

Drug Metabolism and Disposition

META-ANALYSIS OF THE TURNOVER OF INTESTINAL EPITHELIA IN PRE-CLINICAL ANIMAL SPECIES AND HUMAN

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Supplementary data

Table 1. Enterocyte turnover in the small intestine of the rat.

Segment	Turnover (days)	SD	N	Additional information	Method	References
Stomach	2.84	0.84	6		Colchicine, <i>in vivo</i>	Stevens Leblonde 1953
Proximal SI	1.24	NA	2	Postoperative controls, day 2	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Proximal SI	1.97	NA	2	Postoperative controls, day 4	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Proximal SI	1.71	NA	2	Postoperative controls, day 6	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Proximal SI	2.01	NA	2	Postoperative controls, day 10	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Proximal SI	1.86	NA	2	Postoperative controls, day 12	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Duodenum	1.57	NA	4		Histological study	(Leblond and Stevens, 1948)
Duodenum	1.2	0.2	4		C-glycoside, <i>in vivo</i>	(Macallan et al., 1998)
Duodenum	1.59	NA	5		BrdUrd, <i>in vivo</i>	(Qi et al., 2009)

Duodenum	1.78	NA	12	Controls	H-Thymidine, <i>in vivo</i>	(Holt et al., 1983)
Duodenum	1.98	NA	12	Rats aged 4-5 months	H-Thymidine, <i>in vivo</i>	(Holt et al., 1983)
Duodenum	2.5	NA	8		H-Thymidine, <i>in vivo</i>	(Loran and Althausen, 1960)
Duodenum	2.22	NA	6		H-Thymidine, <i>in vivo</i>	(Koldovsky et al., 1966)
Median SI	2.07	NA	2	Postoperative controls, day 2	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Median SI	2.3	NA	2	Postoperative controls, day 4	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Median SI	1.88	NA	2	Postoperative controls, day 6	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Median SI	1.68	NA	2	Postoperative controls, day 10	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Median SI	1.44	NA	2	Postoperative controls, day 12	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Proximal jejunum	1.5	NA	5		BrdUrd, <i>in vivo</i>	(Qi et al., 2009)
Jejunum	2.29	NA	6	Non-diabetic controls	H-Thymidine, <i>in vivo</i>	(Thomson et al., 1994)
Jejunum	2.21	NA	8	Saturated fat diet	H-Thymidine, <i>in vivo</i>	(Thomson et al., 1994)
Jejunum	1.54	NA	6	Resection controls	H-Thymidine, <i>in vivo</i>	(Thomson et al., 1994)
Jejunum	2.33	NA	6	Irradiation controls	H-Thymidine, <i>in vivo</i>	(Thomson et al., 1994)
Jejunum	2.25	NA	7	20% protein	H-Thymidine, <i>in vivo</i>	(King et al., 1983)

				diet	vivo	
Jejunum	2.38	NA	7	5% protein diet	H-Thymidine, <i>in vivo</i>	(King et al., 1983)
Jejunum	2.04	NA	3		H-Thymidine, <i>in vivo</i>	(Cheeseman, 1986)
Jejunum	1.3	NA	NA		NA	(Bertalanffy and Lau, 1962)
Jejunum	2.06	NA	12	Controls	H-Thymidine, <i>in vivo</i>	(Holt et al., 1983)
Jejunum	2.2	NA	12	Rats aged 4-5 months	H-Thymidine, <i>in vivo</i>	(Holt et al., 1983)
Jejunum	2.08	NA	8		H-Thymidine, <i>in vivo</i>	(Loran and Althausen, 1960)
Jejunum	2.22	NA	6		H-Thymidine, <i>in vivo</i>	(Koldovsky et al., 1966)
Jejunum	3.94	0.54	10		H-Thymidine, <i>in vivo</i>	(Shambaugh et al., 1967)
Jejunum	1.3	NaN	16		Colchicine technique, <i>in vivo</i>	(Bertalanffy, 1960)
Jejunum	2.87	NA	21		H-Thymidine, <i>in vivo</i>	Uribe and Johansson 1988
Distal jejunum	1.64	NA	5		BrdUrd, <i>in vivo</i>	(Qi et al., 2009)
Distal SI	2.5	NA	2	Postoperative controls, day 2	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Distal SI	3.05	NA	2	Postoperative controls, day 4	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Distal SI	2.7	NA	2	Postoperative controls, day 6	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)

