Lipid peroxide mediated oxidative rearrangement of the pyrazinone carboxamide core of neutrophil elastase inhibitor AZD9819 in blood plasma samples

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List of nonstandard abbreviations: HMBC, Heteronuclear Multiple Bond Correlation; LC, high performance liquid chromatography; LDL, low density lipoprotein; MeCN, acetonitrile; MS, mass spectrometry; MS/MS, tandem mass spectrometry; m/z, mass-to-charge ratio; P450, cytochrome P450; ROP, rearranged oxidation product; XO, xanthine oxidase
ABSTRACT

This study focused on the mechanistic interpretation of ex vivo oxidation of a candidate drug in blood plasma samples. An unexpected lipid peroxide-mediated epoxidation followed by a dramatic rearrangement led to production of a 5-membered oxazole derivative from the original 6-membered pyrazinone-carboxamide core of a human neutrophil elastase inhibitor, 6-(1-(4-cyanophenyl)-1H-pyrazol-5-yl)-N-ethyl-5-methyl-3-oxo-4-(3-(trifluoromethyl)phenyl)-3,4-dihydropyrazine-2-carboxamide (AZD9819). The rearranged oxidation product 2-(1-(4-cyanophenyl)-1H-pyrazol-5-yl)-5-(N-ethylacetamido)-N-(3-(trifluoromethyl)phenyl)oxazole-4-carboxamide was characterized by accurate mass MS/MS fragmentations, by 2D NMR and X-ray crystallography of an authentic standard, and by incorporation of an 18O atom from molecular 18O2 to the location predicted by our proposed mechanism. The lipid peroxide mediated oxidation was demonstrated by using human low-density lipoprotein (LDL) in pH 7.4 phosphate buffer and by inhibiting the oxidation with ascorbic acid or GSH, two antioxidants effective in both plasma and the LDL incubation. A nucleophilic mechanism for the epoxidation of AZD9819 by lipid hydroperoxides explains the prevention of its ex vivo oxidation by acidification of the plasma samples. The discovery of the lipid peroxide-dependant oxidation of an analyte and the means of prevention could provide valuable information for biotransformation and bioanalysis.
Introduction

Accurate measurement of drug molecule concentrations in blood plasma samples is essential for pharmacokinetic and toxicokinetic studies in drug discovery and development (Li et al., 2011; Rozeta et al., 2011). Correct identification of metabolites in plasma samples is critical for safety assessment of a drug candidate (Ma and Chowdhury, 2011; Penner et al. 2012; Gao et al. 2013). The occurrence of ex vivo instability complicates the analysis by making metabolite identities insecure and measured drug concentrations uncertain. While the instability of some molecules could be predicted by the presence of certain chemically or biologically labile moieties (Li et al., 2011; Mitchell, 2014; Erve 2008), in other cases ex vivo instability can be unexpected.

One such unexpected instability was discovered during the preclinical development of an inhaled drug candidate AZD9819 (Lindsjö, 2011), an inhibitor of human neutrophil elastase for potential treatment of chronic obstructive pulmonary disease (Sjö, 2012; Lucas et al., 2013). A mono-oxidation product of AZD9819 was detected in plasma ex vivo and in vitro. The occurrence of this oxidation was empirically prevented by acidification following plasma sample collection (e.g., with 2M phosphoric acid added at approximately 1.5% of the plasma volume) in preclinical and clinical development studies. Two intriguing findings emerged from our investigation of this otherwise seemingly simple mono-oxidation. First, the mono-oxidation product was in fact a drastically rearranged oxidation product (ROP). Second, we pinpointed lipid peroxides as the cause of the initial epoxidation step in our proposed mechanism of ROP formation. This cause of oxidation of drug molecules in blood plasma has not been reported previously.

The ROP formation is distinct in terms of both the cause of oxidation and subsequent rearrangements from two previous reports of rearranged oxidative metabolites of pyrazinone-containing thrombin inhibitors developed at Merck (Singh et al., 2003; Subramanian et al., 2003). Oxidative rearrangements in the Merck cases occurred via cytochrome P450-mediated epoxidation at the C=C bond in a pyrazinone ring (of 3-amino-6-methyl-pyrazinone and 3-amino-6-chloro-pyrazinone derivatives) (Singh et al., 2003; Subramanian et al., 2003). This is no surprise because P450 enzymes are known to be capable of epoxidation (Parkinson, 2001; Guengerich, 2007). However
ROP was not produced by cytochrome P450 in liver in our case with AZD9819. Our finding of the role of lipid peroxides in the mechanism of ROP formation described in this article could help in troubleshooting and preventing the instability of some other xenobiotic or endogenous organic molecules in plasma samples.
MATERIALS AND METHODS

6-(1-(4-cyanophenyl)-1H-pyrazol-5-yl)-N-ethyl-5-methyl-3-oxo-4-(3-(trifluoromethyl)phenyl)-3,4-
dihydropyrazine-2-carboxamide (AZD9819) was synthesized by a reported procedure (Lindsjö, 2011). MS: [M+H]+ m/z 493.1592 ± 0.0011 (s.d., n = 14; Calc. m/z 493.1594). 1H NMR (600 MHz, CD3SOCD3): δ 1.02 (t, J = 7.2 Hz, 3H), 1.88 (s, 3H), 3.19 (qd, J = 7.0, 6.5 Hz, 2H), 6.77 (d, J = 1.8 Hz, 1H), 7.67 (d, J = 8.7 Hz, 2H), 7.78 (d, J = 8.0 Hz, 1H), 7.88 (t, J = 8.0 Hz 1H), 7.89 (d, J = 8.7 Hz 2H), 7.93 (d, J = 1.8 Hz 1H), 7.95 (s, 1H), 7.96 (d, J = 8.0 Hz 1H), 8.71 (t, J = 8.0 Hz 1H). 13C NMR (150 MHz, CD3SOCD3): 14.5, 18.7, 33.6, 109.3, 111.2, 118.5, 122.5, 123.7 (q, J = 272.5 Hz) 123.9, 124.8 (q, J = 3.5 Hz), 126.6 (q, J = 3.6 Hz), 130.5 (q, J = 32.6 Hz), 131.3, 132.0, 133.4, 137.4, 138.4, 141.2, 142.4, 143.0, 143.3, 154.4, 161.3. (Complete assignment available in Supplemental Figures 1 and 2)

