Metabolic Switching of BILR 355 in the Presence of Ritonavir I: Identifying an
Unexpected Disproportionate Human Metabolite

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HIV-1, human immunodeficiency virus; NNRTI, non-nucleoside reverse transcriptase inhibitor;
BILR 355, 11-ethyl-5,11-dihydro-5-methyl-8-[2-[(1-oxido-4-quinolinyl)oxy]ethyl]-6H-
dipyrido[3,2-b:2’,3’-e] [1,4]diazepin-6-one; $T_{\text{max}}$, mean time to maximum drug concentration;
$T_{1/2}$, terminal half-life; CYP, cytochrome P450; RTV, ritonavir; $C_{\text{max}}$, maximum plasma concentration;
$\text{AUC}_{0-\infty}$, area under the plasma concentration-time curve from time zero to infinity; HLMs, human liver microsomes; LC-MS/MS, high performance liquid chromatography-tandem mass spectrometry; $m/z$, mass to charge ratio; $K_m$, Michaelis-Menten constant; $V_{\text{max}}$, maximum reaction velocity; $\text{CL}_{\text{int}}$, intrinsic clearance; $\text{CL}_{\text{h}}$, hepatic clearance; $Q_h$, hepatic blood flow in human; $\text{AUC}_{0-\tau}$, area under the plasma concentration-time curve over the dosing interval
ABSTRACT:

BILR 355 is an inhibitor of the human immunodeficiency virus-1. BILR 355 exhibited a nonlinear pharmacokinetic profile and low exposure after oral administration to humans. This paper describes the in vitro metabolism of BILR 355, which is correlated with the in vivo nonlinearity findings. Our in vitro studies had demonstrated that BILR 355 was extensively metabolized by cytochrome P450 3A. Consequently BILR 355 was concomitantly administered with ritonavir (RTV) in attempt to boost systemic exposure which did occur in humans. In addition, the expectation was that the overall metabolism of BILR 355 would be decreased with concomitant administration of RTV. Subsequent metabolite profiling was performed using human plasma samples obtained from clinical Phase Ib studies with concomitant administration of BILR 355 and RTV. A total of eighteen metabolites were observed. Their structures were proposed based on high performance liquid chromatography-tandem mass spectrometry technologies and ten metabolites were confirmed by comparison with synthetic standards. Surprisingly, a disproportionate human metabolite, BILR 516, was uncovered during this metabolite profiling study and PK analysis of BILR 516 showed that it had a longer half-life and higher exposure than the parent compound at steady state. Interestingly, BILR 516 was not detected in human plasma when BILR 355 was administered alone. So while RTV boosted the exposure of BILR 355, it resulted in a significant metabolic switching of BILR 355. Overall, this paper demonstrates an unusual example of metabolic switching and raises the concern on the consequence of metabolic switching during drug development.
Introduction

The human immunodeficiency virus (HIV)-1 infection remains a major worldwide health issue. The current antiretroviral therapy has dramatically reduced morbidity and mortality among HIV-infected patients and significantly modified the course of HIV disease into a manageable chronic disease with longer survival and improved quality of life of the patients (Palella et al., 1998; Hogg et al., 1998; Moreno et al., 2010). However, resistance has emerged to almost all of these antiretroviral agents, sometimes rendering them ineffective (Hammer et al., 2008). First generation non-nucleoside reverse transcriptase inhibitors (NNRTIs) are particularly vulnerable, since resistance may emerge with a single resistance codon alteration and the emergence of drug resistant HIV-1 variants is fairly rapid (De Clercq, 1998). In addition, extensive cross-resistance among NNRTIs was observed, resulting in a loss of the efficacy of subsequent NNRTI treatment (de Béthune, 2010). Therefore, it is essential to have additional NNRTIs developed to effectively treat patients with NNRTI-resistant HIV (De Clercq, 2004).

BILR 355 is a second generation NNRTI (Boone, 2006; Bonneau et al., 2005). The in vitro results showed that the EC$_{50}$ of BILR 355 was 0.26 ng/mL (0.59 nM) against wild type HIV-1 and ranged from 1.5 to 13 ng/mL (3.4 nM to 29 nM) against clinically common single and double NNRTI mutations (Bonneau et al., 2005). A clinical study revealed that BILR 355 was rapidly absorbed after a single oral dose in healthy volunteers with the mean time to maximum drug concentration (T$_{max}$) of 0.5 to 1.5 h. However, thereafter the concentrations of BILR 355 rapidly declined, resulting in a short terminal half-life (T$_{1/2}$) of approximately 2 to 4 h (Huang et al., 2008). Our in vitro data demonstrated significant metabolism of BILR 355 by cytochrome P450 (CYP) 3A. Thus, in an attempt to overcome the resulting low exposure in humans, BILR 355 was administered with ritonavir (RTV), a potent CYP 3A inhibitor previously used to...
boost plasma concentrations and to increase T1/2 of HIV protease inhibitors (Hull and Montaner, 2011). With concomitant administration of RTV, the T1/2 of BILR 355 increased to approximately 10 to 16 hours and the maximum plasma concentration (Cmax) increased 2- to 5-fold with the area under the plasma concentration-time curve from time zero to infinity (AUC0-∞) increasing 15- to 30-fold (Huang et al., 2008); a profile more consistent with once daily dosing.

However, this combination created an unexpected problem, a disproportionate human metabolite (BILR 516), as a result of significant metabolic switching of BILR 355. The current manuscript describes the in vitro metabolism of BILR 355 by CYP3A and identification of the metabolites resulting from metabolic switching due to inhibition of CYP3A by the concomitantly administered RTV.
Materials and Methods

Chemicals, Reagents and Other Materials. BILR 355, BILR 402, BILR 516, INLA 1678, BI 211136, BIRK 122, EXLS 1451, INRF105, BILR 564, BI 212580, and BI 212578 were synthesized at Boehringer Ingelheim Pharmaceuticals, Inc (Ridgefield, CT) and their structures are shown in supplemental data. BILR 483 (D₂-BILR 355 with three deuterium on the methyl group) and nevirapine were also synthesized at Boehringer Ingelheim Pharmaceuticals, Inc. NADPH was purchased from Sigma-Aldrich (St. Louis, MO). All other reagents and solvents were of analytical grade or higher purity and were obtained from commercial suppliers. Human liver microsomes (HLMs) from individual donors and pooled HLMs from 15 donors were obtained from Celsis In Vitro Technologies (Baltimore, MD). All rCYPs (produced in baculovirus-infected insect cells) and control insect cell microsomes were obtained from BD Biosciences (Woburn, MA). The microsomes and rCYPs were stored at -80 °C until used.

Kₘ and Vₘₐₓ Determination of BILR 355 Metabolism BILR 355 was incubated with HLMs from four individual donors to assess apparent Kₘ and Vₘₐₓ values. Incubations were carried out in 0.1 M potassium phosphate buffer (containing 5 mM of MgCl₂, pH 7.4). Microsomes were pre-incubated with BILR 355 for 5 minutes at 37°C. The reaction was initiated by the addition of NADPH at a final concentration of 2 mM. The final concentrations of microsomes were 0.5, 1, or 2 mg/mL. The final incubation volume was 0.9 mL and the final organic solvent concentration in each reaction did not exceed 1%. Aliquots (100 µL) were transferred from the incubation mixtures at various time intervals (0, 0.25, 1, 2, 3, 5, 10, 15, 30 min) and quenched with an equal volume of acetonitrile. Subsequently, 100 µL of BILR 483 in acetonitrile at a concentration of 0.5 µM was added as the internal standard to each well. All samples were centrifuged at 1600 g for 10 min and the supernatants were analyzed by LC-
MS/MS (high performance liquid chromatography-tandem mass spectrometry). The experiment was performed in duplicate for each lot of HLMs.

