

**Celius et al. *FMO* induction by dioxin - Supplementary Tables & Figures**

**Supplementary Table 1**

**Primers and probes for real-time RT-PCR**

<b>Gene target (Accession number)</b>	<b>Forward primer '5 → 3'</b>	<b>Reverse primer '5 → 3'</b>	<b>Probe '5 → 3'</b>
FMO1 (NM010231)	ACATTACCACCGCCAAGTGT	TGCAGTAGCACAAGCCAAAC	TGAACGGAAGAAAAACAAGCATAGCGG
FMO2 (NM018881)	ACTCAGAGCAACGGAAAGGA	CCTGGGAATGACTTGAGTGG	ATGGTTTGCAGCGGCCATCA
FMO3 (NM008030)	AAGACTCCTTTCCAGGACTGAACC	TTCCCTTCCATATTCTGGTTCCT	AAAGGCAAATGCTTCCACAGCAGGGACT
FMO4 (NM144878)	CCAGTGGGATGTTGTCACAG	AAATGTGGGCTCAGGAATTG	AAGGCAAACGGGACAGGGCA
FMO5 (NM010232)	ATGACCTGCCCAATCGTATC	CCTGGAGCCATCCTCAAATA	AATTCACAGAGACAGCCGCCG
CYP1A1 (NM012540)	GAATGCCAATGTCCAGCTCTCA	TACCAGGTACATGAGGCTCCAA	AGCAGTTGTGATTGTGTCAAACCCAGCTCC

## Supplementary Table 2

microRNAs that are predicted to target mouse FMO2 or FMO3 mRNAs

Data are from:

<http://microrna.sanger.ac.uk>. Bold blue font indicates microRNA species or their close structural relatives that are significantly upregulated by TCDD in liver of C57BL/6J mice (Moffat et al. *Toxicol Sci* 99:470-487, 2007).

### Mouse FMO3

mmu-miR-15b\*  
 hsa-miR-518d-5p  
 hsa-miR-548c-3p  
 mmu-miR-382  
 mmu-miR-470  
 mmu-miR-677  
 mmu-miR-27a\*  
 mmu-miR-143  
 mmu-miR-24-1\*  
 mml-miR-189  
**mmu-miR-203\***  
 mmu-miR-382  
 mmu-miR-488  
 hsa-miR-626  
 hsa-miR-601  
 mmu-miR-509-3p  
 hsa-miR-583  
**mmu-miR-148a\***  
 mmu-miR-590-3p  
 hsa-miR-581  
 mmu-miR-340-5p  
 mmu-miR-24-2\*  
 mmu-miR-431  
 mmu-miR-145  
 mmu-miR-218-1\*  
 hsa-miR-583  
 mmu-miR-503\*  
 mmu-miR-370  
 mmu-miR-361  
 mmu-miR-218-2\*  
 mmu-miR-27a\*  
 mmu-miR-654-3p  
 mmu-miR-369-5p  
 mmu-miR-590-5p  
 mmu-miR-137  
**hsa-miR-519d**

### Mouse FMO2

mmu-miR-22\*  
 hsa-miR-548a-3p  
**mmu-miR-203**  
 hsa-miR-589  
 mmu-miR-466d-3p  
 mmu-miR-466b-3-3p  
 mmu-miR-684  
 hsa-miR-651  
 mmu-miR-153  
 mmu-miR-590-5p

