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**METABOLISM AND DISPOSITION OF FLUTICASONE
FUROATE, AN ENHANCED-AFFINITY GLUCOCORTICOID, IN
HUMANS**

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Metabolism and disposition of fluticasone furoate in humans

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Abbreviations:

AMS, accelerator mass spectrometry

AP-1, activation protein-1

AUC, area under the curve

CL_p, plasma clearance

HPLC, high-performance liquid chromatography

NFκB, nuclear factor kappa-B

pMC, percent modern carbon

V_{ss}, volume of distribution at steady state

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ABSTRACT

The purpose of this study was to investigate the metabolism and disposition of fluticasone furoate, an enhanced-affinity glucocorticoid receptor agonist, in humans. In a two-part, open-label design study, five healthy male subjects received an oral dose of 2 mg [^{14}C] fluticasone furoate, followed 4 weeks later by an intravenous dose of 0.25 mg [^{14}C] fluticasone furoate (as a 30 min infusion). Oral absorption was rapid and estimated at approximately 30%, although the oral bioavailability was markedly lower at 1.6%, limited by extensive first-pass metabolism. Plasma clearance (CL_p) was 58.3 L/h, with a volume of distribution (V_{ss}) of 642 L and a terminal elimination half-life of 15.3 h. The major circulating component identified in plasma extracts after intravenous and oral dosing was unchanged parent compound, with GW694301X (M10) also being notable after oral administration. Mean recovery of radioactivity was approximately 92% and 102% at 216 and 168 h after intravenous and oral administration, respectively, with most (at least 90%) recovered in the faeces. Fluticasone furoate was extensively metabolised, with only trace amounts of unchanged parent compound observed in faeces following either route of administration. The predominant pathway was removal of the S-fluoromethyl carbothioate group, to yield the 17 β carboxylic acid (GW694301X, M10). Other pathways included oxidative defluorination to yield a hydroxyl at the C6 position. There was no evidence for metabolic loss of the furoate group from fluticasone furoate or any of its metabolites. Evidence presented suggests that enterocytes have a role in the metabolism of unabsorbed fluticasone furoate.

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Introduction

Fluticasone furoate (6 α ,11 β ,16 α ,17 α)-6,9-difluoro-17-(((fluoromethyl)thio)carbonyl)-11-hydroxy-16-methyl-3-oxoandrosta-1,4-dien-17-yl-2-furancarboxylate is a new enhanced-affinity glucocorticoid receptor agonist. It is a synthetic fluorinated corticosteroid that has been developed as an intranasal treatment for patients with symptoms of rhinitis. Fluticasone furoate, otherwise known as GW685698X, is not a salt or prodrug, because the entire molecule is required for pharmacological activity. It has similar or greater potency than other clinically used corticosteroids (including mometasone furoate, budesonide, fluticasone propionate and the active principle of ciclesonide) for the glucocorticoid receptor and against the pro-inflammatory transcription factors nuclear factor κ B (NF κ B), activation protein-1 (AP-1), and tumour necrosis factor-induced interleukin-8 cytokine production (Salter et al., 2007). Agonism of the glucocorticoid receptor is known to suppress the activation of downstream transcription factors, such as NF κ B and AP-1, and to activate the glucocorticoid response element transactivation pathway (Rhen and Cidlowski, 2005). Inhibition of the NF κ B pathway, in particular, is thought to be intimately involved in the anti-inflammatory activity of glucocorticoids, because it is a key pathway in the synthesis of a number of inflammatory cytokines (Karin et al., 2004). It is well documented that topical glucocorticoids interact with many of the inflammatory pathways, and there is a large body of clinical evidence to support their use for the treatment of rhinitis, asthma and chronic obstructive pulmonary disease (Goodman and Gilman, 2006). The purpose of the present study was to investigate the metabolism and disposition of [14 C] fluticasone furoate after oral and intravenous administration to healthy male subjects. These dose routes were used as surrogates for the intranasal and inhalation routes, from which the majority of the dose is likely to be

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swallowed. The oral route acted as a surrogate for the portion of an intranasal or inhaled dose that is swallowed and the intravenous route represented the portion absorbed locally into the systemic circulation.

Materials and Methods

Materials. Fluticasone furoate, [^{14}C] fluticasone furoate (see Fig. 1), and GW694301X, GSK728920A, GSK728921A and GSK728922A (authentic chemicals of possible metabolites of fluticasone furoate) were supplied by Chemical Development, GlaxoSmithKline R&D, Stevenage, UK.

All other solvents and reagents were of analytical grade and were purchased from commercial suppliers.

Formulated Drug. [^{14}C] fluticasone furoate and fluticasone furoate were supplied to the study centre by Pharmaceutical Development, GlaxoSmithKline, Ware, UK, as sterile solutions in propylene glycol. The solutions were at a concentration of 0.25 mg/ml, and with a specific activity of 100 $\mu\text{Ci}/\text{mg}$.

For the oral dose the radiolabelled and non-radiolabelled solutions were mixed in a ratio of 1:1.67 to give a dose of 8 ml volume, equivalent to 2 mg fluticasone furoate and approximately 75 μCi .

Subjects and In Vivo Study Design. The clinical part of the study was conducted at Charles River Laboratories (formerly known as Inveresk), Edinburgh, UK, in accordance with Good Clinical Practice guidelines and the Declaration of Helsinki. All subjects provided written informed consent before participation. The protocol was approved by the investigational centre ethics committee and by the Administration of Radioactive Substances Advisory Committee, UK.

