Drug interactions of thalidomide with midazolam and cyclosporine A: heterotropic cooperativity of human cytochrome P450 3A5

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Cooperativity of P450 3A5 with thalidomide

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**Abbreviations:**

- $K_e$, effector constant
- $K_i$, inhibition constant
- $K_m$, Michaelis constant
- $K_{m\text{Obs}}$, apparent observed $K_m$ value
- $K_s$, substrate inhibition constant
- HPLC, high-performance liquid chromatography
ABSTRACT

There is growing clinical interest of thalidomide because of its immunomodulatory and anti-angiogenic properties, despite its teratogenecity. However, little information about thalidomide has been reported regarding its precise effects on drug metabolizing enzymes. We investigated the effects of thalidomide on cytochrome P450 enzymes in human liver microsomes to clarify the potential for possible drug interactions. Thalidomide inhibited S-mephenytoin 4'-hydroxylation activities of recombinant P450 2C19 and human liver microsomes: the apparent concentration of thalidomide producing 50% inhibition was approximately 270 µM for P450 2C19. Midazolam 4-hydroxylation activities were suppressed by thalidomide but activities of 1'-hydroxylation and total midazolam oxidation and testosterone 6β-hydroxylation were enhanced in the presence of thalidomide. Recombinant P450 3A5 was found to have altered kinetics at clinically relevant concentrations of thalidomide (10-30 µM). P450 3A4 was also affected, but only at higher thalidomide concentrations. Enhanced midazolam hydroxylation by thalidomide was also seen in liver microsomal samples harboring the CYP3A5*1 allele. Similarly enhanced rates of cyclosporine A clearance were observed in P450 3A5 and liver microsomes expressing P450 3A5 in the presence of thalidomide. A proposed effector constant for thalidomide corresponded roughly to its clinical plasma levels. Docking studies with a P450 3A5 homology model, based on the published structure of P450 3A4, revealed close interaction between thalidomide and the heme of P450 3A5. The present results suggest that total midazolam metabolism or cyclosporine A clearance may be increased by thalidomide in a dose-dependent manner. Unexpected drug interactions involving thalidomide might occur via heterotropic cooperativity of polymorphic P450 3A5.
Introduction

Cytochrome P450 (P450 or CYP) comprises a superfamily of enzymes involved in the oxidation of a large number of endogenous and exogenous compounds (Guengerich, 2008). In human liver, P450 3A4 is the major P450 enzyme followed by P450 2C9 (Shimada et al., 1994); however, the importance of polymorphic P450 3A5 in drug oxidations in Asian populations has been recently suggested (Yamaori et al., 2004; Niwa et al., 2008a). P450 2C19 also catalyzes oxidation of many marketed drugs (Williams et al., 2004) but its content in human liver is relatively low (Inoue et al., 1997). Large interindividual variations in the contents and activities of several P450 forms in human livers lead to different roles of P450s in the oxidations of substrates associated with their pharmacological or toxicological actions (Guengerich, 2008).

Thalidomide [α-(N-phthalimido)glutarimide] had been withdrawn from Europe and Japan in the early 1960s because of its teratogenic effects in humans but has been approved by US Food and Drug Administration in 1998 for the treatment of erythema nodosum leprosum, an acute inflammatory reaction (Calabrese and Resztak, 1998) and in 2006 for the treatment of refractory multiple myeloma. In Japan thalidomide was similarly designated for the treatment of refractory multiple myeloma in 2005 as an “orphan” drug (Sembongi et al., 2008) and a decision regarding final approval is expected in 2008. Many clinical trials with thalidomide are ongoing for both anti-inflammatory and antiangiogenic activities (Vogelsang et al., 1992; Macpherson et al., 2003; Breitkreutz and Anderson, 2008; Kamikawa et al., 2006). Drug interactions between thalidomide and hormonal contraceptives have only been negative (Trapnell et al., 1998; Teo et al., 2000), and thalidomide has been considered to undergo very little metabolism by the P450 system, at least P450 3A4. However, at least
two hydroxylated metabolites of thalidomide have been found in human urine and plasma, both of which could be formed at very low concentrations after incubation with human liver microsomes or recombinant P450 2C19 (Ando et al., 2002). In addition, previous findings have shown spontaneous non-enzymatic hydrolysis (Schumacher et al., 1965). To address the inhibitory potential of a drug, it has been considered to be important to determine an inhibition constant ($K_i$) (Ito et al., 2004). However, there is little information about inhibition by thalidomide thus far.

