

DMD #20388

Aldo-keto reductase 1C2 fails to metabolize doxorubicin and daunorubicin *in vitro*

Ryan H. Takahashi, Onkar S. Bains, Tom A. Pfeifer, Thomas A. Grigliatti, Ronald E. Reid and K. Wayne Riggs.

Division of Pharmaceutics and Biopharmaceutics, Faculty of Pharmaceutical Sciences (R.H.T., O.S.B., K.W.R.), University of British Columbia; Life Sciences Institute, Department of Zoology (T.A.P., T.A.G.), Faculty of Science, University of British Columbia; Division of Biomolecular and Pharmaceutical Chemistry (R.E.R.), Faculty of Pharmaceutical Sciences, University of British Columbia

A) Running title: Lack of *in vitro* anthracycline metabolism by AKR1C2

B) Corresponding Author: Dr. K. Wayne Riggs

Faculty of Pharmaceutical Sciences

University of British Columbia

2146 East Mall

Vancouver, BC, Canada, V6T 1Z3

Tel: (604) 822-2061

Fax: (604) 822-3035

E-mail: riggskw@interchange.ubc.ca

C) Number of text pages: 14

Number of tables: 0

Number of figures: 2

Number of references: 23

Number of words in abstract: 205

Number of words in introduction: 298

Number of words in results and discussion: 615

D) List of abbreviations: AKR: aldo-keto reductase; DAUN: daunorubicin; DAUNol: daunorubicinol; DOX: doxorubicin; DOXol: doxorubicinol; HPLC: high-performance liquid chromatography.

Abstract

The anthracycline drugs are important for the treatment of a number of malignancies; however, their clinical use is associated with dose-dependent severe chronic cardiotoxicity. While the mechanism for this side effect has not yet been identified, the alcohol metabolites formed during DAUN and DOX therapies have been implicated. The alcohol metabolites of DAUN and DOX, DAUNol and DOXol, respectively, are generated through reduction of the C-13 carbonyl function, which is reportedly mediated by members of the aldo-keto reductase and carbonyl reductase families of proteins. In our search for potential biomarkers for the occurrence of this side effect, we examined the activity of recombinant aldo-keto reductase enzymes, AKR1A1 and AKR1C2, with DAUN and DOX as substrates. Using purified histidine-tagged recombinant proteins and the direct measurement of metabolite formation with an HPLC-fluorescence assay, we did not observe DAUNol or DOXol generation *in vitro* by AKR1C2, whereas AKR1A1 did catalyze the reduction reactions. DAUNol was generated by AKR1A1 at a rate of 1.71 ± 0.09 pmol/min/mg protein and a low level of DOXol was produced by AKR1A1, however, it was below the limits of quantification for the method. These data suggest that the generation of DAUNol or DOXol by AKR1C2 metabolism *in vivo* is unlikely to occur during anthracycline treatment.

The anthracycline antibiotics, DOX and DAUN, are commonly-used, effective treatments for solid tumors and acute myeloid leukemias (Weiss, 1992). Unfortunately, the widespread use of the anthracyclines is limited by the development of dose-dependent chronic cardiotoxicity in patients. This side effect manifests as an irreversible disorder following long-term treatment with approximately 20% of cases leading to congestive heart failure (Singal and Iliskovic, 1998). Though the mechanism by which this side effect develops is not yet identified, evidence implicates metabolism as an important factor (Olson and Mushlin, 1990). Following DOX or DAUN administration, the C-13 alcohol metabolites, DOXol and DAUNol, are the major identified metabolites (Takanashi and Bachur, 1976), and these metabolites are speculated to be responsible for causing cardiac tissue damage (Del Tacca et al., 1985; Boucek et al., 1987; Cusack et al., 1993; Behnia and Boroujerdi, 1999; Olson et al., 2003; Sacco et al., 2003).

Formation of DOXol and DAUNol is mediated by cytosolic oxidoreductase enzymes of the AKR and short-chain dehydrogenase/reductase families, with the AKR1A1, AKR1B10, AKR1C2, and carbonyl reductase enzymes identified as major catalysts (Licata et al., 2000; Martin et al., 2006; Jin and Penning, 2007; Oppermann, 2007). Inhibition of the anthracycline-metabolizing enzymes is suggested as a strategy for avoiding treatment-related cardiotoxicity (Behnia and Boroujerdi, 1999) and chemical inhibitors for reductases and anthracycline derivatives that are metabolically less labile have been explored for their potential roles as safer alternatives to the currently-used anthracycline drug treatments. In part, the success of these strategies relies on accurately identifying the enzymes responsible for anthracycline metabolism.