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Five healthy male Caucasian subjects, aged 50–56 years and with a body mass index within the range of 19–30 kg/m² were enrolled into this study. The study design was an open-label two-period cross-over, with an oral administration followed by intravenous administration. The subjects were in good health as shown by medical examination, clinical chemistry, haematology and urine analysis. The subjects were non-smokers with no history of drug or alcohol abuse, who were taking no other medication at the time of the study, and had taken no prescribed medication within 14 days of the study commencing.

All subjects received a single oral dose of [¹⁴C] fluticasone furoate at 2 mg (3 MBq, 75 µCi) and a dose volume of 8 ml. On a separate occasion each subject received an intravenous infusion of 0.25 mg of [¹⁴C] fluticasone furoate (1 MBq, 25 µCi) over 30 min using a dose volume of 1 ml. The two dosing occasions were separated by at least 28 days.

Blood samples were collected via an indwelling cannula or by direct venepuncture into lithium heparin-containing polypropylene tubes.

Following oral dosing, blood samples (10 ml) were collected at 10, 15, 30 and 45 min and 1, 1.5, 2, 3, 5, 9, 12, 24, 48, 72, 96 and 168 h after dosing, with larger samples (30 ml) for metabolic investigations being collected at 0.5, 2 and 12 h. After the start of the intravenous infusion, samples (10 ml) were collected at 15, 30, 32, 35 and 45 min and 1, 1.5, 2, 3, 4, 6, 9, 12, 24, 48, 72, 96 and 168 h after dosing, with the larger samples being collected at 0.75, 1.5 and 12 h. The samples were processed within an hour of collection by centrifugation for 10 min at 1500g at nominally 4°C to yield plasma, which was then stored frozen at nominally –20°C or processed for the determination of radioactivity levels in plasma. Urine samples were collected at 0–6,

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6–12 and 12–24 h and then at 24-h intervals up to at least 168 h. Faecal samples were collected over 24-h periods up to at least 168 h post-dose. Both urine and faecal collections continued beyond the 168-h time point following intravenous administration, until 90% of the total radioactive dose administered was recovered from all matrices, or less than 1% of the radioactive dose had been excreted over a 24-h period. The faecal and urine samples were stored frozen at nominally -20°C prior to analysis.

Radioassay of samples. After measurement of the total sample volume or weight (as appropriate) of excreta, the levels of radioactivity in samples were determined by liquid scintillation counting (Beckman LS series, Bucks, UK or Perkin Elmer Life Sciences instruments, Bucks, UK) with quench correction performed by an automatic external standard ratio method, which was established using sealed ^{14}C standards. Aliquots of liquid samples (e.g. urine, dose dilutions) or extracts of samples were mixed with scintillation fluid. Faecal samples were homogenised with an appropriate amount of water. Aliquots of homogenised faecal material were combusted using a Packard model 307 oxidiser prior to radioassay by scintillation counting.

The plasma had low levels of radioactivity; therefore, these samples were diluted appropriately with water and the level of radioactivity determined using accelerator mass spectrometry (AMS).

Determination of Radiochemical Purity. The radiochemical purity of [^{14}C] fluticasone furoate was confirmed by high-performance liquid chromatography (HPLC). The chromatographic instrument used consisted of an HP1100 series quaternary pump, column oven (50°C), UV detector (λ 245 nm) and auto-sampler using a Zorbax SB-C8 column (150 x 4.6 mm, 3.5 micron particle size). The mobile phase consisted of aqueous 0.1% trifluoroacetic acid (solvent A) and a mixture of

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acetonitrile/methanol (75/25, solvent B) at a flow rate of 1.5 ml/min. A gradient was used, starting at 60% A with a linear change to 80% B over 30 min. The column was re-equilibrated following each injection. Using on-line radiodetection, the column eluate was combined with scintillant (Ultima-Flo M supplied by Fluorochem, 4.5 ml/min) prior to detection using a Radiomatic Flo-one 150TR radioactivity monitor.

Dose analysis showed that all oral and intravenous doses had a radiochemical purity of 100%.

Quantification of Fluticasone Furoate and GW694301X in Plasma.

Concentrations of fluticasone furoate and GW694301X (M10) in plasma samples were determined using a validated analytical method using in-line solid phase extraction, followed by HPLC and tandem mass spectrometry (HPLC-MS/MS). The validation of this method was performed as detailed for the quantification of fluticasone furoate and GW694301X (Scott et al., 2007) using similar methodology. [¹³C₃]-CC118781 (fluticasone propionate) and GW819063 (10 µl in 20% acetonitrile in water with both at a concentration of 10 ng/ml) were added to plasma samples (0.15 ml) as internal standards. An aliquot of each sample (0.1 ml) was applied to conditioned Prospect C2 cartridges (10 x 1 mm), which were washed with 5% aqueous methanol containing 0.025% formic acid (2 ml) prior to being eluted in-line for 30 seconds onto the analytical column. The cartridge eluant was applied to a HyPurity C18 column (100 x 4.6mm, 5 µM, ThermoHypersil, Runcorn, Cheshire, UK) eluted at a flow rate of 0.5 ml/min using a linear gradient from 100% solvent A to 100% solvent B over 4.3 min, held until 5.15 min prior to returning to 100% solvent A by 6 min. Solvent A was a mixture of methanol and 10 mM ammonium formate (pH 5, 77.5/22.5, v/v) and solvent B was acetonitrile. The column eluant was