The purpose of this study was to clarify the inhibitory potential of thalidomide with human P450 enzymes, for understanding of possible drug interactions. We investigated the effects of thalidomide on P450 activities and found that thalidomide inhibited P450 2C19-dependent S-mephenytoin 4'-hydroxylation at high concentrations but enhanced P450 3A5-dependent midazolam hydroxylation and cyclosporine A clearance at clinically relevant concentrations. Although there are reports on ligand cooperativity with P450 3A5 (Niwa et al., 2008b), we report a proposed effector constant for thalidomide in the heterotropic cooperativity of P450 3A5 in human liver microsomes, adapting $1 + [I]/K_i$ treatment theory for drug interaction studies. A proposed model for heterotropic cooperativity of P450 3A5 is presented.

Materials and Methods

Chemicals. (+)-Thalidomide was purchased from Sigma-Aldrich (St. Louis, MO). Cyclosporine A and midazolam were obtained from Wako Pure Chemicals (Osaka, Japan). Other chemicals and reagents used in this study were obtained from the sources described.
previously (Yamazaki et al., 2002; Yamazaki et al., 2006) or were of the highest qualities commercially available.

**Enzyme preparations.** Human liver microsomes were prepared in 10 mM Tris-HCl buffer (pH 7.4) containing 0.10 mM EDTA and 20% (v/v) glycerol as described previously (Yamazaki et al., 2006). The use of the human livers for this study was approved by the Ethics Committees of Vanderbilt University and Showa Pharmaceutical University. Each recombinant human cytochrome P450, coexpressed in *E. coli* membranes with human NADPH-P450 reductase, was prepared as described earlier (Yamazaki et al., 2002). Genomic DNA samples from livers were genotyped for the *CYP2C19* and *CYP3A5* genes as described previously (Inoue et al., 1997; Yamaori et al., 2004).

**Enzyme assays.** Midazolam 1'- and 4-hydroxylation activities were determined using high-performance liquid chromatography (HPLC) (Emoto et al, 2008). Cyclosporine A oxidation was determined by the disappearance of parent compound (Dai et al., 2004). Briefly, a typical incubation mixture (total volume of 0.25 ml) contained microsomal protein (0.25 mg/ml) or recombinant P450 (0.06 μM), an NADPH-generating system (0.25 mM NADP⁺, 2.5 mM glucose 6-phosphate, and 0.25 unit/ml glucose 6-phosphate dehydrogenase), and substrate and/or thalidomide in 0.10 M potassium phosphate buffer (pH 7.4), unless otherwise specified. For P450 activity determinations, incubations were carried out at 37 °C for 10-30 min. Incubations were terminated by adding 0.25 ml of ice-cold acetonitrile. The aqueous supernatant was centrifuged at 2,000 g for 10 min and subjected to HPLC using with an analytical octadecylsilane (C₁₈) column (4.6 mm × 150 mm, 5 μm).

Ethoxyresorufin *O*-deethylation, 7-ethoxycoumarin *O*-deethylation, diclofenac 4'-hydroxylation, *S*-mephénytoin 4'-hydroxylation, and testosterone 6β-hydroxylation...
activities were determined as described previously (Yamazaki et al., 2002). Microsomal protein concentrations were estimated by using a bicinchoninic acid (BCA) protein assay kit (Pierce, Rockford, IL). Concentrations of total P450 (Omura and Sato, 1964) and NADPH-P450 reductase (EC 1.6.2.4) (Yamazaki et al., 2002), and P450 3A4 and 3A5 contents in liver microsomes (Yamaori et al., 2005) were determined as described previously. The liver microsomal samples HL-1 and HL-3 contained 18 pmol P450 3A4 and 6.1 pmol P450 3A5 per mg protein and 17 pmol P450 3A4 and 24 pmol P450 3A5 per mg protein, respectively (Yamaori et al., 2005).