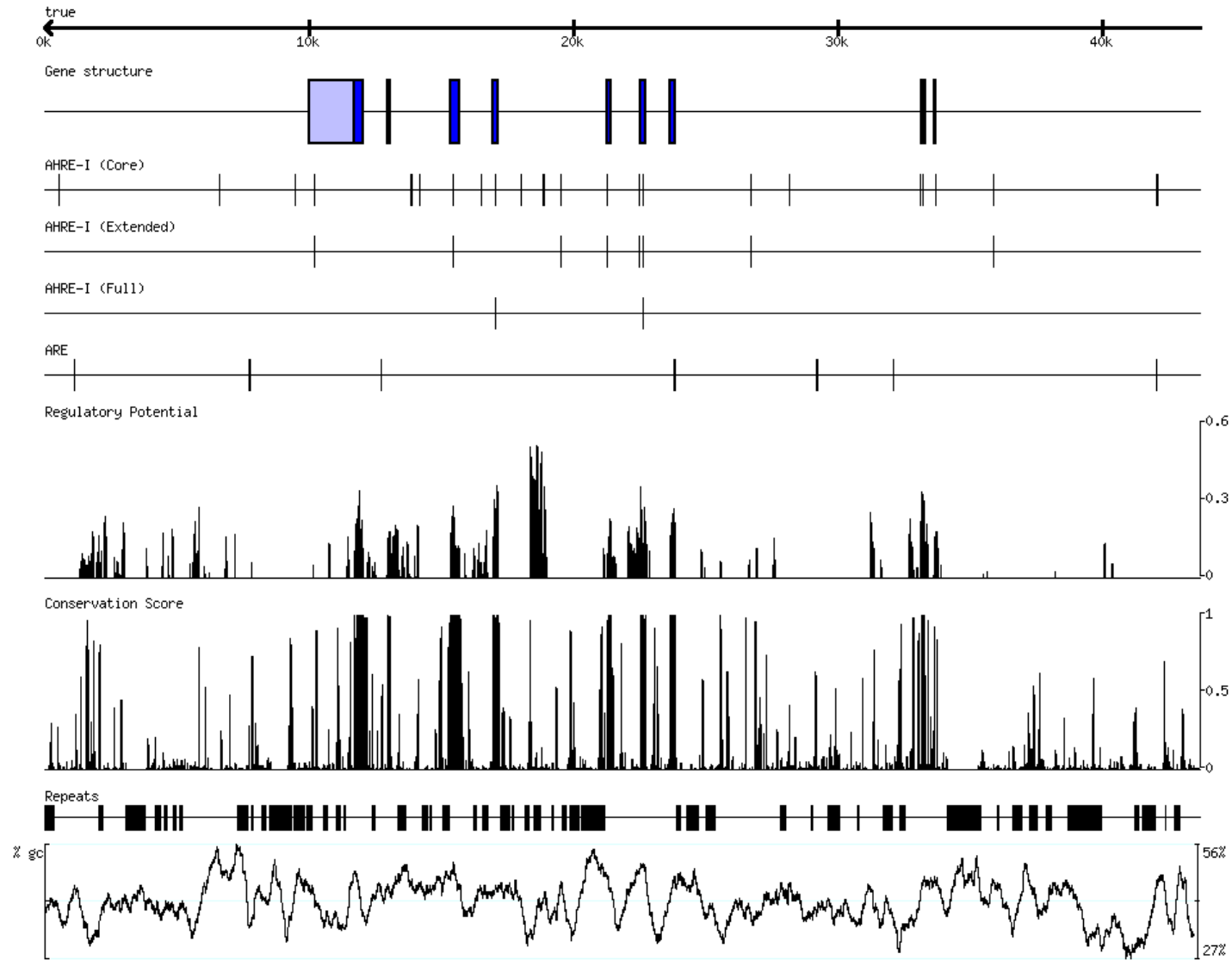
### **Legend for Supplementary Figures 1 to 4**

Sequences for rat and mouse FMO2 and FMO3 genes were downloaded from the UCSC Genome Browser Database (builds rn4 and mm8, respectively). Sequence 10 kbp 3' and 5' to the gene was included for each gene. Also extracted from the same resource were the Regulatory Potential (mouse only) and phyloHMM Conservation scores. The Regulatory Potential indicates the likelihood that any given base plays a regulatory role, while the phyloHMM score gives the posterior probability that a particular base is evolutionarily conserved. In both cases, higher scores indicate higher probabilities. On top of these structures, we also mapped the location of five distinct transcription factor binding motifs: the core, extended, and full AHRE-I motifs; the AHRE-II motif; and the ARE motif. All these motifs and their mappings have been described in detail previously (Boutros et al. *Biochem Biophys Res Comm* 321:707-715, 2004).