The synthetic standard of 2-(1-(4-cyanophenyl)-1H-pyrazol-5-yl)-5-(N-ethylacetamido)-N-(3-
(trifluoromethyl)phenyl)oxazole-4-carboxamide (ROP): AZD9819 (1g, 0.002 mol) was slurried in acetonitrile (50 mL) and water (50 mL) at 25 °C. Sodium perborate tetrahydrate (0.38 g, 0.0024 mol) was dissolved in water (20 mL) and added slowly to the reaction mixture over approximately 3 hours. During the addition, the pH of the reaction mixture was maintained at 8 by the addition of 0.1 M hydrochloric acid solution. After addition of the perborate, the reaction was stirred for 20 hours, diluted with water (100 mL) and extracted with dichloromethane (2 x 50 mL). The combined dichloromethane extracts were washed with water (2 x 50 mL), dried over anhydrous sodium sulfate and evaporated under reduced pressure to leave the product ROP as a white powder. Yield = 0.85 g 83%. MS: [M+H]+ m/z 509.1544 ± 0.0011 (s.d., n = 8; Calc. m/z 509.1543). 1H NMR (600 MHz, CD3SOCD3) δ 0.90 (t, J = 6.9 Hz, 3H), 1.93 (br s, 3H), 3.55 (q, J = 6.9 Hz, 2H), 7.32 (d, J = 1.9 Hz, 1H), 7.47 (d, J = 7.9 Hz, 1H), 7.59 (t, J = 7.9 Hz, 1H), 7.83 (d, J = 8.6 Hz, 2H), 8.03 (d, J = 8.6 Hz, 2H), 8.05 (d, J = 7.9 Hz, 1H), 8.05 (d, J = 1.9 Hz, 1H), 8.26 (s, 1H), 10.53 (s, 1H). 13C NMR (151 MHz, CD3SOCD3) δ 13.1, 21.8, 42.6, 111.2, 112.2, 116.8 (q, J = 3.8 Hz), 118.2, 120.6 (q, J = 3.5 Hz), 124.1 (q, J = 272.2 Hz), 124.3, 125.9, 126.5, 129.4 (q, J = 31.6 Hz), 129.8, 129.9, 133.5, 138.8, 141.9, 142.7, 148.5, 149.8, 158.2, 169.3. (Complete assignment available in
Supplemental Figures 3 and 4). The crystal structure of the ROP synthetic standard was determined by X-ray crystallography (details available in the Supplemental Data) and the data (CCDC number 1039312) can be obtained free of charge from the Cambridge Crystallographic Data Centre at http://www.ccdc.cam.ac.uk/data_request/cif

**Blood plasma.** Blank plasma used for the study with $[^{14}\text{C}]$AZD9819 included Han Wistar rat (K2-EDTA, male, pooled) and human plasma (K2-EDTA, mixed gender, 4 donor pool) prepared at AstraZeneca R&D, Lund, Sweden, and Beagle dog plasma (K2-EDTA, male, 3 donor pool) obtained from Harlan-Winkelmann (Borchen, Germany). Blank plasma used for the study with non-radiolabeled AZD9819 included Han Wistar rat plasma (K3-EDTA, male) and Beagle dog plasma (K3-EDTA) obtained from Biorelamation (Westbury, NY), and human plasma (K3-EDTA, mixed gender, pooled) obtained from BioChemed Services (Winchester, VA).

**Chemicals, biochemicals, and proteins.** Most were purchased from Sigma-Aldrich (St. Louis, MO), e.g., allopurinol, L-ascorbic acid (vitamin C, Cat No 255564), bovine liver catalase (Cat No C40), reduced L-glutathione (GSH, Cat No G4251), hemoglobin human (Cat No H7379), hydrogen peroxide solution 30 wt.% (Cat No 216763), $^{18}\text{O}_2$ gas (97 atom % $^{18}\text{O}$, Cat No 490474), $^{18}\text{O}$-Water (97 atom % $^{18}\text{O}$, Cat No 329878), potassium cyanide (KCN), and sodium azide (NaN$_3$).

Human low density lipoprotein (LDL) lyophilized powder (Cat No L8292, Purity: > 95%) was also bought from Sigma-Aldrich. The LDL is composed of 20–25% protein and 75–80% lipid. The lipid portion can be further described as the following: 9% free cholesterol, 42% cholesteryl ester, 20–24% phospholipid, and 5% triglyceride.

**Plasma incubation of $[^{14}\text{C}]$AZD9819:** Plasma samples were incubated at 37°C with 14 μM $[^{14}\text{C}]$AZD9819. As a negative control, the stability of $[^{14}\text{C}]$AZD9819 was investigated in 0.1 M phosphate buffer pH 7.4 supplemented with 5 mM MgCl$_2$ for 2 h. Aliquots of samples were taken at 0 and 120 min. and the reaction was terminated with an equal volume of pre-chilled MeCN/formic acid (99/1; v/v).
Incubations of AZD9819 with human low density lipoprotein (LDL). Deionized water was added to lyophilized LDL, resulting in a 5 mg/mL LDL suspension containing approximately 150 mM NaCl and 0.01% EDTA at pH 7.4. Incubation mixtures were prepared with 500 mM pH 7.4 phosphate buffer solution to reach final concentrations of 15 µM AZD9819 and 1 mg/mL LDL, in the absence/presence of 50 mM GSH or 50 mM ascorbic acid. The total incubation volume was 250 µL. Additional incubations at pH 4.5 and pH 10.5 were conducted for 15 µM AZD9819 with LDL in 100 mM ammonium acetate pH 4.5 buffer and 100 mM glycine pH 10.5 buffer, respectively. Negative control experiments were also conducted with blank buffer solutions. In all incubations, AZD9819 (2 mM in methanol) was added last to allow approximately 40 min. pre-incubation of LDL at room temperature in the presence/absence of GSH or ascorbic acid. All LDL incubations were repeated in triplicates and were terminated after at 37 °C for 2 h by adding twice volume of MeCN containing 1% formic acid.

Incorporation of $^{18}$O from molecular $^{18}$O$_2$. Prepared 15 µM AZD9819 in 250 µL rat plasma was sealed in 1-dram glass vial with a rubber septum at room temperature. Two stainless steel syringe needles were inserted through the septum, with one into the plasma mixture for purging and the other above the liquid level for venting. The plasma mixture and the vial were slowly purged with $^{18}$O$_2$ for approximately 5 min. Some plasma mixture was lost through the needle venting to the air due to foaming. Both needles were removed from the septum and the vial was then incubated at 37 °C for 2 h. The reaction was stopped with MeCN (approx. 2:1; v:v).

