

**Impact of Basolateral Mrp3 (Abcc3) and Mrp4 (Abcc4) on the Hepatobiliary
Disposition of Fexofenadine in Perfused Mouse Livers**

Xianbin Tian, Brandon Swift, Maciej J. Zamek-Gliszczynski, Martin G. Belinsky, Gary
D. Kruh and Kim L. R. Brouwer

School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599,
USA (X.T., B.S., M.J.Z-G., K.L.R.B)

Medical Science Division, Fox Chase Cancer Center, Philadelphia, Pennsylvania
(M.G.B., G.D.K.)

Running title:

Role of Mrps in fexofenadine hepatic disposition

Address correspondence to:

Kim L. R. Brouwer, Pharm.D., Ph.D.

Division of Pharmacotherapy and Experimental Therapeutics

School of Pharmacy, C.B. #7360 Kerr Hall

University of North Carolina at Chapel Hill, Chapel Hill, N.C. 27599-7360

Phone (919) 962-7030; Fax (919) 962-0644; email: kbrouwer@unc.edu

Number of Pages: 25

Number of Tables: 2

Number of Figures: 3

Number of References: 28

Number of Words:

Abstract: 207

Introduction: 632

Discussion: 834

List of Nonstandard Abbreviations:

Mrp, multidrug resistance-associated protein; organic anion transporting polypeptide (Oatp); WT, wild-type C57BL/6; *Abcc2*(-/-), Mrp2 gene knockout; *Abcc3*(-/-), Mrp3 gene knockout; *Abcc2*(-/-)/*Abcc3*(-/-), Mrp2 and Mrp3 double-knockout; *Abcc4*(-/-), Mrp4 gene knockout.

Abstract

The disposition of fexofenadine, a commonly-used antihistamine drug, is governed primarily by active transport. Biliary excretion of the parent compound is the major route of systemic clearance. Previous studies demonstrated that fexofenadine hepatic uptake is mediated by organic anion transporting polypeptides. Recently, we showed that in mice fexofenadine is excreted into bile primarily by Mrp2 (Abcc2). In the present study, the role of Mrp3 (Abcc3) and Mrp4 (Abcc4) in the hepatobiliary disposition of fexofenadine was examined in knockout mice using *in situ* liver perfusion. Compared to wild-type mice, basolateral excretion of fexofenadine was impaired resulting in a ~50% decrease in perfusate recovery in *Abcc3(-/-)* mice; in contrast, fexofenadine hepatobiliary disposition was unaltered in *Abcc4(-/-)* mice. As expected, in *Abcc2(-/-)* mice, fexofenadine was redirected from the canalicular to the basolateral membrane for excretion. In *Abcc2(-/-)/Abcc3(-/-)* double knock-out mice, fexofenadine biliary excretion was impaired, but perfusate recovery was similar to wild-type mice, and more than 2-fold higher than in *Abcc3(-/-)* mice, presumably due to compensatory basolateral transport mechanism(s). These results demonstrate that multiple transport proteins are involved in the hepatobiliary disposition of fexofenadine. In addition to Mrp2 and Mrp3, other transport proteins play an important role in the biliary and hepatic basolateral excretion of this zwitterionic drug.

Introduction

Fexofenadine is an H1 receptor antagonist commonly used in the treatment of seasonal allergies and chronic urticaria. The intestinal absorption of fexofenadine is mediated by organic anion transporting polypeptides (Oatps), a process that is counteracted by P-glycoprotein-mediated efflux (Cvetkovic et al., 1999). Metabolism accounts for less than 5% of total fexofenadine clearance in humans, and the oral and intravenous exposure of fexofenadine in rats is not altered by the pan-cytochrome P450 mechanism-based inactivator, aminobenzotriazole (Strelevitz et al., 2005). However, specific information regarding fexofenadine metabolism in mice is not available. Biliary excretion of unchanged fexofenadine is the primary route of systemic clearance (Tahara et al., 2005). Although fexofenadine intestinal absorption and blood-brain barrier penetration are limited by P-glycoprotein (Cvetkovic et al., 1999; Tahara et al., 2005), the biliary excretion of fexofenadine is mediated by multiple transport mechanisms (Tian et al., 2008). The unusual dispositional properties of fexofenadine, i.e. low metabolism and extensive transport, have led to its use as a probe for the analysis of both Oatp and P-glycoprotein modulation by various co-administered drugs and herbal products (Banfield et al., 2002; Zhou et al., 2004; Shon et al., 2005; Lemma et al., 2006).

Following uptake from sinusoidal blood into the hepatocyte, drugs and derived metabolites, may undergo excretion across the canalicular membrane into bile or across the basolateral membrane into blood. For example, carboxydichlorofluorescein is taken up into hepatocytes via basolateral Oatps, and is subsequently excreted unchanged into bile by multidrug resistance-associated protein (Mrp) 2 and also undergoes basolateral export into blood by Mrp3 (Zamek-Gliszczyński et al., 2003; Nezasa et al., 2006).