### **Legend for Supplementary Figure 5**

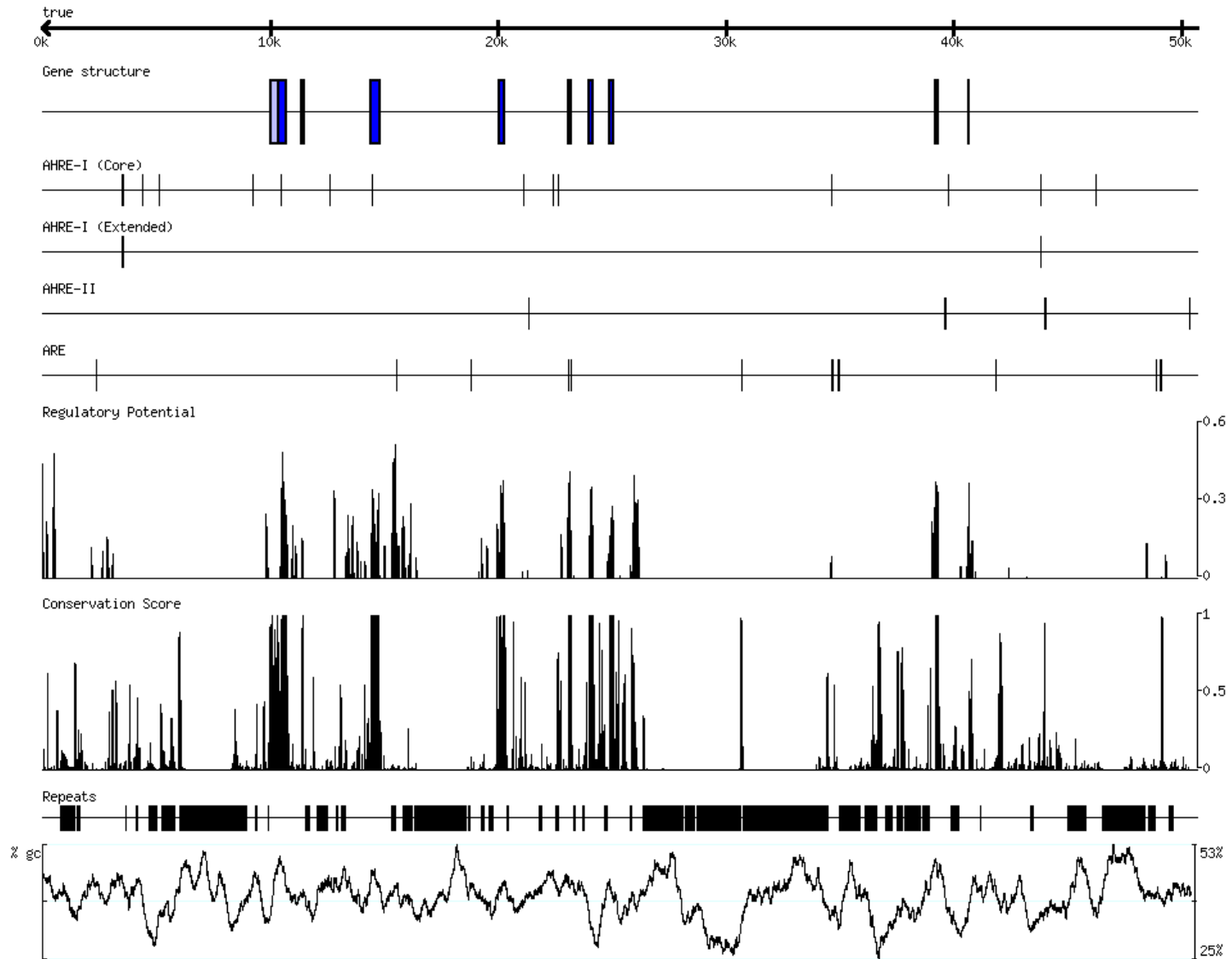
To assess if the region of the FMO3 promoter that bound the AHR in mouse represents a conserved regulatory module we employed a sequence alignment approach. This fragment of the mouse *Fmo3* promoter was extracted from the mouse genome (build mm9). It was then aligned to the rat genome (build rn4) and human genome (build hg18) using the BLAST-Like Alignment Tool (BLAT; Kent, WJ *Genome Research* 12:656-664, 2002), which is specialized for aligning small fragments to genomes. The best-matching alignment for each species was selected. For the human genome this best alignment involved 26 of 78 base-pairs in the mouse region assessed, while for the rat genome it involved 10 of 78 base-pairs. In the human genome the aligned region represents an intron of the *OAZ2* gene and in the rat genome it represents a gene-desert.