Plasma incubations of AZD9819 in the presence of NaN$_3$, allopurinol, KCN, ascorbic acid, or reduced L-glutathione (GSH). AZD9819 at 15 µM was incubated at 37 °C in rat or dog plasma in the presence of NaN$_3$ (up to 50 mM), in rat, dog or human plasma in the presence of 100 µM allopurinol, in rat plasma in the presence of 50 mM KCN, in rat or dog plasma with ascorbic acid added (up to 50 mM), and in rat, dog, and human plasma with GSH added (up to 50 mM). The pH of plasma samples with added GSH or ascorbic acid was verified to be still above pH 7 with pH test paper. As control, 15 µM AZD9819 was incubated in the plasma of respective species. The incubation volume was 250 µL. The incubations were terminated at 2 h by adding twice volume of MeCN.
Heat inactivated rat plasma. A tube of several mL of rat plasma was inactivated at 56 °C for 1 h. In separate vials, 15 µM AZD9819 and 15 µM ROP standard were incubated with 500 µL of inactivated rat plasma at 37 °C. AZD9819 and ROP standard were also incubated in untreated rat plasma at the same concentration as respective positive controls. The reaction was terminated at 2 h by adding an equal volume of MeCN. Additionally, AZD9819 was incubated at 37 °C for 2h in denatured rat plasma by mixing with an equal volume of MeCN at time zero.

Catalase pretreated plasma. 250-uL rat plasma was pre-incubated with 1000 unit/mL bovine liver catalase at 37 °C for 30 min. Then AZD9819 was added at 15 µM and the incubation was continued for 2 h before being terminated by adding twice volume of MeCN. As a control, 15 µM AZD9819 was incubated in untreated rat plasma, including a pre-incubation at 37 °C for 30 min. without catalase. The effectiveness of the catalase was confirmed by incubation with 500 µM H₂O₂ in pH 7.4 buffer when the reactivity of the H₂O₂ was completely eliminated.

Incubation of AZD9819 with human hemoglobin in the absence/presence of H₂O₂. AZD9819 at 15 µM was incubated at 37 °C with 100 µM human hemoglobin in a 100 mM pH 7.4 phosphate buffer, with the 100 µM hemoglobin in the presence of 500 µM H₂O₂, or with 500 µM H₂O₂ alone in the buffer. After 2 h, bovine liver catalase was added to the incubation mixture at 1000 unit/mL, followed by additional 15-minute incubation. As a negative control, a 500 µM H₂O₂ in the buffer was pre-incubated with 1000 unit/mL for 15 minute before AZD9819 was added. At the end of all incubations, twice the volume of MeCN containing 1% formic acid was added to the incubation mixture.

Radiochromatography and LC-MS and MS/MS of [¹⁴C]AZD9819 incubation products. After centrifugation of terminated incubation mixtures to pellet proteins, supernatants were recovered and diluted with one volume of Milli-Q water. Aliquots of 100 µL samples were injected into an LC-radioactivity detector system, comprising of an Agilent 1100 series LC system (Palo Alto, CA) coupled to a Flo-One 500 series on-line radioactivity flow detector with a 500 µL liquid cell (Perkin Elmer). LC separation utilized an ACE C18, 3 µm, 150x4.6 mm
column (Aberdeen, Scotland, UK). The mobile phases consisted of (A) MeCN:H2O (5:95) and (B) MeCN:H2O (95:5). Both solvents contained 0.1% formic acid. The LC gradient started at 5% B, increased linearly to 25% B during the first minute and then to 65% B over the next 24 minutes. It was then increased to 100% B at 26 min and retained for 3 minutes, finally decreased to 5% B at 30 min. The flow rate was 1 mL/min of LC mobile phase and 3 mL/min of liquid scintillation cocktail.

An LC system of the same type as above was used in line with a Q-ToF Premier mass spectrometer (Waters, Wilmslow, UK) equipped with LockSpray interface and electrospray probe. The LC-MS and MS/MS analysis utilized the same type but a smaller LC column (50 x 2.1 mm), and the same LC mobile phases as above with similar but short gradients at the flow rate of 0.25 mL/min. MS/MS was performed at 30-eV collision energy with argon as collision gas.

**LC-UV-MS and MS/MS of non-radiolabeled AZD9819 incubation products.** After centrifugation of terminated incubation mixtures to pellet proteins, aliquots of 10 or 15 µL supernatant samples were injected into an LC-UV-MS system, consisting of an Acquity UPLC system (Waters, Milford, MA) and an LTQ-Orbitrap XL high resolution mass spectrometer (Thermo Scientific, San Jose, CA). An ACE C18, 3 µm, 150x2.1 mm column was used for LC separation. The mobile phases at the flow rate of 0.2 mL/min consisted of (A) water and (B) acetonitrile. Both solvents contained 0.1% formic acid. The LC gradient started at 5% B, increased linearly to 25% during the first minute and then to 65% B over the next 24 minutes. It was then increased to 100% B at 26 min and retained for 3 minutes, finally decreased to 5% B at 30 min. The UV wavelength at 268 nm with 6nm resolution was monitored in acquisition of UV chromatograms and photodiode array UV spectra of 200 to 500 nm were also acquired.

LC-MS spectra were acquired at the resolution of 30,000. Collision-induced dissociation (CID) with helium as collision gas in the linear Ion trap, or the instrument manufacturer’s named “high energy collision-induced dissociation” (HCD) with nitrogen as collision gas in the HCD cell, was used in acquisition of MS/MS spectra by
the Orbitrap analyzer at the resolution of 15,000. The normalized collision energy at 45 and 35 of the manufacturer's respective units were used in HCD and CID experiments. The fragmentation by the HCD is similar to that by the QToF CID used for the radiolabeled study, both of which produce more fragments from [M+H]+ ions of AZD9819 or its products. However, the linear ion trap CID was used for most of experiments in the present study, due to higher signals of the ion trap CID.

**Nuclear Magnetic Resonance Spectroscopy.** 1H NMR, 13C NMR, 2D-HSQC and HMBC spectra were recorded at 298 K in DMSO-d6 on a Bruker Avance Spectrometer at a proton frequency of 599.7 MHz and carbon frequency of 150.8 MHz. Referencing is reported relative to deuterated solvent signals at 2.500 ppm (proton) and 39.52 ppm (carbon).