Similarly, glucuronide conjugates of acetaminophen and morphine also are subject to bidirectional excretion into bile and blood by Mrp2 and Mrp3, respectively (Xiong 2000, 2002, van de Wetering 2007, Zelcer 2005). Basolateral Mrp3 is similar to canalicular Mrp2 in terms of amino acid sequence, membrane topology, and substrate specificity (Konig et al., 1999; Ogawa et al., 2000). Mrp3 is highly upregulated in obstructive cholestasis (e.g. bile duct ligation) and hereditary conjugated hyperbilirubinemia in rats and humans (e.g. Mrp2-deficient animals, Dubin-Johnson patients), acting as an overflow pump for hepatic excretion of Mrp2 substrates and conjugated bile acids when biliary excretion is impaired (Hirohashi et al., 1998; Konig 1999). Although expression levels of Mrp3 protein are relatively high in mice, a 2-fold up-regulation of Mrp3 protein is still observed in Mrp2-knockout mice (Nezasa et al., 2005). Based upon recent findings that the biliary excretion of fexofenadine is mediated in part by Mrp2 in mice (Tian et al., 2008), we hypothesized that fexofenadine may also undergo hepatic basolateral excretion via Mrp3. In addition to Mrp3, Mrp4 functions as an excretory mechanism on the hepatic basolateral membrane (Rius et al., 2003). Limited evidence of functional overlap between Mrp3 and Mrp4 exists. For example, both Mrp3 and Mrp4 are upregulated in response to obstructive cholestasis (Hirohashi et al., 1998; Denk et al., 2004). Furthermore, both Mrp3 and Mrp4 mediate the hepatic basolateral excretion of the sulfate metabolites of acetaminophen, 4-methylumbelliferone, and harmol in mice (Zamek-Gliszczynski et al., 2006). While these features suggest that Mrp3 and Mrp4 may be involved in the hepatic basolateral excretion of fexofenadine, the extent to which these pumps function in this capacity has not been investigated.

Studies outlined in this manuscript tested the hypothesis that fexofenadine undergoes basolateral excretion from the liver via Mrp3 and Mrp4. Perfused livers from relevant transporter-gene knockout mice were utilized to examine the role of Mrp2, Mrp3 and Mrp4 in the hepatobiliary disposition of fexofenadine. Furthermore, the combined role of Mrp2 and Mrp3 in the overall hepatic excretion of this zwitterion was studied in livers from *Abcc2(-)/Abcc3(-)* double-knockout mice. The data presented herein demonstrate the importance of Mrp2 and Mrp3 in the biliary and hepatic basolateral excretion, respectively, of fexofenadine in mice.

Material and Methods

Chemicals. Fexofenadine, cetirizine, taurocholate, and Krebs-Henseleit buffer packets were purchased from Sigma Chemical Co. (St. Louis, MO). All other chemicals were of reagent grade and were readily available from commercial sources.

Animals. Male C57BL/6 wild-type (WT) (age-matched heterozygotes), *Abcc2*(-/-), *Abcc3*(-/-), *Abcc2*(-/-)/*Abcc3*(-/-), and *Abcc4*(-/-) mice (23-31g) were generated as described previously (Belinsky et al., 2005; Belinsky et al., 2007). *Abcc2*(-/-) mice have been generated recently by gene-targeting methods in the laboratory of Dr. Gary Kruh (unpublished). *Abcc2*(-/-), *Abcc3*(-/-) and *Abcc4*(-/-) mice employed in this study were derived from mixed 129/C57BL/6 animals and were backcrossed to C57BL/6 for eight generations. *Abcc2*(-/-) and *Abcc3*(-/-) mice on the C57BL/6 background were used to breed the *Abcc2*(-/-)/*Abcc3*(-/-) mice. Mice were maintained on a 12-h light/dark cycle with free access to water and rodent chow. Experiments were performed at Fox Chase Cancer Center and approved by their Institutional Animal Care and Use Committee.

In Situ Liver Perfusion. All experimental procedures were performed under full anesthesia induced with ketamine/xylazine (140/8 mg/kg i.p.). The liver perfusion procedure was modified slightly from the previous report of Tian et al (2007). Briefly, the common bile duct was ligated above the duodenum to prevent bile from entering the intestine, and the gallbladder was cannulated with PE-10 tubing (Becton Dickinson, Parsippany, NJ). The portal vein was cannulated with a 20-gauge catheter (B. Braun Medical Inc., Bethlehem, PA), the abdominal vena cava below the liver was severed immediately by incision, and the inferior vena cava above the liver was cannulated with a 20-gauge catheter. The liver was perfused (5 ml/min with fexofenadine-free continually

oxygenated Krebs-Henseleit buffer containing 5 μM taurocholate) for an equilibration period of approximately 15 min. Subsequently, the inferior vena cava was ligated between the liver and kidney to direct all perfusate outflow through the cannulated inferior vena cava above the liver. Following the pre-perfusion period to allow for equilibration of temperature and bile flow, the liver was perfused for 30 min with buffer containing 0.5 μM fexofenadine, followed by a washout with fexofenadine-free buffer for 45 min. Bile was collected in 7-min intervals; outflow perfusate was collected at designated time points *in toto*. Perfusion pressure and bile flow were monitored to assess liver viability (Chandra et al., 2005).

Analytical Methods. Bile and outflow perfusate samples were analyzed by liquid chromatography with detection by tandem mass spectrometry (Applied Biosystems API 4000 triple quadrupole with TurboIonSpray interface, MDS Sciex, Concord, ON, Canada). Fexofenadine and cetirizine (internal standard) were eluted from an Aquasil C18 column (5 μm , 50mm x 2.1mm, Thermo-Electron, Waltham, MA) using a mobile phase gradient (A: 0.1% formic acid in water, B: 0.1% formic acid in methanol); 0-0.8 min hold at 10% B, 0.8-3.5 min linear increase to 85% B, 3.5-4.0 min hold at 85% B, 4.0-4.5 min return to 10% B, 4.5-5 min hold at 10% B; flow rate was 0.75 mL/min (Shimadzu solvent delivery system, Columbia, MD). Fexofenadine and the internal standard, cetirizine, were detected in positive ion mode using multiple reaction monitoring: fexofenadine: 502.3 \rightarrow 466.4 m/z, cetirizine: 389.0 \rightarrow 201.0 m/z. Fexofenadine was quantified with standard curves prepared in the appropriate matrix.