**Kinetic analysis.** Kinetic analysis was done using nonlinear regression analysis programs (KaleidaGraph, Synergy Software, Reading, PA and Prism, GraphPad Software, San Diego, CA) using the following these equations. Inhibition by thalidomide (concentration [I]) of midazolam 4-hydroxylation activity (v) of P450 3A as a function of substrate concentration [S] with a maximum velocity (V_max) was analyzed for competitive inhibition with the usual equation (1)

\[ v = \frac{V_{\text{max}} \cdot [S]}{(K_{\text{mObs}} + [S])} \]  

(1)

and an apparently observed \( K_m \) value (\( K_{m\text{Obs}} \)), namely

\[ K_{m\text{Obs}} = K_m \cdot (1 + [I] / K_i) \]  

(2)

where \( K_i \) is the inhibition constant (Ito et al., 2004).

On the other hand, midazolam 1'-hydroxylation activity (v) shows a substrate-inhibition manner with the equation (3) including the substrate inhibition constant \( K_s \) (Dai et al., 2004):

\[ v = \frac{V_{\text{max}} \cdot [S]}{(K_{m\text{Obs}} + [S] + [S]^2/K_s)} \]  

(3)
When the enzyme activities ($v$) are increased by thalidomide (concentration $[E]$), resulting apparent $V_{\text{max}}$ values multiplied by $(1 + [E]/K_e)$ or deceased $K_{\text{mObs}}$ divided by $(1 + [E]/K_e)$ are proposed using the equation (3). In the present study, apparent $V_{\text{max}}$ values were not affected, but $K_{\text{mObs}}$ values were decreased in the presence of thalidomide, therefore the equations (3) and (4) including a novel parameter for effector concentration $K_e$ was proposed in this study:

$$K_{\text{mObs}} = K_m / (1 + [E] / K_e) \quad (4)$$

**Docking simulation of thalidomide into reported structure of P450 3A4 and a homology model of P450 3A5.** The human P450 3A5 primary sequence was aligned with human P450 3A4 (Protein Data Bank code 1TQN) in the MOE software (ver. 2007.09, Chemical Computing Group, Montreal, Canada) for modeling of a three-dimensional structure (Pearson et al., 2007). Prior to docking, the energy of the P450 3A4 or 3A5 structure was minimized using the CHARM22 force field. Docking simulations were carried out for thalidomide binding to the reported P450 3A4 or a homology model of P450 3A5 using the MMFF94x force field distributed in the MOE Dock software. Twenty solutions were generated for each docking experiment and ranked according to total interaction energy (S value).

**Results**

**Inhibitory or enhancing effects of thalidomide on activities of human P450 enzymes.** Because P450 2C19 has already been shown to be the major enzyme involved in the oxidative metabolism of thalidomide (Ando et al., 2002), we first investigated the
inhibitory effects of thalidomide on the typical marker activities of a serious of P450 enzymes. The oxidation activities measured with the substrates ethoxyresorufin (10 µM) (recombinant P450 1A1 and 1A2), 7-ethoxycoumarin (100 µM) (recombinant P450 2B6), and diclofenac (50 µM) (recombinant P450 2C9) were not affected by thalidomide (up to 500 µM) under these conditions (results not shown). P450 2C19 S-mephenytoin 4'-hydroxylation activity was suppressed by thalidomide in a concentration-dependent manner (Fig. 1A): 50% inhibition was observed with a thalidomide concentration of 270 µM. Similar inhibitory effects of thalidomide on S-mephenytoin hydroxylase activity were seen in liver microsomal preparations (genotyped as CYP2C19*1/*1) (Fig. 1B).

On the contrary, testosterone 6β-hydroxylation (Figs. 2A,2B) by P450 3A4 or P4503A5 was enhanced by thalidomide. Midazolam 1'-hydroxylation activities (Fig. 2D) of P450 3A5 were highly enhanced in the presence of low concentrations of thalidomide. These activities were less affected by thalidomide with P450 3A4 than P450 3A5 (Fig. 2C). The minor pathway, midazolam 4-hydroxylation, was slightly suppressed by thalidomide (with P450 3A4 or 3A5), but rates of formation of the major product (1'-hydroxylation) and total oxidative products of midazolam were stimulated in the presence of thalidomide.

