same enzyme. The relative k_{cat} and relative k_{cat}/K_m values for 1-hydroxypyrene, 4-nitrophenol, scopoletin, 4-methylumbelliferone, and entacapone with eight UGT1A isoforms are

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presented in Table 4. Relative k_{cat} values, and relative specificity constants, k_{cat}/K_m , were calculated on the basis of the relative expression levels of the individual UGT1A isoforms. The k_{cat} and k_{cat}/K_m values for the glucuronidation of entacapone by UGT1A9 were arbitrarily assigned as 1.0, and the values for other UGTs were compared with those for UGT1A9. In general, all UGT1A isoforms were capable of conjugating phenolic substrates, and the highest relative k_{cat} values for all isoforms, with the exception of 1A4, were closely similar. UGT1A4 also conjugated phenolic substrates but at lower efficiency. Nevertheless, examination of the relative k_{cat}/K_m values revealed that, regardless of the relative k_{cat} values *per se*, all eight UGTs showed the highest relative k_{cat}/K_m values with 1-hydroxypyrene, a large polycyclic aromatic planar phenol.

Catalytic constant k_{cat} . A k_{cat} value of 1.9 s^{-1} was estimated from kinetic experiments with purified UGT1A9 (Fig. 6). A consistent means of determining k_{cat} requires knowledge of the absolute enzyme concentration and, hence, the total UGT1A9 concentration was calculated by means of the the total amount of protein in the purified enzyme. The apparent homogeneity of the purified UGT1A9 was demonstrated by SDS-Page and Western blotting (Kurkela et al., 2003).

In the case of a compulsory ordered bi bi mechanism, a unique relationship exists between the kinetic parameters and the rate constants (Fig. 7). Hence, the individual rate constants k_1 and k_{-1} could be calculated from the initial rate measurements (Cornish-Bowden, 1995).

The rate constants for the glucuronidation of scopoletin by purified UGT1A9 are presented in Fig. 6. The first-order rate constant for the dissociation of the enzyme–UDPGA complex, $k_{-1}=2.1 \text{ s}^{-1}$, equals the rate constant for the product formation, k_{cat} , in accordance with the steady-state assumption.

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Discussion

The bisubstrate kinetics of eight recombinant human UGTs was analyzed. The glucuronidation reaction was well described by the rate equations derived for mechanisms involving the formation of a ternary complex. Nevertheless, the bisubstrate kinetics alone did not allow us to determine whether there was a compulsory order of binding of substrates. An unambiguous differentiation of the random and compulsory ordered mechanisms was established by the observed essentially uncompetitive inhibition by 1-naphthol with respect to UDPGA (Fig. 4A). This result indicates that 1-naphthol preferably binds to the enzyme–UDPGA complex, that is, the reaction follows a compulsory ordered kinetic mechanism in which UDPGA is the first-binding substrate. The observed competitive inhibition by UDP with respect to UDPGA (Fig. 5B) and noncompetitive inhibition with respect to entacapone (Fig. 5A) provided further evidence for the compulsory ordered mechanism with UDPGA as the first-binding substrate.

Previous studies on the enzyme kinetic mechanism have provided different and sometimes conflicting results (Potrepka and Spratt, 1972; Vessey and Zakim, 1972; Sanchez and Tephly, 1975; Rao et al., 1976; Koster and Noordhoek, 1983; Falany et al., 1987; Yin et al., 1994). The ambiguity of the results from product and dead-end inhibition experiments in earlier work may well be explained by the presence of several different UGT isoforms in the tissue samples, a fact that was not fully appreciated in the past. The presence of several isoforms would severely complicate the interpretation of inhibition kinetic data, especially when analyzed by means of linearized plots. In addition, those studies that relied on partially purified rat enzymes probably suffered from the presence of UGTs that were partially detergent-inactivated during the membrane solubilization and the purification process (Kurkela et al., 2003).

At high concentrations of both substrates, substrate inhibition was frequently observed with all UGT1A isoforms except 1A4. The only human UGT of the 1A subfamily for which we have not

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detected substrate inhibition was 1A4. This enzyme preferably catalyzes the N-glucuronidation of aglycones containing amino groups, and only a few phenolic compounds are glucuronidated by UGT1A4 efficiently. An attempt was made to determine the enzyme kinetic parameters for the glucuronidation of 4-aminobiphenyl, which is a widely accepted model substrate for UGT1A4. Although it was a good substrate for UGT1A4 when considering V_{\max} , the bisubstrate kinetics for 4-aminobiphenyl could not be determined reliably due to the combination of low solubility and poor method sensitivity. A hydroxylated PAH compound, 1-hydroxypyrene, exhibited a considerably lower K_m but showed no substrate inhibition in the concentration range that was studied. The apparent substrate inhibition was used to exclude the rapid equilibrium random ordered bi bi mechanism. In the case of a compulsory ordered bi bi mechanism, substrate inhibition allows the identification of the second-binding substrate. The occurrence of substrate inhibition for UGTs 1A1, 1A3, and 1A6-1A10 suggests, therefore, that glucuronidation catalyzed by these isoforms does not involve rapid equilibrium, and that the reaction probably follows the scheme presented in Fig. 2B. The K_{siB} values (Table 3.) were significantly higher than the corresponding K_m values for all UGT1A isoforms, however, which means that of substrate inhibition is probably of negligible significance in physiological environment. As reported previously (Houston and Kenworthy, 2000; Uchaipichat et al., 2004), substrate inhibition may also arise from the binding of a second substrate molecule to the enzyme–substrate complex. Nevertheless, the kinetic data fitted well to equations 3 and 4, and incorporation in the kinetic equation of a parameter describing the binding of a second substrate molecule resulted in poorer statistics of the fits.

Kinetic constants derived from bisubstrate reactions have not been reported previously for individually expressed UGT isoforms. The K_m values for UDPGA have been determined at saturating aglycone substrate concentrations with recombinant human UGTs 1A1 (400-1810 μM) (Senafi et al., 1994; Court et al., 2001), 1A3 (250 μM), 1A4 (300 μM) (Green et al., 1995), 1A6 (110-200 μM) (Battaglia et al., 1998; Ouzzine et al., 2000; Court et al., 2001), and 1A9 (190-500

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μM) (Senafi et al., 1994; Court et al., 2001). The K_m values obtained in this work (Table 2.) are within the same range as most of those results. The physiological concentration of UDPGA in human liver is ca. $300 \mu\text{M}$ ($301 \mu\text{mol/kg}$ wet weight) and at least one magnitude lower than this in human kidney (Cappiello et al., 2000). The physiological concentrations of UDPGA only rarely exceed the K_m values obtained in this study and speculations that a decrease in the availability of UDPGA would impair the capacity for hepatic glucuronide conjugation (Howell et al., 1986; Evdokimova et al., 2001) may be justified. A comparison of the aglycone K_m values in Table 2 with the few published results for the same combinations of UGT isoform and substrate shows our values to be mostly within the same order of magnitude as the earlier ones, for example, those reported for UGT1A1 with ethinylestradiol (Williams et al., 2002; Soars et al., 2003), UGT1A3 with scopoletin (Green et al., 1998), and UGT1A6 with 1-naphthol (Uchaipichat et al., 2004), as well as those reported for eight UGT1A isoforms with 4-methylumbelliferone (Uchaipichat et al., 2004).