Data Analysis. All data were reported as mean \pm SD (n = 3-4, per group). Statistical significance was assessed by one-way analysis of variance (ANOVA) with Tukey's *post-*

hoc test. In cases where the normality test failed, data were log-transformed prior to statistical analysis. In all cases, $p < 0.05$ was considered to be statistically significant.

Results

Body weight, liver weight, and bile flow. Body weight was comparable between WT and knockout mice (Table 1). Liver weight (normalized for body weight) was significantly increased (~30%) in *Abcc2*(-/-) and (~46%) in *Abcc2*(-/-)/*Abcc3*(-/-) relative to WT mice. *Abcc3*(-/-) and *Abcc4*(-/-) mouse liver weights were comparable to WT controls (Table 1). The bile flow rate was decreased ~30% in *Abcc2*(-/-) and *Abcc2*(-/-)/*Abcc3*(-/-) mouse livers relative to WT, but was normal in livers from *Abcc3*(-/-) and *Abcc4*(-/-) mice (Table 1).

Recovery of fexofenadine in outflow perfusate. Fexofenadine concentrations in outflow perfusate from WT and knockout mouse livers are presented in Figure 1. For clarity, only mean data are plotted; standard deviations were within ~50% of the mean data. The total recovery of fexofenadine at the end of the washout period as a percentage of the infused dose was $73 \pm 10\%$ in WT, $62 \pm 30\%$ in *Abcc3*(-/-), $68 \pm 9\%$ in *Abcc4*(-/-), $77 \pm 12\%$ in *Abcc2*(-/-), and $71 \pm 15\%$ in *Abcc2*(-/-)/*Abcc3*(-/-) mouse livers. In order to quantify the impact of Mrp3 and Mrp4 genetic ablation on the basolateral efflux of fexofenadine from the liver, fexofenadine recovery in perfusate during the washout phase was expressed as a percentage of liver fexofenadine content at the end of the infusion (determined as the difference between infused dose and the cumulative mass of fexofenadine excreted in outflow perfusate and bile through 30 min; Table 2). The fraction of fexofenadine excreted into outflow perfusate was significantly decreased (~50%) in livers from *Abcc3*(-/-) mice; in contrast, significantly increased perfusate recovery (~60%) was noted in *Abcc2*(-/-) relative to WT mouse livers. Interestingly, perfusate recovery of fexofenadine in livers from *Abcc2*(-/-)/*Abcc3*(-/-) mice was similar

to WT control livers. In contrast to livers from *Abcc3*(*-/-*) mice, perfusate recovery of fexofenadine in livers from *Abcc4*(*-/-*) mice was unchanged.

Recovery of fexofenadine in bile. Fexofenadine biliary excretion rates and cumulative biliary excretion in livers from WT and knockout mice are presented in Figures 2A and B, respectively; standard deviations were within ~50% of the mean data and are not included in the figures for clarity. The biliary recovery of fexofenadine was not significantly different in *Abcc3*(*-/-*) and *Abcc4*(*-/-*) mouse livers relative to WT controls (Figure 2B; Table 2). Fexofenadine biliary excretion rates and cumulative biliary excretion were significantly decreased in livers from *Abcc2*(*-/-*) and *Abcc2*(*-/-*)/*Abcc3*(*-/-*) mice (Figure 2). The biliary recovery of fexofenadine during the washout phase, normalized for liver content of fexofenadine at the end of the infusion, was ~61% and ~53% lower in *Abcc2*(*-/-*) and *Abcc2*(*-/-*)/*Abcc3*(*-/-*) relative to WT mouse livers, respectively (Table 2). Interestingly, liver fexofenadine concentrations at the end of the washout period were not altered in *Abcc3*(*-/-*), *Abcc4*(*-/-*) and *Abcc2*(*-/-*)/*Abcc3*(*-/-*) mice (Table 2). However, consistent with our previous report (Tian et al., 2008), liver fexofenadine concentrations decreased ~2-fold in *Abcc2*(*-/-*) mice after the washout phase (Table 2).

Discussion

Previous studies utilizing transporter gene knockout mice identified Mrp2 as the predominant protein responsible for fexofenadine biliary excretion (Tian et al., 2008). In contrast, fexofenadine biliary excretion was not impaired in mice lacking the breast cancer resistance protein or P-glycoprotein (Tahara et al., 2005). In agreement with previous work, Mrp2 was responsible for ~60% of fexofenadine biliary excretion in the current study. However, the present data also establish the importance of Mrp3 in the hepatic basolateral excretion of fexofenadine. Approximately 50% of fexofenadine basolateral excretion may be attributed to Mrp3-mediated transport when outflow perfusate concentrations are in the range of 300 ng/ml based on the 2-fold decrease in perfusate recovery of fexofenadine in livers from *Abcc3(-/-)* mice. These data highlight the functional interplay between Mrp2 and Mrp3 in fexofenadine hepatobiliary disposition. However, the basolateral excretion of fexofenadine in *Abcc2(-/-)/Abcc3(-/-)* mice was not impaired. Fexofenadine concentrations in *Abcc3(-/-)* and *Abcc2(-/-)/Abcc3(-/-)* mouse livers were similar to those in WT mouse livers. Mechanism(s) responsible for the Mrp2- and Mrp3-independent component of fexofenadine hepatobiliary clearance remain to be elucidated.