**Modifications of kinetic parameters for P450 3A-dependent drug oxidation activities by thalidomide.** It was of interest to describe the enhancing effects of thalidomide in terms of an effector concentration. Midazolam 1'- and 4-hydroxylation activities were plotted against substrate concentrations in the presence of various concentrations of thalidomide (Fig. 3). Only small effects of thalidomide on the midazolam hydroxylation activities of P450 3A4 were observed (Fig. 3A). In the contrast, midazolam 1'-hydroxylation activities of P450 3A5 at low substrate concentrations were enhanced in the
presence of thalidomide; however, midazolam 4-hydroxylation were suppressed (Fig. 3B). Human liver microsomes were also used as the enzyme source. The midazolam hydroxylation activities of liver microsomal sample HL-3 (Fig. 3C), genotyped as CYP3A5*3/*3, were not extensively affected, as in the case of recombinant P450 3A4. Midazolam 1'-hydroxylation activities in liver microsomal sample HL-1 (Fig. 3D), genotyped as CYP3A5*1/*3, were also enhanced by the low concentrations of thalidomide.

Apparent $K_m$ values ($K_{m_{\text{Obs}}}$) for midazolam 1'-hydroxyation activities of P450 3A5 using the Michaelis-Menten equation were reduced from 5.5 $\mu$M to 3.0, 1.8, 1.2, 0.97, and 0.77 $\mu$M in the presence of 10, 50, 100, 250, and 500 $\mu$M thalidomide, respectively (Fig. 3B). We adopted a new parameter, $K_e$, for modified $K_{m_{\text{Obs}}}$ vales as $K_m / (1 + [E] / K_e)$, in the similar way as $K_i$ for $K_m \cdot (1 + [I] / K_i)$ (Table 1). The $K_e$ value of thalidomide enhancement of midazolam 1'-hydroxyation catalyzed by recombinant P450 3A5 was calculated to be 40 ± 7 $\mu$M. Similar reduced $K_{m_{\text{Obs}}}$ values for midazolam 1'-hydroxyation activities by increasing concentrations of thalidomide were obtained in the human liver microsomes expressing P450 3A5 protein (CYP3A5*1/*3): $K_e$ value was 62 ± 24 $\mu$M using the equation of $K_{m_{\text{Obs}}} = K_m / (1 + [E] / K_e)$. In the case of P450 3A4, > 10-fold higher $K_e$ values of thalidomide were obtained (Table 1).

To confirm the enhancing effects of thalidomide on liver microsomal P450 3A5, the oxidation of cyclosporine A was also used as a test reaction (Fig. 4). P450 3A4-catalyzed cyclosporine A oxidation was not affected by thalidomide (Fig. 4A). P450 3A5-catalyzed cyclosporine A oxidation (substrate concentrations of 2 and 10 $\mu$M) was enhanced by increasing concentrations (2-30 $\mu$M) of thalidomide (Fig. 4B). Similar enhancement of cyclosporine A oxidation by thalidomide was seen in liver microsomes expressing P450 3A5.
and 3A4 (Fig. 4D) but not in other liver microsomes mainly expressing P450 3A4 (Fig. 4C). Thalidomide (10-30 µM) enhanced cyclosporine A oxidation mediated by P450 3A5 in human liver microsomes approximately 2-fold.

**Docking simulation of thalidomide into P450s 3A4 and 3A5.** The human P450 3A4 crystal structures allowed generation of a homology model of P450 3A5 using the MOE program. The top-rank docking-model of thalidomide in P450 3A4 or 3A5 was adopted. In the P450 3A4 model, the aromatic ring of thalidomide was found far from the heme of P450 3A4 (Fig. 5A). In contrast, in the P450 3A5 model, the cycloalkane ring of thalidomide was closely orientated to the center of the heme of P450 3A5 (Fig. 5B).

**Discussion**

In the course of screening of the inhibitory potential of thalidomide with human P450 enzymes, thalidomide inhibition of P450 2C19-mediated activity and enhancement of P450 3A activities were seen (Figs. 1,2). Thalidomide inhibited P450 2C19-dependent S-mephenytoin 4'-hydroxylation activity only at high concentrations but enhanced P450 3A5-depedent midazolam 1'-hydroxylation activity at low concentrations. P450s 2C9 and 2C19 have both been shown to have smaller substrate pocket than P450 3A4 (Yano et al., 2004; Locuson and Wahlstrom, 2005; Ahlstrom and Zamora, 2008). The thalidomide inhibition constant for P450 2C19 was similar to the substrate concentration used for S-mephenytoin. Thus, the active site area of P450 2C19 may be blocked by a similar concentration of thalidomide, resulting in half-maximal inhibition of P450 2C19 function.