**Metabolism of BILR 355 by Recombinant CYP Isoforms.** BILR 355 (1 or 10 µM) was incubated with recombinant CYP isoforms (rCYP1A2, rCYP2B6, rCYP2C9, rCYP2C19, rCYP2D6, and rCYP3A4) in a 0.1 M potassium phosphate buffer with 5 mM of MgCl₂ at pH 7.4. The final rCYP concentrations were 50 pmol/mL and the total incubation volume was 800 µL. The reactions were pre-incubated at 37 ºC for 5 min before the addition of NADPH at a final concentration of 2 mM. Control samples were prepared identically, except that non-transfected insect cell microsomal protein was substituted for individual recombinant CYP isoforms. The depletion of BILR 355 was monitored over a full time course of 30 min. Aliquots of incubation mixtures were transferred at 0.5, 2, 5, 10, 15, 20, 30 min and quenched with acetonitrile containing BILR 483 as internal standard. The plates were centrifuged and the supernatant was analyzed by LC-MS/MS. The experiment was performed in duplicate.

**Inhibition of BILR 355 Metabolism in HLMs by Isoform-Selective Inhibitors.** BILR 355 (1 µM) was incubated with pooled HLMs in the presence and absence of isoform-selective chemical inhibitors. The chemical inhibitors were furafylline (30 µM) for CYP1A2, sulfaphenazole (5 µM) for CYP2C9, tranylcypromine (100 µM) for CYP2C19, quinidine (5 µM) for CYP2D6, and ketoconazole (1 µM and 3 µM) for CYP3A4 (Chauret et al., 1997; Bourrié et al., 1996). The incubation conditions and sample analysis were as described above for HLMs. The experiment was performed in triplicate.

**Metabolite Profiling of BILR 355 in Clinical Samples.** Human plasma samples were obtained from six healthy male volunteers administered 150 mg BILR 355 with 100 mg RTV
Steady state levels of BILR 355 were achieved by Day 7 and only one dose of BILR 355 and RTV was given on day 7 to obtain a full PK profile. Intensive PK samples were taken on Day 1 at 0, 0.25, 0.5, 0.75, 1, 1.5, 2, 2.5, 3, 4, 6, 9, and 12 h and on Day 7 at 0, 0.25, 0.5, 0.75, 1, 1.5, 2, 2.5, 3, 4, 6, 9, 12, 16, 24, 48, and 72 h following drug administration. Plasma samples from different subjects were pooled at Tmax (1.5 h) and two later time points (6 h and 24 h) on day 7. The zero hour samples at Day 1 were also pooled and used as control samples for metabolite identification. The pooled plasma samples were vortexed thoroughly and 300 µL aliquots were transferred to Eppendorf tubes. Nine hundred microliters of methanol was added to the plasma aliquot. The mixture was vortexed thoroughly and centrifuged at 17,000 g for 5 min. The supernatant was then dried down under nitrogen at room temperature. The residue was reconstituted in 200 µL water/methanol (50:50; v:v). The reconstituted sample was centrifuged at 17,000 g for 5 min and the supernatant was used for metabolite identification by LC-MS/MS. After the metabolites were identified, nine metabolites with authentic standards, including INLA 1678, BI 211136, BIRK 122, EXLS 1451, INRF 105, BILR 564, BILR 516, BI 212578, and BI 212580, were quantitated by LC-MS/MS using the plasma samples at selected time points. Nevirapine was used as the internal standard at a final concentration of 0.1 µM for metabolite quantitation.

Equipment and Chromatographic Conditions for Quantitation and Metabolite Identification of BILR 355. For quantitation of BILR 355 in in vitro incubations, an LC-MS/MS system consisting of a Gilson 215 liquid handler autosampler (Gilson, Inc., Middleton, WI), two Series 200 Micro pumps (Perkin Elmer, Waltham, MA), and a Applied Biosystems MDS Sciex API365 triple quadrupole mass spectrometer (Applied Biosystems/Sciex, Thornhill, Ontario, Canada) was used. HPLC column used was a Waters Symmetry C18 (2 x 50 mm, 3.5
μm particle size, Waters, Milford, MA). Mobile phase compositions were mobile phase A: water/acetonitrile/acetic acid (95:5:0.05 v/v/v) and mobile phase B: water/acetonitrile/acetic acid (5:95:0.05 v/v/v). BILR 355 and the isotope labelled internal standard, BILR 483, eluted at 1.7 min with a gradient of mobile phase B (maintain at 15% B for 0.5 min, increase from 15% B to 85% B in 3 min, and then keep at 85% B for 0.5 min) at a flow rate of 0.25 mL/min. The mass spectrometer was optimized for BILR 355 with an IonSpray voltage of 5.2 kV, an ion source temperature of 350 ºC, an orifice plate voltage of 35 V, a ring voltage of 300 V, and nebulizer gas of 13 L/min. The multiple reaction monitoring transitions requested for BILR 355 and the internal standard BILR 483 were \( m/z \) (mass to charge ratio) \( 442 \rightarrow 281 \) and \( m/z \) \( 445 \rightarrow 284 \), respectively.

Different LC-MS/MS instruments were used for identification and quantitation of metabolites of BILR 355. The instruments consisted of an SIL-5000 autosampler and two LC-10AD vp pumps (Shimadzu Scientific Instrument, Norwell, MA) connected with a Applied Biosystems MDS Sciex 4000 QTrap mass spectrometer (Applied Biosystems/Sciex, Thornhill, Ontario, Canada). An Atlantis dC18 column (Waters, Milford, MA) was used (3.9 × 150 mm, 3 μm particle size). Mobile phases were similar to previously described except that the concentration of acetic acid was 0.1%. For metabolite identification, the gradients were the following: mobile phase B 0-1% over 10 min, 1-3% over 1 min, 3-12% over 19 min, 12-20% over 20 min, 20-29% over 25 min, 29-70% over 8 min, 70-100% over 2 min at a flow rate of 0.7 mL/min. For metabolite quantitation of INLA 1678, BI 211136, BIRK 122, EXLS 1451, INRF 105, BILR 564, BILR 516, a 20-min gradient with mobile phase B increasing from 15% to 55% was used at a flow rate of 0.7 mL/min. The multiple reaction monitoring transitions requested and the retention times for metabolites are listed below: INLA 1678 (\( m/z \) 412→267; 7.3 min), BI 211136 (\( m/z \) 299...
→271; 7.6 min), BIRK 122 (m/z 299→271; 9.0 min), EXLS 1451 (m/z 414→253; 9.7 min), INRF 105 (m/z 313→285; 9.9 min), BILR 564 (m/z 428→267; 10.9 min), BILR 516 (m/z 442→281; 16.1 min). The internal standard nevirapine was monitored at m/z 267→226 (8.0 min). For metabolite quantitation of BI 212580 and BI 212578, a 5-min gradient was used (mobile phase B increased from 45 to 55% in 5 min at a flow rate of 0.7 mL/min). The multiple reaction monitoring transitions for BI 212580, BI 212578, and the internal standard nevirapine were m/z 458→253, m/z 428→267, and m/z 267→226, respectively and their retention times were 2.5, 3.5, and 2.8 min, respectively. The mass spectrometer parameters for above analyses included an IonSpray voltage at 4.5 kV, a declustering potential at 50 V, an ion source temperature at 550ºC, and collision energy at 30 V.