Distal SI	2.48	NA	2	Postoperative controls, day 10	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Distal SI	1.45	NA	2	Postoperative controls, day 12	H-Thymidine, <i>in vivo</i>	(Menge et al., 1982)
Proximal ileum	1.62	NA	5		BrdUrd, <i>in vivo</i>	(Qi et al., 2009)
Ileum	1.88	NA	6		H-Alanine and lysine uptake	(Menge et al., 1983)
Ileum	1.35	NA	4		Histological study	(Leblond and Stevens, 1948)
Ileum	1.64	NA	12	Controls	H-Thymidine, <i>in vivo</i>	(Holt et al., 1983)
Ileum	1.37	NA	12	Rats aged 4-5 months	H-Thymidine, <i>in vivo</i>	(Holt et al., 1983)
Ileum	1.6	NA	25		Vincristine, <i>in vivo</i>	(Alam et al., 1994)
Ileum	2.82	NA	8		H-Thymidine, <i>in vivo</i>	(Loran and Althausen, 1960)
Ileum	2.5	NA	6		H-Thymidine, <i>in vivo</i>	(Koldovsky et al., 1966)
Ileum	2.00	NA	NA		H-Thymidine, <i>in vivo</i>	Holle 1991
Distal ileum	1.43	NA	5		BrdUrd, <i>in vivo</i>	(Qi et al., 2009)
NA	2.1	NA	5		Colchicine technique, <i>in vivo</i>	(Altmann and Enesco, 1967)
Caecum	1.04	NA	4		Vincristine, <i>in vivo</i>	Sunter 1979
Colon	3.00	NA	NA		H-Thymidine, <i>in</i>	Messier 1960

				<i>vivo</i>	
Colon	1.34	NA		Proximal colon, apical fold BrdU, <i>in vivo</i>	Qi 2009
Colon	1.77	NA		Proximal colon, basal fold BrdU, <i>in vivo</i>	Qi 2009
Colon	1.40	NA		Mid colon BrdU, <i>in vivo</i>	Qi 2009
Colon	NA	4		Descending colon Vincristine, <i>in vivo</i>	Sunter 1979
Colon	NA	4		Segment 2 Vincristine, <i>in vivo</i>	Sunter 1979
Colon	NA	4		Segment 3 Vincristine, <i>in vivo</i>	Sunter 1979
Colon	10.00	NA	16	Colchicine, <i>in vivo</i>	Bertalanffy 1960
Colon	2.58	NA	10	H-Thymidine, <i>in vivo</i>	Finney 1989
Colon	5.42	4.20- 7.50	12	H-Thymidine, <i>in vivo</i>	Sunter 1978
Colon	1.33	NA	50	Vinblastine, <i>in vivo</i>	Tutton and Barkla 1976
Rectum	6.2	NA	16	Colchicine, <i>in vivo</i>	Bertalanffy 1960
NA=Not applicable, SI=Small intestine.					

Table 2. Enterocyte turnover in the small intestine of the mouse.