In this paper, we characterize the involvement of two recombinant aldo-keto reductase enzymes, AKR1A1 and AKR1C2, in mediating the *in vitro* production of DAUNol and DOXol from DAUN and DOX using a specific and sensitive HPLC-fluorescence assay for direct measurement of the alcohol metabolites.

MATERIALS AND METHODS

Chemicals and enzymes. 1-acenaphthenol, *p*-nitrobenzaldehyde, DL-glyceraldehyde, DAUN, DOX, idarubicin, potassium phosphate, and NADPH were purchased from Sigma-Aldrich (St. Louis, MO). HPLC-grade acetonitrile and formic acid were purchased from Fisher Scientific (Fair Lawn, NJ). DNA modifying and restriction enzymes were obtained from Fermentas (Burlington, ON) and New England Biolabs, Inc. (Ipswich, MA). An authentic chemical standard for DOXol was obtained from Qventas Inc. (Branford, CT, USA).

Cloning of human AKR1A1 and 1C2. The *AKR1A1* and *AKR1C2* genes were sub-cloned from lab vector constructs (p2ZOp2N-*AKR1A1* and p2ZOp2N-*AKR1C2*) using standard molecular cloning techniques to the prokaryotic expression vector pET28a (Novagen, Madison, WI) as previously described (Bains, O.S.B. et al., 2008). The expression constructs encoded an AKR with a six-histidine affinity tag separated by a 23-amino acid residue linker attached to the amino terminus of the expressed protein.

Expression of recombinant enzymes. The AKR proteins were expressed and purified from *Escherichia coli* cultures. Briefly, pET28a-AKR constructs were transformed by heat-shock into BL21 (DE3) pLysS and grown in 500 ml low-salt LB media supplemented with 50 µg/ml kanamycin sulfate and 25 µg/ml chloramphenicol at 37°C. Expression was induced by addition of isopropyl β-D-1-thiogalactopyranoside administration to a final concentration of 1 mM and the cultures grown for a further 3 hours. Bacterial pellets were collected by centrifugation (4000 rpm for 20 mins at 4°C), then resuspended at 5 ml per gram wet weight with 300 mM NaCl, 50 mM NaH₂PO₄, (pH 8.0). Lysis of the cell suspensions and purification of the AKR proteins using Ni-NTA agarose were carried out according to the manufacturer's recommendations (QIAGEN, Mississauga, Ontario). Glycerol was added to the tagged purified protein to a final concentration of 20% and the samples stored at -20°C.

The recombinant proteins were detected by Western blot analysis using the Odyssey Infrared Imaging System (LI-COR[®] Biosciences, Lincoln, NE) with the primary monoclonal AKR1A1 and AKR1C2 antibodies (Abnova Corporation, Taipei City, Taiwan) diluted 1:5000 and 1:3000, respectively, and secondary IRDye 800CW goat anti-mouse IgG antibody (LI-COR[®] Biosciences), diluted 1:5000. Protein concentrations were determined by the Bradford method using bovine serum albumin as a standard (BioRad, Hercules, CA). Protein purity was assessed by SDS-PAGE stained with Sypro Ruby (Invitrogen, Burlington, ON).

Measurement of AKR enzyme activities. AKR activity was measured by monitoring the initial rate of NADP(H) oxidation/reduction reactions using standard conditions (Penning, 2004). Fluorescence measurements of reduced cofactor were made with excitation and emission wavelengths of 355 and 460 nm, respectively, using a Fluoroskan Ascent FL (Thermo Fisher Scientific, Waltham, MA). The assays were carried out with 200-275 ng purified protein, cofactor (180 μ M NADPH for AKR1A1; 2.3 mM NADP⁺ for AKR1C2) and 1 mM test substrate (*p*-nitrobenzaldehyde or DL-glyceraldehyde for AKR1A1; 1-acenaphthenol for AKR1C2) in a reaction mixture of 150 μ l 100mM potassium phosphate, pH 7, at 25°C. Maximal rates (min^{-1}) were calculated from the Ascent[®] program (version 2.6) using a 5 min interval. Enzymatic activities (μ mol cofactor consumed/min/mg purified protein) were calculated based on a standard curve constructed from fluorescence measurements for known NADPH concentrations. Organic solvents at a concentration below 4% (v/v) in the final mixture were used to solubilize some substrates and were not observed to affect enzyme function. All incubations were conducted in quadruplicate for each purified protein preparation.