Mrp2 plays a critical role as an organic anion excretory system in liver detoxification. Unlike human MRP3, MRP2 protein is highly expressed in humans under normal conditions (Stöckel et al., 2000). Protein expression of Mrp3, the “Mrp2 backup system”, is increased markedly in rats and humans that are deficient in Mrp2 (Hirohashi et al., 1998; König et al., 1999; Xiong et al., 2002). Mrp3 constitutive expression levels in mice are already high, but in Mrp2-knockout mice, Mrp3 is further upregulated (Nezasa

et al., 2006). In contrast, this functional compensation is not reciprocal in mouse liver; the absence of Mrp3 does not result in a notable upregulation of Mrp2 protein or function in mouse liver, as demonstrated by this and other studies (Belinsky et al., 2005; Zamek-Gliszczynski et al., 2006). This suggests that hepatic protection mechanisms other than Mrp2-mediated biliary excretion exist in mice when Mrp3 function is ablated. Mrp4 is another efflux transport protein located on the hepatic basolateral membrane (Rius et al., 2003). Limited evidence of functional overlap between Mrp3 and Mrp4 (Hirohashi et al 1999; Denk et al., 2004; Zamek-Gliszczynski et al., 2006) prompted the exploration of fexofenadine disposition in livers from *Abcc4*(-/-) mice. Interestingly, fexofenadine basolateral excretion was not affected by the absence of Mrp4. The impact of genetic ablation of Mrp4 has been associated with impaired Sulfotransferase 2a1 expression and function (Assem et al., 2004), but an appreciable effect on the function or expression of other hepatic transport proteins has not been observed (Assem et al., 2004; Mennone et al., 2006; Zamek-Gliszczynski et al., 2006). Whether Mrp4 serves as the backup system for Mrp3-mediated fexofenadine basolateral excretion requires further investigation.

The existence of multiple transport and backup transport systems on both the basolateral and canalicular membranes complicates the extrapolation of transport studies from single-knockout to double-knockout animals (van de Wetering et al., 2007). The biliary and basolateral excretion of the Mrp2 and Mrp3 substrate, morphine-3-glucuronide in *Abcc2*(-/-)/*Abcc3*(-/-) mice following morphine administration was severely impaired, resulting in considerable hepatic accumulation of the glucuronide metabolite. However, residual plasma concentrations of morphine-3-glucuronide in *Abcc2*(-/-)/*Abcc3*(-/-) mice were substantial, and the concentrations of morphine-3-

glucuronide were similar between *Abcc2*(-/-)/*Abcc3*(-/-) and WT mice 24 hrs post morphine administration, which resulted in near-normal urinary excretion of morphine-3-glucuronide, suggesting that a low-capacity backup transport system was present for the basolateral excretion of morphine-3-glucuronide into plasma in *Abcc2*(-/-)/*Abcc3*(-/-) mice (van de Wetering et al., 2007). In the present study, fexofenadine biliary excretion in livers from *Abcc2*(-/-)/*Abcc3*(-/-) mice was impaired to the same extent as in mice only lacking Mrp2; fexofenadine basolateral excretion in livers from these double-knockout mice was similar to WT mice, and was more than two-fold higher than in Mrp3 single-knockout mice. The unchanged liver concentrations of fexofenadine in *Abcc3*(-/-) mice, and considerable basolateral excretion of fexofenadine in the livers of *Abcc2*(-/-)/*Abcc3*(-/-) mice, are consistent with the hypothesis that additional clearance mechanisms compensate for the loss of Mrp3 function. Basolateral Mrps (other than Mrp3), the bidirectional organic anion transport proteins, or other basolateral export proteins may contribute to increased basolateral clearance of fexofenadine in *Abcc2*(-/-)/*Abcc3*(-/-) mouse livers. The possibility of altered fexofenadine metabolism in *Abcc3*(-/-) and *Abcc2*(-/-)/*Abcc3*(-/-) mouse livers cannot be ruled out because no detailed information on fexofenadine metabolism in mouse livers has been published.

In summary, fexofenadine is taken up into the hepatocyte and undergoes biliary as well as basolateral excretion. Using perfused livers from relevant transporter gene knockout mice, Mrp2 and Mrp3 were identified as important transport proteins mediating fexofenadine biliary and hepatic basolateral excretion, respectively. Mrp4 did not appear to contribute to hepatic basolateral excretion of this zwitterionic drug in the intact mouse liver in the presence of Mrp3 and/or Mrp2 proteins. Mechanism(s) responsible for the

residual, Mrp2- and Mrp3-independent, biliary and basolateral excretion of fexofenadine remain to be elucidated. Once again, this study exemplified the importance of using physiologically relevant systems such as the intact liver to evaluate the contributions of hepatic transport proteins in drug pharmacokinetic studies.

Acknowledgment

The authors would like to thank Arlene S. Bridges, Ph. D. for her analytical support and Peijin Zhang, Ph. D. for helpful scientific discussions.

References

- Assem M, Schuetz EG, Leggas M, Sun D, Yasuda K, Reid G, Zelcer N, Adachi M, Strom S, Evans RM, Moore DD, Borst P and Schuetz JD (2004) Interactions between hepatic Mrp4 and Sult2a as revealed by the constitutive androstane receptor and Mrp4 knockout mice. *J Biol Chem* **279**:22250-22257.
- Banfield C, Gupta S, Marino M, Lim J and Affrime M (2002) Grapefruit juice reduces the oral bioavailability of fexofenadine but not desloratadine. *Clin Pharmacokinet* **41**:311-318.
- Belinsky MG, Dawson PA, Shchaveleva I, Bain LJ, Wang R, Ling V, Chen ZS, Grinberg A, Westphal H, Klein-Szanto A, Lerro A and Kruh GD (2005) Analysis of the in vivo functions of Mrp3. *Mol Pharmacol* **68**:160-168.
- Belinsky MG, Guo P, Lee K, Zhou F, Kotova E, Grinberg A, Westphal H, Shchaveleva I, Klein-Szanto A, Gallo JM, Kruh GD (2007) Multidrug resistance protein 4 protects bone marrow, thymus, spleen, and intestine from nucleotide analogue-induced damage. *Cancer Res* **67**:262-268.
- Chandra P, Johnson BM, Zhang P, Pollack GM, Brouwer KL (2005) Modulation of hepatic canalicular or basolateral transport proteins alters hepatobiliary disposition of a model organic anion in the isolated perfused rat liver. *Drug Metab Dispos* **33**:1238-43.
- Cvetkovic M, Leake B, Fromm MF, Wilkinson GR and Kim RB (1999) OATP and P-glycoprotein transporters mediate the cellular uptake and excretion of fexofenadine. *Drug Metab Dispos* **27**:866-871.