Segment	Turnover (days)	SD	N	Additional information	Method	References
Stomach	2.60	NA	NA		H-Thymidine, <i>in vivo</i>	Karam Leblond 1993
Stomach	2.98	NA	4		H-Thymidine, <i>in vivo</i>	Lee 1985
Stomach	3.11	NA	14		H-Thymidine, <i>in vivo</i>	Karam Leblond 1993b
Duodenum	2.81	0.23	16	High- carbohydrate diet	H-Thymidine, <i>in vivo</i>	(Ferraris et al., 1992)
Duodenum	3.09	0.31	16	Low- carbohydrate diet	H-Thymidine, <i>in vivo</i>	(Ferraris et al., 1992)
Duodenum	2.43	0.13	NA		H-Thymidine, <i>in vivo</i>	(Cheng and Bjerknes, 1982)
Duodenum	2.35	0.56	5		H-Thymidine, <i>in vivo</i>	(Merzel and Leblond, 1969)
Duodenum	3.3	NA	8		H-Thymidine, <i>in vivo</i>	(Cheng and Leblond, 1974)
Duodenum	2	NA	1		H-Thymidine, <i>in vivo</i>	(Walker and Leblond, 1958)
Duodenum	2.08	NA	6		H-Thymidine, <i>in vivo</i>	(Grey, 1968)
Duodenum	1.71	NA	20	Age: 93 days	H-Thymidine, <i>in vivo</i>	(Lesher et al., 1961)
Duodenum	2	NA	20	Age: 372 days	H-Thymidine, <i>in vivo</i>	(Lesher et al., 1961)
Duodenum	2.21	NA	20	Age: 940 days	H-Thymidine, <i>in vivo</i>	(Lesher et al., 1961)
Jejunum	3.17	0.33	16	High- carbohydrate diet	H-Thymidine, <i>in vivo</i>	(Ferraris et al., 1992)
Jejunum	2.85	0.31	16	Low- carbohydrate diet	H-Thymidine, <i>in vivo</i>	(Ferraris et al., 1992)

Jejunum	2.59	0.18	NA		H-Thymidine, <i>in vivo</i>	(Cheng and Bjerknes, 1982)
Jejunum	3.4	NA	8		H-Thymidine, <i>in vivo</i>	(Cheng and Leblond, 1974)
Jejunum	4	NA	20		H-Thymidine, <i>in vivo</i>	(Thompson et al., 1990)
Jejunum	1.83	NA	34	Age: 93 days	H-Thymidine, <i>in vivo</i>	(Fry et al., 1961)
Jejunum	2.17	NA	34	Age: 372 days	H-Thymidine, <i>in vivo</i>	(Fry et al., 1961)
Jejunum	2.21	NA	52	Age: 940 days	H-Thymidine, <i>in vivo</i>	(Fry et al., 1961)
Ileum	2.6	0.38	16	High-carbohydrate diet	H-Thymidine, <i>in vivo</i>	(Ferraris et al., 1992)
Ileum	2.56	0.35	16	Low-carbohydrate diet	H-Thymidine, <i>in vivo</i>	(Ferraris et al., 1992)
Ileum	2.43	0.12	NA		H-Thymidine, <i>in vivo</i>	(Cheng and Bjerknes, 1982)
Ileum	2	NA	NA		H-Thymidine, <i>in vivo</i>	(Quastler and Sherman, 1959)
Ileum	1.29	NA	10	Age: 93 days	H-Thymidine, <i>in vivo</i>	(Fry et al., 1962)
Ileum	1.38	NA	8	Age: 372 days	H-Thymidine, <i>in vivo</i>	(Fry et al., 1962)
Ileum	1.29	NA	10	Age: 940 days	H-Thymidine, <i>in vivo</i>	(Fry et al., 1962)
Ileum	1.00	NA	23		H-Thymidine, <i>in vivo</i>	Creamer 1961
Colon	2.51	1.61	5		H-Thymidine, <i>in vivo</i>	Cheng and Bjerknes 1983
Colon	2.17	NA	5		Histological examination	Aluwihare 1971
Colon	4.85	0.94	4		H-Thymidine, <i>in vivo</i>	Tsubouchi 1981
Colon	2.42	0.11	NA		H-Thymidine, <i>in vivo</i>	Cheng and Bjerknes 1982
Colon	3.79	NA	8		H-Thymidine, <i>in vivo</i>	Chang Leblond 1971