Polymorphic P450 3A5 have been suggested to contribute as much as 50% of the
liver P450 3A in one-third of Caucasians and one-half of African-Americans (Kuehl et al., 2001; Koch et al., 2002) or Japanese (Yamaori et al., 2004). Since P450 3A5 is 83% identical to P450 3A4, it is generally believed that the substrate specificity of P450 3A5 is similar to that of P450 3A4, although some differences in catalytic properties have been found (Daly, 2006; Niwa et al., 2008a; Niwa et al., 2008b). Less activation of P450 3A4 by thalidomide was observed under our conditions. On the other hand, midazolam 4-hydroxylation (minor pathway) was inhibited but 1'-hydroxylation and total disappearance of midazolam (as well as cyclosporine A) were enhanced by thalidomide, presumably mediated via a cooperativity mechanism in P450 3A5 (Figs. 3,4). In our preliminary experiments, preincubation of thalidomide with P450 3A5 in the presence of NADPH resulted in decreases in enhanced midazolam 1'-hydroxylation (results not shown), suggesting the involvement of thalidomide itself, not a product (tentatively 5'-hydroxythalidomide), in the activation. Similar activation of P450 3A5-mediated midazolam 1'-hydroxylation were also observed in another liver microsomal sample, HL-130, harboring CYP3A5*1/*3 but not in HL-127 (CYP3A5*3/*3) (results not shown). Because the apparent $K_{\text{m,Obs}}$ values for midazolam 1'-hydroxylation in recombinant P450 3A5 and human liver microsomes were similarly decreased in the presence of thalidomide, the proposed $K_e$ values of thalidomide (approximately 40 µM) for P450 3A5 and liver microsomes are consistent and are one order of magnitude less than those for P450 3A4 (Table 1). These results suggest the hypothesis that thalidomide preferentially binds to P450 3A5. To further address this hypothesis, docking simulations of thalidomide into P450 3A4 and 3A5 models were performed (Fig. 5). Thalidomide could closely dock to the P450 3A5 heme region in silico analysis (Fig. 5B). These results and findings may be collectively relevant to a different preferable substrate orientation for P450 3A5, explaining the P450 3A5 activation.
This activation phenomenon is generally termed heterotropic cooperativity, involving two different ligands in the active site of a P450 enzyme (Mcconn II et al., 2004; Isin and Guengerich, 2006). A proposed model for heterotropic cooperativity of P450 3A5 is shown in Fig. 5C. When thalidomide docks near the heme iron and/or suitable substrate binding area (Roberts and Atkins, 2007; Skopalik et al., 2008), the proposed substrate pocket of P450 3A5 would be affected resulting in large active site. On the other hand, because thalidomide apparently does not dock closely to the heme of the P450 3A4 structure (Fig. 5A), the substrate binding area might not be strongly affected (Fig. 5C). This proposed mechanism could be derived from the several differences in reported important amino residues (positions at 210-214) of the F-helix of P450 3A4, remote from the heme of the P450 3A4 catalytic center (Szklarz and Halpert, 1997; Fowler et al., 2002; Harlow and Halpert, 1998), and a characterized Ser239 residue in substrate recognition site-3 (Pearson et al., 2007) (Fig. 5D). The Leu211 residue in both P450 3A4 and 3A5 has been suggested to be an important residue for homotropic cooperativity in P450 3A4 (Szklarz and Halpert, 1997; Fowler et al., 2002; Harlow and Halpert, 1998). Lys212 of P450 3A5 seems to have a similar property, corresponding to Arg212, in P450 3A4 forming the roof of the active site (Harlow and Halpert, 1998). Amino acid differences at positions Leu210, Asp214, and Ser239 in P450 3A4 and Phe210, Gly214, and Cys239 in P450 3A5 might produce more flexible site and higher lipophilicity in the P450 3A5 active site, to show different phenomena with P450 3A4 for thalidomide. These residues might be key determinant factors for the specificity of thalidomide activation of P450 3A5. Consequently, a thalidomide molecule shifts P450 3A5 into a conformation, such that the proximal binding niche exists near the heme, leading to the stimulation of the cyclosporine and midazolam oxidative activation observed in the present study.
In terms of the clinical consequences of drug interactions with thalidomide, the present results suggest that a more rapid drug clearance might be due to thalidomide via a P450 3A5 contribution. The reported plasma concentrations of thalidomide (Vogelsang et al., 1992) are similar to the proposed $K_e$ constants estimated in the present study. Adapting $1 + [I]/K_i$ theory to the $1/(1 + [E]/K_e)$ equation, thalidomide concentrations could reach levels similar to calculated $K_e$ constant and, in principal, approximately 1.5-fold enhanced elimination of other drugs mediated by P450 3A5. We propose the $K_e$ value as a general constant, whereas it is highly likely to be dependent upon an exact pairing of effector and substrate. Vincristine has been reported to be a P450 3A5 substrate (Dennison et al., 2007). Therefore, we investigated interactions between thalidomide and vincristine in vitro. In preliminary experiments, however, low concentrations of thalidomide did not activate the oxidative cleavage of vincristine (catalyzed by P450 3A5 or liver microsomes with $CYP3A5*1/*3$) (results not shown). The $K_m$ values (for the major oxidative cleavage of vincristine catalyzed by P450 3A5) have been reported to be 13-17 µM (Dennison et al., 2006). Because vincristine is a large molecule and has high P450 3A5 affinity, these present results may be interpreted that thalidomide could not dock in P450 3A5 prior to vincristine.