- Denk GU, Soroka CJ, Takeyama Y, Chen WS, Schuetz JD and Boyer JL (2004)
Multidrug resistance-associated protein 4 is up-regulated in liver but down-regulated in kidney in obstructive cholestasis in the rat. *J Hepatol* **40**:585-591.
- Hirohashi T, Suzuki H, Ito K, Ogawa K, Kume K, Shimizu T and Sugiyama Y (1998)
Hepatic expression of multidrug resistance-associated protein-like proteins maintained in eisai hyperbilirubinemic rats. *Mol Pharmacol* **53**:1068-1075.
- Hirohashi T, Suzuki H and Sugiyama Y (1999) Characterization of the transport properties of cloned rat multidrug resistance-associated protein 3 (MRP3). *J Biol Chem* **274**:15181-15185.
- Konig J, Rost D, Cui Y and Keppler D (1999) Characterization of the human multidrug resistance protein isoform MRP3 localized to the basolateral hepatocyte membrane. *Hepatology* **29**:1156-1163.
- Lemma GL, Wang Z, Hamman MA, Zaheer NA, Gorski JC and Hall SD (2006) The effect of short- and long-term administration of verapamil on the disposition of cytochrome P450 3A and P-glycoprotein substrates. *Clin Pharmacol Ther* **79**:218-230.
- Mennone A, Soroka CJ, Cai SY, Harry K, Adachi M, Hagey L, Schuetz JD and Boyer JL (2006) Mrp4^{-/-} mice have an impaired cytoprotective response in obstructive cholestasis. *Hepatology* **43**:1013-1021.
- Nezasa K, Tian X, Zamek-Gliszczynski MJ, Patel NJ, Raub TJ and Brouwer KL (2006) Altered hepatobiliary disposition of 5 (and 6)-carboxy-2',7'-dichlorofluorescein in Abcg2 (Bcrp1) and Abcc2 (Mrp2) knockout mice. *Drug Metab Dispos* **34**:718-723.

Ogawa K, Suzuki H, Hirohashi T, Ishikawa T, Meier PJ, Hirose K, Akizawa T, Yoshioka M and Sugiyama Y (2000) Characterization of inducible nature of MRP3 in rat liver. *Am J Physiol Gastrointest Liver Physiol* **278**:G438-446.

Rius M, Nies AT, Hummel-Eisenbeiss J, Jedlitschky G and Keppler D (2003) Cotransport of reduced glutathione with bile salts by MRP4 (ABCC4) localized to the basolateral hepatocyte membrane. *Hepatology* **38**:374-384.

Shon JH, Yoon YR, Hong WS, Nguyen PM, Lee SS, Choi YG, Cha IJ and Shin JG (2005) Effect of itraconazole on the pharmacokinetics and pharmacodynamics of fexofenadine in relation to the MDR1 genetic polymorphism. *Clin Pharmacol Ther* **78**:191-201.

Stöckel B, König J, Nies AT, Cui Y, Brom M, Keppler D. 2000. Characterization of the 5'-flanking region of the human multidrug resistance protein 2 (MRP2) gene and its regulation in comparison with the multidrug resistance protein 3 (MRP3) gene. *Eur J Biochem.* **267**:1347-58.

Strelevitz TJ, Foti RS and Fisher MB (2006) In vivo use of the P450 inactivator 1-aminobenzotriazole in the rat: varied dosing route to elucidate gut and liver contributions to first-pass and systemic clearance. *J Pharm Sci* **95**:1334-1341.

Tahara H, Kusuhara H, Fuse E and Sugiyama Y (2005) P-glycoprotein plays a major role in the efflux of fexofenadine in the small intestine and blood-brain barrier, but only a limited role in its biliary excretion. *Drug Metab Dispos* **33**:963-968.

Tian X, Li J, Zamek-Gliszczynski MJ, Bridges AS, Zhang P, Patel NJ, Raub TJ, Pollack GM, Brouwer KL (2007) Roles of P-glycoprotein, Bcrp, and Mrp2 in biliary excretion of spiramycin in mice. *Antimicrob Agents Chemother.* **51**:3230-4.