**X-ray crystallography.** Single crystals of appropriate size and quality for crystal structure determination were grown with the free interface diffusion method. Acetonitrile was used as solvent and diisopropylether as antisolvent. Crystal data for the ROP synthetic standard: C25H19F3N6O3, Mw=508.46 g/mol, colorless, triclinic, P-1 (No 2), a=11.3364(4)Å, b=12.3259(4)Å, c=18.0013(7)Å, α=99.870(2)°, β=95.648(2)°, γ=107.680(2)°, V=2330.53(14) Å³, Z=4, ρcalc=1.449 gcm⁻¹, μ=0.115 mm⁻¹, R1=0.047, wR2=0.154, GOF=1.08. The data were collected at 200 K on a Bruker ApexII diffractometer with graphite-monochromated MoK(α) radiation and employing a 0.22×0.20×0.04 mm crystal (Rint=0.048). The crystal structure was determined by direct methods and refined by full matrix least-squares analyses with anisotropic temperature factors for all atoms except hydrogen atoms. Hydrogen atom positions were calculated using known molecular geometries, refined in riding mode with fixed isotropic temperature factors. The fluorine atoms of the -CF₃ group in one of the molecules in the asymmetric unit are disordered. CCDC-1039312 contains the supplementary crystallographic data for the ROP synthetic standard. The data can be obtained free of charge from The Cambridge Crystallographic Data Centre at http://www.ccdc.cam.ac.uk/data_request/cif
RESULTS

Figure 1 shows radiochromatographic profiles of rat, dog, and human plasma generated products of [14C]AZD9819 following a 2 h incubation at 37 °C. Chemical structures of [14C]ROP and its deacetylation product are provided in the figure for easy reading. During our investigations, a great number of LC-UV-MS chromatograms demonstrating ROP formation in plasma samples were acquired. We have chosen to present the radiochromatograms here as they convincingly demonstrate that ROP is the sole direct product of the parent compound AZD9819. The deacetyl-ROP that occurred mainly in rat plasma was shown to be an enzymatic hydrolysis product of ROP (described later). Interestingly we were able to synthesize an authentic standard of ROP before we even knew what the structure was. The choice of synthetic method resulted from preliminary work using a commercial peroxidase in the presence of H2O2 (Coprinus cinereus peroxidase recombinant in an aspergillus, supplied by Novozymes, Denmark), to make a similar oxidation product of an analog compound (details of the analog compound not provided for proprietary reasons). ROP was made from AZD9819 at high yield by reaction with sodium perborate in MeCN:H2O (Experimental Section). The synthetic standard was unambiguously proven to be identical to ROP generated in plasma, by a perfect match in LC-UV-MS/MS (retention time, UV spectra, and high resolution MS/MS spectra showing the accurate mass of fragment ions. Supplemental Figure 5). It was also verified that the plasma-generated [14C]ROP and non-radiolabeled ROP at our different laboratories have the same chemical structure, by matching all LC-MS/MS fragmentations (Supplemental Figure 6).

Even after a high-purity standard was synthesized, it was still challenging to elucidate the structure. ROP does not share any similar fragmentation with AZD9819 in MS/MS (Supplemental Figures 5 and 6) and the lack of protons at the center core of ROP means that the presence of a new oxazole ring cannot be proven directly by NMR. Hence a number of putative structures (Supplemental Figure 7), including those we could envisage arose from possible rearrangement mechanisms following an initial oxidation, were proposed to generate hypothetical products to match NMR and MS/MS data (Supplemental Figures 3, 4, 5, and 6). The majority of these putative
structures deliberately contained an oxazole substructure, because preliminary assignment of a MS/MS fragment ion of ROP observed in biological samples had hinted at this possibility (Supplemental Figure 6).

Scheme 1 shows the deduced structure for ROP and the proposed oxidative rearrangement mechanism for its formation, by putting all the information together. The cause of the initial oxidation and other details of the mechanism will be discussed later. A key part of structural elucidation process was the use of predicted $^{13}$C chemical shifts, and a comparison is shown in Table 1 illustrating the change in chemical shift of carbon atoms which map from AZD9819 to ROP (Scheme 1), and how well they match predicted values. Heteronuclear Multiple Bond Correlation (HMBC) NMR correlation data was also useful. For example, Figure 2 shows a portion of the HMBC spectrum of the ROP synthetic standard. The correlations observed, specifically between H33 and the carbonyl C34 of the acetyl, prove the position of the acetyl group in ROP and led us to propose the transfer of this group in the mechanism (4 to 5 in Scheme 1). We were subsequently able to obtain an X-ray structure from a single crystal of the synthetic standard to further corroborate the structural identification (Figure 3).

Figure 4 illustrates that an $^{18}$O atom from molecular $^{18}$O$_2$ is incorporated to the location predicted by the proposed oxidative rearrangement mechanism. In the LC-MS spectrum of rat plasma generated ROP (Figure 4A), the naturally occurring isotopic peak at m/z 511 (i.e. 2 Da higher than the monoisotopic peak at m/z 509) has a low relative abundance of less than 5%. When $^{18}$O$_2$ was added to the rat plasma incubation of AZD9819, $^{18}$O-incorporated [M+H]$^+$ ions at m/z 511 were observed by a clear increase in the relative abundance of the m/z 511 peak (Figures 4B vs. A). MS/MS fragmentation of $^{18}$O-[M+H]$^+$ ions (m/z 511) vs. the monoisotopic [M+H]$^+$ ions (m/z 509) are compared in Figures 4D vs. 4C. Two major fragment ions occur in both spectra. The fragment ion formed by the cleavage of the acetyl amide bond remains unchanged at m/z 467.14 after the incorporation of an $^{18}$O atom, while the fragment ion formed by the neutral loss of a 3-(trifluoromethyl)aniline shifts by 2 Da from m/z 348.11 to 350.11 (Figure 4D vs. 4C). This indicates that the incorporated $^{18}$O is indeed located at the acetyl carbonyl as expected by the proposed mechanism (Scheme 1). It is worth noting that naturally occurring isotopic ions at m/z 511 (Figure 4A) consist of a mixture of one $^{13}$C + one $^{15}$N atom, two $^{13}$C or two $^{15}$N atoms, or one $^{18}$O
atom with random distribution across the whole molecule. This would result in both 1-Da and 2-Da shifts to higher m/z for both fragments (Supplemental Figure 8).

ROP was not produced by cytochrome P450 in liver, as it was absent from incubations of AZD9819 in rat, dog, and human hepatocytes (data not shown). In search for the cause of the oxidation of AZD9819 in plasma, a number of experiments applying different reagents or experimental conditions to the incubations in plasma were performed and are summarized in Table 2. The percentages in the table were estimated from LC-UV peak areas with consideration of the different UV response of ROP vs. AZD9819 (with an absorbance ratio of approximately 1.5 at 268 nm). Additionally, it was assumed that deacetyl-ROP observed mainly in rat plasma has the same UV absorbance as ROP. The percentage values in Table 2 are intended for qualitative comparison and along with the other experiments described here, helped guide us to the cause of the oxidation.

The formation of ROP was not a simple chemical degradation process, because ROP was not produced in any of the control incubations of AZD9819 in blank buffer solutions. It was also not a photo oxidative process, because the use of an amber vial or wrapping a vial in aluminum foil gave the same yield of ROP as the control incubation in a clear vial under ambient light (Table 2).