The current study also has implications for the structure-function relationships of the UGTs. A close inspection of the K_{AX} values in Table 2 reveals distribution to low and high values. UGTs 1A4, 1A6, 1A7, and 1A9 exhibited low K_{AX} values ($163\text{-}316 \mu\text{M}$), while UGTs 1A1, 1A3, 1A8, and 1A10 exhibited higher K_{AX} values ($693\text{-}1264 \mu\text{M}$) for UDPGA. A similar distribution of UGTs appears in the K_m values, with the exception of UGT1A1, which had low K_m and higher K_{AX} (Table 2). These differences in K_{AX} values suggest that the 3-D structure of the UDPGA binding site is not fully conserved, not even within the UGT1A subfamily. Furthermore, UGTs 1A7-1A10 are highly homologous throughout the entire protein, and this is reflected in similar K_m values towards entacapone. Nevertheless, UGT1A9 has very low K_{AX} and low K_m for UDPGA, while UGT1A8 exhibits the highest K_{AX} and K_m values in this study (Table 2.). These results suggest that, in addition to the C-terminal half of the UGTs, the UDPGA binding is affected by the amino acid residues of the N-terminal half of the protein.

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The relative k_{cat} values (Table 4) demonstrate that all the UGT1A isoforms are capable of conjugating phenolic substrates, and all isoforms with the exception of UGT1A4 exhibit similar k_{cat} values for their best phenolic substrates. All the UGT1A isoforms were efficient catalysts of 1-OH-pyrene and scopoletin glucuronidation, but UGT1A6 was the only isoform that could conjugate 4-nitrophenol with high efficiency. Entacapone, with its large non-planar side chain, was a poor substrate for UGTs 1A1-1A6 and a good substrate for UGTs 1A7-1A10. The highest relative specificity constants with all isoforms were observed for the glucuronidation of 1-OH-pyrene, a large polycyclic aromatic planar phenol. These results suggest, contrary to the general assumption, that UGT1A6 is not restricted to the glucuronidation of small planar phenols.

The individual rate constants for the binding of UDPGA to the enzyme, the dissociation of the enzyme-UDPGA complex, and the product formation for the glucuronidation of scopoletin by purified UGT1A9 were calculated from the enzyme kinetic parameters (Fig. 6A) assuming the enzyme follows compulsory ordered mechanism. The purified UGT1A9 presented a lower affinity for scopoletin and UDPGA than the native protein, as evidenced by significant increases in the K_m and K_{AX} values. Nevertheless, the specific activity of the purified enzyme was ca. 1000-fold higher than the value of the membrane-bound UGT1A9.

When $[S] \ll K_m$, the upper limit for the reaction rate is defined by the diffusional rate of encounter of the free enzyme with substrate, which is characterized by k_1 (typically $10^8 - 10^9 \text{ M}^{-1}\text{s}^{-1}$). When $[S] \gg K_m$, in turn, the formation of [ES] complex is rapid and often not rate-limiting. Here, the formation of the enzyme-UDPGA complex was rapid ($3026 \text{ M}^{-1}\text{s}^{-1}$) compared to the formation of scopoletin glucuronide (1.9 s^{-1}) (Fig. 6). The k_{cat} value (1.9 s^{-1}) for the formation of scopoletin glucuronide was 1-2 orders of magnitude higher than previously reported for the N-glucuronidation of N-hydroxy-PhIP by UGT1A9 (Malfatti and Felton, 2004). However, UGT1A1 catalyzed the glucuronidation of N-hydroxy-PhIP at the highest rate, $k_{\text{cat}}=1.9 \text{ s}^{-1}$, which is in line with our

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observation that UGTs 1A1, 1A3, and 1A6-1A10 show similar k_{cat} values for their best phenolic substrates.

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References

- Battaglia E, Terrier N, Mizeracka M, Senay C, Magdalou J, Fournel-Gigleux S and Radominska-Pandya A (1998) Interaction of periodate-oxidized UDP-glucuronic acid with recombinant human liver UDP-glucuronosyltransferase 1A6. *Drug Metab Dispos* **26**:812-817.
- Cappiello M, Giuliani L, Rane A and Pacifici GM (2000) Uridine 5'-diphosphoglucuronic acid (UDPGlcUA) in the human fetal liver, kidney and placenta. *Eur J Drug Metab Pharmacokin* **25**:161-163.
- Copeland RA (2000) *Enzymes. A Practical introduction to structure, mechanism and data analysis*. Wiley-VCH, Inc., New York.
- Cornish-Bowden A (1995) *Fundamentals of enzyme kinetics*. Portland Press, London.
- Court MH, Duan SX, von Moltke LL, Greenblatt DJ, Patten CJ, Miners JO and Mackenzie PI (2001) Interindividual variability in acetaminophen glucuronidation by human liver microsomes: identification of relevant acetaminophen UDP-glucuronosyltransferase isoforms. *J Pharmacol Exp Ther* **299**:998-1006.
- Evdokimova E, Taper H and Buc Calderon P (2001) Role of ATP and glycogen reserves in both paracetamol sulfation and glucuronidation by cultured precision-cut rat liver slices. *Toxicol in Vitro* **15**:683-690.
- Falany CN, Green MD and Tephly TR (1987) The enzymatic mechanism of glucuronidation catalyzed by two purified rat liver steroid UDP-glucuronosyltransferases. *J Biol Chem* **262**:1218-1222.
- Gong QH, Cho JW, Huang T, Potter C, Gholami N, Basu NK, Kubota S, Carvalho S, Pennington MW, Owens IS and Popescu NC (2001) Thirteen UDPglucuronosyltransferase genes are encoded at the human UGT1 gene complex locus. *Pharmacogenetics* **11**:357-368.
- Green MD, Bishop WP and Tephly TR (1995) Expressed human UGT1.4 protein catalyzes the formation of quaternary ammonium-linked glucuronides. *Drug Metab Dispos* **23**:299-302.
- Green MD, King CD, Mojarrabi B, Mackenzie PI and Tephly TR (1998) Glucuronidation of amines and other xenobiotics catalyzed by expressed human UDP-glucuronosyltransferase 1A3. *Drug Metab Dispos* **26**:507-512.
- Houston JB and Kenworthy KE (2000) In vitro-In vivo scaling of CYP kinetic data not consistent with the classical michaelis-menten model. *Drug Metab Dispos* **28**:246-254.
- Howell SR, Hazelton GA and Klaassen CD (1986) Depletion of hepatic UDP-glucuronic acid by drugs that are glucuronidated. *J Pharmacol Exp Ther* **236**:610-614.
- Johnson LP and Fenselau C (1978) Enzymatic conjugation and hydrolysis of [18O]isoborneol glucuronide. *Drug Metab Dispos* **6**:677-679.
- Kaivosaaari S, Salonen JS, Mortensen J and Taskinen J (2001) High-performance liquid chromatographic method combining radiochemical and ultraviolet detection for determination of low activities of uridine 5'-diphosphate-glucuronosyltransferase. *Anal Biochem* **292**:178-187.
- Koster AS and Noordhoek J (1983) Kinetic properties of the rat intestinal microsomal 1-naphthol:UDP-glucuronosyl transferase. Inhibition by UDP and UDP-N-acetylglucosamine. *Biochim Biophys Acta* **761**:76-85.
- Kurkela M, Garcia-Horsman JA, Luukkanen L, Mörsky S, Taskinen J, Baumann M, Kostainen R, Hirvonen J and Finel M (2003) Expression and characterization of recombinant human UDP glucuronosyltransferases (UGTs); UGT1A9 is more resistant to detergent inhibition than the other UGTs, and was purified as an active dimeric enzyme. *J Biol Chem* **278**:3536-3544.
- Kurkela M, Hirvonen J, Kostainen R and Finel M (2004) The interactions between the N-terminal and C-terminal domains of the human UDP-glucuronosyltransferases are partly isoform-specific, and may involve both monomers. *Biochem Pharmacol* **68**:2443-2450.