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introduced into a TurboIonSpray source of a Sciex API 4000 mass spectrometer (Applied Biosystems, Warrington, Cheshire, UK) operated in the most sensitive ionisation mode for each analyte, with the monitored fragmentation ions allowing quantification at the required sensitivity. The system was run in negative ion mode initially for 4.4 min, prior to being switched to positive ion mode. The temperature of the probe was maintained at 600°C, with a curtain gas setting of 25 and collision gas setting of 6. GW694301X and GW819063 were monitored in negative ion mode by multiple reaction monitoring of 489 → 111 and 520 → 142, respectively. Fluticasone furoate and [¹³C₃]-CCI18781 were monitored in positive ion mode by multiple reaction monitoring of 539 → 313 and 504 → 313, respectively. The appropriate mass adjustments were performed to monitor for radiolabelled material. The dynamic range of the assay for all analytes was from 10 to 1000 pg/ml.

Determination of Metabolite Profiles. Representative faecal samples from each subject were obtained by pooling across sampling times on a total sample weight basis, to generate sample pools where possible containing 90% or greater of the radioactivity excreted in the faeces. Plasma samples from individual subjects were pooled using equal volumes to produce a single representative sample per time point. No further analysis of urine samples was conducted, because of the limited amount of radioactive drug-related material in urine.

Radioactive material was extracted from plasma samples using acetonitrile (5 ml per ml of sample, rotary mixed at ambient temperature for approximately 1 h), three times with the supernatants being combined. The extracts were then evaporated to near dryness under a stream of nitrogen prior to being reconstituted in de-ionised water (0.5 or 1 ml). Radioactive material was extracted from faecal samples using alkaline and acidified methanol as described for plasma, and the reconstituted extracts from

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each extraction method were combined using equal volumes prior to analysis by radio-HPLC. Radio-metabolite profiles were determined by analysis of appropriate aliquots of plasma and faecal extracts by radio-HPLC using on-line or off-line radiodetection.

HPLC method 1 (used for analysis of all matrices.) The chromatographic instrument used consisted of an Agilent (South Queensferry, Scotland, UK) 1100 binary pump, column oven (40°C), UV detector (λ 245 nm) and auto-sampler (CTC Analytics, or LC PAL injector, both Presearch, Basingstoke, Hampshire, UK) using a ThermoHypersil Fluophase RP column (250 x 4.6 mm, 5 micron particle size, Runcorn, Cheshire, UK). The mobile phase consisted of 50 mM ammonium acetate (BDH, Poole, Dorset, UK) (pH unadjusted) (solvent A) and acetonitrile (solvent B, supplied by Fisher Scientific, Loughborough, Leicestershire, UK) at a flow rate of 1 ml/min. A gradient was used, starting at 100% A with a linear change to 25% B over 30 min, followed by a linear increase to 40% B by 40 min and then a further increase to 100% by 45 min, with these conditions being held for a further 5 min. The column was re-equilibrated after each injection. Using on-line radiodetection, the column eluate was combined with scintillant (FlowLogic MaxCount supplied by Lablogic, 3 ml/min) prior to detection using a β -Ram (Lablogic, Sheffield, UK) radioactivity monitor. For off-line radiodetection, fractions were collected using a Gilson 222XL fraction collector (supplied by Gilson, Villiers, France) on 4 x 96 Deepwell LumaPlates containing yttrium silicate solid scintillant (Perkin-Elmer, Beaconsfield, Bucks, UK) or into standard 96-well plates. Radioactivity determination was performed either by scintillation counting (TopCount MXT counter, Packard/Perkin Elmer, Beaconsfield, Bucks, UK) or by AMS analysis of the fractions. HPLC column recoveries were determined on selected samples by collecting the total

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HPLC column eluate for the appropriate run and assaying the radioactivity, to assess recovery of injected radioactivity.

Full recoveries of radioactivity were obtained from the HPLC eluant collected.

HPLC method 2 (used for analysis of human plasma samples). The chromatographic instrument is as listed for method 1, with the exception of a different column being used: a ThermoHypersil Gold column (250 x 4.6 mm, 5 micron particle size). The mobile phase consisted of 50 mM ammonium acetate pH 5, adjusted using acetic acid (supplied by Fisher Scientific, Loughborough, Leicestershire, UK), (solvent A) and acetonitrile (solvent B) at a flow rate of 1 ml/min. A gradient was used, starting at 25% B with a linear increase to 40% B over 8 min, followed by a linear increase to 45% B by 12 min and then a further increase to 75% by 13 min, held at these conditions for 2 min before increasing to 90% by 18 min. The column was washed and re-equilibrated after each injection.

Identification of Metabolites. Structural characterisation was performed on selected samples using HPLC-Mass Spectrometry using Quattro Micro triple quadrupole (Micromass), LCT time of flight (Micromass, Manchester, UK) and Deca XP^{plus} ion trap (Thermo-Finnigan, San Jose, CA, USA) mass spectrometers. Electrospray ionisation, in positive or negative modes, was used. The HPLC flow was split (1:25 to 1:50) between the mass spectrometer and on-line or off-line radiodetector.

Metabolites of fluticasone furoate were identified based on charged molecular ions and their collision-induced disassociation fragmentation. Authentic standards, where available, were used to compare chromatographic retention time to metabolites that were structurally characterised. Supporting data obtained from metabolites identified from preclinical species or from in vitro incubations with human-derived tissue preparations were also used in the assignment of metabolite structures.