In conclusion, the present study suggests that total midazolam metabolism or cyclosporine clearance may be increased by thalidomide with clinical relevant concentrations through the heterotropic cooperativity of human P450 3A5. Because of the high frequency of polymorphic P450 3A5 expression in Asians and Africans, a relatively high frequency of unexpected drug interactions involving thalidomide might occur via P450 3A5 contribution in drug metabolism.
Acknowledgments

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Footnote

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Legends for figures

Fig. 1. Effects of thalidomide on S-mephenytoin 4'-hydroxylation activity by recombinant P450 2C19 (A) and human liver microsomes (B).

S-Mephenytoin (200 μM) was incubated with P450 2C19 and human liver microsomal sample HL-1 (genotyped as CYP2C19*1/*1) in the absence or the presence of thalidomide. Results are presented as means and ranges of duplicate determinations.

Fig. 2. Effects of thalidomide on testosterone 6β-hydroxylation activity (●, A, B) and midazolam 1'- (▲) and 4- (■) hydroxylation activity (C, D) by recombinant P450 3A4 (A,C) and P450 3A5 (B,D).

Testosterone or midazolam (100 μM) was incubated with P450 3A4 or 3A5 in the absence or the presence of thalidomide. Results are presented as means and ranges of duplicate determinations.

Fig. 3. Effects of substrate concentrations (0.25-200 μM) on midazolam 1'- and 4-hydroxylation activity of recombinant P450 3A4 (A) and P450 3A5 (B) and human liver microsomes (C,D) in the absence (○) or the presence of 10 (▲), 25 (□), 50 (◇), 100 (●), 250 (▲), and 500 μM (■) thalidomide.

Human liver samples HL-3 (C) and HL-1 (D) were genotyped as CYP3A5*3/*3 and CYP3A5*1/*3, respectively. Results are presented as means of duplicate determinations.
Fig. 4.  Enhanced cyclosporine oxidation activity with recombinant P450 3A4 (A) and 3A5 (B) and human liver microsomes (C,D) in the absence (□) or the presence of 2 (■), 10 (□), and 30 µM (■) thalidomide.

Human liver samples HL-3 (C) and HL-1 (D) were genotyped for the CYP3A5*3/*3 and CYP3A5*1/*3, respectively. Results are presented as means ± SD from triplicate determinations.

Fig. 5.  Docking simulation of thalidomide into a reported structure of P450 3A4 (A) and a homology model of P450 3A5 (B).

Thalidomide was docked in orientations with interaction energies (S values) of 38.5 (A) and 38.1 (B), respectively. Proposed working models for heterotropic binding to P450 3A4 and 3A5 (C) and comparison of key amino acid residues in the reported F-helix and SRS-3 between P450 3A4 and P450 3A5 (D) are shown. SRS-3, substrate recognition site-3.
Table 1. Effect of thalidomide on midazolam 1'- and 4-hydroxylation activities of P450 3A4 and 3A5 and human liver microsomes with the kinetic parameter $K_e$.