“DMD 4093”

- Kuuranne T, Kurkela M, Thevis M, Schänzer W, Finel M and Kostianen R (2003) Glucuronidation Of Anabolic Androgenic Steroids By Recombinant Human UDP-Glucuronosyltransferases. *Drug Metab Dispos* **31**:1117-1124.
- Luukkanen L, Kilpelainen I, Kangas H, Ottoila P, Elovaara E and Taskinen J (1999) Enzyme-assisted synthesis and structural characterization of nitrocatechol glucuronides. *Bioconj Chem* **10**:150-154.
- Luukkanen L, Mikkola J, Forsman T, Taavitsainen P, Taskinen J and Elovaara E (2001) Glucuronidation of 1-hydroxypyrene by human liver microsomes and human UDP-glucuronosyltransferases UGT1A6, UGT1A7, and UGT1A9: development of a high-sensitivity glucuronidation assay for human tissue. *Drug Metab Dispos* **29**:1096-1101.
- Mackenzie P (1990) Expression of chimeric cDNAs in cell culture defines a region on UDP-glucuronosyltransferase involved in substrate selection. *J Biol Chem* **265**:3432-3435.
- Mackenzie PI, Owens IS, Burchell B, Bock KW, Bairoch A, Belanger A, Fournel-Gigleux S, Green M, Hum DW, Iyanagi T, Lancet D, Louisot P, Magdalou J, Chowdhury JR, Ritter JK, Schachter H, Tephly TR, Tipton KF and Nebert DW (1997) The UDP glycosyltransferase gene superfamily: recommended nomenclature update based on evolutionary divergence. *Pharmacogenetics* **7**:255-269.
- Malfatti MA and Felton JS (2004) Human UDP-Glucuronosyltransferase 1A1 Is the Primary Enzyme Responsible for the N-Glucuronidation of N-Hydroxy-PhIP in Vitro. *Chem Res Toxicol* **17**:1137-1144.
- Matern H, Lappas N and Matern S (1991) Isolation and characterization of hyodeoxycholic acid UDP-glucuronosyltransferase from human liver. *Eur J Biochem* **200**:393-400.
- Ouzzine M, Antonio L, Burchell B, Netter P, Fournel-Gigleux S and Magdalou J (2000) Importance of histidine residues for the function of the human liver UDP-glucuronosyltransferase UGT1A6: evidence for the catalytic role of histidine 370. *Mol Pharmacol* **58**:1609-1615.
- Pillot T, Ouzzine M, Fournel-Gigleux S, Lafaurie C, Tebbi D, Treat S, Radomska A, Lester R, Siest G and Magdalou J (1993) Determination of the human liver UDP-glucuronosyltransferase 2B4 domains involved in the binding of UDP-glucuronic acid using photoaffinity labeling of fusion proteins. *Biochem. Biophys. Res. Commun.* **197**:785-791.
- Potrepka RF and Spratt JL (1972) A study on the enzymatic mechanism of guinea-pig hepatic-microsomal bilirubin glucuronyl transferase. *Eur J Biochem* **29**:433-439.
- Rao ML, Rao GS and Breuer H (1976) Investigations on the kinetic properties of estrone glucuronyltransferase from pig kidney. *Biochim Biophys Acta* **452**:89-100.
- Sanchez E and Tephly TR (1975) Morphine metabolism. IV. Studies on the mechanism of morphine: uridine diphosphoglucuronyltransferase and its activation by bilirubin. *Mol Pharmacol* **11**:613-620.
- Senafi SB, Clarke DJ and Burchell B (1994) Investigation of the substrate specificity of a cloned expressed human bilirubin UDP-glucuronosyltransferase: UDP-sugar specificity and involvement in steroid and xenobiotic glucuronidation. *Biochem J* **303**:233-240.
- Senay C, Battaglia E, Chen G, Breton R, Fournel-Gigleux S, Magdalou J and Radomska-Pandya A (1999) Photoaffinity labeling of the aglycon binding site of the recombinant human liver UDP-glucuronosyltransferase UGT1A6 with 7-azido-4- methylcoumarin. *Arch Biochem Biophys* **368**:75-84.
- Soars MG, Ring BJ and Wrighton SA (2003) The effect of incubation conditions on the enzyme kinetics of UDP-glucuronosyltransferases. *Drug Metab Dispos* **31**:762-767.
- Tukey RH and Strassburg CP (2000) Human UDP-glucuronosyltransferases: metabolism, expression, and disease. *Annu Rev Pharmacol Toxicol* **40**:581-616.
- Uchaipichat V, Mackenzie PI, Guo X-H, Gardner-Stephen D, Galetin A, Houston JB and Miners JO (2004) Human UDP-glucuronosyltransferases: Isoform selectivity and kinetics of 4-

“DMD 4093”

- methylumbelliferone and 1-naphthol glucuronidation, effects of organic solvents, and inhibition by diclofenac and probenecid. *Drug Metab Dispos* **32**:413-423.
- Vessey DA and Zakim D (1972) Regulation of microsomal enzymes by phospholipids. V. Kinetic studies of hepatic uridine diphosphate-glucuronyltransferase. *J Biol Chem* **247**:3023-3028.
- Williams JA, Ring BJ, Cantrell VE, Campanale K, Jones DR, Hall SD and Wrighton SA (2002) Differential modulation of UDP-glucuronosyltransferase 1A1 (UGT1A1) catalyzed estradiol-3-glucuronidation by the addition of UGT1A1 substrates and other compounds to human liver microsomes. *Drug Metab Dispos* **30**:1266-1273.
- Yin H, Bennett G and Jones JP (1994) Mechanistic studies of uridine diphosphate glucuronosyltransferase. *Chem Biol Interact* **90**:47-58.

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Legends for figures

Fig. 1. The bisubstrate kinetics for eight human recombinant UGTs. The initial rate data was fitted to the rate equation for ternary complex mechanism (Eq. 2) at concentration ranges with negligible substrate inhibition; the R^2 values varied from 0.959 to 0.998. **A** UGT1A1 (10, 25, 50, and 100 μM ethinylestradiol), **B** UGT1A3 (25, 50, 100, 250, 500, and 1000 μM scopoletin), **C** UGT1A4 (10, 25, 50, 100, 150, and 250 μM 1-hydroxypyrene), **D** UGT1A6 (50, 100, 250, 400, 500, and 1000 μM scopoletin), **E** UGT1A7 (5, 20, 50, 100, 200, and 250 μM entacapone), **F** UGT1A8 (10, 25, 50, 100, and 250 μM entacapone), **G** UGT1A9 (2.5, 10, 50, 100, and 250 μM entacapone), **H** UGT1A10 (2.5, 10, 50, 100, and 250 μM entacapone). Each data point represents the average of at least two replicates. Experimental details are presented in Materials and methods. For kinetic parameters see Table 2.

Fig. 2. Scheme **A** presents a random ordered bi bi mechanism and scheme **B** a compulsory ordered bi bi mechanism. E=UGT, AX=UDP-glucuronic acid, B=aglycone substrate.