Data Analysis for Kinetic Parameters and Clearance Determination  Apparent kinetic parameters for BILR 355 were determined using substrate depletion. The initial rate of BILR 355 depletion in HLMs was estimated using a linear regression of compound remaining at several initial time points demonstrating an acceptable linearity with r²>0.9. The apparent kinetic parameters for the depletion of BILR 355 by each HLM lot were determined by non-linear regression analysis of the rate based on the Michaelis-Menten equation using the statistical program GraFit Version 5 (Erithacus Software Ltd., Horley, UK). The K_m and V_max values obtained were used to calculate the intrinsic clearance (Cl_int, in vitro) according to Cl_int, in vitro = V_max/K_m, where V_max is the normalized maximum reaction velocity in pmol/min/mg of HL microsomal protein and K_m is the apparent Michaelis-Menten constant (Obach et al., 1997). The following constants were used to estimate Cl_int, in vivo from Cl_int, in vitro: a value of 45 mg of microsomal protein/g of liver (Houston, 1994), 1800 g of human liver weight, and average 70 kg of human body weight (Davies and Morris, 1993). To calculate the hepatic clearance, the well
stirred model was applied and $CL_{h}$ (mL/min/kg) was calculated using the following equation (Pang and Rowland, 1977):

$$CL_{h} = \frac{Q_{h} \times f_{u} \times CL_{int}}{Q_{h} + f_{u} \times CL_{int}}$$

$Q_{h}$ = hepatic blood flow in human (20.7mL/min/kg) (Davies and Morris, 1993), $f_{u}$ = fraction of unbound drug (assume $f_{u} = 1$).
Results

Apparent Michaelis-Menten Kinetic Parameters of BILR 355 Metabolism in HLMs.

Four lots of pre-characterized HLMs were selected based on their testosterone 6β-hydroxylase activities representing a range from low to high CYP3A4 activities from available individual lots of human liver microsomes. Protein linearity was evaluated. Rates of metabolism were linear with concentrations of HLMs over the range of 0.5 mg/mL to 2 mg/mL microsomal protein. Therefore, the amount of microsomal protein in the $K_m$ and $V_{max}$ assay was adjusted within the range of 0.5 to 2 mg/mL, so that the initial rate of the metabolism could be practically measured for each lot of HLMs. The initial rate of BILR 355 depletion was measured at various substrate concentrations (0.25, 0.5, 1, 2, 4 and 8 µM). The apparent $K_m$ and $V_{max}$ values for each HLM lot were calculated using non-linear regression analysis based on the Michaelis-Menten equation. The mean $K_m$ and $V_{max}$ values are 0.79 µM and 111 pmol/min/mg, respectively, as summarized in Table 1. Intrinsic clearance and hepatic clearance values calculated from each lot of HLMs are also listed in Table 1.

CYP Reaction Phenotyping. The metabolism of BILR 355 by various rCYP isoforms (CYP1A2, CYP2B6, CYP2C9, CYP2C19, CYP2D6, and CYP3A4) was demonstrated by measuring the time-dependent compound depletion in the presence of NADPH. BILR 355 was significantly metabolized only by recombinant human CYP3A4. The initial rate of BILR 355 depletion was calculated from a linear regression of compound remaining at several initial time points demonstrating an acceptable linearity with $r^2 > 0.9$. The average initial rate of BILR 355 depletion by rCYP3A4 was $2.50 \pm 0.21$ and $2.59 \pm 0.02$ nmol/min/nmol CYP at initial substrate concentrations of 1 and 10 µM, respectively. Isoform-selective inhibitors of the five major drug metabolizing CYP isoforms were used to investigate their inhibitory effects on BILR 355
metabolism by HLMs. In the presence of a selective inhibitor of CYP3A4, ketoconazole at 1 and 3 µM, the metabolism of BILR 355 was inhibited by about 70% and 100%, respectively. The other chemical inhibitors did not cause any significant inhibitory effects. Furthermore, as shown in Table 1, \( V_{\text{max}} \) of BILR 355 metabolism with HLMs ranged from 35 to 230 pmol/min/mg microsomal protein. The \( V_{\text{max}} \) values correlated well with the testosterone 6β-hydroxylase activities (Table 1) of each HLM lot reported from the vendor \((r^2=0.96)\). There were no correlations with other reported CYP-selective probe activities, including phenacetin O-deethylation activity (CYP1A2), coumarin 7-hydroxylase activity (CYP2A6), mephenytoin 4-hydroxylase activity (CYP2C19), and dextromethorphan O-demethylation activity (CYP2D6) (data not shown).

**Identification of Metabolites of BILR 355 in Clinical Samples.** A total of eighteen metabolites were detected and a representative chromatogram of the samples is shown in Fig. 1, following molecular ion extraction of the identified metabolites from an enhanced MS scan chromatogram. Only eight metabolites are shown in Fig. 1. The peak intensities of the other metabolites are too low and thus their peaks are not visible in the ion extraction chromatogram. The metabolites are labelled M1 through M18 according to their respective order of elution from the HPLC column (Table 2). The possible structures of the metabolites are proposed based on their molecular ions and product ion spectra. Due to space limitation, only the product ion spectra of the parent and the three metabolites of interest are shown in Fig. 2. The fragmentation of the parent and the rationale to elucidate the structures of the three metabolites are described below.

BILR 355: BILR 355 eluted from the HPLC column with a retention time of 70.0 min. The product ion MS/MS spectrum of BILR 355 (Fig. 2A) was used as a reference to compare with
the metabolite spectra. The protonated molecular ion of BILR 355 had \( m/z \) of 442. The product ion spectrum contains base ions at \( m/z \) 281 and 253. The product ion at \( m/z \) 281 arises from the loss of the quinoline N-oxide from the protonated parent ion. This is followed by a subsequent loss of the ethyl group from the diazepinone ring, resulting in an ion at \( m/z \) 253.

M18: The peak of the metabolite M18 eluted at approximately 79.9 min. This metabolite had a protonated molecular ion at \( m/z \) 442, which was the same as that of the parent compound. However, this metabolite eluted later than the parent compound, suggesting it was an isomer of BILR 355. The product ion spectrum of M18 yielded the same product ions at 281 and 253 as the parent compound (Fig. 2B), which indicated the molecular composition on the dipyridodiazepinone moiety and the quinoline moiety remained unchanged. It is known that N-oxides are susceptible to reduction; therefore, it is possible that the N-oxide is reduced and further oxidation could occur at a position other than the nitrogen on the quinoline moiety, resulting in the formation of an isomer of BILR 355. The structure of M18 was further confirmed as BILR 516 based on the comparison with an authentic standard. The metabolite and the authentic standard had the same retention time, molecular weight, and MS/MS pattern.