- Tian X, Zamek-Gliszczynski MJ, Li J, Bridges AS, Nezasa K, Patel NJ, Raub TJ and Brouwer KL (2008) Multidrug Resistance-Associated Protein 2 (Mrp2) is Primarily Responsible for the Biliary Excretion of Fexofenadine (FEX) in Mice. *Drug Metab Dispos*, 36:61-4.
- van de Wetering K, Zelcer N, Kuil A, Feddema W, Hillebrand M, Vlaming ML, Schinkel AH, Beijnen JH and Borst P (2007) Multidrug resistance protein 2 and 3 provide alternative routes for hepatic excretion of morphine-glucuronides. *Mol Pharmacol*. **72**:387-94.
- Xiong H, Suzuki H, Sugiyama Y, Meier PJ, Pollack GM and Brouwer KL (2002) Mechanisms of impaired biliary excretion of acetaminophen glucuronide after acute phenobarbital treatment or phenobarbital pretreatment. *Drug Metab Dispos* **30**:962-969.
- Xiong H, Turner KC, Ward ES, Jansen PL and Brouwer KL (2000) Altered hepatobiliary disposition of acetaminophen glucuronide in isolated perfused livers from multidrug resistance-associated protein 2-deficient TR(-) rats. *J Pharmacol Exp Ther* **295**:512-518.
- Zamek-Gliszczynski MJ, Nezasa K, Tian X, Bridges AS, Lee K, Belinsky MG, Kruh GD and Brouwer KL (2006) Evaluation of the role of multidrug resistance-associated protein (Mrp) 3 and Mrp4 in hepatic basolateral excretion of sulfate and glucuronide metabolites of acetaminophen, 4-methylumbelliferone, and harmol in Abcc3^{-/-} and Abcc4^{-/-} mice. *J Pharmacol Exp Ther* **319**:1485-1491.

Zamek-Gliszczyński MJ, Xiong H, Patel NJ, Turncliff RZ, Pollack GM and Brouwer KL

(2003) Pharmacokinetics of 5 (and 6)-carboxy-2',7'-dichlorofluorescein and its diacetate promoiety in the liver. *J Pharmacol Exp Ther* **304**:801-809.

Zelcer N, van de Wetering K, Hillebrand M, Sarton E, Kuil A, Wielinga PR, Tephly T,

Dahan A, Beijnen JH and Borst P (2005) Mice lacking multidrug resistance protein 3 show altered morphine pharmacokinetics and morphine-6-glucuronide antinociception. *Proc Natl Acad Sci U S A* **102**:7274-7279.

Zhou S, Chan E, Pan SQ, Huang M and Lee EJ (2004) Pharmacokinetic interactions of

drugs with St John's wort. *J Psychopharmacol* **18**:262-276.

Footnotes

This work was supported by the National Institutes of Health (GM41935 to K.L.R.B. and CA73728 to G.D.K.) and by the National Cancer Institute (Core Grant CA06927 to Fox Chase Cancer Center).

Current address:

Wyeth, Discovery Pharmacokinetics, Andover, MA 01810, USA (X. T.)

Eli Lilly and Company, Drug Disposition, Indianapolis, IN 46285, USA (M.J.Z-G)

Department of Medicine, University of Illinois at Chicago, Chicago, IL 60612 (G.D.K.)

Figure Legends

Figure 1. Fexofenadine concentrations in outflow perfusate from wild-type C57BL/6 (●), *Abcc3*(-/-) (Δ), *Abcc4*(-/-) (∇), *Abcc2*(-/-) (□), and *Abcc2*(-/-)/*Abcc3*(-/-) (◇) mouse livers (mean data are plotted; n = 3-4 per group). Livers were perfused (5 ml/min) with fexofenadine (0.5 μM) for 30 min followed by a 45-min washout phase.

Figure 2. Biliary excretion rate (A) and cumulative biliary excretion (B) of fexofenadine in perfused livers from wild-type C57BL/6 (●), *Abcc3*(-/-) (Δ), *Abcc4*(-/-) (∇), *Abcc2*(-/-) (□), and *Abcc2*(-/-)/*Abcc3*(-/-) (◇) mice (mean data are plotted; n = 3-4 per group). Livers were perfused (5 ml/min) with fexofenadine (0.5 μM) for 30 min followed by a 45-min washout phase.

Table 1. Body weight, liver weight normalized for body weight, and bile flow rate in wild-type and transporter gene knockout mice. Mean \pm SD (n = 3-4 per group). * p < 0.05 knockout vs. wild type.

Mice	Body Weight (g)	Liver Weight / Body Weight	Bile Flow Rate (μ L/min/g liver)
Wild-Type	27.5 \pm 1.4	0.041 \pm 0.006	1.3 \pm 0.3
<i>Abcc3</i> (-/-)	27.9 \pm 3.1	0.043 \pm 0.001	1.3 \pm 0.2
<i>Abcc4</i> (-/-)	26.1 \pm 2.4	0.041 \pm 0.005	1.3 \pm 0.4
<i>Abcc2</i> (-/-)	29.4 \pm 1.9	0.053 \pm 0.001 *	0.87 \pm 0.02 *
<i>Abcc2</i> (-/-)/ <i>Abcc3</i> (-/-)	27.7 \pm 1.5	0.060 \pm 0.002 *	0.85 \pm 0.15 *

Table 2. Recovery of fexofenadine in perfusate and bile during the washout phase of mouse liver perfusions, and liver concentrations after the washout phase. Values are expressed as percentage of fexofenadine liver content after the 30-min infusion. Mean \pm SD (n = 3-4 per group). * p< 0.05 knockout vs. wild type.

Mice	Biliary Recovery (% liver content at 30 min)	Recovery in Perfusate (% liver content at 30 min)	Liver Concentration (ng/g liver)
Wild-Type	15.3 \pm 3.8	25.8 \pm 4.1	1500 \pm 330
<i>Abcc3</i> (-/-)	13.5 \pm 6.1	12.7 \pm 6.1*	1500 \pm 500
<i>Abcc4</i> (-/-)	12.3 \pm 3.3	26.2 \pm 7.6	1300 \pm 420
<i>Abcc2</i> (-/-)	5.9 \pm 3.4*	42.0 \pm 2.7 *	850 \pm 400
<i>Abcc2</i> (-/-)/ <i>Abcc3</i> (-/-)	7.1 \pm 2.9 *	28.6 \pm 12.3	1600 \pm 420