Fig. 3. The substrate inhibition kinetics for seven human recombinant UGTs. The initial rate data was fitted to the rate equation for ternary complex mechanism taking into account substrate inhibition (Eq. 3 and 4). Initial rate data was either from the bisubstrate kinetic analysis (25, 50, 100, 250, 500, 1000, 2500, and 5000 μM UDPGA), or determined at 1 mM UDPGA. The R^2 values varied from 0.864 to 0.994. **A** UGT1A1 (ethinylestradiol), **B** UGT1A3 (entacapone), **C** UGT1A6 (4-nitrophenol), **D** UGT1A7 (4-hydroxyestrone), **E** UGT1A8 (entacapone), **F** UGT1A9 (entacapone), **G** UGT1A10 (entacapone). Each data point represents the average of at least two replicates. Experimental details are presented in Materials and methods. For kinetic parameters see Table 3.

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Fig. 4. Effect of 1-naphthol on entacapone glucuronidation. Inhibition by a competing substrate was determined for UGT1A9 at 100 μM entacapone when UDPGA was the varied substrate (0, 0.05, 0.2, and 1 μM 1-naphthol) and at 1000 μM UDPGA when entacapone was the varied substrate (0, 0.5, 2.5, and 10 μM 1-naphthol). **A** 1-Naphthol was an uncompetitive inhibitor with respect to UDPGA ($K_i=0.169 \pm 0.004 \mu\text{M}$), and **B** a competitive inhibitor with respect to entacapone ($K_i=0.183 \pm 0.0033 \mu\text{M}$). Each data point represents the average of two replicates.

Fig. 5. Inhibition of entacapone glucuronidation by UDP. Product inhibition by UDP (0, 25, 50, 100, 250, and 500 μM) was determined for UGT1A9 at 100 μM concentration of the fixed substrate. **A** UDP was a noncompetitive inhibitor with respect to entacapone ($K_i=20.3 \pm 1.03 \mu\text{M}$, $\alpha K_i=11.5 \mu\text{M}$) and **B** competitive with respect to UDPGA ($K_i=6.03 \pm 0.135 \mu\text{M}$). Each data point represents the average of two replicates.

Fig. 6. Kinetics of purified and membrane-bound UGT1A9. **A** Bisubstrate kinetics of purified UGT1A9 at 10, 50, 100, 250, 500, 750, 1000, and 2000 μM scopoletin as aglycone substrate. The kinetic parameters were : K_m (UDPGA)= $623 \pm 2.37 \mu\text{M}$, K_m (scopoletin)= $673 \pm 2.37 \mu\text{M}$, $V_{\max}=1885 \pm 3.33 \text{ nmol/min/mg}$, $K_{iA}=689 \pm 2.68 \mu\text{M}$, $K_{iB}=745 \pm 3.15 \mu\text{M}$, $k_{\text{cat}}=1.9 \text{ s}^{-1}$, $k_1=3026 \text{ M}^{-1}\text{s}^{-1}$, $k_{-1}=2.1 \text{ s}^{-1}$. All data points represent a single determination. **B** Bisubstrate kinetics of membrane-bound UGT1A9 at 5, 10, 25, 100, and 250 μM scopoletin as aglycone substrate. See Table 2. for kinetic parameters. All data points are the average of two replicates.

Fig. 7. A scheme presenting a compulsory ordered bi bi mechanism and individual rate constants.

AX, UDPGA; B, aglycone substrate; A, UDP; BX, glucuronide; $k_{\text{cat}} = \frac{k_3 k_4}{(k_3 + k_4)}$

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Table 1. Analytical conditions in the kinetic analysis of eight UGT1A isoenzymes

	Column	Eluent	Flow	Inj. vol.	Detection	RT, gluc.	RT, aglycon	Quantitation
EED ^a	Chromolith SpeedRod	50 mM PB ^c pH 3 / MeOH ^d , 60/40	1 ml/min (40°C)	100 µl	Fluorescence, λ _{ex} = 280 nm, λ _{em} = 305 nm	4.1 min	19.1 min	Radiolabelled [¹⁴ C]UDPGA
Scopoletin	Chromolith SpeedRod	50 mM PB ^c pH 3 / MeOH ^d , 90/10	0.8 ml/min (5-11 min 2.5 ml/min) (40°C)	80 µl	Fluorescence, λ _{ex} = 335 nm, λ _{em} = 455 nm	4.2 min	8.5 min	External standard
4-ABP ^b	Waters Symmetry C ₁₈ , 5 µm, 3.9 x 50 mm	20 mM PB ^c pH 6.8 / MeOH ^d , gradient 0-8 min (35-40%), 8-20 min (40-80%), 20-23 min (80-35%), 23-35 min (35% MeOH ^d)	1 ml/min (40°C)	100 µl	UV 275 nm	5.8 min	15.6 min	Radiolabelled [¹⁴ C]UDPGA
1-Hydroxypyrene	Hypersil BDS C ₁₈ , 5 µm, 4 x 150 mm	0.5% aqueous acetic acid / ACN ^e , 40/60	0.9 ml/min (20°C)	20 µl	Fluorescence, λ _{ex} = 237 nm, λ _{em} = 388 nm	2.3 min	6.8 min	External standard
1-Naphthol	Waters Symmetry C ₁₈ , 5 µm, 3.9 x 150 mm	50 mM PB ^c pH 3 / MeOH ^d , 58/42	1 ml/min (40°C)	40 µl	Fluorescence, λ _{ex} = 285 nm, λ _{em} = 335 nm	4.2 min	13.2 min	External standard
Entacapone	Hypersil BDS C ₁₈ , 5 µm, 4 x 150 mm	0.5% aqueous acetic acid/ACN ^e , 65/35	1 ml/min (40°C)	50 µl	UV 310 nm	2.6 min	6.5 min	External standard
4-Hydroxyestrone	Chromolith SpeedRod	50 mM PB ^c pH 3 / MeOH ^d , gradient 0-8 min (15-40%), 8-11 min (40%), 11-12 min (40-15%), 12-15 min (15% MeOH ^d)	2 ml/min (40°C)	100 µl	UV 220 nm	6.7 min	8.2 min	Radiolabelled [¹⁴ C]UDPGA
4-Nitrophenol	Chromolith SpeedRod	50 mM PB ^c pH 3 / MeOH ^d , 85/15	1 ml/min	50 µl	UV 300 nm	1.5 min	6.0 min	External standard
Umbelliferone	Chromolith SpeedRod	50 mM PB ^c pH 3 / MeOH ^d , 85/15	1.5 ml/min	25 µl	Fluorescence, λ _{ex} = 316 nm, λ _{em} = 382 nm	1.3 min	5.3 min	External standard

^aEED ethinylestradiol, ^b4-ABP 4-aminobiphenyl, ^cPB phosphate buffer, ^dMeOH methanol, ^eACN acetonitrile.

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Table 2. The enzyme kinetic parameters for eight UGT1A isoenzymes with appropriate aglycone substrates. The initial rate data was fitted to Eq. 2 omitting the high substrate concentrations with obvious substrate inhibition.