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Accelerator Mass Spectrometry. The ^{14}C content of human plasma, plasma extracts and HPLC fractions were measured by AMS, which measures the radiocarbon content in a sample through separation of the isotopes of carbon present by their different mass-to-charge ratios. Before AMS analysis, the samples were graphitised via a two-step process of oxidation and reduction (Vogel, 1992). The graphite, containing a cobalt catalyst, was packed into an aluminium cathode and loaded into a sample wheel that was then placed into the ion source of the AMS instrument (NEC 15SDH-2 Pelletron AMS system, Wisconsin, USA). A generic value of 4.14%, based on historical data, for the carbon content of plasma was used for any neat plasma samples analysed, and values were adjusted appropriately to allow for any dilution. The carbon content for the HPLC fractions was deemed to be insignificant and only the carbon content of the liquid paraffin carrier (84.17%) was used in calculations to determine the ^{14}C content based on the determined ^{14}C to ^{12}C ratio in the samples. The AMS data, which are expressed as percent modern carbon (pMC) were used to calculate the dpm/ml of sample, where 100 pMC equals 0.01356 dpm/mg carbon.

Pharmacokinetic Calculations. Pharmacokinetic parameters were calculated by noncompartmental methods using WinNonLin Professional v4.1 (Pharsight, Mountain View, CA). Area under the curve, $\text{AUC}_{(0-t)}$, was calculated using the log-linear trapezoidal method and extrapolated to infinity according to $\text{AUC}_{(0-\infty)} = \text{Ct} \cdot t_{1/2}/\ln 2 + \text{AUC}_{(0-t)}$ where Ct is the last observed concentration. The $t_{1/2}$ values were obtained by the ratio of $\ln 2/\lambda_z$, where λ_z was the terminal phase rate constant estimated by linear regression analysis of the log-transformed concentration-time data during the terminal elimination phase. Mean residence time was calculated based on the last measurable concentration. Plasma clearance (CLp) and volume of distribution at steady state (Vss) were calculated following intravenous administration only. Oral bioavailability

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was calculated using the dose normalised $AUC_{(0-\infty)}$ oral/dose normalised $AUC_{(0-\infty)}$ intravenous.

Results

Demographic, Safety, and Tolerability Data from Humans. Five healthy male Caucasian subjects were enrolled and completed the study. The subjects were of a mean age of 54 years (range 50–56 years), a height of 169 cm (range 163–181 cm) and weight 79.2 kg (range 71.2–93.0 kg) with a mean body mass index of 27.6 (range 25.1–30.1). Fluticasone furoate was well tolerated, with no drug-related adverse events or clinically significant changes in vital signs observed. No concomitant medication was reported for any subject during the study.

Pharmacokinetics of Fluticasone Furoate and GW694301X. Pharmacokinetic data for fluticasone furoate, its carboxylic acid metabolite (GW694301X, M10), and total radioactivity are listed in Table 1. After oral administration of [14 C] fluticasone furoate at a dose of 2 mg, maximum mean concentrations (C_{max}) of fluticasone furoate occurred within 30 min (T_{max}) of administration, indicating rapid oral absorption of the dose. Concentrations declined rapidly thereafter, such that they were below the limit of quantification beyond 9 h (Fig. 2). After a 30 min intravenous infusion of [14 C] fluticasone furoate at a dose of 0.25 mg, CL_p was 58.3 L/h with a mean V_{ss} of 642 L. In contrast to post-oral administration, concentrations of fluticasone furoate were measurable up to 24 h post-infusion and mean terminal half-life was estimated at 15.3 h. Mean concentrations of GW694301X peaked at 0.75 h post-oral dose of fluticasone furoate and declined rapidly thereafter, with a systemic exposure (based on either $AUC_{(0-\infty)}$ or $AUC_{(0-t)}$) approximately 5-to 9-fold lower than for fluticasone

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furoate. No GW694301X was quantifiable in plasma after intravenous administration of fluticasone furoate. Comparison of the $AUC_{(0-t)}$ values for radioactivity following oral and intravenous administration (normalising for dose) indicates that oral absorption was approximately 30% (range 18–37%). The mean bioavailability of fluticasone furoate from the oral solution (comparing oral and intravenous $AUC_{(0-t)}$ values for fluticasone furoate and normalising for dose) was considerably lower at 1.6% (range 0.5–2.8%). The $AUC_{(0-\infty)}$ for total radioactivity in plasma was approximately 36- and 5-fold higher than that for fluticasone furoate following oral or intravenous administration, respectively. Furthermore, the mean half-life of total radioactivity (at 35.6 h) was notably longer than that of fluticasone furoate (15.3 h) after intravenous dosing.

Excretion of [14 C] Fluticasone Furoate Drug-Related Material. The mean total recovery of radioactive drug-related material in excreta collected after intravenous administration of [14 C] fluticasone furoate to humans was high (92%) and was complete post-oral administration, following collections up to 240 and 168 h, respectively (Table 2). Most of the dose was excreted in the faeces for both routes of administration; urinary excretion was a very minor route (< 3%). Radioactivity was eliminated rapidly: most was excreted within the first 72 h post-dose, with approximately 71% and 91% of the dose recovered over that period following intravenous and oral administration, respectively.