Measurement of anthracycline reduction. Identification and quantification of DOXol and DAUNol were performed using high-performance liquid chromatography (HPLC; Waters

Alliance 2695, Milford, MA) and fluorescence detection with excitation and emission wavelengths of 460 and 550 nm, respectively (Waters 2475 Multi λ Detector). Quantification was based on peak area ratio of DOXol to the internal standard. Linear calibration was determined over the concentration range 25-500 nM using a $1/x^2$ weighting scheme. Because an authentic chemical standard for DAUNol could not be obtained, DAUNol was quantified as DOXol equivalents using a response ratio of 1.0. The limit of quantification for DOXol was estimated as 25 nM. All data processes were performed with Waters Empower software (ver 2.0).

Purified tagged AKR1A1 and AKR1C2 proteins were incubated in 150 μ l reaction mixtures containing 25 mM KH_2PO_4 , pH 7.4; and 1 μ M of DOX or DAUN at 37°C. Reaction conditions were selected to reflect a human physiological environment. Reactions were initiated with the addition of 1 mM NADPH. Incubations were conducted with up to 1 μ g of purified protein for 120 and 240 minutes for DAUN and DOX, respectively, to maximize metabolite generation given the apparent low activity of these enzymes. At the end of the incubations, reductase activity was stopped with the addition of 150 μ l of ice-cold acetonitrile that contained idarubicin as an internal standard. Protein was removed from the sample by vortex mixing and centrifugation at 10,000 \times g for 10 minutes at 4°C, and the supernatant was removed for HPLC analysis. All incubations were conducted in triplicate for each purified protein preparation.

RESULTS AND DISCUSSION

The expression of the recombinant tagged AKR1A1 and AKR1C2 was confirmed by Western blot analysis showing bands with mobility corresponding to the molecular weight of the tagged AKRs (~41-42 kDa) (Figure 1). Total protein staining of a SDS-PAGE gel demonstrated that the only detectable bands for the purified preparations corresponded to the AKR1A1 and AKR1C2 proteins identified by Western blotting (Figure 1).

Activities of the recombinant AKR1A1 using the test substrates *p*-nitrobenzaldehyde and DL-glyceraldehyde were 5.39 ± 0.35 and 1.56 ± 0.16 $\mu\text{mol}/\text{min}/\text{mg}$ purified protein, respectively, while the activity of the recombinant AKR1C2 using 1-acenaphthenol was 2.23 ± 0.08 $\mu\text{mol}/\text{min}/\text{mg}$. These activities are in good agreement with data reported for purified recombinant and native proteins by other laboratories: AKR1A1, *p*-nitrobenzaldehyde, 6.0 $\mu\text{mol}/\text{min}/\text{mg}$ (Palackal et al., 2001); DL-glyceraldehyde, 1.26 $\mu\text{mol}/\text{min}/\text{mg}$ (O'Connor et al., 1999); AKR1C2, 1-acenaphthenol, 2.5 $\mu\text{mol}/\text{min}/\text{mg}$ (Burczynski et al., 1998), providing confidence that the recombinant proteins have retained full reductase function.

Formation of DAUNol and DOXol by AKR1A1 and absence of metabolite generation by AKR1C2 were measured by HPLC-fluorescence (Figure 2). The C-13 metabolites were positively identified by identical chromatographic retention times to an authentic chemical standard of DOXol and to the metabolites generated in incubations with human liver cytosolic fractions. Mordente et al. (2001) measured the formation of DAUNol and DOXol by human cardiac cytosol reductases with average rates of 27.5 and 2.3 $\text{pmol}/\text{min}/\text{mg}$ protein, respectively. The high substrate specificity of AKR1A1 was also demonstrated in our results from recombinant enzymes where DAUN was metabolized to a much greater extent than DOX. The levels of DOXol generated were too low for quantification using the HPLC assay, whereas specific reductase activities of 1.71 ± 0.09 $\text{pmol}/\text{min}/\text{mg}$ were determined for DAUN reduction.

Our findings of a lack of AKR1C2 involvement in DOX and DAUN metabolism differ from previously published results (Ohara et al., 1995). This discrepancy may be due to contaminating proteins in previous enzyme preparations or non-specific measurement of reductase activities. SDS-PAGE gel staining for our enzyme preparations shows absence of any detectable contaminating proteins following affinity chromatography. Sypro Ruby total protein staining