We considered a few possible enzymatic causes. Peroxidases are known to catalyze epoxidation of C=C bonds by peroxides such as H₂O₂ (Adam et al., 1999; Ortiz de Montellano, 1992). Indeed, as stated earlier, we made a similar oxidation product of an analog compound using a commercial peroxidase. However, the oxidation of AZD9819 in plasma is not inhibited by sodium azide (NaN₃, Table 2), a known inhibitor of peroxidases (Ortiz de Montellano, 1992; Pozdnyakova, 2013). We also excluded xanthine oxidase (XO). Kalgutkar and co-workers have reported a XO mediated oxidation of a quinoxaline derivative in rodent plasma (Sharma et al., 2011), involving incorporation of an ¹⁸O atom from H₂¹⁸O but not from molecular ¹⁸O₂. However, the addition of the XO inhibitor allopurinol (Sharma et al., 2011) to plasma did not inhibit formation of ROP (Table 2). Moreover, ROP incorporated an ¹⁸O from molecular ¹⁸O₂ (Figure 4) but not from H₂¹⁸O (data not shown) as would be expected. Additionally, we ruled out the involvement of hemoglobin. Literature reports about hemoglobin catalyzed
epoxidation of styrenes (Ortiz de Montellano and Catalano, 1985; Tscherret-Guth and Ortiz de Montellano, 1996) and retinoic acid (Iwahashi et al., 1985) offered a possible cause if hemolysis had occurred and released hemoglobin into the plasma. In both cases the reaction was effectively inhibited by cyanide ions (Ortiz de Montellano et al., 1985; Iwahashi et al., 1985). However in our case, KCN enhanced rather than inhibited ROP formation (Table 2). Furthermore, incubation of 15 µM AZD9819 with 100 µM human hemoglobin did not produce any ROP, and the incubation of AZD9819 in 100 µM hemoglobin in the presence of 500 µM H2O2 did not catalyze the reaction (data not shown).

The formation of ROP from AZD9819 was most likely an ex vivo and in vitro phenomenon. During preclinical and clinical development, metabolite identification was conducted in rat/dog plasma, urine, feces and bile samples collected post an iv or oral dose of [14C]AZD9819, and human plasma & urine samples collected following inhaled doses of AZD9819. Among all in vivo samples, we did not observe any ROP. Furthermore, complete recovery of total radioactivity in preclinical mass balance studies post a dose of [14C]AZD9819 (virtually 100% in rat, >93% in dog) suggested that it was unlikely that ROP or its intermediates were formed but then bound to macromolecules in vivo.

The effect of temperature on ROP formation was revealing (Table 2). The drop in yield from 37°C to room temperature might not be as great as would be expected for an enzymatic reaction and ROP formation was not completely eliminated even on ice. Inactivation of rat plasma at 56 °C for 1 h did not prevent the ROP formation from AZD9819 (Table 2), but abolished formation of deactyl-ROP from ROP synthetic standard suggesting deacetyl-ROP is an enzymatic product of ROP (Supplemental Figure 9). In contrast, denaturing the plasma with MeCN did eliminate ROP formation (Table 2). Collectively, these observations suggested the likely involvement of heat-resistant proteins. However, we did not realize that these could be lipoproteins until lipid peroxides emerged to be the probable cause from subsequent experiments.

ROP did not appear to be produced by H2O2 oxidation in plasma, because the incubation of 15 µM AZD9819 with up to 500 µM H2O2 in pH7.4 buffer only produced a trace amount of ROP (less than 1% of parent, data not
shown). Moreover, pre-treatment of rat plasma with bovine liver catalase at 1000 unit/mL, which would decompose any H₂O₂ present in the plasma, did not abolish the oxidation of AZD9819 (Table 2). We also ruled out the formation of ROP by hydroxyl radical or hydroperoxyl radical from H₂O₂, because ROP was not formed by the Fenton reaction of AZD9819 with 100 µM H₂O₂ and 100 µM ferrous ion (Fe²⁺) in a pH4.5 buffer (within the optimal pH 3-6 range of the Fenton reaction). Intriguingly, during these experiments we made the fortuitous discovery that ROP could be produced from AZD9819, albeit alongside other oxidative products, using a very high concentration of H₂O₂ (90 mM) in pH 7.4 phosphate buffer (Supplemental Figure 10). This hinted that even if H₂O₂ was not involved in ROP formation in plasma, perhaps other peroxides might be.

Ascorbic acid (one form of vitamin C) was tested to determine whether a radical mechanism could be involved. Inhibition was modest at 1 mM, but significant at 50 mM (Table 2). We were unaware that ascorbic acid is “an outstanding antioxidant in blood plasma” against lipid peroxidation, until during the manuscript preparation we learnt this from two highly cited papers that will be discussed later (Frei et al., 1988 & 1989).

In an effort to trap the epoxide presumed in our mechanism, GSH was added to rat plasma at 5 mM resulting in a clear decrease in ROP (Table 2). Importantly, GSH conjugates were not found, but the disappearance of parent was also attenuated (data not shown). This suggests that GSH probably prevents the reaction at the first epoxidation stage. Additional experiments with 50 mM GSH in plasma showed even greater inhibition on both ROP formation (Table 2) and AZD9819 depletion (data not shown). GSH is an important antioxidant in biological systems preventing damage by reactive oxygen species such as free radicals and peroxides, in addition to being a strong nucleophile which reacts well with soft electrophiles. GSH-dependent selenoperoxidases catalyze two-electron reduction of lipid peroxides (LOOH) to inert lipid alcohols (LOH), while two reduced GSH molecules become one oxidized GSSG (Girotti, 1998).

Collectively, these observations led us to consider lipid peroxides as the probable cause for the oxidation of AZD9819 in plasma. Moreover, a lipid peroxide mediated oxidation would incorporate molecular oxygen during the lipid peroxidation chain propagation (Frei et al., 1989; Girotti, 1998) and be consistent with our observation.
on the involvement of $^{18}$O$_2$ (Figure 4).

Finally we succeeded in generating ROP using human low-density lipoprotein (LDL, 1 mg/mL) in pH 7.4 buffer (Figure 5B). The level of ROP formation was diminished in the presence of 50 mM GSH or ascorbic acid (Figure 5C and 5D). Percentages of ROP formation derived from LC-UV peak areas and UV signal response ratio are compared in Figure 5F.
DISCUSSION

The inhibitive effects of ascorbic acid and GSH on AZD9819 oxidation in both the LDL and plasma incubation provided the supporting evidence to the involvement of lipid peroxides in the oxidation. Frei et al. have previously stated that ascorbate is the only plasma antioxidant that can completely protect plasma lipids against detectable peroxidative damage induced by aqueous peroxy radicals (Frei et al., 1988 & 1989). Additionally, they have demonstrated that protein thiol groups are the second in line for antioxidant defense, while naturally occurring GSH in isolated plasma is present only at a very low concentration (<0.5 µM. Frei et al., 1988).