# Mouse – FMO2



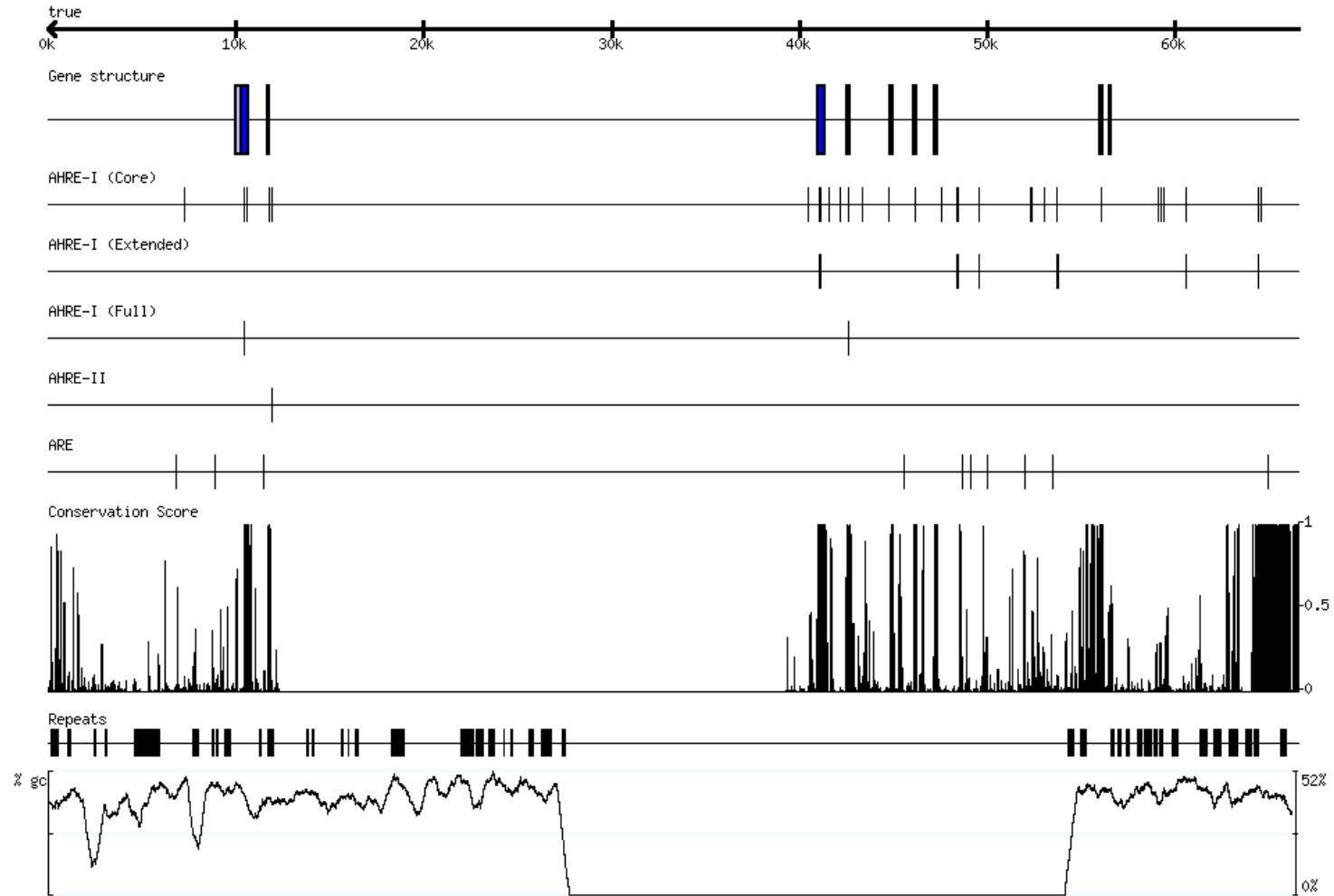
Supp Fig 1

# Mouse – FMO3