Colon	4.96	1.24	18	Flat mucosa	Colchicine, <i>in vivo</i>	Baril 1982
Colon	3.11	0.65	18	Mucosal fold tops	Colchicine, <i>in vivo</i>	Baril 1982
Colon	0.67	NA	35		H-Thymidine, <i>in vivo</i>	Lipkin and Quastler 1962
Colon	0.65	NA	36		H-Thymidine, <i>in vivo</i>	De Rodriguez 1979
Colon	0.79	NA	25		H-Thymidine, <i>in vivo</i>	Thrasher 1967
Colon	1.06	NA	20		H-Thymidine, <i>in vivo</i>	Richards 1977
Colon	0.97	NA	16		H-Thymidine, <i>in vivo</i>	Chang Nadler 1975
Colon	0.97	NA	22		H-Thymidine, <i>in vivo</i>	Kovacs and Potten 1973
Colon	0.79	NA	4		H-Thymidine, <i>in vivo</i>	Potten 1974
Colon	0.67	NA	12		H-Thymidine, <i>in vivo</i>	Bottomley 1973
Colon	0.90	0.22	7		H-Thymidine, <i>in vivo</i>	Novacki 1993
Jejunum and ileum	3	NA	NA		H-Thymidine, <i>in vivo</i>	(Leblond and Messier, 1958)
Duodenum and jejunum	2-3	NA	23		H-Thymidine, <i>in vivo</i>	Creamer 1961
NA	2.67	NA	7		H-Thymidine, <i>in vivo</i>	(Smith et al., 1984)

NA=Not applicable.

Table 3. Enterocyte turnover in the human gastrointestinal tract.

Segment	Turnover (Days)	SD	N	Additional information	Method	References
Oesophagus	6.35	2.08	1		H-Thymidine, <i>in vivo</i>	(Bell et al., 1967)
Stomach	5	1	2		H-Thymidine, <i>in vivo</i>	(Macdonald et al., 1964)
Stomach	3.5	0.5	3		H-Thymidine, <i>in vivo</i>	(Lipkin et al., 1963b)
Stomach	3.46	1.21	16	Gastric body	BrdUrd, <i>in vivo</i>	(Patel et al., 1993)
Stomach	2.58	1.17	10	Gastric antrum	BrdUrd, <i>in vivo</i>	(Patel et al., 1993)
Stomach	3.43	1.14	4		H-Thymidine, <i>in vivo</i>	(Wright et al., 1977)
Duodenum	5.42	NA	1	Individual 1	H-Thymidine, <i>in vivo</i>	(Macdonald et al., 1964)
Duodenum	5.5	0.5	1	Individual 2	H-Thymidine, <i>in vivo</i>	(Macdonald et al., 1964)
Duodenum	2	NA	56		Histological study	(Bertalanffy and Nagy, 1961)
Duodenum	1.26	0.17	2		H-Thymidine, <i>in vivo</i>	(Weinstein, 1974)
Jejunum	5	NA	3		H-Thymidine, <i>in vivo</i>	(Shorter et al., 1964)
Ileum	1.4	NA	6		Histological study	(Bullen et al., 2006)
Ileum	3	NA	3		H-Thymidine, <i>in vivo</i>	(Lipkin et al., 1963b)
Colon	1	NA	2		H-Thymidine, <i>in vivo</i>	(Lipkin et al., 1963a)
Colon	0.83	NA	3		H-Thymidine, <i>in vivo</i>	(Lipkin et al., 1963b)
Colon	3.41	NA	66		BrdUrd, <i>in vivo</i>	(Potten et al., 1992)
Colon	1.63	NA	1		H-Thymidine, <i>in vivo</i>	(Lipkin, 1969)
Colon	3.04	0.25	8		H-Thymidine, <i>in vitro</i>	(Bleiberg and Galand, 1976)
Rectum	5.5	0.5	2		H-Thymidine, <i>in vivo</i>	(Macdonald et al., 1964)
Rectum	7	1	1		H-Thymidine, <i>in vivo</i>	(Cole and Mc, 1961)