Enzyme	V_{\max} ($\text{pmol min}^{-1} \text{mg}^{-1}$)	K_m UDPGA (μM)	K_{AX} UDPGA (μM)	K_m substrate (μM)	Substrate
UGT1A1	920 \pm 35	409 \pm 40	1402 \pm 235	12.2 \pm 1.77	Ethinylestradiol
UGT1A3	6946 \pm 149	371 \pm 22.3	699 \pm 25.7	500 \pm 17.7	Scopoletin
UGT1A4	5.89 \pm 0.065	201 \pm 6.20	342 \pm 25.9	16.6 \pm 0.738	1-Hydroxypyrene
UGT1A6	1261 \pm 24.3	51.5 \pm 3.26	163 \pm 8.87	3.36 \pm 0.134	1-Naphthol
	5386 \pm 72.0	166 \pm 6.63	252 \pm 13.2	194 \pm 6.42	Scopoletin
UGT1A7	13821 \pm 72.6	248 \pm 3.17	316 \pm 8.54	23.0 \pm 0.355	Entacapone
UGT1A8	2554 \pm 87.3	1256 \pm 75.5	1264 \pm 130	20.7 \pm 1.79	4-Hydroxyestron
	2344 \pm 84.6	875 \pm 70.6	1159 \pm 111	63.3 \pm 4.97	Entacapone
UGT1A9	10171 \pm 51.9	388 \pm 5.26	186 \pm 6.90	20.7 \pm 0.266	Entacapone
	1849 \pm 44.1	87.9 \pm 5.97	111 \pm 11.2	36.7 \pm 2.09	Scopoletin
UGT1A10	4716 \pm 70.5	670 \pm 23.1	693 \pm 48.7	28.9 \pm 1.20	Entacapone

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Table 3. The enzyme kinetic parameters for seven UGT1A isoforms showing substrate inhibition. The initial rate data was fitted to Eq. 3 or 4.

Enzyme	V_{\max} ($\text{pmol min}^{-1} \text{mg}^{-1}$)	K_m UDPGA (μM)	K_{AX} UDPGA (μM)	K_m substrate (μM)	K_{sIB} (μM)	Substrate
UGT1A1	2234 \pm 209 574 \pm 54	1217 \pm 128	755 \pm 71	49.5 \pm 6.9 391 \pm 55	80 \pm 10 3716 \pm 1139	Ethinylestradiol Umbelliferone*
UGT1A3	1015 \pm 112 68 \pm 14			508 \pm 78 19.3 \pm 8.1	1302 \pm 271 469 \pm 324	Entacapone* Ethinylestradiol ^a
UGT1A6	22910 \pm 602			644 \pm 22	1419 \pm 70	4-Nitrophenol*
UGT1A7	664 \pm 47			5.99 \pm 1.3	320 \pm 107	4-Hydroxyestrone ^a
UGT1A8	3641 \pm 332	1759 \pm 183	796 \pm 74.5	119 \pm 16	612 \pm 131	Entacapone
UGT1A9	10722 \pm 158	299 \pm 5.55	145 \pm 11.0	26.3 \pm 1.1	2406 \pm 155	Entacapone
UGT1A10	5469 \pm 107	944 \pm 28.1	496 \pm 34.8	37.4 \pm 1.6	2800 \pm 262	Entacapone

^a The kinetic parameters were determined at 1 mM UDPGA using Eq. 4.

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Table 4. Relative catalytic efficiency of eight recombinant human UGTs of the 1A subfamily in glucuronidation of phenolic compounds. Relative catalytic constant $k_{cat} = 1.0$ and relative specificity constant $k_{cat}/K_m = 1.0$ were arbitrarily assigned for the glucuronidation of entacapone by UGT1A9.

	1-Hydroxypyrene			4-Nitrophenol			Scopoletin			4-Methylumbelliferone			Entacapone		
	k_{cat}	K_m , μM	k_{cat}/K_m	k_{cat}	K_m , μM	k_{cat}/K_m	k_{cat}	K_m , μM	k_{cat}/K_m	k_{cat}	K_m , μM	k_{cat}/K_m	k_{cat}	K_m , μM	k_{cat}/K_m
1A1	0.08	8	0.2	0.09	1039	0.002	0.4	89	0.09	0.03	96	0.01	0.004	42	0.002 ^c
													0.002	17	0.003 ^d
1A3	0.01 ^a		0.02 ^b	0.06	1063	0.002	0.2	500	0.02	0.06	1165	0.003	0.03	259	0.004
1A4	0.002	17	0.001	0.0		0.0	0.0		0.0	0.0		0.0	0.0		
1A6	0.1	10	0.2	0.6	304	0.2	0.9	194	0.1	0.7	127	0.1	0.0		
1A7	0.1	2	4	0.1	240	0.03	0.2	16	0.5	0.3	38	0.3	0.7	23	1.4
1A8	0.1 ^a		0.8 ^b	0.1	2327	0.005	0.2	24	0.6	0.2	330	0.02	0.2	63	0.2
1A9	0.03	0.3	4	0.1	74	0.03	0.1	37	0.1	0.1	16	0.2	1.0	21	1.0
1A10	0.1	0.5	15	0.3	231	0.1	0.6	14	1.6	0.5	30	0.5	0.3	29	0.5

^a k_{cat} calculated from the maximal observed reaction velocity, ^b k_{cat}/K_m determined from the slope of the linear portion of Michaelis Menten curve, ^c3-*O*-glucuronide, ^d4-*O*-glucuronide