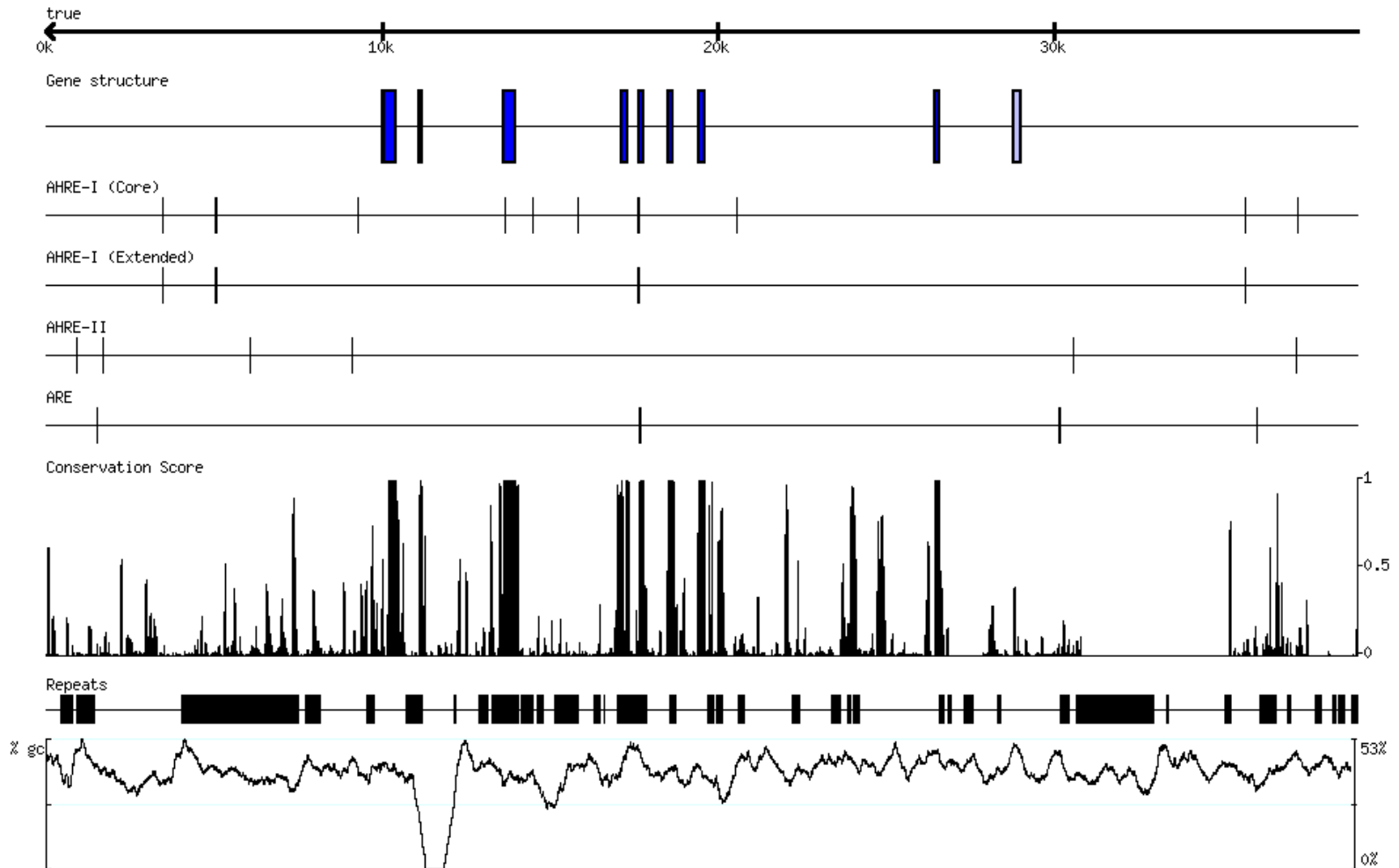
Supp Fig 2

# Rat – FMO2



Supp Fig 3

# Rat – FMO3



Supp Fig 4