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>Midazolam hydroxylation</th>
<th>Midazolam $V_{max}$</th>
<th>Midazolam $K_m$, µM</th>
<th>Midazolam $K_s$, µM</th>
<th>Thalidomide $K_i$, µM</th>
<th>Thalidomide $K_e$, µM</th>
<th>Goodness of fit, $r^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>P450 3A4</td>
<td>1'-</td>
<td>8.6 ± 0.3</td>
<td>1.8 ± 0.2</td>
<td>87 ± 10</td>
<td>—</td>
<td>850 ± 370</td>
<td>0.98</td>
</tr>
<tr>
<td></td>
<td>4-</td>
<td>3.4 ± 0.1</td>
<td>25 ± 2</td>
<td>—</td>
<td>1500 ± 620</td>
<td>—</td>
<td>0.99</td>
</tr>
<tr>
<td>P450 3A5</td>
<td>1'-</td>
<td>18 ± 1</td>
<td>5.5 ± 0.5</td>
<td>1100 ± 200</td>
<td>—</td>
<td>40 ± 7</td>
<td>0.98</td>
</tr>
<tr>
<td></td>
<td>4-</td>
<td>2.2 ± 0.1</td>
<td>59 ± 6</td>
<td>—</td>
<td>160 ± 20</td>
<td>—</td>
<td>0.99</td>
</tr>
<tr>
<td>HL-3 microsomes</td>
<td>1'-</td>
<td>0.83 ± 0.02</td>
<td>3.9 ± 0.3</td>
<td>700 ± 100</td>
<td>—</td>
<td>770 ± 300</td>
<td>0.98</td>
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<td>(CYP3A5*3/*3)</td>
<td>4-</td>
<td>0.30 ± 0.01</td>
<td>52 ± 4</td>
<td>—</td>
<td>850 ± 240</td>
<td>—</td>
<td>0.98</td>
</tr>
<tr>
<td>HL-1 microsomes</td>
<td>1'-</td>
<td>1.7 ± 0.1</td>
<td>12 ± 3</td>
<td>670 ± 290</td>
<td>—</td>
<td>62 ± 24</td>
<td>0.94</td>
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<tr>
<td>(CYP3A5*1/*3)</td>
<td>4-</td>
<td>0.35 ± 0.01</td>
<td>66 ± 5</td>
<td>—</td>
<td>1100 ± 300</td>
<td>—</td>
<td>0.94</td>
</tr>
</tbody>
</table>

Kinetic parameters were calculated with non-linear regression analysis with SE values. $K_e$, effector constant; $K_i$, inhibition constant; $K_m$, Michaelis constant, $K_s$, substrate inhibition constant.
Fig. 1

S-Mephenytoin 4'-hydroxylation

(A) P450 2C19

(B) Human liver microsomes

S-Mephenytoin 4'-hydroxylation

(nmol/min/nmol P450)

Thalidomide (µM)

(nmol/min/mg protein)

0 250 500

0 0.025 0.050
Fig. 3

(A) P450 3A4

(B) P450 3A5

(C) Liver microsomes (CYP3A5*3/*3)

(D) Liver microsomes (CYP3A5*1/*3)

Midazolam concentration (µM)

Midazolam 1'- and 4-hydroxylation

(nmol/min/nmol P450) (nmol/min/mg protein)
Fig. 4

(A) P450 3A4

(C) Liver microsomes
  \(\text{CYP3A5}^{*3/*3}\)

(B) P450 3A5

(D) Liver microsomes
  \(\text{CYP3A5}^{*1/*3}\)

Cyclosporine A disappearance
(nmol/min/mg protein) (nmol/min/nmol P450)

Cyclosporine A concentration (µM)
Fig. 5

(A) P450 3A4

(B) P450 3A5

(C) 3A4

(D) 3A4:

Leu Leu Arg Phe Asp

3A5:

Phe Leu Lys Phe Gly

Thalidomide

Heme

Thalidomide

Heme

■, Thalidomide; ○, Substrate.

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<th>Position</th>
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