provides detection of proteins at 1-2 ng (Berggren et al., 2000) and therefore we estimate protein contamination in our preparations at less than 0.01%. Monitoring NADPH absorbance is the standard assay to characterize reductase enzyme function; however, this method cannot distinguish contaminating protein-catalyzed or non-enzyme catalyzed cofactor usage, potentially leading to erroneous identification of reductase function or substrate use. In our attempts to use the spectroscopic method to measure cofactor usage in AKR1C2-catalyzed DOX and DAUN metabolism, we could not distinguish enzyme and non-enzyme catalyzed signal decrease with substrate concentrations of 10, 50, and 250 μM (data not shown). To specifically measure AKR activities for the anthracycline drugs DOX and DAUN, we have used a direct and sensitive HPLC-fluorescence assay and do not identify any alcohol metabolites formed by AKR1C2 that exceed the limit of quantification of the assay (25 nM). In our *in vitro* system, the initial anthracycline concentrations are 1 μM , a physiologically-relevant concentration for patient cardiac tissue (Stewart et al., 1993).

The failure of AKR1C2 to generate alcohol metabolites *in vitro* suggests that it is unimportant for the generation of DOX or DAUN metabolites *in vivo*. We recognize that *in vivo* studies are needed to clearly define the role of AKR1C2 in anthracycline metabolism; however, the conduct of definitive studies will be difficult. Several reductase enzymes are involved in the metabolism of these drugs and distinguishing their individual contributions to the total metabolism will be a challenge using the traditional approaches of correlation studies and chemical or antibody inhibition. It is with these considerations that we present our *in vitro* findings to focus future work on anthracycline metabolism on the other AKR isoforms and carbonyl reductases.

ACKNOWLEDGMENTS

The authors extend their gratitude to Dr. R.C. Mottus, Dr. J.W. Hodgson, Mr. J.G. Doheny, Mr. O. Toub, Ms. P. Kalas and Ms. M.A. Earp for their advice and technical expertise.

REFERENCES

- Bains OS, Takahashi RH, Pfeifer TA, Grigliatti TA, Reid RE and Riggs KW (2008) Two allelic variants of aldo-keto reductase 1A1 exhibit reduced in vitro metabolism of daunorubicin. *Drug Metab Dispos* **in press**.
- Behnia K and Boroujerdi M (1999) Inhibition of aldo-keto reductases by phenobarbital alters metabolism, pharmacokinetics and toxicity of doxorubicin in rats. *J Pharm Pharmacol* **51**:1275-1282.
- Berggren K, Chernokalskaya E, Steinberg TH, Kemper C, Lopez MF, Diwu Z, Haugland RP and Patton WF (2000) Background-free, high sensitivity staining of proteins in one- and two-dimensional sodium dodecyl sulfate-polyacrylamide gels using a luminescent ruthenium complex. *Electrophoresis* **21**:2509-2521.
- Boucek RJ, Jr., Olson RD, Brenner DE, Ogunbunmi EM, Inui M and Fleischer S (1987) The major metabolite of doxorubicin is a potent inhibitor of membrane-associated ion pumps. A correlative study of cardiac muscle with isolated membrane fractions. *J Biol Chem* **262**:15851-15856.
- Burczynski ME, Harvey RG and Penning TM (1998) Expression and characterization of four recombinant human dihydrodiol dehydrogenase isoforms: oxidation of trans-7, 8-dihydroxy-7,8-dihydrobenzo[a]pyrene to the activated o-quinone metabolite benzo[a]pyrene-7,8-dione. *Biochemistry* **37**:6781-6790.
- Cusack BJ, Mushlin PS, Voulelis LD, Li X, Boucek RJ, Jr. and Olson RD (1993) Daunorubicin-induced cardiac injury in the rabbit: a role for daunorubicinol? *Toxicol Appl Pharmacol* **118**:177-185.
- Del Tacca M, Danesi R, Ducci M, Bernardini C and Romanini A (1985) Might adriamycinol contribute to adriamycin-induced cardiotoxicity? *Pharmacol Res Commun* **17**:1073-1084.