Similar to the relationship between BILR 355 and BILR 516, two other pairs of metabolites were identified, i.e. M7-M14 and M13-M17. Each pair of metabolites had the same protonated molecular ions, the same product ions, but different retention times. Therefore, it was proposed that M7 and M13, which eluted earlier, had the quinoline N-oxide structure similar to BILR 355 and M14 and M17, which eluted later, had the quinolone structure similar to BILR 516.

M14: This metabolite had a protonated molecular ion at \( m/z \) 458, which is 16 amu higher than that of BILR 516, suggesting an addition of oxygen. Its product ion spectrum showed
abundant product ions at m/z 253, 279, 297, and 440 (Fig. 2C). The product ion at m/z 297 indicated possible oxidation on the dipyridodiazepinone moiety. Two abundant product ions at m/z 440 and m/z 279 could originate from the loss of H₂O from the molecular ion and the product ion at m/z 297, respectively. Similar to BILR 516, a product ion at m/z 253 was detected for M14, which suggested the dipyridodiazepinone ring was unchanged. So the likely oxidative position would be the ethyl group connected to the diazepinone ring. In addition, oxidation at the terminal carbon is relatively more stable compared to the oxidation at the α-position. Therefore, M14 was tentatively identified as a hydroxylated BILR 516 with oxidation at the terminal carbon of the ethyl group on the diazepinone moiety.

M17: This metabolite had a protonated molecular ion at m/z 428, corresponding to a loss of 14 amu form BILR 516. The product ion spectrum of M17 showed abundant product ions at m/z 239 and 267 (Fig. 2D), which were also 14 amu less than the corresponding product ions of BILR 516. So it is likely that M17 is formed after demethylation on the diazepinone ring from BILR 516.

The retention times and characteristic product ions of the parent and all metabolite are listed in Table 2. In addition to BILR 516, standards of nine metabolites were also synthesized, including INLA 1678, BI 211136, BIRK 122, EXLS 1451, INRF 105, BILR 564, BILR 402, BI 212580 (M14), and BI 212578 (M17) based on proposed structures. Both ¹H and ¹³C NMR data of the parent and synthetic standards of the metabolites are listed in supplemental data. The structures of the ten metabolites were confirmed by comparison with synthetic standards based on the same retention times, molecular weight, and MS/MS patterns. The overall metabolic pathways of BILR 355 are proposed in Fig. 3. The names of the standards are also listed in the final column in Table 2 and next to the metabolite names in Fig. 3.
Quantitation of Metabolites of BILR 355 in Clinical Samples. The levels of the nine metabolites with authentic standards, including INLA 1678, BI 211136, BIRK 122, EXLS 1451, INRF 105, BILR 564, BILR 516, BI 212580, and BI 212578, were measured in clinical samples from individual subjects after the last dose of BILR 355 and RTV on day 7 (steady state). The quantitation result showed that INLA 1678, BI 211136, BIRK 122, EXLS 1451, INRF 105, BILR 564, BI 212580, and BI 212578 were minor metabolites of BILR 355, with plasma concentration of the tested metabolites less than 0.22 µM in all clinical samples. BILR 402 was also monitored in a separate study and has been shown to be a minor metabolite of BILR 355 with C_max of 0.18 µM (Boehringer Ingelheim Pharmaceuticals, Inc., Data on file).

The levels of BILR 516 were surprisingly high at steady state, exceeding levels of parent drug. The mean plasma concentration-time profiles of BILR 516 on day 1 and steady state are plotted in Fig. 4. The BILR 355 profiles are also shown in Fig. 4 for comparison purposes (Huang et al., 2009). The calculated C_max, area under the plasma concentration-time curve over the dosing interval (AUC0-τ), and T_1/2 of BILR 516 and BI 212578, and comparison of these values with those obtained for BILR 355 (Huang et al., 2009) are shown in Table 3.
Discussion

BILR355 is a second generation NNRTI developed to treat HIV-1 infection. BILR 355 has been shown to be a good substrate of CYP3A with a relatively low apparent $K_m$ of 0.79 µM and a high turnover in HLM. Extensive first pass metabolism by intestinal and hepatic CYP3A was proposed as an explanation for the low systemic exposure and short half-life following oral administration to humans (Huang et al., 2008). A favorable PK profile, with increased systemic exposure, was achieved after concomitant administration of the CYP3A inhibitor RTV (Huang et al., 2008) as seen with other HIV drugs (Hull and Montaner, 2011). An unexpected consequence of the inhibition of this primary clearance pathway was a metabolic switching leading to the formation of a disproportionate human metabolite.

First, the \textit{in vitro} metabolism of BILR 355 was evaluated in HLMs. As summarized in Table 1, the calculated intrinsic clearance of BILR 355 by HLMs appears to be fairly high. The average $CL_h/Q_h$ from four HLM lots was 82%, indicating that BILR 355 may go through extensive Phase I metabolism \textit{in vivo}. The apparent $K_m$ values for BILR 355 metabolism ranged from 0.63 µM to 1.14 µM with a mean value of 0.79 µM. In the clinical Phase Ia study with administration of single doses of BILR 355 from 12.5 mg to 100 mg, $C_{\text{max}}$ ranged from 14.8 ng/mL (0.03 µM) to 937 ng/mL (2.12 µM)(Huang et al., 2008), which bracketed the range of apparent $K_m$ values shown above. Therefore, it was not surprising to see the mean exposure ($AUC_{0-\infty}$) increase super-proportionally (71.8 ng.h/mL to 1,310 ng.h.mL) and the mean apparent clearance ($CL/F$) decrease (246 to 79.2 L/h) over this range of doses in the Phase Ia study (Huang et al., 2008). The observed nonlinearity in $AUC_{0-\infty}$ and $CL/F$ is likely due to saturation of the metabolism of BILR 355.
CYP3A4 was identified as the major CYP responsible for the metabolism of BILR 355, of the six major human CYP isoforms tested, as only rCYP3A4 caused a detectable depletion of BILR 355 and only ketoconazole (CYP3A4 specific inhibitor) resulted in significant inhibition of the metabolism of BILR 355 by HLMs (complete inhibition with 3 µM ketoconazole). In addition, the apparent $V_{\text{max}}$ values of BILR 355 metabolism correlated well with the reported CYP3A4 activity of the different lots of HLMs, but no correlation was observed with activities of CYP1A2, CYP2A6, CYP2C19, or CYP2D6. The depletion rates of BILR 355 in the presence of rCYP3A4 were similar at substrate concentrations of 1 µM and 10 µM. This finding suggests that the apparent $K_m$ for the CYP3A4-catalyzed metabolism of BILR 355 is less than 1 µM, which is consistent with the apparent $K_m$ values for BILR 355 metabolism in HLMs. Overall, the combination of the results from rCYP profiling, chemical inhibition, and correlation analysis supports that CYP3A4 plays an important role in the metabolism of BILR 355. RTV, a potent CYP3A inhibitor, was therefore used as a boosting agent for BILR 355 and effectively elevated the exposure of BILR 355 in humans (Huang et al., 2008), which further confirms BILR 355 is extensively metabolized by CYP3A. The involvement of CYP3A5 was not specifically tested in these studies. However, since there is typically an overlap in substrate specificity between CYP3A4 and CYP3A5, it is possible that CYP3A5 is also involved in the metabolism of BILR 355. Overall, the current phenotyping results support the concept that the increase of BILR 355 systemic exposure with concomitant administration of RTV is primarily due to inhibition of CYP3A.