Rectum	3.66	0.42	18	Normal subjects	Histological study	(Shorter et al., 1966)
Rectum	4.73	0.59	9	Chronic ulcerative colitis in remission	Histological study	(Shorter et al., 1966)
Rectum	5.45	0.61	17	Active chronic ulcerative colitis	Histological study	(Shorter et al., 1966)
Rectum	0.83	NA	3		H-Thymidine, <i>in vivo</i>	(Lipkin et al., 1963b)
Rectum	3	NA	3		H-Thymidine, <i>in vivo</i>	(Shorter et al., 1964)
Rectum	3.5	0.5	16		H-Thymidine, <i>in vitro</i>	(Deschner et al., 1963)
Rectum	3.75	NA	8		H-Thymidine, <i>in vitro</i>	(Bleiberg et al., 1970)
NA=Not applicable						

Methods: Statistical analysis

Weighted arithmetic means (WX) were calculated utilising reported means (x_i) and sample size (n_i) of the individual studies (Equation 1). Studies reporting mean only data where n_j was not clearly stated were penalised, assuming $n_j = 1$; whereas for studies where the mean and standard deviation were reported without clearly stating n_j were assigned $n_j = 3$.

$$WX = \frac{\sum_{i=1}^n n_i \cdot X_i}{\sum_{i=1}^n n_i}$$

Equation 1

The combined standard deviation (overall SD) was obtained through calculating the total sum of squares (overall SS) (Equation 2 and Equation 3).

$$Overall\ SS = \sum_{i=1}^n [(SD_i^2 + X_i^2) + n_i] - N \cdot WX^2$$

Equation 2

$$Overall\ SD = \sqrt{\frac{Overall\ SS}{N}}$$

Equation 3

The geometric mean (GM) and standard deviation (GSD) of the enterocyte turnover was calculated by calculating σ (lnSD; Equation 4 and Equation 5) and using the reported mean and variance and μ (lnGM; Equation 6 and Equation 7).

$$\sigma = \sqrt{\ln\left(\frac{variance}{X^2} + 1\right)}$$

Equation 4

$$GSD = e^\sigma$$

Equation 5

$$\mu = \ln X - \frac{1}{2} \cdot \ln\left(\frac{1}{2} \cdot \sigma^2\right)$$

Equation 6

$$GM = e^\mu$$

Equation 7

Statistical analysis was carried out using Welch's t test ($P<0.05$) with post-hoc Dunn-Šidák correction in Matlab® R2010a (Mathworks, Natick, USA), using the sample means (X), size (n) and standard deviations (s^2), where data allowed (Equation 8).