Metabolite Profiles in Plasma and Faeces. Proposed structure and spectral data supporting metabolite identification are shown in Table 3. Radioactive drug-related material in plasma is expressed as % sample radioactivity. Radioactive drug-related material in faeces is expressed as % administered dose. Where the sample preparation step resulted in some loss of radioactivity, if the recovery was <90%, the

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chromatogram data have been multiplied by the percentage recovered to calculate % sample radioactivity and % dose.

Metabolites of [¹⁴C] Fluticasone Furoate in Plasma. It was established that [¹⁴C] fluticasone furoate spiked into plasma was stable when stored under the same conditions as the test samples.

Low levels of radioactivity were present in the human plasma extracts; therefore, the profiling of these samples was performed using AMS as the radioactivity detection method. Representative reconstructed HPLC radiochromatograms of plasma extracts following intravenous and oral administration, at selected sampling times, are shown in Fig. 3. The extraction efficiency of radioactivity from plasma was in excess of 70% at early time points, but notably lower (9–35%) at later time points.

The principal radiolabelled component in plasma following a single intravenous infusion of [¹⁴C] fluticasone furoate was unchanged fluticasone furoate (P), which accounted for at least 52% of radioactive material in plasma samples up to 1.5 h after dosing. The 17 β -carboxylic acid metabolite (GW694301X, M10) of fluticasone furoate was assigned based on retention time and represented up to 4% of plasma radioactivity. Three other minor components were also observed, each representing less than 2% of plasma radioactivity.

After oral administration, M10 and fluticasone furoate were the major components in plasma, comprising 28% and 17% of sample radioactivity, respectively, at 0.5 h, and 6% and 7%, respectively, at 2 h post-oral administration. Metabolites M21 (6 hydroxyl of parent), M26 (6 hydroxyl of M10) and M32 (an hydroxylated product of M10) were observed in human plasma, based on their co-elution with either authentic standards or identified metabolites from in vitro incubations; all accounted for less

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than 4% of plasma radioactivity. The proposed structures and supporting spectral data for the metabolites are shown in Table 3.

Metabolites of [¹⁴C] Fluticasone Furoate in Faeces. It was established that [¹⁴C] fluticasone furoate spiked into faeces was stable when stored under the same conditions as the test samples.

The mean extraction efficiency of radioactivity from the faecal samples was 90% (oral) and 84% (intravenous). The extraction efficiency of radioactivity from control human faecal samples spiked with [¹⁴C] fluticasone furoate was 95%. Representative HPLC radiochromatograms of human faecal extracts following either intravenous or oral administration are shown in Fig. 3.

The predominant identified drug-related component in the faecal extracts from four out of five subjects following dosing by either route was M10, representing 32% to 40% of the oral dose and 24% to 31% of the intravenous dose. However, in one subject, M10 was a minor component of the total drug-related material excreted in the faeces. Other drug-related components assigned by co-chromatography with authentic standards were the 6-hydroxy metabolite of M10 (M26) and parent compound, each representing less than 8% of an oral or intravenous dose.

Although most of the dose was excreted in faeces, the absolute amounts of drug-related material present in the faecal samples were very low. A number of drug-related components remained insensitive to detection by mass spectrometry and their structures were not elucidated.

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Discussion

Fluticasone furoate, administered as an intranasal aqueous suspension via a unique side-actuated device, is a new chemical entity for the treatment of nasal and ocular symptoms of allergic rhinitis (Martin et al., 2007). As such, it is important to gain an understanding of the metabolism and disposition of fluticasone furoate in humans.

The pharmacokinetic profile of fluticasone furoate following intravenous administration to humans is characterised by a high plasma clearance at 58.3 L/h, which is approximately two-thirds human liver blood flow (Davies and Morris, 1993), and a very large volume of distribution of 642 L, which equates to approximately fifteen times total body water (Davies and Morris, 1993), indicating extensive tissue uptake. These properties in combination resulted in a moderate plasma half-life of 15.3 h. In contrast, the plasma half-life observed following oral administration was shorter, at approximately 4 h; however, this is likely to be representative of the distribution phase, with plasma concentrations falling below the assay limit of quantification before the true elimination phase was observed. This is consistent with a large first-pass effect resulting in a very low mean oral bioavailability, calculated at 1.6%. The absolute bioavailability of intranasally administered fluticasone furoate in healthy volunteers has been determined to be 0.5% (Allen et al., 2007). The lower bioavailability observed with the clinical intranasal formulation is likely as a result of differences in dose, dose route and formulation compared with those used in this study.

Systemic exposure to radioactivity was much greater than systemic exposure to fluticasone furoate for both routes of administration, with the observed difference being greater following oral administration. In addition, the elimination half-life of

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radioactivity was greater than that of fluticasone furoate. These observations are consistent with the presence of one or more circulating metabolites with a longer elimination half-life than parent, and of extensive first-pass metabolism after oral administration. Oral absorption was estimated to be only moderate (approximately 30%), despite the high tissue permeability suggested by the high volume of distribution. The estimate of oral absorption (obtained by comparison of the $AUC_{(0-t)}$ values for total plasma radioactivity following oral and intravenous administration) may be an over-estimate, because the profiles of drug-related material following the two routes of administration differ; however, it is consistent with the ratios of urinary excretion, although these are based on very low values. A similar extent of absorption was also observed in rats and dogs (unpublished data), where 29% and 19% (30% of the recovered radioactivity) of an oral radioactive dose was recovered in the bile and urine of bile-duct cannulated rats and a dog, respectively.