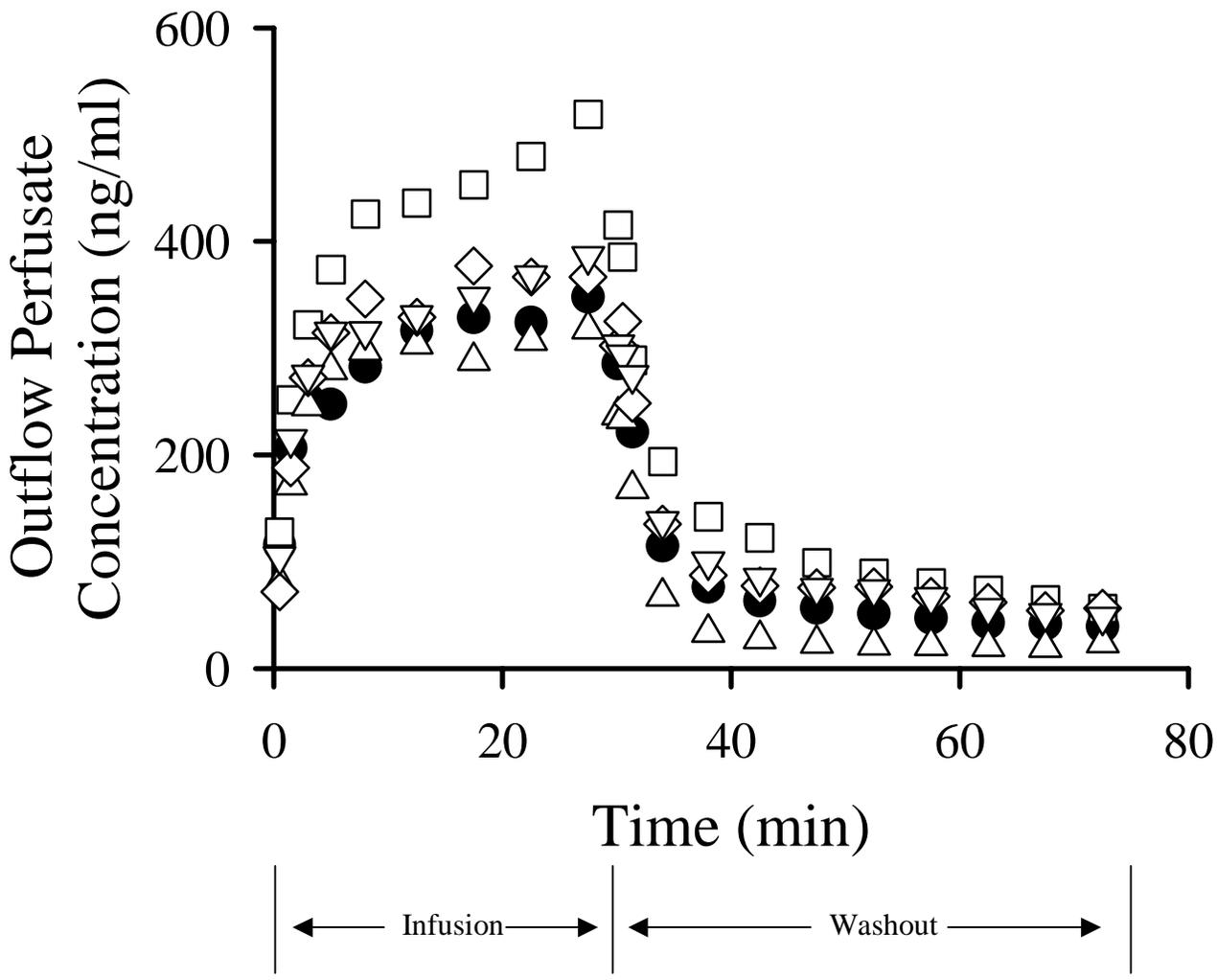


Figure 1

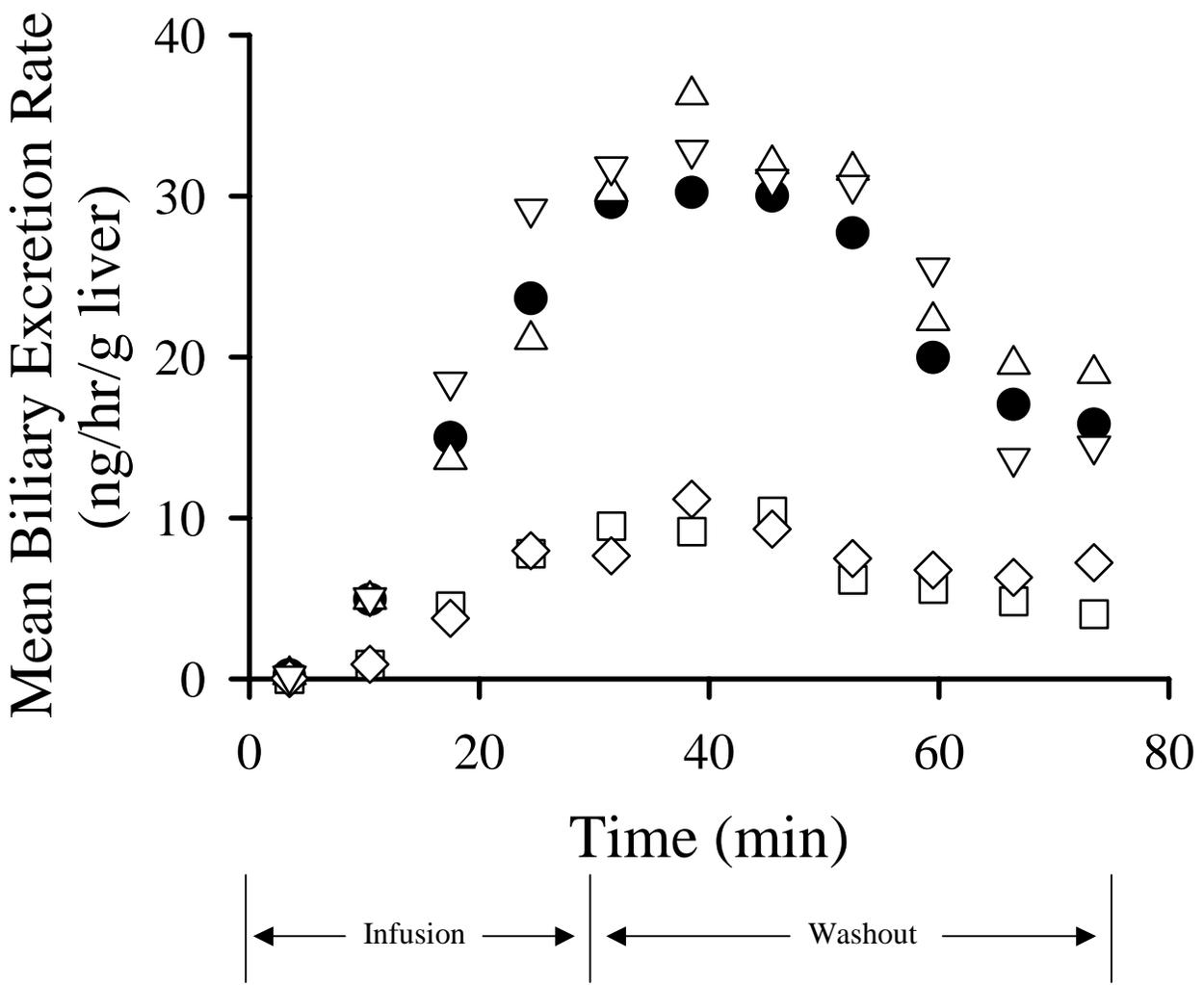