Fig. 1

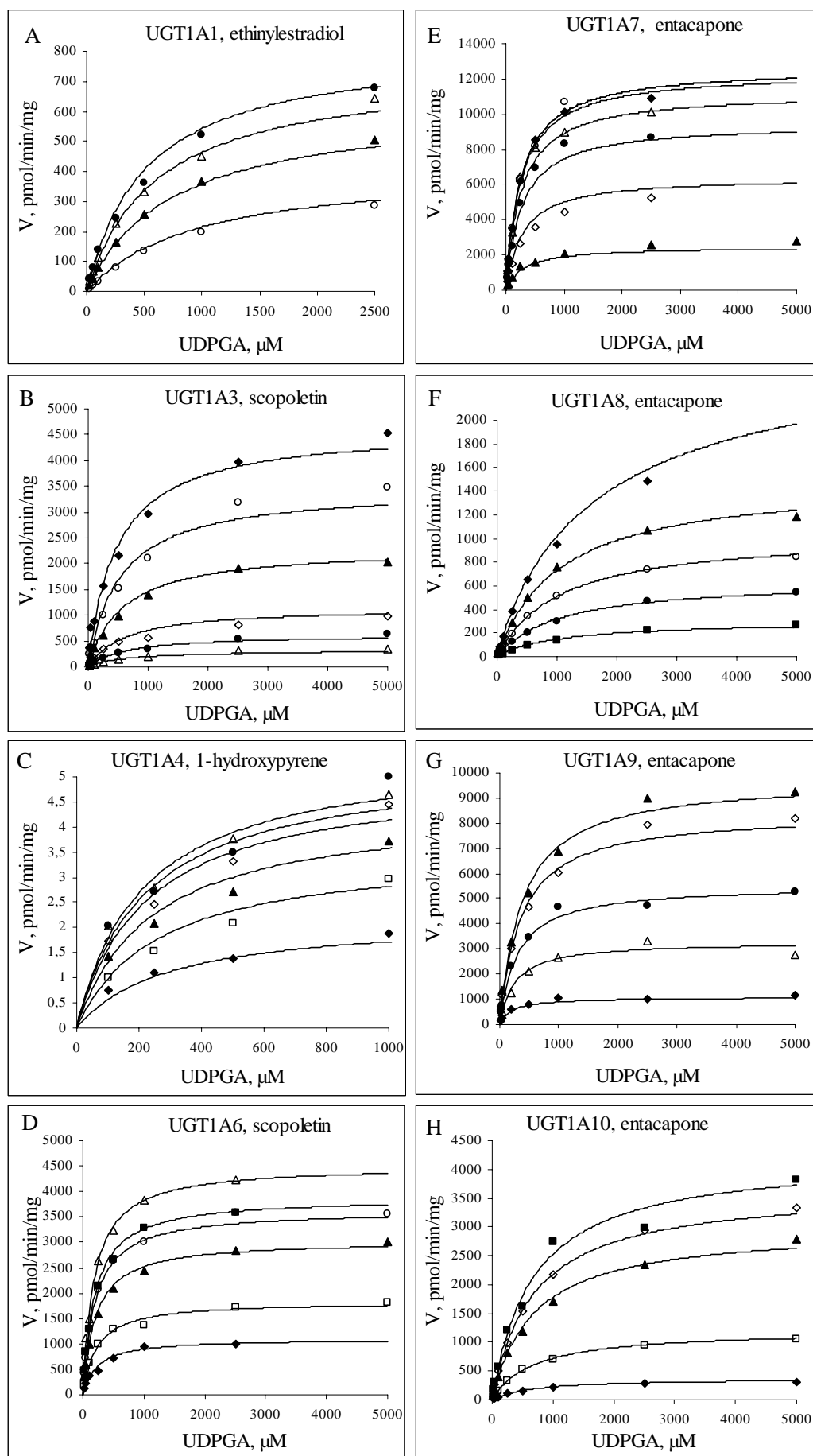


Fig. 2

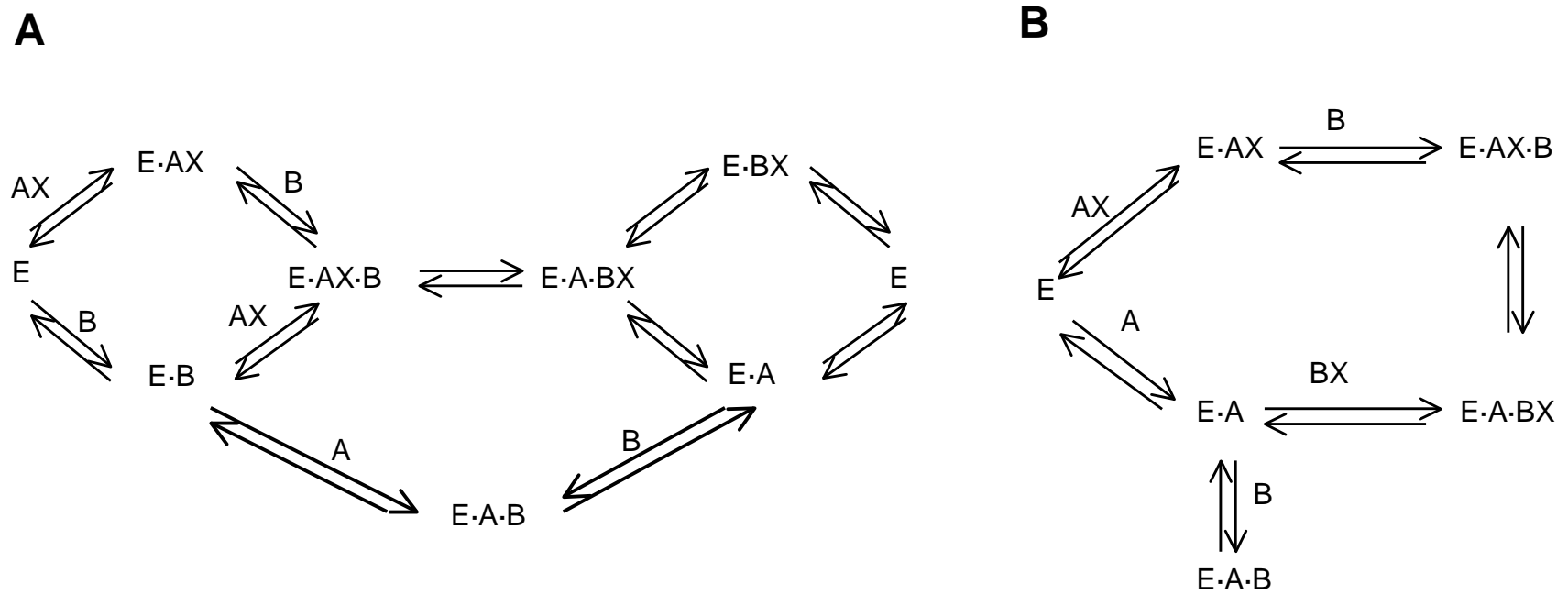


Fig. 3