The principal component in plasma extracts following intravenous administration of [^{14}C] fluticasone furoate (0.25 mg) to healthy male subjects was fluticasone furoate, with low levels of the 17 β carboxylic acid metabolite (M10) of fluticasone furoate also observed. In contrast, following oral administration (2 mg) to the same male subjects, M10 and fluticasone furoate were both principal components in the plasma extracts. Three other minor drug-related components (each less than 5% of plasma radioactivity) were observed and were assigned as products of oxidative metabolism. The greater contribution of metabolites to plasma radioactivity after oral than after intravenous administration is consistent with high first-pass metabolism. The remainder of the plasma radioactivity following intravenous or oral administration was either unextracted or unassigned because of the low levels of drug-related material in the plasma. The unextracted radioactivity represented a very low amount

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of material, less than 1 ng equivalents/ml, which equates to less than 30 femtomoles per mg of plasma protein.

AMS was used to quantify radiolabelled drug-related components in plasma off-line following separation by HPLC because of the low levels of radioactivity present in human plasma following intravenous or oral administration of [¹⁴C] fluticasone furoate. AMS is a very sensitive method for the measurement of radiocarbon (Vogel et al., 1995). The assignment of metabolite structures by chromatographic retention time was performed by comparison to retention times of authentic reference standards and a metabolite isolated from in vitro incubations followed by definitive identification. The use of AMS resulted in a clearer definition of the circulating components than would have been achieved by conventional off-line radiodetection using a solid yttrium silicate scintillant, which was employed to profile plasma samples from non-clinical species. In the non-clinical species, only fluticasone furoate and the metabolite M10 were identified in plasma after intravenous and/or oral administration (unpublished data). As a result of the difference in sensitivity between AMS and the solid scintillant detection method, it is considered that a number of the minor metabolites (less than 5% of radioactive drug-related material) observed in human plasma may not have been observed in non-clinical species because of the difference in limits of detection.

The mean recovery of the radioactive dose following intravenous administration of [¹⁴C] fluticasone furoate to male subjects was greater than 92%. Similar recovery was obtained in rats (86%) and dogs (81%) following an intravenous dose (unpublished data). After oral administration to male subjects, the recovery of radioactivity was complete. Most radioactivity was excreted in the faeces after both routes of administration. This excretion pattern is similar to that observed for other synthetic

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corticosteroids, such as ciclesonide (Nave et al., 2004) and mometasone furoate (Affrime et al., 2000).

The major route of fluticasone furoate metabolism in humans is ester hydrolysis leading to formation of the 17 β carboxylic acid (M10) with loss of the S-fluoromethyl carbothioate moiety. The loss of this moiety is also the significant route of metabolism of fluticasone propionate, with the biotransformation being catalysed by cytochrome P450 3A4 (Pearce et al., 2006). Oxidative defluorination of fluticasone furoate at the C6 position also occurred and resulted in the formation of M21 and M26; this biotransformation is known to occur in a number of fluorine-containing drugs (Park et al., 2001). Oxidative defluorination is also a metabolic pathway for fluticasone propionate in vitro in human-derived preparations (Shenoy et al., 1993). Oxidative defluorination of fluticasone furoate resulted in hydroxylation at the C6 position; C6 hydroxylation is a well-precedented route of metabolism for corticosteroids and has been observed in human-derived in vitro systems for budesonide (Jonsson et al., 1995), mometasone furoate (Teng et al., 2003b) and ciclesonide (Peet et al., 2005). A summary of the proposed oxidative pathways for fluticasone furoate in humans is shown in Fig. 4. There was no evidence for metabolic loss of the furoate group from fluticasone furoate or any of its metabolites; this differs to mometasone furoate, where loss of the furoate group is observed with the formation of mometasone in human-derived fluids and tissue preparations (Teng et al., 2003b) and rat-derived fluids and tissue preparations (Teng et al., 2003a).

Although limited data were obtained from in vivo excreta because of the low amounts of drug-related material present, it was observed that, although oral absorption of fluticasone furoate in human was at most approximately 30%, little or no unchanged parent drug was observed in the faeces after oral administration. A similar pattern was

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also observed in orally dosed bile-duct cannulated rats and, furthermore, fluticasone furoate is stable in a preparation of high-density anaerobic microbiota from either human or rat (unpublished data). These data, together with evidence that glucocorticoids are substrates for the P-glycoprotein efflux transporter (Yates et al., 2003), suggest that enterocytes may have a major role in the metabolism of fluticasone furoate in the gastro-intestinal tract through a cycle of absorption, metabolism and efflux, in addition to limiting absorption of this highly permeable moiety.

The major routes of fluticasone furoate metabolism in humans have been defined following a single intravenous and/or oral administration of [¹⁴C] fluticasone furoate, within the constraints of the very low levels of drug-related material in the plasma and excreta. No notable differences in the routes of metabolism between humans and the non-clinical species were observed.

Most of an intranasal or inhaled dose is likely to be swallowed; therefore, the oral and intravenous routes of administration were considered adequate surrogate routes from which to obtain samples to define the major routes of metabolism of fluticasone furoate in humans. The oral route acted as a surrogate for the portion of an intranasal or inhaled dose that is swallowed, and the intravenous route represented the drug that is absorbed locally into the systemic circulation.

The routes of fluticasone furoate metabolism identified are consistent with those preceded for other marketed corticosteroids, such as fluticasone propionate, mometasone furoate, ciclesonide and budesonide.