It was expected that the levels of metabolites would decrease with concomitant administration of the CYP3A inhibitor RTV. Metabolite profiling studies for the Phase Ib plasma samples, in which RTV was administered with BILR 355, showed a total of eighteen metabolites. The
structures of ten of these metabolites were confirmed based on comparison with authentic standards. While almost all of these metabolites were minor based on quantitation using authentic standards or peak area estimations for metabolites without authentic standards, there was one metabolite found at substantial levels, BILR 516. This metabolite was not observed in human plasma when BILR 355 was administered along (Boehringer Ingelheim Pharmaceuticals, Inc., Data on file).

BILR 516 accumulated in humans over time (Fig. 4) which was attributable to its long T$_{1/2}$ (54.5 h) (Table 3). The T$_{1/2}$ of BILR 516 is significantly longer than the T$_{1/2}$ of the parent compound (14.7 h) (Table 3). As a result, the exposure of BILR 516 was even higher than the exposure of the parent drug at steady-state (Fig. 4). Since the exposure of BILR 516 was greater than 10% of the total drug-related materials, it was identified as a major human metabolite (FDA, 2008).

Based on both the FDA guidance for Industry: Safety Testing of Drug Metabolites and the later published 2009 ICH guidance M3 (R2)(FDA, 2008; EMEA, 2009), the coverage of BILR 516 in Toxicology species needed to be assessed. Additional studies showed that the levels of BILR 516 were very low in Sprague Dawley rats and Beagle dogs, the two general Toxicology species for BILR 355 (Boehringer Ingelheim Pharmaceuticals, Inc., data on file). As such, BILR 516 was defined as a disproportionate human metabolite. A better understanding of the formation of BILR 516 was needed in order to help select an appropriate Toxicology species (Ref. companion paper).

Similarly, BI 212580 and BI 212578 have the same quinolone structure as BILR 516. It is possible that both metabolites may have longer half-lives similar to BILR 516 with a potential to
accumulate in the body over time and become major metabolites. Therefore, both metabolites were synthesized based on the proposed structures and their exposure was assessed using the authentic standards and compared with the exposure of the parent. The pharmacokinetic parameters of BI 212578 are included in Table 3. The concentrations of BI 212580 were lower than the Limit of Quantitation (0.0426 µM) at most time points and thus its pharmacokinetic parameters could not be calculated. BI 212578 does have a long T1/2 of 33 h, as anticipated. Both BI 212580 and BI 212578 were confirmed as minor metabolites and no further assessment was warranted.

Overall, the exposure of BILR 355 was significantly improved by concomitant administration of RTV, mainly due to suppression of CYP3A-mediated metabolism of BILR 355. However, as a consequence, a disproportionate human metabolite, BILR 516, emerged, which was not detected in human plasma samples when BILR 355 was administered alone. The resulting consequence of a new major metabolite posed several challenges for drug development. This paper raises a concern on potential metabolic switching with concomitant administration of a potent boosting agent or any potent inhibitors of major metabolic pathways. The metabolic pathways for the formation of BILR 516 from BILR 355 turned out to be rather complicated as outlined in the accompanying publication (Ref. companion paper). Briefly, BILR 516 is formed by sequential reactions mediated by two non-CYP enzymes. First, BILR 355 is transformed to BILR 402 by gut bacteria through reduction of the N-oxide and then BILR 402 is metabolized to BILR 516 by aldehyde oxidase.
Acknowledgements:

We thank Dr. Fenglei Huang for providing the $C_{\text{max}}$ data of BILR 402 and thank Dr. Timothy S. Tracy for scientific advice and review of the manuscript.
Authorship Contributions:

Participated in research design: Li, Lai, Tweedie

Conducted experiments: Li, Lai, Whitcher-Johnstone

Contributed new reagents or analytic tools: Busacca, Eriksson, Lorenz

Performed data analysis: Li, Lai,

Wrote or contributed to the writing of the manuscript: Li, Tweedie
References


Footnotes:

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This work was presented at the 2009 ISSX meeting.


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W. George Lai (Current address: Drug Disposition, Eisai Research Institute, Andover, MA)
Legends for Figures:

FIG. 1. Representative LC-MS chromatogram with extracted ions of the identified metabolites

FIG. 2. Product ion spectra of BILR 355 (A), and three metabolites, M18 (B), M14 (C), and M17 (D)

FIG. 3. Proposed metabolite structures and metabolic pathways of BILR 355 in humans

FIG. 4. Plasma concentration – time profiles of BILR 355 and BILR 516 on Day 1 and Day 7 (steady state) in humans
Table 1 Michaelis-Menten kinetic parameters for BILR 355 metabolism as measured by compound depletion in human liver microsomes (n=2).

<table>
<thead>
<tr>
<th>HLM lot</th>
<th>Testosterone 6β-hydroxylase activities (nmol/min/mg)</th>
<th>HLM protein conc. (mg/mL)</th>
<th>Apparent $K_m$ (µM)</th>
<th>$V_{max}$ (pmol/min/mg of HLM protein)</th>
<th>$CL_{int, in vitro}$ (mL/min/mg of HLM protein)</th>
<th>$CL_{int, in vivo}$ (mL/min/kg of body weight)</th>
<th>$CL_h$ (mL/min/kg of body weight)</th>
<th>$CL_h/Q_h$ (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1028</td>
<td>0.7</td>
<td>2</td>
<td>0.73</td>
<td>46.5</td>
<td>0.0637</td>
<td>73.7</td>
<td>16.2</td>
<td>78.1</td>
</tr>
<tr>
<td>1013</td>
<td>1.8</td>
<td>2</td>
<td>1.14</td>
<td>35.0</td>
<td>0.0307</td>
<td>35.5</td>
<td>13.1</td>
<td>63.2</td>
</tr>
<tr>
<td>1008</td>
<td>4.6</td>
<td>1</td>
<td>0.64</td>
<td>133</td>
<td>0.208</td>
<td>240</td>
<td>19.1</td>
<td>92.1</td>
</tr>
<tr>
<td>1003</td>
<td>7.9</td>
<td>0.5</td>
<td>0.63</td>
<td>230</td>
<td>0.365</td>
<td>422</td>
<td>19.7</td>
<td>95.3</td>
</tr>
<tr>
<td>Mean ± SD</td>
<td></td>
<td></td>
<td>0.79±0.24</td>
<td>111±91</td>
<td>0.167±0.153</td>
<td>193±177</td>
<td>17.0±3.0</td>
<td>82.2±14.7</td>
</tr>
</tbody>
</table>
Table 2 List of retention times, molecular ions, and characteristic product ions of BILR 355 and its metabolites