- Jin Y and Penning TM (2007) Aldo-keto reductases and bioactivation/detoxication. *Annu Rev Pharmacol Toxicol* **47**:263-292.
- Licata S, Saponiero A, Mordente A and Minotti G (2000) Doxorubicin metabolism and toxicity in human myocardium: role of cytoplasmic deglycosidation and carbonyl reduction. *Chem Res Toxicol* **13**:414-420.
- Martin HJ, Breyer-Pfaff U, Wsol V, Venz S, Block S and Maser E (2006) Purification and characterization of akr1b10 from human liver: role in carbonyl reduction of xenobiotics. *Drug Metab Dispos* **34**:464-470.
- Mordente A, Meucci E, Martorana GE, Giardina B and Minotti G (2001) Human heart cytosolic reductases and anthracycline cardiotoxicity. *IUBMB Life* **52**:83-88.
- O'Connor T, Ireland LS, Harrison DJ and Hayes JD (1999) Major differences exist in the function and tissue-specific expression of human aflatoxin B1 aldehyde reductase and the principal human aldo-keto reductase AKR1 family members. *Biochem J* **343 Pt 2**:487-504.
- Ohara H, Miyabe Y, Deyashiki Y, Matsuura K and Hara A (1995) Reduction of drug ketones by dihydrodiol dehydrogenases, carbonyl reductase and aldehyde reductase of human liver. *Biochem Pharmacol* **50**:221-227.
- Olson LE, Bedja D, Alvey SJ, Cardounel AJ, Gabrielson KL and Reeves RH (2003) Protection from doxorubicin-induced cardiac toxicity in mice with a null allele of carbonyl reductase 1. *Cancer Res* **63**:6602-6606.
- Olson RD and Mushlin PS (1990) Doxorubicin cardiotoxicity: analysis of prevailing hypotheses. *Faseb J* **4**:3076-3086.

- Oppermann U (2007) Carbonyl reductases: the complex relationships of mammalian carbonyl- and quinone-reducing enzymes and their role in physiology. *Annu Rev Pharmacol Toxicol* **47**:293-322.
- Palackal NT, Burczynski ME, Harvey RG and Penning TM (2001) The ubiquitous aldehyde reductase (AKR1A1) oxidizes proximate carcinogen trans-dihydrodiols to o-quinones: potential role in polycyclic aromatic hydrocarbon activation. *Biochemistry* **40**:10901-10910.
- Penning TM (2004) Aldo-keto reductases and formation of polycyclic aromatic hydrocarbon o-quinones. *Methods Enzymol* **378**:31-67.
- Sacco G, Giampietro R, Salvatorelli E, Menna P, Bertani N, Graiani G, Animati F, Goso C, Maggi CA, Manzini S and Minotti G (2003) Chronic cardiotoxicity of anticancer anthracyclines in the rat: role of secondary metabolites and reduced toxicity by a novel anthracycline with impaired metabolite formation and reactivity. *Br J Pharmacol* **139**:641-651.
- Singal PK and Iliskovic N (1998) Doxorubicin-induced cardiomyopathy. *N Engl J Med* **339**:900-905.
- Stewart DJ, Grewaal D, Green RM, Mikhael N, Goel R, Montpetit VA and Redmond MD (1993) Concentrations of doxorubicin and its metabolites in human autopsy heart and other tissues. *Anticancer Res* **13**:1945-1952.
- Takanashi S and Bachur NR (1976) Adriamycin metabolism in man. Evidence from urinary metabolites. *Drug Metab Dispos* **4**:79-87.
- Weiss RB (1992) The anthracyclines: will we ever find a better doxorubicin? *Semin Oncol* **19**:670-686.

Footnotes

A) These studies were supported by funding from the Canadian Institutes of Health Research (CIHR). R.H.T. was supported by an Rx&D/CIHR scholarship. O.S.B. was supported by a University of British Columbia Graduate Fellowship.

B) Corresponding Author: Dr. K. Wayne Riggs

Faculty of Pharmaceutical Sciences, University of British Columbia, 2146 East Mall