The routes of metabolism for fluticasone furoate and fluticasone propionate are similar, with no evidence that either is metabolised to fluticasone. This demonstrates

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that they act as different chemical entities. The oral bioavailability of fluticasone furoate is very low, limited by absorption and first-pass metabolism and, therefore, any of the drug that is swallowed is unlikely to be active systemically.

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Footnotes

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Figure Legends

FIG. 1. The structure of fluticasone furoate, with position of ^{14}C shown.

FIG. 2. Mean plasma concentrations of fluticasone furoate, GW694301X and total radioactivity as indicated on semi-log plots following an oral administration (A) or intravenous infusion over 30 min (B) of [^{14}C] fluticasone furoate to healthy male subjects ($n = 5$).

FIG. 3. HPLC radiochromatograms of plasma and faecal extract from healthy male subjects following intravenous or oral administration of [^{14}C] fluticasone furoate. The plasma chromatograms are reconstructed from fractions analysed by accelerator mass spectrometry.

FIG. 4. Summary of the identified oxidative pathways of metabolism for fluticasone furoate.

Table 1**Pharmacokinetic parameters in humans for fluticasone furoate, GW694301X and ¹⁴C radioactivity in plasma**

Mean pharmacokinetic parameters of fluticasone furoate, a carboxylic acid metabolite GW694301X and ¹⁴C radioactivity in plasma of healthy male subjects following a single oral (2 mg) or a single intravenous (0.25 mg) administration of [¹⁴C] fluticasone furoate are shown. Mean ± standard deviation, *n* = 5

Treatment	Parameter	Fluticasone furoate	GW694301X	Radioactivity ^a
2 mg oral	AUC _(0-t) (pg.h/ml)	511 ± 358	102 ± 61	26400 ± 14400
	AUC _(0-∞) (pg.h/ml)	1080 ^b	120 ± 59	39300 ± 11700 ^c
	C _{max} (pg/ml)	213 ± 177	71 ± 42	1380 ± 239
	T _{max} (h) ^e	0.50	0.75	0.75
	t _{1/2} (h)	4.05 ^b	1.04 ± 0.39	20.8 ± 5.30 ^c
0.25 mg intravenous	AUC _(0-t) (pg.h/ml)	4000 ± 846	ND	11300 ± 5150

AUC _(0-∞) (pg.h/ml)	4410 ± 834	ND	21400 ± 15200 ^d
C _{max} (pg/ml)	2980 ± 1140	ND	5060 ± 1330
T _{max} (h) ^e	0.50	ND	0.50
t _{1/2} (h)	15.3 ± 2.74	ND	35.6 ± 33.3 ^d
Cl (L/h)	58.3 ± 10.9	ND	NA
V _{ss} (L)	642 ± 215	ND	NA

a Radioactivity concentrations are expressed in terms of pg equivalents of fluticasone furoate

b *n* = 2, due to inadequate definition of the terminal elimination phase these pharmacokinetic parameters could not be defined in all subjects

c *n* = 3, due to inadequate definition of the terminal elimination phase these pharmacokinetic parameters could not be defined in all subjects

d *n* = 4, due to inadequate definition of the terminal elimination phase these pharmacokinetic parameters could not be defined in all subjects

e T_{max} quoted as median

ND: Not determined, all concentrations below lower limit of quantification (10 pg/ml);

NA: Not applicable.

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Table 2

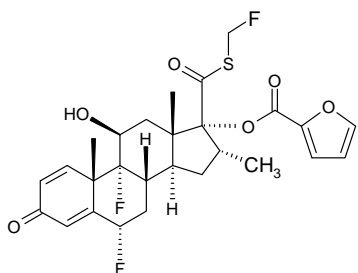
Mean percentage cumulative recovery of radioactivity following oral or intravenous administration of [¹⁴C] fluticasone furoate to humans

Time (h)	Oral Administration (2 mg)			Intravenous Administration (0.25 mg)		
	Faeces	Urine	Total	Faeces	Urine	Total
0–6	NA	0.49	0.49	NA	1.27	1.27
6–12	NA	0.69	0.69	NA	1.59	1.59
0–24	5.03	0.82	5.85	2.26	1.84	4.10
0–48	52.52	0.92	53.44	32.57	2.14	34.71
0–72	90.44	0.97	91.41	69.06	2.31	71.37
0–96	94.66	0.98	95.64	78.78	2.46	81.24
0–120	100.38	0.99	101.37	82.45	2.50	84.95
0–144	100.96	1.00	101.96	86.40	2.54	88.94
0–168	101.15	1.00	102.15	87.96	2.57	90.53
0–240	NA	NA	NA	89.78	2.61	92.39

NA: Not applicable

Table 3**Proposed structure, chromatographic retention times and mass spectral data for metabolites of fluticasone furoate in humans**

Metabolite	Retention time (min)	Matrix	Positive (M + H ⁺) or Negative (M-H) ion MS Parent and fragment ions (m/z)
Fluticasone furoate	46.3	Authentic standard	P: 539 (M+H ⁺) F1: 519 Loss of HF F2: 473 F3: 333 F4: 313

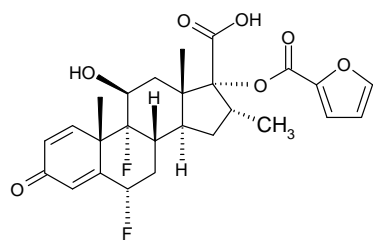


F5: 293

M10

37.2

Plasma

P: 491 (M+H⁺)