Lipid peroxide mediated oxidation explains why ROP was only formed ex vivo and in vitro, since a number of scavengers of lipid peroxy radicals and lipid peroxides maintain a very low level of lipid peroxides in a living biological system (Frei et al., 1988 & 1989; Girotti, 1998). However, those scavengers can be depleted ex vivo allowing the buildup of lipid peroxides by chain propagation. Lipid peroxide mediated oxidation also rationalizes why protein precipitation by acetonitrile abolishes the ROP formation in acetonitrile denatured plasma (Table 2). Frei et al. reported that after the depletion of ascorbate in plasma, peroxidation occurred first to lipids transported in lipoproteins, whereas nonesterified fatty acids were still protected from oxidation by binding to plasma albumin (Frei et al., 1988). In the LDL incubation without plasma albumin, acetonitrile containing 1% formic was used as presumably a better quench solution. Differing levels of pre-existing lipid peroxides would rationalize our observation of variations in ROP formation in different plasma samples of the same species, e.g., number of free/thaw cycles, length of storage time at -18 °C (data not shown). Future work could include quantitative analysis of lipid peroxides, e.g., to study the relationship between the level of lipid peroxides and the ROP formation, and to investigate factors affecting the level of peroxides in plasma samples. Finally, ROP formation via epoxidation of AZD9819 in plasma is not contradictory to the chemical preparation of ROP using sodium perborate, since sodium perborate functions mainly as a convenient source of mildly alkaline hydrogen peroxide and is a known agent for epoxidation of a wide range of alkenes while also being a known agent of heteroatom oxidation (Mckillop and Sanderson, 2000; Bortolini, 2009).
The level of ROP generated by LDL was enhanced several fold when incubated at pH 10.5 relative to that at pH 7.4 (Figure 5E vs. 5B; and Figure 5F). Furthermore, ROP was absent from the LDL incubation in pH 4.5 buffer (data not shown), consistent with the empirical use of acidification to prevent ex vivo oxidation of AZD9819 in plasma samples. No ROP was formed when AZD9819 was incubated in the individual buffer solutions as a control. The calculated pKa (ACD/pKa DB 2012) for an allylic hydroperoxide of a polyunsaturated fatty acid is approximately 11.3 and thus the pH-dependence suggests that a nucleophilic epoxidation of the C=C bond in the pyrazinone ring of AZD9819 (Scheme 2) is highly plausible. It is known that nucleophilic epoxidation by hydroperoxide can occur at alkenes that are conjugated with electron withdrawing groups (Apeloig et al., 1983).

Interestingly, the addition of 50 mM KCN to the LDL incubation at pH 7.4 approximately doubled the formation of ROP (data not shown, but similar to that observed in rat plasma, Table 2). A cyanide conjugate with [M+H]+ at m/z 494.1547 was detected in the presence of 50 mM KCN in both the LDL and rat plasma incubations of AZD9819. The measured accurate mass in LC-MS indicated that the cyanide conjugate had exactly the same elemental composition as that of a HCN adduct of deacetyl-ROP. However, the LC-MS/MS fragmentation does not support the same skeleton structure as deacetyl-ROP (spectra not shown). We assume that CN− adds to the imine of intermediate 1, ultimately leading to the loss of HCN and formation of oxazole as an analogous mechanism to that when HO− adds to intermediate in Scheme 1. Additionally, an unknown rearrangement mechanism after the CN− addition might have led the formation to the rearranged cyanide conjugate detected at m/z 494.1547.

The hydration of the imine in our proposed mechanism is necessary to change the rigid sp2 carbon to a more flexible sp3 centre, thereby allowing intramolecular nucleophilic attack of the carboxamide oxygen on the epoxide (1 to 2, Scheme 1). We also note that an oxygen anion is naturally formed for the proposed ring closure (2, Scheme 1). Previously reported mechanisms for P450-mediated oxidation of a pyrazinone ring are related in that they involve either hydrolysis or GSH addition to an epoxide (Singh et al., 2003; Subramanian et al., 2003; Zhou et al., 2010), presumably catalyzed by epoxide hydrolases or glutathione S-transferases present in liver (Parkinson, 2001). These led to the pyrazinone ring opening by the breakage of one or both C-N bonds resulting
in multiple products. Our proposed mechanism, involving a non-enzymatic epoxidation of AZD9819 followed by imine-hydration and subsequent elimination of a water molecule (Scheme 1) leads to a single rearranged product and is therefore consistent with the radiochromatographic data (Figure 1).

The final rearrangement step proposed for intermediates 4 to 5 in Scheme 1 can be viewed as the intramolecular transfer of the negative charge from an apparently high energy anion to a reasonably low energy anion and therefore the formation of a more stable species. The negative charge in 5 can be delocalized by adjacent carbonyl and phenyl groups, whereas the charge in 4 would be delocalized only by the oxazole. Interestingly, intermediate 4 was briefly assumed to be likely structure of ROP before the characterization of ROP by HMBC 2D NMR as shown in Figure 2. At that time, we had to postulate an acetyl migration rearrangement in gas-phase MS/MS fragmentation in order to interpret a major fragment ion by the neutral loss of a 3-(trifluoromethyl)aniline (m/z 348.11). After which, the HMBC data made us to quickly realize that the rearrangement of acetyl migration had already happened in the formation of ROP.

The deacetyl-ROP that occurred mainly in rat plasma has been shown to be an enzymatic hydrolysis product of ROP (Supplemental Figure 9A). It is known that hydrolysis of carboxamide in plasma can be mediated by carboxylesterases (Parkinson 2001). The species difference in deacetyl-ROP between rat and dog/human (Figure 1A vs. B & C) can be readily rationalized by the differences in carboxylesterase expression and the hydrolase activity in plasma (Bahar et al., 2012).

To the best of our knowledge, this is the first reported case of epoxidation of a pharmacological agent by lipid peroxides in plasma. We acknowledge that epoxidation by lipid peroxides is neither a new concept nor a new phenomenon. It is known that lipid epoxides can be generated as secondary oxidation products of lipid peroxidation (Scheick and Spiteller, 1993; Chen and Chung, 1996). Scheick and Spiteller (1993) reported the epoxidation at enol ethers R1-O-CH=CH-R2 of a number of plasmalogens by reaction with a lipid hydroperoxide that was produced from linoleic acid by tomato homogenate. Incorporation of an $^{15}$O from molecular $^{18}$O$_2$ occurred to the lipid peroxide first, then into final trapped products derived from the epoxides of a number of
plasmalogens. Chen and Chung (1996) reported that two linoleic-acid derived hydroperoxides were capable of epoxidizing a lipid peroxidation product 4-hydroxy-2-nonenal to a mutagenic epoxy aldehyde. Lipid hydroperoxides produced during the peroxidase cycle are also known to be capable of epoxidizing polycyclic aromatic hydrocarbon (PAHs) as an alternative pathway to P450-mediated activation of PAHs to mutagenic epoxides (Parkison, 2001; Reed, 1987). Additionally, non-P450-catalyzed epoxidation of the terminal furan of aflatoxin B1 (a hepatocarcinogen and hepatotoxin produced by fungal metabolism) was shown to be dependent on the preoxidation of arachidonic acid by prostaglandin H synthase (Parkinson, 2001; Reed, 1987; Battista and Mament, 1985). The feasibility of epoxidation by lipid hydroperoxides has therefore been demonstrated in the literature.