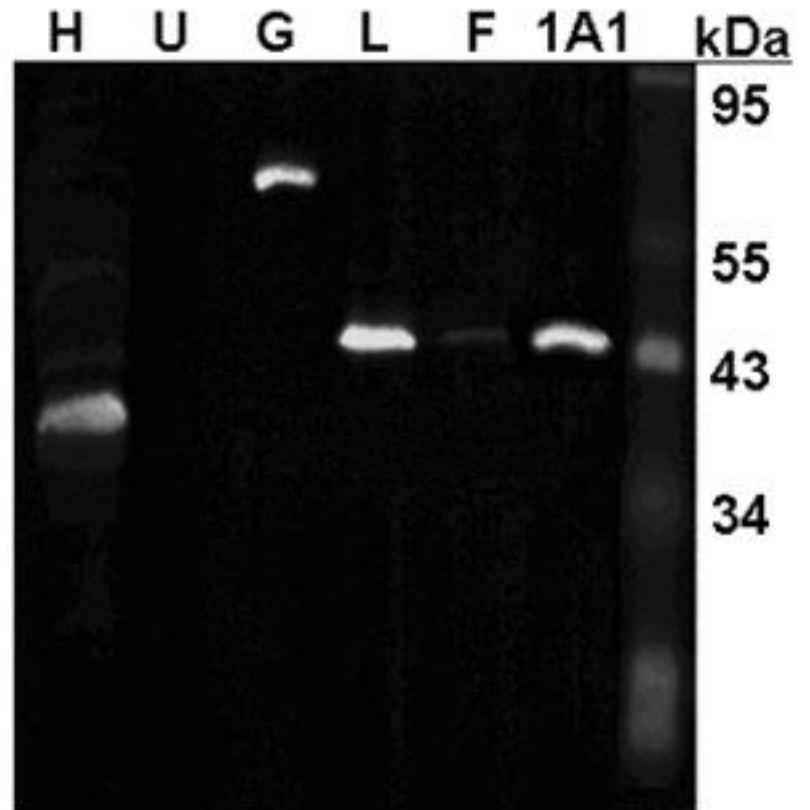
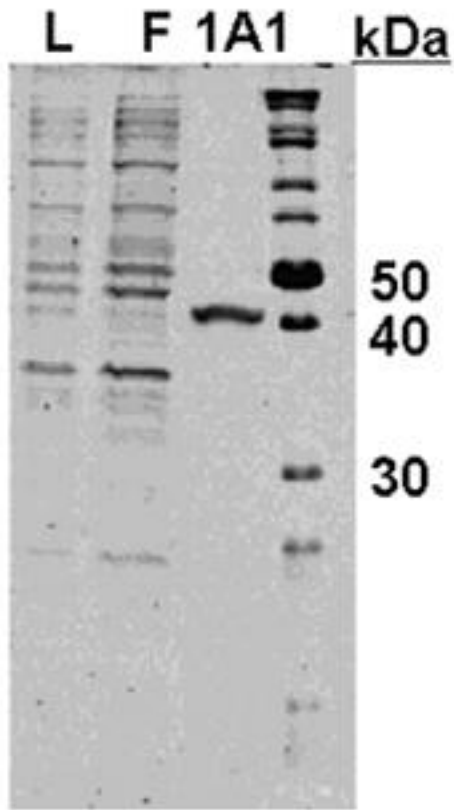
Vancouver, BC, Canada, V6T 1Z3. Email : riggskw@interchange.ubc.ca

FIGURE LEGENDS

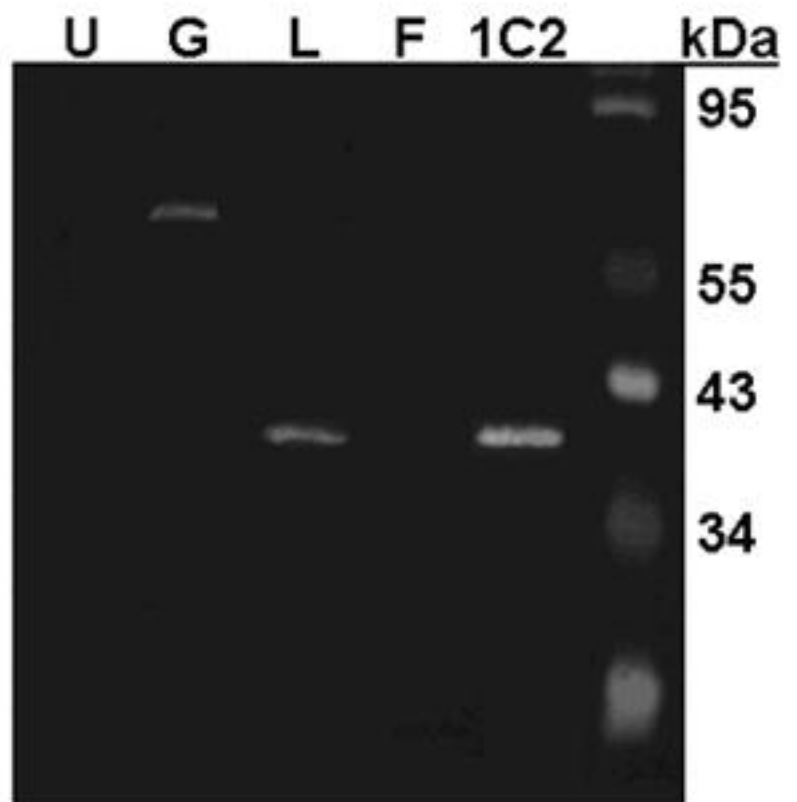
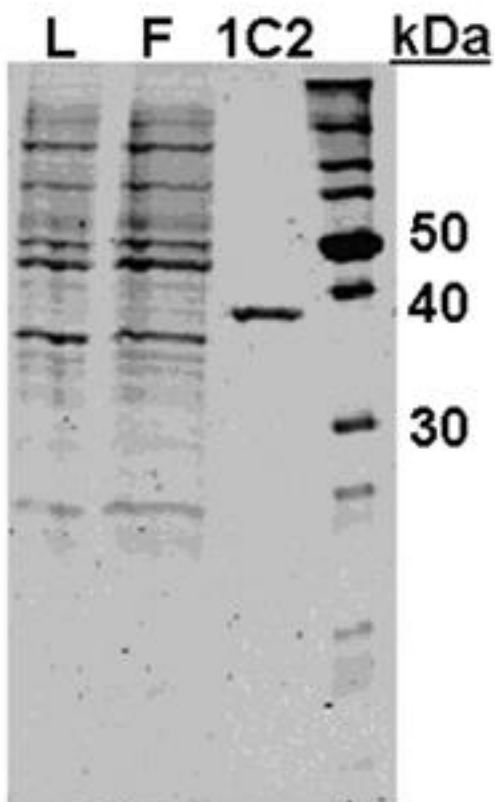
Figure 1—Purification of recombinant histidine-tagged human (A) AKR1A1 and (B) AKR1C2. (*Left*) SDS-PAGE stained with Sypro Ruby shows purified protein samples, 1A1 or 1C2 (700 ng), free of contaminating proteins from bacterial lysates (L) (20 μ g total protein). Removal of contaminating proteins is observed in fractions from Qiagen purification procedures (Ni-NTA column flow through (F), 15 μ g total protein). (*Right*) Western blotting of purified protein samples, 1A1 or 1C2 (700 ng), confirms expression of the desired AKR protein with mobility at expected molecular weight (~41-42 kDa). Positive controls for antibody immunoreactivity are human liver cytosol (H) and GST-tagged purified human recombinant AKR enzymes (G; 700 ng). No antibody immunoreactivity is observed for untransformed bacterial lysate (U; 20 μ g total protein).