$$t = \frac{X_1 - X_2}{\sqrt{\frac{s_1^2}{N_1} + \frac{s_2^2}{N_2}}}$$

Equation 8

References

- Alam M, Midtvedt T, and Uribe A (1994) Differential cell kinetics in the ileum and colon of germfree rats. *Scand J Gastroenterol* **29**:445-451.
- Altmann GG and Enesco M (1967) Cell number as a measure of distribution and renewal of epithelial cells in the small intestine of growing and adult rats. *Am J Anat* **121**:319-336.
- Bell B, Almy TP, and Lipkin M (1967) Cell proliferation kinetics in the gastrointestinal tract of man. 3. Cell renewal in esophagus, stomach, and jejunum of a patient with treated pernicious anemia. *Journal of the National Cancer Institute* **38**:615-628.
- Bertalanffy FD (1960) Mitotic rates and renewal times of the digestive tract epithelia in the rat. *Acta anatomica* **40**:130-148.
- Bertalanffy FD and Lau C (1962) Cell Renewal. *International Journal of Cytology* **13**:357-366.
- Bertalanffy FD and Nagy KP (1961) Mitotic activity and renewal rate of the epithelial cells of human duodenum. *Acta anatomica* **45**:362-370.
- Bleiberg H and Galand P (1976) In vitro autoradiographic determination of cell kinetic parameters in adenocarcinomas and adjacent healthy mucosa of the human colon and rectum. *Cancer Res* **36**:325-328.
- Bleiberg H, Mainguet P, Galand P, Chretien J, and Dupont-Mairesse N (1970) Cell renewal in the human rectum. In vitro autoradiographic study on active ulcerative colitis. *Gastroenterology* **58**:851-855.
- Bullen TF, Forrest S, Campbell F, Dodson AR, Hershman MJ, Pritchard DM, Turner JR, Montrose MH, and Watson AJ (2006) Characterization of epithelial cell shedding from human small intestine. *Lab Invest* **86**:1052-1063.
- Cheeseman CI (1986) Expression of amino acid and peptide transport systems in rat small intestine. *Am J Physiol* **251**:G636-641.
- Cheng H and Bjerknes M (1982) Whole population cell kinetics of mouse duodenal, jejunal, ileal, and colonic epithelia as determined by radioautography and flow cytometry. *Anat Rec* **203**:251-264.
- Cheng H and Leblond CP (1974) Origin, differentiation and renewal of the four main epithelial cell types in the mouse small intestine. V. Unitarian Theory of the origin of the four epithelial cell types. *Am J Anat* **141**:537-561.

- Cole JW and Mc KA (1961) Observations of cell renewal in human rectal mucosa in vivo with thymidine-H3. *Gastroenterology* **41**:122-125.
- Deschner E, Lewis CM, and Lipkin M (1963) In Vitro Study of Human Rectal Epithelial Cells. I. Atypical Zone of H3 Thymidine Incorporation in Mucosa of Multiple Polyposis. *J Clin Invest* **42**:1922-1928.
- Ferraris RP, Villenas SA, and Diamond J (1992) Regulation of brush-border enzyme activities and enterocyte migration rates in mouse small intestine. *Am J Physiol* **262**:G1047-1059.
- Fry RJ, Lesher S, and Kohn HI (1961) Age effect on cell-transit time in mouse jejunal epithelium. *Am J Physiol* **201**:213-216.
- Fry RJ, Lesher S, and Kohn HI (1962) Influence of age on the transit time of cells of the mouse intestinal epithelium. III. Ileum. *Lab Invest* **11**:289-293.
- Grey RD (1968) Epithelial cell migration in the intestine of the young mouse. *Developmental biology* **18**:501-504.
- Holt PR, Kotler DP, and Pascal RR (1983) A simple method for determining epithelial cell turnover in small intestine. Studies in young and aging rat gut. *Gastroenterology* **84**:69-74.
- King IS, Paterson JY, Peacock MA, Smith MW, and Syme G (1983) Effect of diet upon enterocyte differentiation in the rat jejunum. *J Physiol* **344**:465-481.
- Koldovsky O, Sunshine P, and Kretchmer N (1966) Cellular migration of intestinal epithelia in suckling and weaned rats. *Nature* **212**:1389-1390.
- Leblond CP and Messier B (1958) Renewal of chief cells and goblet cells in the small intestine as shown by radioautography after injection of thymidine-H3 into mice. *Anat Rec* **132**:247-259.
- Leblond CP and Stevens CE (1948) The constant renewal of the intestinal epithelium in the albino rat. *Anat Rec* **100**:357-377.
- Lesher S, Fry RJ, and Kohn HI (1961) Influence of age on transit time of cells of mouse intestinal epithelium. I. Duodenum. *Lab Invest* **10**:291-300.
- Lipkin M (1969) Cell proliferation in gastrointestinal disease. *National Cancer Institute monograph* **30**:199-207.
- Lipkin M, Bell B, and Sherlock P (1963a) Cell Proliferation Kinetics in the Gastrointestinal Tract of Man. I. Cell Renewal in Colon and Rectum. *J Clin Invest* **42**:767-776.
- Lipkin M, Sherlock P, and Bell B (1963b) Cell Proliferation Kinetics in the Gastrointestinal Tract of Man. II. Cell Renewal in Stomach, Ileum, Colon, and Rectum. *Gastroenterology* **45**:721-729.
- Loran MR and Althausen TL (1960) Cellular proliferation of intestinal epithelia in the rat two months after partial resection of the ileum. *The Journal of biophysical and biochemical cytology* **7**:667-672.
- Macallan DC, Fullerton CA, Neese RA, Haddock K, Park SS, and Hellerstein MK (1998) Measurement of cell proliferation by labeling of DNA with stable isotope-labeled glucose: studies in vitro, in animals, and in humans. *Proc Natl Acad Sci U S A* **95**:708-713.
- Macdonald WC, Trier JS, and Everett NB (1964) Cell Proliferation and Migration in the Stomach, Duodenum, and Rectum of Man: Radioautographic Studies. *Gastroenterology* **46**:405-417.
- Menge H, Hopert R, Alexopoulos T, and Riecken EO (1982) Three-dimensional structure and cell kinetics at different sites of rat intestinal remnants during the early adaptive response to resection. *Research in experimental medicine Zeitschrift fur die gesamte experimentelle Medizin einschliesslich experimenteller Chirurgie* **181**:77-94.