It seems unlikely that AZD9819 and related compounds are the only xenobiotic analytes susceptible to lipid peroxide-dependant oxidation. In the present study, we have demonstrated two practical, effective means of preventing the oxidation: Lowing the pH to stop the nucleophilic reaction of epoxidation by lipid hydroperoxides, or adding the antioxidant ascorbic acid to inhibit the radical reaction of lipid peroxidation. We hope that knowledge of this new possible cause of analyte instability in plasma samples and the means of prevention will prove useful for other scientists in biotransformation and bioanalysis.
ACKNOWLEDGMENT

We thank Prof. Neal Castagnoli of Virginia Tech, VA, and Dr. Richard Thompson, Dr. Matthew Perry and Dr. Per-Ola Norrby of AstraZeneca at Mölndal, Sweden, for helpful discussions. We thank our chemistry colleagues Peter R. Hansen for original synthesis of AZD9819, Grigorios Nikitidis for the scaled-up synthesis, and Tobias Persson for the synthesis of $[^{14}\text{C}]$AZD9819 at AstraZeneca at Lund in Sweden. We thank Ann Jönsson and Kristina Olsmar for the bioanalytical stability testing, Helena Nilsson and Dr. Per Brunmark for LC-MS work on in vitro metabolites at AstraZeneca at Lund, Sweden. We also thank a number of biotransformation experts from biotech and other pharmaceutical companies for stimulating discussions regarding partial results and a partial mechanism presented at two conferences.
AUTHORSHIP CONTRIBUTIONS

Participated in research design: Gu, Lewis, Wells, Hosagrahara, Johnsson, and Hallström

Conducted experiments: Gu, Lewis, Wells, Svensson, Johnsson, and Hallström

Contributed new reagents or analytic tools: Wells

Performed data analysis: Gu, Lewis, Svensson, Johnsson, and Hallström

Wrote or contributed to the writing of the manuscript: Gu, Lewis, Wells, Svensson, Hosagrahara, Johnsson, and Hallström
REFERENCES


spectrometry, and NMR. *Drug Metab Dispos* **31**:1437-1447


SCHEME LEGENDS

**Scheme 1.** Proposed mechanism for the lipid peroxide-dependant oxidation of the pyrazinone-carboxamide core of AZD9819 and subsequent rearrangement.

**Scheme 2.** Postulated mechanism for nucleophilic epoxidation of AZD9819 by lipid hydroperoxides.

FIGURE LEGENDS

**Figure 1.** Radiochromatograms showing products formed by incubation of 14 μM [14C]AZD9819 in (A) rat, (B) dog, and (C) human plasma at 37°C for 2 h.

**Figure 2.** HMBC 2D NMR of the ROP synthetic standard. The highlighted correlation suggests the migration of the acetyl group from 4 to 5 in Scheme 1.

**Figure 3.** X-ray crystallography molecular structure of ROP synthetic standard (50% probability ellipsoids). One of the molecules in the asymmetric unit is shown (Z’=2).

**Figure 4.** LC-MS spectra of protonated molecular ion regions showing (A) rat plasma generated ROP, and (B) the incorporation of an 18O atom from 18O2 added to the incubation. LC-MS/MS CID product ions spectra acquired for (C) [M+H]+ ions at m/z 509, and (D) 18O-[M+H]+ ions at m/z 511, indicating 18O incorporation at the location predicted by the proposed mechanism in Scheme 1.

**Figure 5.** LC-MS accurate mass extracted ion chromatograms following a 2 h incubation of 15 μM AZD9819 at 37 °C in (A) phosphate pH 7.4 buffer, in (B) 1 mg/mL human LDL in phosphate pH 7.4 buffer, (C) B +50 mM GSH, (D) B +50 mM ascorbic acid, and (E) 1 mg/mL human LDL prepared in glycine pH 10.5 buffer. Also displayed are (F) yields of ROP with different incubations, according to LC-UV peak areas and relative UV signal response of two compounds. Error bars are plotted in the standard deviation of triplicate incubations.
Table 1. Comparison of carbon NMR chemical shifts of selected equivalent carbons as numbered in Scheme 1 in AZD9819 and ROP comparing with predicted shifts (ACD labs carbon predictor 2012 build 60488)

<table>
<thead>
<tr>
<th>Carbon</th>
<th>AZD9819 δObs./ppm</th>
<th>AZD9819 δPred./ppm</th>
<th>ROP δObs./ppm</th>
<th>ROP δPred./ppm</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>161.3</td>
<td>162.0</td>
<td>149.8</td>
<td>147.2</td>
</tr>
<tr>
<td>2</td>
<td>143.3</td>
<td>141.7</td>
<td>126.5</td>
<td>127.5</td>
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<tr>
<td>3</td>
<td>154.4</td>
<td>152.8</td>
<td>158.2</td>
<td>159.9</td>
</tr>
<tr>
<td>4</td>
<td>18.7</td>
<td>16.3</td>
<td>21.8</td>
<td>23.2</td>
</tr>
<tr>
<td>5</td>
<td>142.4</td>
<td>135.4</td>
<td>169.3</td>
<td>168.2</td>
</tr>
<tr>
<td>6</td>
<td>122.5</td>
<td>126.6</td>
<td>148.5</td>
<td>147.5</td>
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</table>
Table 2. Qualitative comparison in the effect of different agents or experimental conditions on the formation of ROP from AZD9819. All incubations and controls are conducted at 37 °C for 2 h, unless specified otherwise.