<table>
<thead>
<tr>
<th>Metabolite No.</th>
<th>Retention time (min)</th>
<th>Molecule ion (m/z)</th>
<th>Characteristic product ions (m/z)</th>
<th>Structure confirmed</th>
<th>Name of the synthetic standard*</th>
</tr>
</thead>
<tbody>
<tr>
<td>M1</td>
<td>39.5</td>
<td>285</td>
<td>226, 240, 257</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M2</td>
<td>42.6</td>
<td>299</td>
<td>271</td>
<td>Yes</td>
<td>BI 211136</td>
</tr>
<tr>
<td>M3</td>
<td>46.0</td>
<td>412</td>
<td>239, 267</td>
<td>Yes</td>
<td>INLA 1678</td>
</tr>
<tr>
<td>M4</td>
<td>46.9</td>
<td>442</td>
<td>269, 297</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M5</td>
<td>48.2</td>
<td>299</td>
<td>271</td>
<td>Yes</td>
<td>BIRK 122</td>
</tr>
<tr>
<td>M6</td>
<td>50.4</td>
<td>442</td>
<td>253, 281</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M7</td>
<td>50.5</td>
<td>458</td>
<td>253, 279, 297, 440</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M8</td>
<td>51.4</td>
<td>313</td>
<td>285</td>
<td>Yes</td>
<td>INRF 105</td>
</tr>
<tr>
<td>M9</td>
<td>52.6</td>
<td>426</td>
<td>253, 281</td>
<td>Yes</td>
<td>BILR 402</td>
</tr>
<tr>
<td>M10</td>
<td>52.9</td>
<td>634</td>
<td>253, 269, 297, 458, 473</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M11</td>
<td>56.0</td>
<td>634</td>
<td>269, 297, 279, 458, 473</td>
<td>Yes</td>
<td>EXLS 1451</td>
</tr>
<tr>
<td>M12</td>
<td>56.5</td>
<td>634</td>
<td>279, 458, 440</td>
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<td></td>
</tr>
<tr>
<td>M13</td>
<td>61.6</td>
<td>634</td>
<td>297, 458, 440</td>
<td>Yes</td>
<td>BILR 564</td>
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<tr>
<td>M14</td>
<td>61.7</td>
<td>634</td>
<td>253, 279, 297, 440</td>
<td>Yes</td>
<td>BI 212580</td>
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<tr>
<td>M15</td>
<td>63.8</td>
<td>634</td>
<td>297, 440, 458</td>
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<td></td>
</tr>
<tr>
<td>M16</td>
<td>65.9</td>
<td>634</td>
<td>253, 281, 458</td>
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<td></td>
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<tr>
<td>M17</td>
<td>75.2</td>
<td>428</td>
<td>239, 267</td>
<td>Yes</td>
<td>BI 212578</td>
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<td>M18</td>
<td>79.9</td>
<td>442</td>
<td>253, 281</td>
<td>Yes</td>
<td>BILR 516</td>
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<tr>
<td>BILR 355</td>
<td>70.0</td>
<td>442</td>
<td>253, 281</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*1H NMR and 13C NMR data of the synthetic standards are listed in the supplemental data.
Table 3: Pharmacokinetic parameters of BILR 355, BILR 516, and BI 212578 after 7 Days dosing of BILR 355 (150 mg)/RTV (100mg) (b.i.d.) in humans

<table>
<thead>
<tr>
<th>Parameters</th>
<th>BILR 355 Mean ± SD</th>
<th>BILR 516 Mean ± SD</th>
<th>BILR 516 /BILR 355 (%)</th>
<th>BI 212578 Mean ± SD</th>
<th>M17/BILR 355 (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C&lt;sub&gt;max&lt;/sub&gt;, µM</td>
<td>3.40 ± 0.870</td>
<td>5.37 ± 1.27</td>
<td>158</td>
<td>0.200 ± 0.042</td>
<td>5.88</td>
</tr>
<tr>
<td>AUC&lt;sub&gt;0-τ&lt;/sub&gt;, µmol.h/L</td>
<td>28.2 ± 6.46</td>
<td>57.1 ± 14.6</td>
<td>202</td>
<td>2.15 ± 0.441</td>
<td>7.62</td>
</tr>
<tr>
<td>Half life, h</td>
<td>14.7 ± 0.80</td>
<td>54.5 ± 23.3</td>
<td>-</td>
<td>33.2 ± 7.25</td>
<td>-</td>
</tr>
</tbody>
</table>
Fig. 2
Fig. 3
Fig. 4

Day 1

Day 7
Supplemental Data

Journal: Drug Metabolism & Disposition

Metabolic Switching of BILR 355 in the Presence of Ritonavir I: Identifying an Unexpected Disproportionate Human Metabolite

Yongmei Li, W. George Lai, Andrea Whitcher-Johnstone, Carl A. Busacca, Magnus C. Eriksson, Jon C. Lorenz, Donald J. Tweedie

$^1$H and $^{13}$C NMR Data of BILR 355 and ten metabolites

BILR 355:

$^1$H NMR (400 MHz, CDCl$_3$) $\delta$: 8.69 (d, $J = 7.0$Hz, 1H, ), 8.62 (d, $J = 8.7$Hz, 1H), 8.39 (d, $J = 2.4$Hz, 1H), 8.21 (d, $J = 8.4$Hz, 1H), 8.18 (dd, $J = 4.7$, 1.6Hz, 1H), 7.8 (dd, $J = 5.4$, 5.4Hz, 1H), 7.70 (dd, $J = 7.2$, 7.2Hz, 1H), 7.47 (dd, $J = 8.0$, 1.6Hz, 1H), 7.07 (dd, $J = 7.9$, 4.7Hz, 1H), 6.78 (d, $J = 7.0$Hz, 1H), 4.42 (dd, $J = 6.2$, 6.2Hz, 2H), 4.18 (q, $J = 6.9$ Hz, 2H), 3.50 (s, 3H), 3.22 (dd, $J = 6.2$, 6.2Hz, 2H), 1.24 (t, $J = 7.0$Hz, 3H). $^{13}$C NMR (125 MHz, DMSO-d$_6$) $\delta$: 166.3 (C6), 157.7 (C15), 154.0 (C12), 151.3 (C21), 151.1 (C9), 144.0 (C2), 141.2
(C7), 140.5 (C30), 135.4 (C23), 131.7 (C4), 131.1 (C13), 130.6 (C26), 128.9 (C8), 128.0 (C27), 122.3 (C28), 122.1 (C29), 120.2 (C3 + C14), 119.2 (C25), 101.9 (C22), 68.9 (C20), 40.4 (C17), 36.7 (C16), 30.6 (C19), 13.4 (C18). Anal. Calcd for C_{25}H_{23}N_{5}O_{3}; [10.7% H_{2}O]: C, 60.60; H, 5.90; N, 14.13. Found C, 60.46; H, 5.45; N, 14.22.