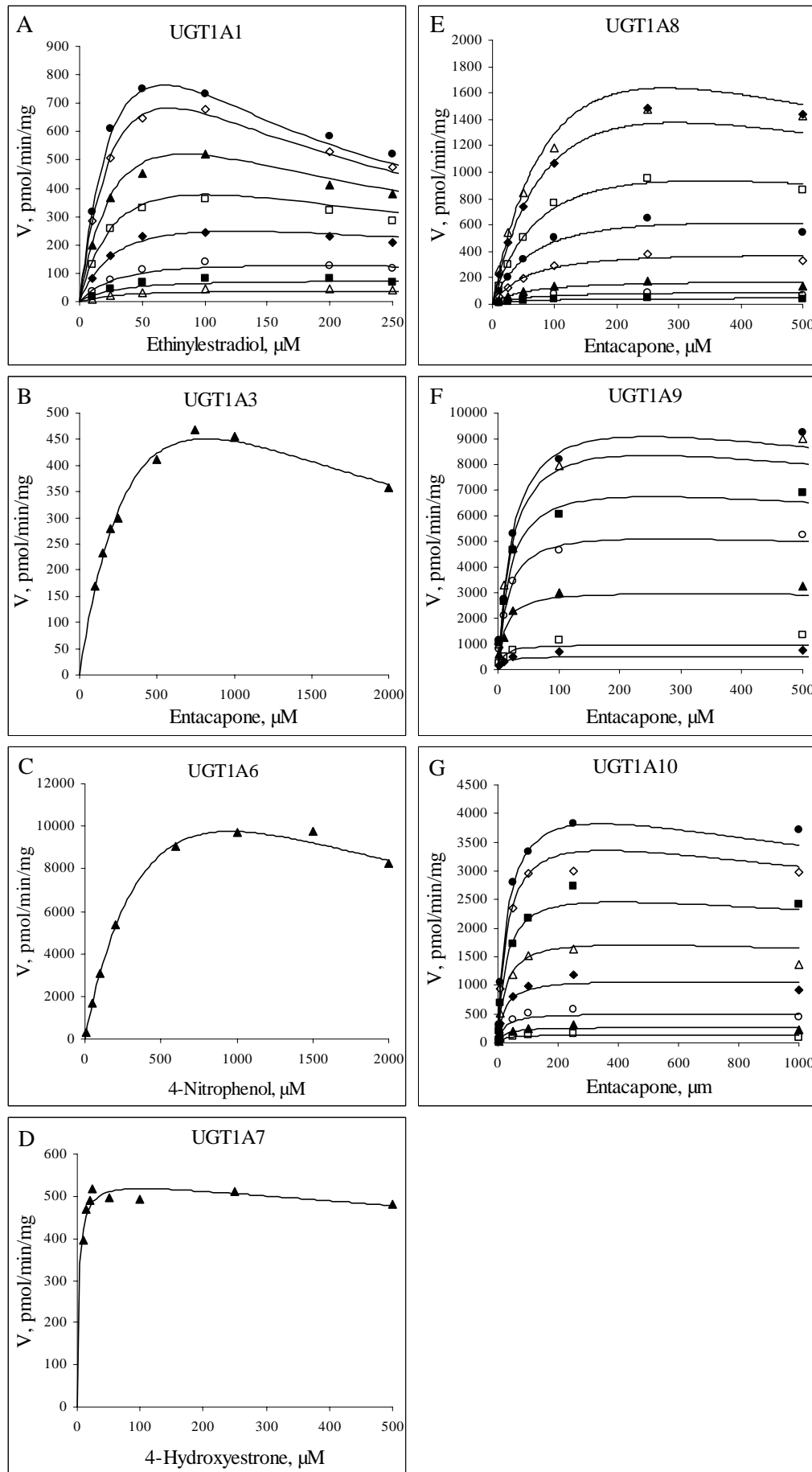
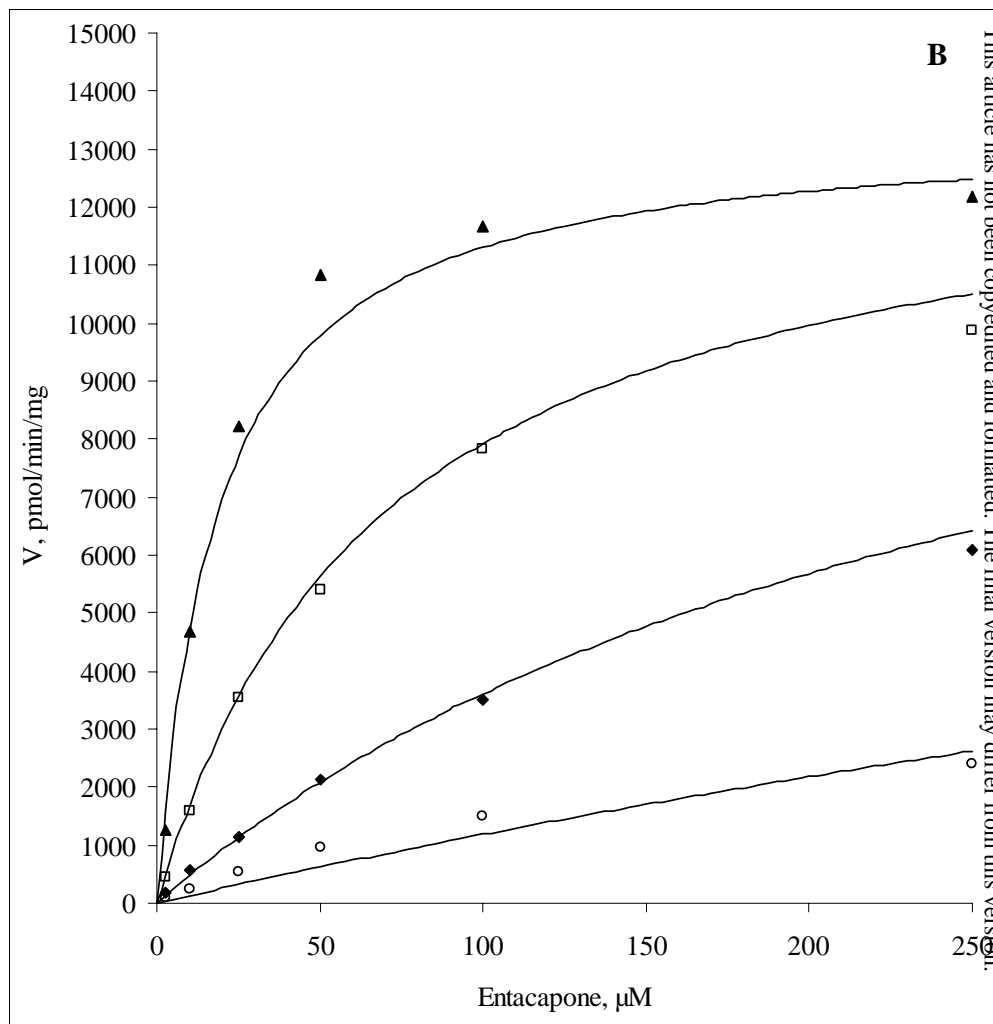
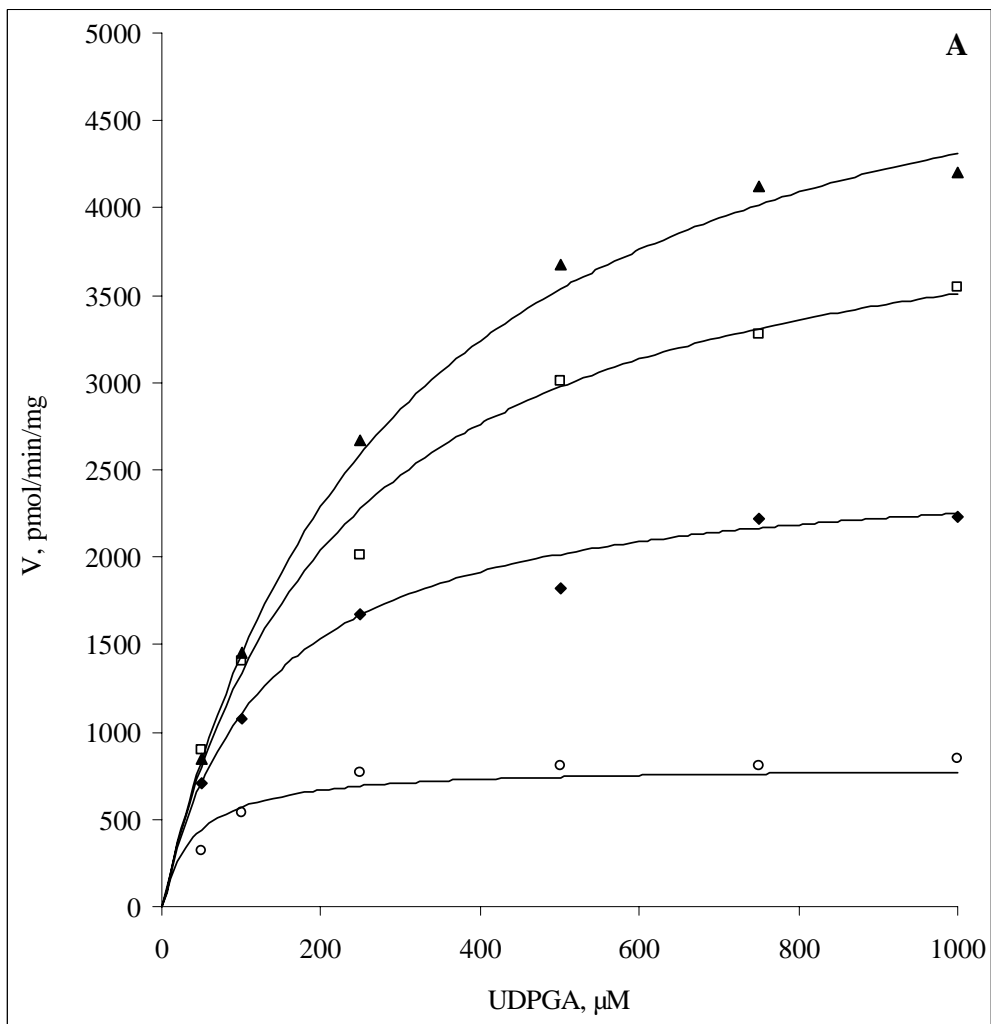