Figure 2A

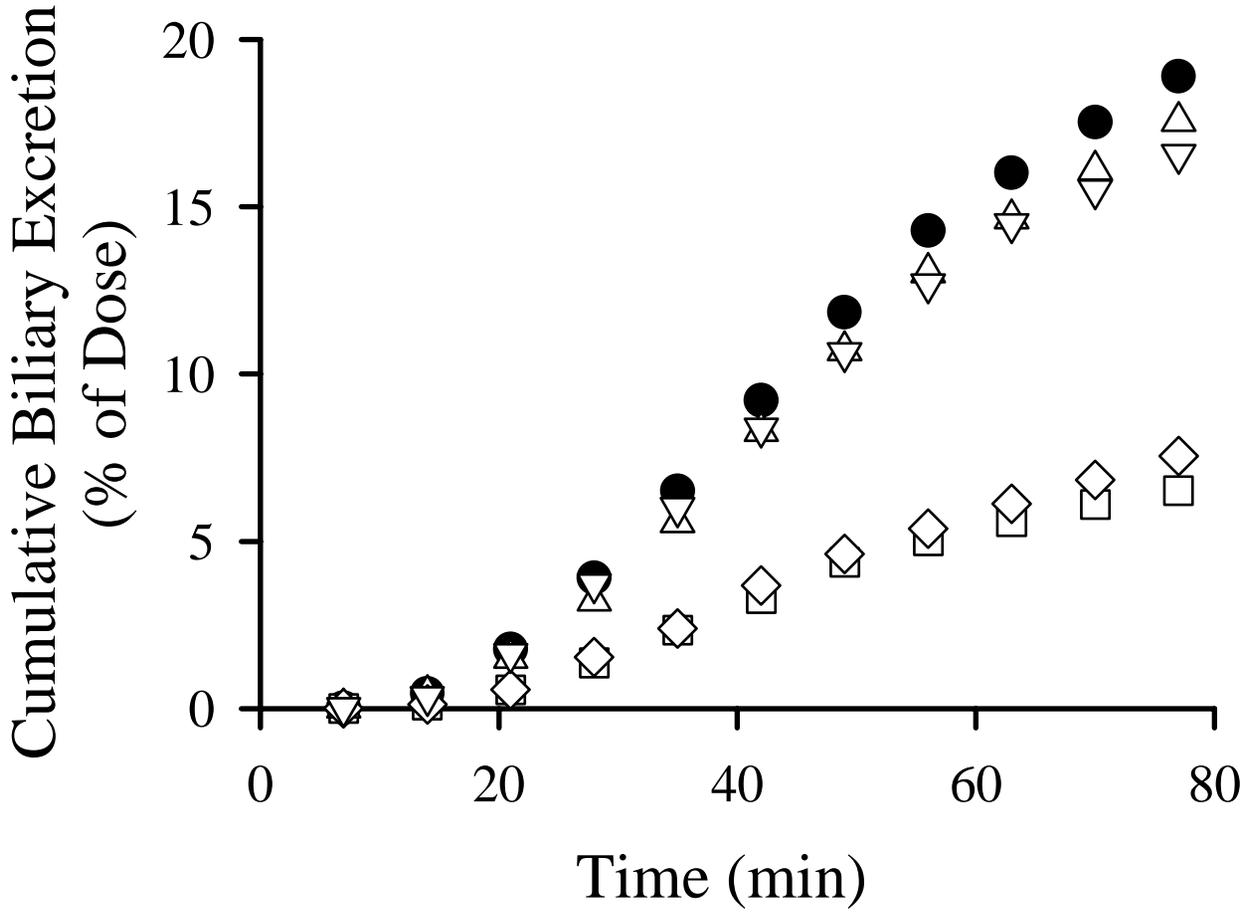


Figure 2B