Figure 2—Generation of DAUNol and DOXol *in vitro* by purified recombinant AKR1A1 incubated with (A) DAUN and (B) DOX. Measurement of DAUNol and DOXol was performed using HPLC-fluorescence. Representative chromatograms show clear resolution of DAUNol and DOXol from DAUN, DOX, and idarubicin (internal standard). Retention times observed for DOXol, DOX, DAUNol, DAUN, and idarubicin (internal standard) are 4.5, 5.5, 6.0, 6.8, and 7.3 minutes, respectively.

Figure 1

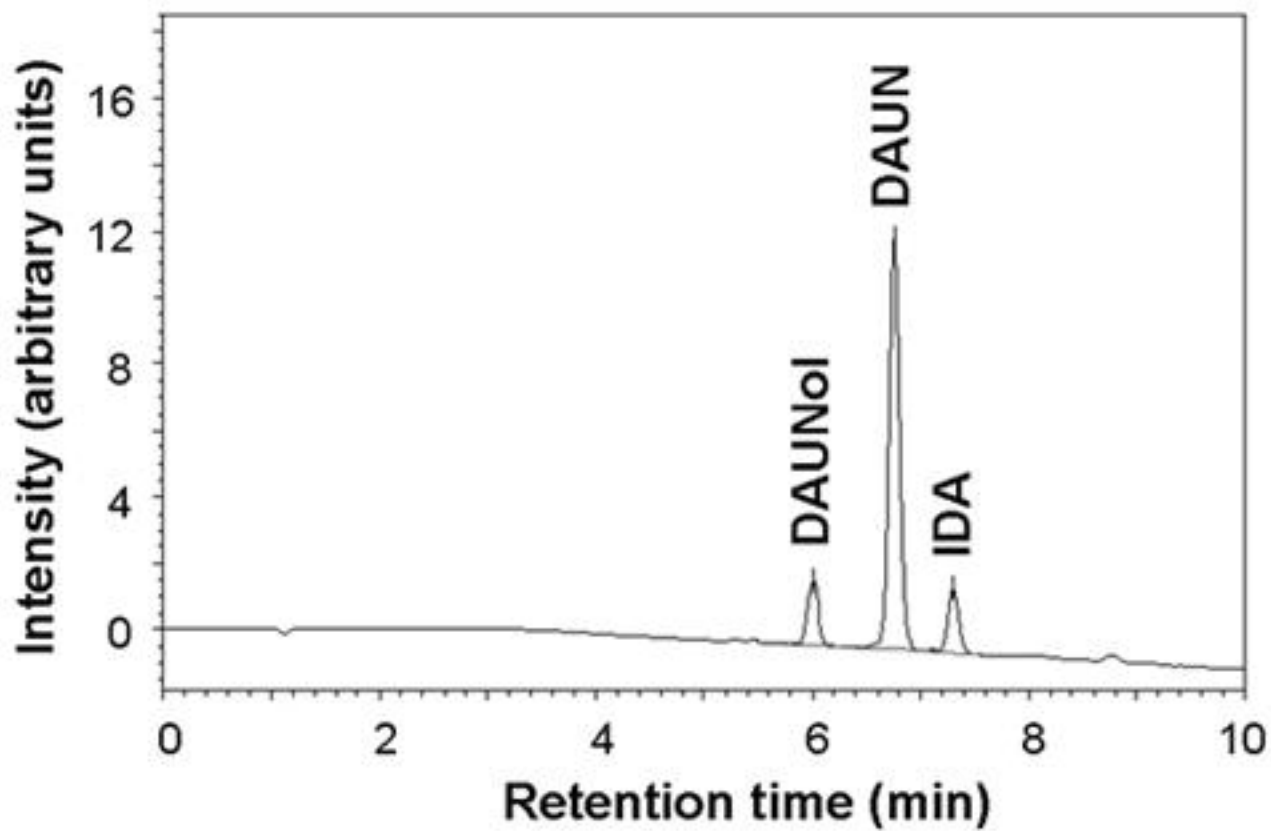


(A)

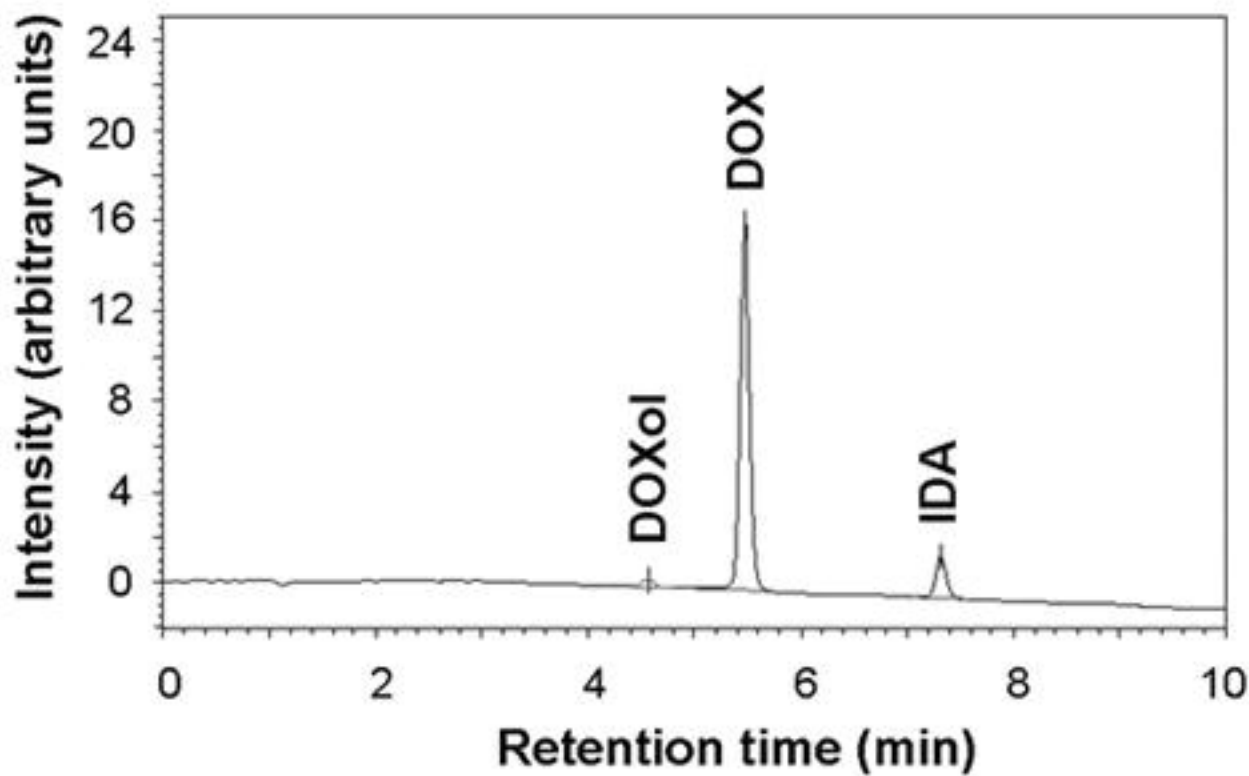


(B)

Figure 2



(A)



(B)