**BI 211136, M2:**

\[ \text{\textit{H}} \text{ NMR (500 MHz, DMSO-}d_{6} \text{)} \delta: 10.44 \text{ (s, 1H),} \\
8.35 \text{ (d, } J = 2.2 \text{ Hz, 1H),} 8.15 \text{ (dd, } J = 1.0, 4.5 \text{ Hz,} \\
1H), 7.97 \text{ (d, } J = 2.2 \text{ Hz, 1H),} 7.47 \text{ (dd, } J = 1.1, 7.8 \text{ Hz, 1H),} \\
7.14 \text{ (dd, } J = 4.7, 7.8 \text{ Hz, 1H),} 4.08 \text{ (q, } J = \\
7.0 \text{ Hz, 2H),} 3.61 \text{ (s, 2H),} 1.15 \text{ (t, } J = 7.0 \text{ Hz, 3H).} \]

\[ \text{\textit{C}} \text{ NMR (125 MHz, DMSO-}d_{6} \text{)} \delta: \\
172.5 \text{ (s),} 166.9 \text{ (s),} 157.3 \text{ (s),} 151.8 \text{ (d),} 151.7 \text{ (s),} 143.4 \text{ (d),} 141.3 \text{ (d),} 129.8 \text{ (d),} 127.0 \text{ (s),} 126.2 \text{ (s),} 120.1 \text{ (x),} 120.0 \text{ (x),} 40.8 \text{ (x),} 36.6 \text{ (x),} 13.6 \text{ (x).} \]

**INLA 1678, M3:**

\[ \text{\textit{H}} \text{ NMR (500 MHz, CDCl}_{3} \text{)} \delta: 9.07 \text{ (s,} \\
1H), 8.71 \text{ (d, } J = 5.2 \text{ Hz, 1H),} 8.49 \text{ (d, } J = \\
2.4 \text{ Hz, 1H),} 8.16-8.21 \text{ (m, 3H),} 8.01 \text{ (d, } J \\
= 8.5 \text{ Hz, 1H),} 7.66 \text{ (dt, } J = 7.6, 1.4 \text{ Hz,} \\
1H), 7.47 \text{ (t, } J = 7.6 \text{ Hz, 1H),} 7.33 \text{ (dd, } J = 7.8, 1.4 \text{ Hz, 1H),} \\
6.98 \text{ (dd, } J = 7.6, 4.8 \text{ Hz,} \\
1H), 6.68 \text{ (d, } J = 5.2 \text{ Hz, 1H),} 4.37 \text{ (t, } J = 6.4 \text{ Hz, 2H),} 4.23 \text{ (q, } J = 7.0 \text{ Hz, 2H),} 3.23 \text{ (t, } J \\
= 6.3 \text{ Hz, 2H),} 1.25 \text{ (t, } J = 7.0 \text{ Hz, 3H).} \]
$^{13}$C NMR (125 MHz, CDCl$_3$) $\delta$: 169.1, 161.1, 158.5, 152.5, 152.3, 151.2, 149.2, 144.5, 129.9, 129.3, 128.9, 128.2, 126.5, 125.8, 121.8, 121.2, 120.0, 119.7, 100.6, 68.2, 41.7, 31.7, 13.8.

**BIRK 122, M5:**

$^1$H NMR (400 MHz, CDCl$_3$) $\delta$: 8.21 (d, $J = 2.4$ Hz, 1H), 8.15 (dd, $J = 4.6$, 1.6 Hz, 1H), 7.91 (d, $J = 2.4$ Hz, 1H), 7.43 (dd, $J = 8.0$, 1.6 Hz, 1H), 7.05 (dd, $J = 8.0$, 4.6 Hz, 1H), 4.12 (br s, 2H), 3.75 (br s, 2H), 3.46 (s, 3H), 2.76 (t, $J = 6.5$ Hz, 2H), 2.63 (br s, 1H), 1.21 (t, $J = 7.0$ Hz, 3H). $^{13}$C NMR (100 MHz, CDCl$_3$) $\delta$: 168.0 (s), 158.4 (s), 155.2 (s), 151.5 (d), 144.5 (d), 141.3 (d), 131.8 (s), 130.9 (d), 129.5 (s), 120.8 (s), 119.8 (d), 62.9 (t), 41.2 (t), 37.6 (q), 35.3 (t), 13.7 (q). Anal. Calcd for C$_{16}$H$_{18}$N$_4$O$_2$: C, 64.41; H, 6.08; N, 18.78. Found C, 64.26; H, 6.15; N, 18.58.

**INRF 105, M8:**

$^1$H NMR (500 MHz, DMSO-d$_6$) $\delta$: 12.47 (s, 1H), 8.32 (d, $J = 2.0$ Hz, 1H), 8.20 (dd, $J = 4.6$, 1.5 Hz, 1H), 7.96 (d, $J = 2.3$ Hz, 1H), 7.81 (dd, $J = 8.0$, 1.6 Hz, 1H), 7.24 (dd, $J = 4.6$, 8.0 Hz, 1H), 4.08 (q, $J = 7.0$ Hz, 2H), 3.63 (s, 3H), 3.43 (s, 3H), 1.17 (t, $J = 7.0$ Hz, 3H). $^{13}$C NMR (125 MHz, DMSO-d$_6$) $\delta$: 172.2 (s), 166.3 (s), 157.5 (s), 154.0 (s), 151.5 (d), 144.5 (d), 141.3 (d), 131.8 (s), 130.9 (d), 129.5 (s), 120.8 (s), 119.8 (d), 62.9 (t), 41.2 (t), 37.6 (q), 35.3 (t), 13.7 (q).
151.3 (d), 144.1 (d), 141.6 (d), 131.7 (d), 131.1 (s), 126.0 (s), 120.3 (d), 120.1 (s), 40.5 (t), 36.7 (q), 36.3 (t), 13.4 (q). Anal. Calcd for C_{16}H_{16}N_{4}O_{3}: C, 61.53; H, 5.16; N, 17.94. Found: C, 61.28; H, 4.94; N, 17.56.

**BILR 402, M9:**

![BILR 402](image)

$^1$H NMR (400 MHz, MeOH-d$_4$, HCl salt) δ:
8.95 (d, $J = 6.7$ Hz, 1H), 8.48 (d, $J = 2.4$ Hz, 1H), 8.40 (d, $J = 8.5$ Hz, 1H), 8.15 (m, 2H), 8.07 (d, $J = 3.5$ Hz, 2H), 7.84 (m, 1H), 7.75 (dd, $J = 1.5$, 8.0 Hz, 1H), 7.47 (d, $J = 6.7$ Hz, 1H), 7.19 (dd, $J = 4.7$, 8.0 Hz, 1H), 4.76 (t, $J = 5.6$ Hz, 2H), 4.10 (m, 2H), 3.49 (s, 3H), 3.34 (t, $J = 6.1$ Hz, 2H), 1.18 (t, $J = 7.1$ Hz, 3H). $^{13}$C NMR (100 MHz, MeOH-d$_4$) δ: 170.2 (s), 169.2 (s), 160.2 (s), 156.0 (s), 152.8 (d), 147.6 (d), 145.9 (d), 142.8 (d), 140.5 (s), 136.1 (d), 133.3 (s), 133.1 (d), 130.3 (d), 130.0 (s), 124.7 (d), 122.3 (s), 122.2 (s), 121.7 (d), 121.4 (d), 103.8 (d), 72.9 (t), 42.18 (t), 37.8 (q), 32.2 (t), 14.0 (q). Anal Calcd for C$_{25}$H$_{24}$ClN$_{5}$O$_{2}$: C, 65.00; H, 5.24; N, 15.16. Found: C, 64.86; H, 5.09; N, 14.87.
EXLS 1451, M11:

1H NMR (500 MHz, DMSO-d$_6$) δ: 8.75 (s, 1H), 8.50 (d, J = 8.7 Hz, 1H), 8.45 (d, J = 6.7 Hz, 1H), 8.40 (s, 1H), 8.17 (s, 1H), 8.14 (d, J = 8.3 Hz, 1H), 8.06 (d, J = 4.2 Hz, 1H), 7.84 (t, J = 7.9 Hz, 1H), 7.76 (d, J = 7.7 Hz, 1H), 7.70 (t, J = 7.5 Hz, 1H), 7.18 (dd, J = 4.6, 7.8 Hz, 1H), 6.98 (d, J = 6.7 Hz, 1H), 4.41 (br t, J = 6.2 Hz, 2H), 3.37 (s, 3H), 3.18 (br t, J = 6.0 Hz, 2H). 13C NMR (125 MHz, DMSO-d$_6$) δ: 166.5 (s), 156.81 (s), 152.4 (d), 151.7 (s), 144.4 (d), 142.2 (d), 140.5 (s), 136.2 (d), 132.1 (d), 131.2 (d), 128.6 (s), 128.41 (s), 128.37 (d), 122.5 (d), 122.2 (s), 120.1 (d), 119.4 (d), 116.9 (s), 102.0 (d), 69.3 (t), 37.6 (q), 30.8 (t).