Fig. 4



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Fig. 5

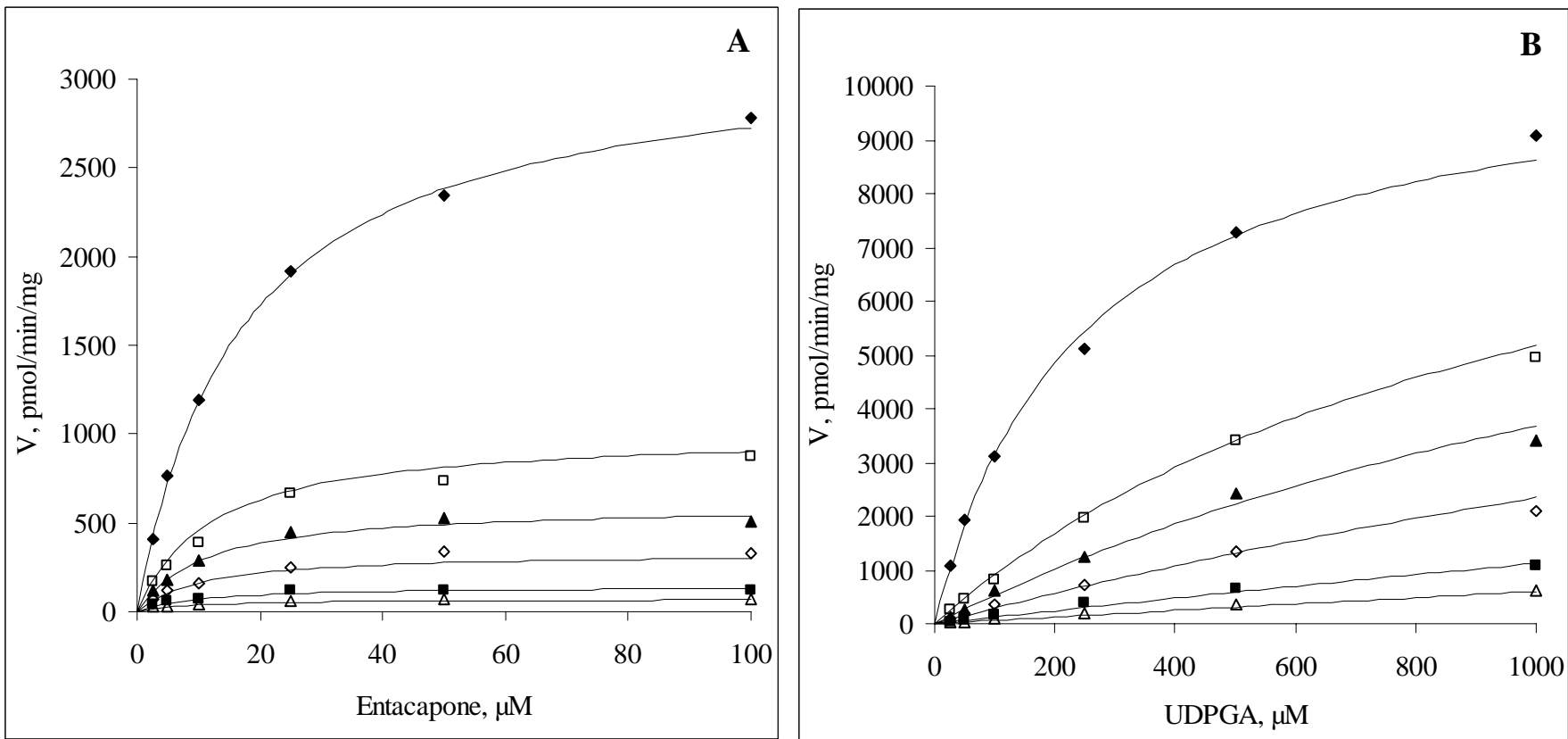


Fig. 6

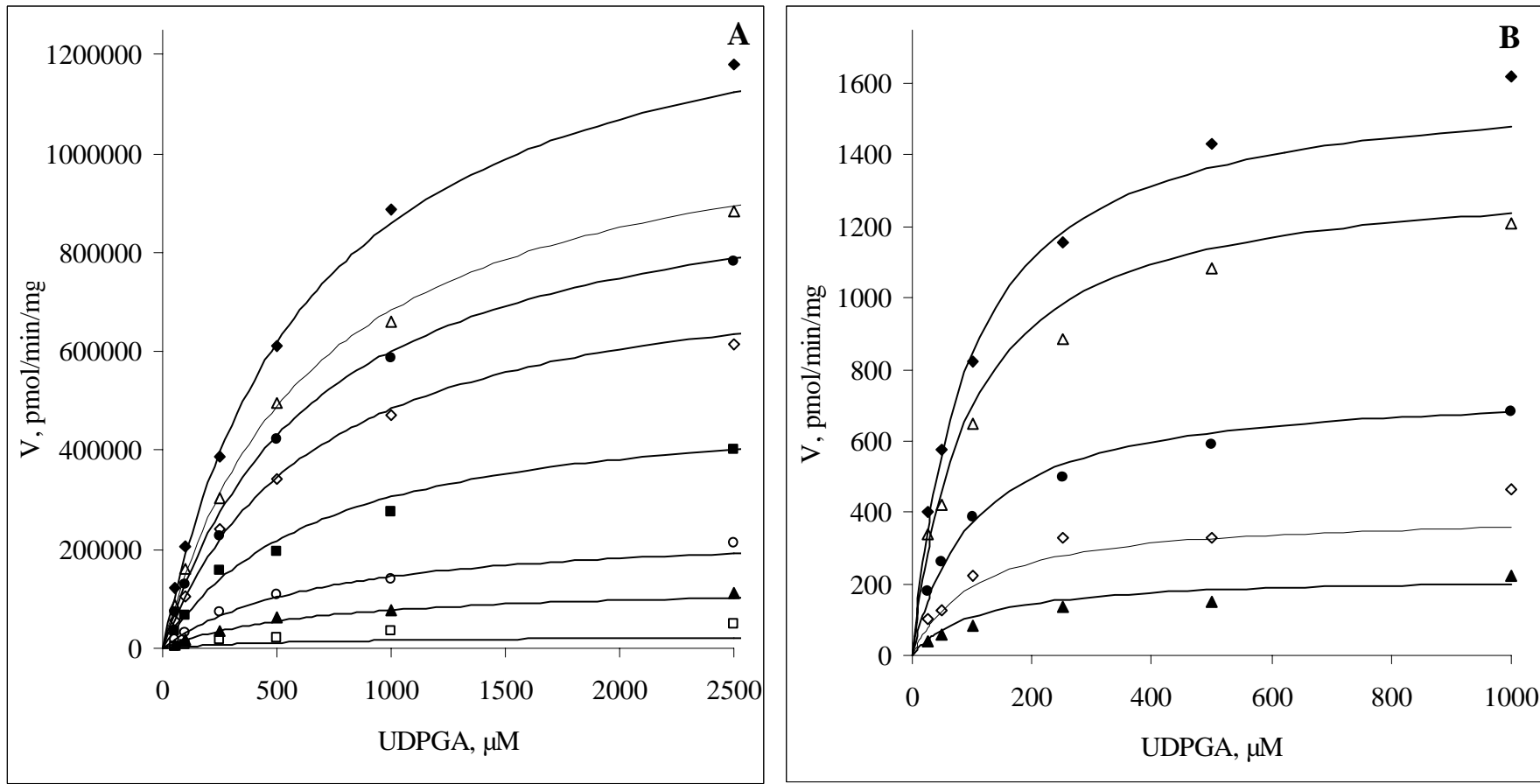


Fig. 7