<table>
<thead>
<tr>
<th>Agent or condition</th>
<th>Conc. of agent</th>
<th>Control</th>
<th>ROP (% of control)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Control plasma</td>
<td>Rat plasma</td>
</tr>
<tr>
<td>Amber vial</td>
<td>N/A</td>
<td>Clear vial</td>
<td>95</td>
</tr>
<tr>
<td>Completely dark</td>
<td>N/A</td>
<td>Ambient light</td>
<td>101</td>
</tr>
<tr>
<td>NaN₃</td>
<td>1 &amp; 50 mM</td>
<td>w/o NaN₃</td>
<td>94 &amp; 84</td>
</tr>
<tr>
<td></td>
<td>10 mM</td>
<td>w/o NaN₃</td>
<td>119</td>
</tr>
<tr>
<td>Allopurinol</td>
<td>100 µM</td>
<td>w/o Allopurinol</td>
<td>99</td>
</tr>
<tr>
<td></td>
<td>100 µM</td>
<td>w/o Allopurinol</td>
<td>102</td>
</tr>
<tr>
<td>KCN</td>
<td>50 mM</td>
<td>w/o KCN</td>
<td>209</td>
</tr>
<tr>
<td>Room temp.</td>
<td>N/A</td>
<td>37 °C</td>
<td>57</td>
</tr>
<tr>
<td></td>
<td>N/A</td>
<td>37 °C</td>
<td>74</td>
</tr>
<tr>
<td>On ice</td>
<td>N/A</td>
<td>37 °C</td>
<td>9</td>
</tr>
<tr>
<td>Preheated plasma at 56°C for 1 h</td>
<td>N/A</td>
<td>Plasma w/o preheating</td>
<td>61</td>
</tr>
<tr>
<td>Denatured plasma</td>
<td>1:1 MeCN</td>
<td>Plasma</td>
<td>3</td>
</tr>
<tr>
<td>Catalase</td>
<td>1000 unit/mL</td>
<td>w/o catalase</td>
<td>82</td>
</tr>
<tr>
<td>Ascorbic acid</td>
<td>1 mM</td>
<td>w/o added ascorbic acid</td>
<td>76</td>
</tr>
<tr>
<td></td>
<td>50 mM</td>
<td>w/o added ascorbic acid</td>
<td>25</td>
</tr>
<tr>
<td>GSH</td>
<td>5 mM</td>
<td>w/o added GSH</td>
<td>33</td>
</tr>
<tr>
<td></td>
<td>50 mM</td>
<td>w/o added GSH</td>
<td>16</td>
</tr>
</tbody>
</table>
Scheme 2
Figure 1
Figure 2
Figure 4

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Figure 5

[Diagram showing chromatograms labeled A to E, with peak annotations for AZD9819 and ROP at different time points.]

Figure 5
Lipid peroxide mediated oxidative rearrangement of drug molecules in blood plasma samples

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Supplemental Figure 2. Carbon NMR spectrum of AZD9819 (CD$_3$SOCD$_3$, 151 MHz, 298 K). Assignments on the spectrum correspond to the arbitrarily numbered atoms on the structure.
Supplemental Figure 3. Proton NMR spectrum of ROP (CD$_3$SOCD$_3$, 600 MHZ, 298 K). Assignments on the spectrum correspond to the arbitrarily numbered atoms on the structure.
Supplemental Figure 4. Carbon NMR spectrum of ROP (CD$_3$SOCD$_3$, 151 MHz, 298 K). Assignments on the spectrum correspond to the arbitrarily numbered atoms on the structure.
Supplemental Figure 5. Verification that synthetic ROP standard is identical to plasma generated ROP by LC-UV- high resolution MS/MS: UV chromatograms of (A) AZD9819 and ROP standards in a mixed solvent, and (B) dog plasma incubate of AZD9819, and UV spectra and LTQ-Orbitrap CID MS/MS spectra for (C) AZD9819, (D) ROP synthetic standard, (E) dog plasma generated ROP. Unassigned low-abundance fragment ions are noise in the LC-MS/MS and not produced from respective [M+H]+ ions. Two assigned +H₂O ions are ion-molecule reaction products with water vapor in the vacuum, which is normal for these fragment ion structures.
Supplemental Figure 6. Verification that plasma generated ROP and \[^{14}\text{C}]\text{ROP} at our different laboratories have the same chemical structure by high resolution MS/MS: Product ion spectra acquired in a Q-ToF Premier instrument for (A) \[^{14}\text{C}]\text{AZD9819}, and (B) rat plasma generated \[^{14}\text{C}]\text{ROP}; in comparison with MS/MS product ion spectra acquired with “HCD” in a LTQ-Orbitrap XL instrument for (C) AZD9819, (D) rat plasma generated ROP, and (E) synthetic ROP standard. Additionally, an incorrectly assigned fragment ion structure during the preliminary MS/MS data interpretation is provided, as it correctly hinted an oxazole new core for ROP. Several assigned \(^{+}\text{H}_2\text{O}\) ions are ion-molecule reaction products with water vapor in the vacuum, which is normal for these fragment ion structures.
Supplemental Figure 7. “Road map” with various tentative structures examined during the structural elucidation for ROP. Two challenging facts about the structural characterization were (i) no proton at the center core of AZD9819 and ROP making NMR interpretation complicated; and (ii) not a single fragment of ROP resembling that of AZD9819 in MS/MS. Hence, putative rearrangement mechanisms were proposed to generate hypothetical products to match to $^{13}$C NMR and MS/MS data.
Supplemental Figure 8. LC-MS/MS product ion spectrum of naturally occurring isotopic ions at m/z 511 of protonated ROP. The fragmentation pattern is distinct from that shown for $^{18}$O-incorporated ROP ions at m/z 511 in Figure 4D, because the naturally occurring isotopic ions of ROP at m/z 511 consist of mainly the mixture of $^{13}$C and $^{15}$N isotopic atoms (one each or both the same atom); only approximately 1/8 consists of $^{18}$O.
Supplemental Figure 9. Heat inactivation of rat plasma proving deacetyl-ROP to be an enzymatic reaction product of ROP, but not preventing the ROP production from AZD9819. Bar charts showing the product formation following the incubation of (A) 15 μM ROP synthetic standard, and (B) 15 μM AZD9819 at 37°C for 2 h in rat plasma or in pretreated rat plasma. Pre-heating rat plasma at 56 °C for 1 h abolished the enzymatic hydrolysis of ROP synthetic standard (panel A). Whereas, ROP was still produced from parent drug AZD9819 in the pre-heated plasma (panel B). In contrast, addition of acetonitrile at t = 0 stopped the ROP formation from AZD9819 in the denatured plasma (panel B).
Supplemental Figure 10. Accidental discovery that ROP could be produced from 15 µM AZD9819 by high concentration H₂O₂ as shown by (A) LC-UV chromatogram, and LC-MS accurate mass extract ion chromatograms of (B) [M+H]⁺ and (C) [M+Na]⁺ ions, following a mistaken incubation of AZD9819 in pH7.4 phosphate buffer with 90 mM H₂O₂, instead of 90 µM H₂O₂ as planned (data not shown). Measured accurate mass m/z values of [M+H]⁺ and [M+Na]⁺ ions of AZD9819, ROP, and two hydration products are labeled. Based on LC-MS/MS spectra of [M+H]⁺ ions, the peak preceding AZD9819 is probably the oxidative hydrolysis product of ROP, with the nitrile group of ROP being converted to an amide by the high concentration H₂O₂. The peak after AZD9819 has not yet been identified.