BILR 564, M13:

1H NMR (500 MHz, DMSO-d$_6$) δ: 10.50 (s, 1H), 8.54 (d, J = 2.4 Hz, 1H), 8.50 (d, J = 8.7 Hz, 1H), 8.46 (d, J = 6.9 Hz, 1H), 8.14 (d, J = 2.3 Hz, 2H), 8.13 (m, 1H), 7.83 (dt, J = 1.1, 8.2 Hz, 1H), 7.69 (t, J = 7.8 Hz, 1H), 7.46 (dd, J = 1.5, 7.8 Hz, 1H), 7.14 (dd, J = 4.7, 7.8 Hz, 1H), 6.99 (d, J = 7.0 Hz, 1H), 4.44 (t, J = 6.3 Hz, 2H), 4.08 (q, J = 7.1 Hz, 2H), 3.20 (t, J = 6.3 Hz, 2H), 1.14 (t, J = 7.0 Hz, 3H). 13C NMR (125 MHz, DMSO-d$_6$) δ: 167.0 (s), 157.3 (s), 151.72 (d), 151.70 (s), 151.4 (s), 143.4 (d), 141.0 (d), 140.5 (s), 135.4 (d), 130.6 (d), 129.7 (d), 128.8
BI 212580, M14:

\[
\begin{align*}
^1H \text{ NMR (}500 \text{ MHz, DMSO-}d_6\text{)} \delta: 11.32 \text{ (br s, 1H), 8.46 (d, } J = 2.3 \text{ Hz, 1H), 8.18 (dd, } J = 1.4, 4.6 \text{ Hz, 1H), 8.10 (d, } J = 2.3 \text{ Hz, 1H), 7.83 (dd, } J = 1.3, 7.9 \text{ Hz, 1H), 7.72 (d, } J = 7.8 \text{ Hz, 1H), 7.49 (dt, } J = 1.1, 8.2 \text{ Hz, 1H), 7.27-7.24 \text{ (m, 2H), } 7.13 \text{ (t, } J = 7.7 \text{ Hz, 1H), 5.87 \text{ (s, 1H), 4.64 (br s, 1H), 4.30 (t, } J = 5.8 \text{ Hz, 2H), 4.19 (br s, 2H), 3.59 (t, } J = 6.1 \text{ Hz, 2H), 3.42 (s, 3H), 3.15 \text{ (t, } J = 6.1 \text{ Hz, 2H).} \\
^13C \text{ NMR (}125 \text{ MHz, DMSO-}d_6\text{)} \delta: 166.3 \text{ (s), 163.1 \text{ (s), 162.0 \text{ (s), 157.7 \text{ (s), 154.0 \text{ (s), 151.1 \text{ (d), 143.9 \text{ (d), 141.4 \text{ (d), 138.6 \text{ (s), 131.9 \text{ (d), 131.1 \text{ (s), 130.9 \text{ (d), 129.0 \text{ (s), 122.2 \text{ (d), 121.3 \text{ (d), 120.4 \text{ (d), 120.1 \text{ (s), 115.1 \text{ (d), 114.4 \text{ (s), 97.3 \text{ (d), 68.3 \text{ (t), 58.7 \text{ (t), 48.4 \text{ (t), 36.9 \text{ (q), 30.5 \text{ (t).}}} 
\end{align*}
\]

BI 212578, M17:

\[
\begin{align*}
^1H \text{ NMR (}500 \text{ MHz, }1:1 \text{ DMSO-}d_6+\text{MeOH-}d_4\text{)} \delta: 8.50 \text{ (d, } J = 2.2 \text{ Hz, 1H), 8.14 (d, } J = 1.8 \text{ Hz, 1H), 8.12 \text{ (m, 1H), 7.79 (d, } J = 7.9 \text{ Hz, 1H), 7.50 (t, } J = 7.9 \text{ Hz, 1H), 7.46 (dd, } J = 1.2, 7.8 \text{ Hz, 1H), 7.29 (d, } J = 8.2 \text{ Hz, 1H), 7.16 \text{ (t, } J = 7.7 \text{ Hz, 1H), 7.11 (dd, } J = 4.7, 7.8 \text{ Hz, 1H), 5.90 \text{ (s, 1H), 4.33 (t, } J =
\end{align*}
\]
6.2 Hz, 2H), 4.14 (m, 2H), 4.11 (q, J = 7.1 Hz, 2H), 1.16 (t, J = 7.0 Hz, 3H). $^{13}$C NMR (125 MHz, 1:1 DMSO-$d_6$ + MeOH-$d_4$) δ: 167.7 (s), 164.2 (s), 163.1 (s), 158.0 (s), 152.4 (s), 152.2 (d), 143.9 (d), 141.5 (d), 138.9 (s), 131.5 (d), 130.1 (d), 129.5 (s), 127.5 (d), 122.8 (s), 122.1 (d), 120.9 (s), 120.3 (d), 115.7 (d), 115.2 (s), 97.3 (d), 68.9 (t), 41.4 (t), 31.1 (t), 13.8 (q).

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$^1$H NMR (400 MHz, DMSO-$d_6$) δ: 8.45 (d, J = 1.8 Hz, 1H, H9), 8.18 (d, J = 4.0 Hz, H2), 8.08 (d, J = 1.8 Hz, H7), 7.82 (d, J = 7.8 Hz, H4), 7.71 (d, J = 7.8 Hz, H28), 7.47 (t, J = 7.8 Hz, H26), 7.25 (m, H3 + H25), 7.11 (t, J = 7.8 Hz, H27), 5.85 (s, H22), 4.30 (br s, H20), 4.08 (br s, H17), 3.27 (s, H16), 3.16 (t, J = 6.6 Hz, H19), 1.21 (t, J = 7.0 Hz, H18). $^{13}$C NMR (100 MHz, DMSO-$d_6$) δ: 166.3 (C15), 163.1 (C23), 163.0 (C21), 157.6 (C15), 153.9 (C12), 151.1 (C9), 144.0 (C2), 141.2 (C7), 138.6 (C30), 131.7 (C4), 131.1 (C13), 130.8 (C26), 128.9 (C8), 122.1 (C28), 121.2 (C27), 120.3 (C14), 120.2 (C3), 115.1 (C25), 114.4 (C29), 97.2 (C22), 68.3 (C20), 40.4 (C17), 36.7 (C16), 30.5 (C19), 13.4 (C18). Anal. Calcd for C$_{25}$H$_{23}$N$_5$O$_3$.[0.5 H$_2$O]: C, 66.65; H, 5.37; N, 15.55. Found C, 66.47; H, 5.02; N, 15.33.