Faeces

471: loss of HF

333: F3

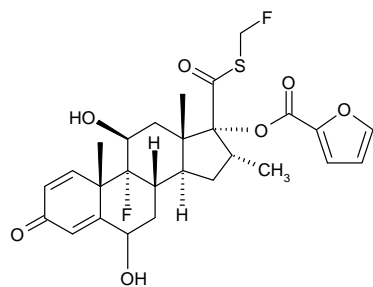
313: F4

293: F5

M21

44.6

Plasma

P: 537 (M+H⁺)(based on co-
elution)

311: F4-2

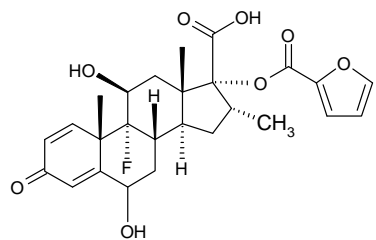
Information on metabolite in
hepatocytes

M26

27.0

Plasma

P: 489 (M+H⁺)

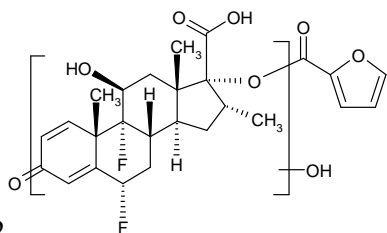


Faeces

313: F4

293: F5

M32



38.1

Human plasma

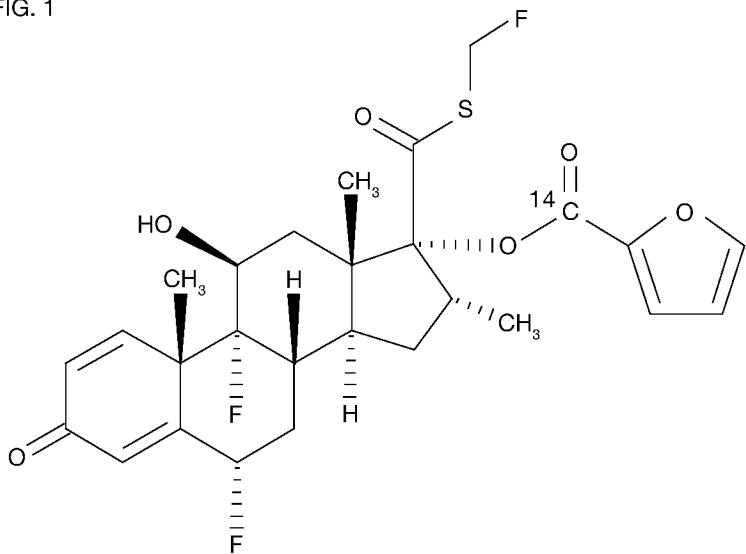
Negative ion (M-H)

Based on co-
elution with
previously
identified
metabolite

P: 505

393

FIG. 1



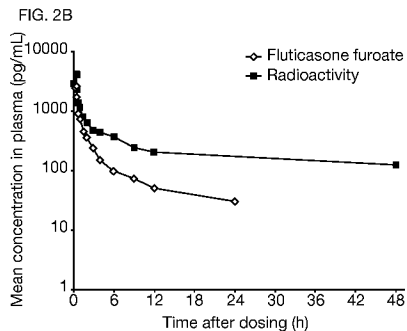
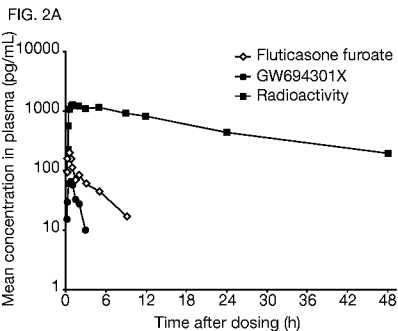


FIG. 3A

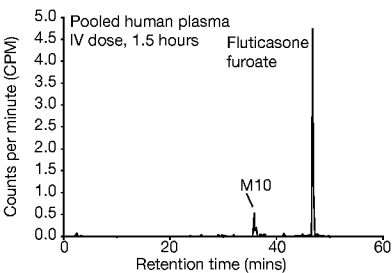


FIG. 3B

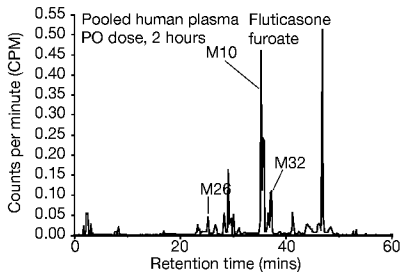


FIG. 3C

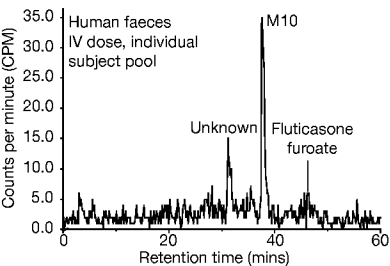


FIG. 3D

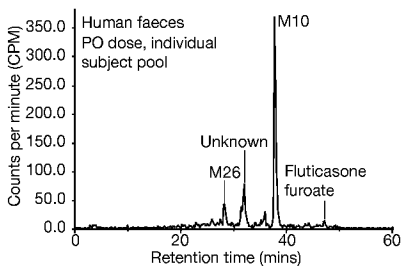


FIG. 4