- Menge H, Sepulveda FV, and Smith MW (1983) Cellular adaptation of amino acid transport following intestinal resection in the rat. *J Physiol* **334**:213-223.
- Merzel J and Leblond CP (1969) Origin and renewal of goblet cells in the epithelium of the mouse small intestine. *Am J Anat* **124**:281-305.
- Patel S, Rew DA, Taylor I, Potten CS, Owen C, and Roberts SA (1993) Study of the proliferation in human gastric mucosa after in vivo bromodeoxyuridine labelling. *Gut* **34**:893-896.
- Potten CS, Kellett M, Rew DA, and Roberts SA (1992) Proliferation in human gastrointestinal epithelium using bromodeoxyuridine in vivo: data for different sites, proximity to a tumour, and polyposis coli. *Gut* **33**:524-529.
- Qi WM, Yamamoto K, Yokoo Y, Miyata H, Inamoto T, Udayanga KG, Kawano J, Yokoyama T, Hoshi N, and Kitagawa H (2009) Histoplanimetric study on the relationship between the cell kinetics of villous columnar epithelial cells and the proliferation of indigenous bacteria in rat small intestine. *The Journal of veterinary medical science / the Japanese Society of Veterinary Science* **71**:463-470.
- Quastler H and Sherman FG (1959) Cell population kinetics in the intestinal epithelium of the mouse. *Experimental cell research* **17**:420-438.
- Shambaugh GE, MacNair DS, and Beisel WR (1967) Small-bowel epithelial migration during a generalized nonenteric infection in the rat. *Am J Dig Dis* **12**:403-408.
- Shorter RG, Moertel CG, Titus JL, and Reitemeier RJ (1964) Cell Kinetics in the Jejunum and Rectum of Man. *Am J Dig Dis* **9**:760-763.
- Shorter RG, Spencer RJ, and Hallenbeck GA (1966) Kinetic studies of the epithelial cells of the rectal mucosa in normal subjects and patients with ulcerative colitis. *Gut* **7**:593-596.
- Smith MW, Patterson JY, and Peacock MA (1984) A comprehensive description of brush border membrane development applying to enterocytes taken from a wide variety of mammalian species. *Comparative biochemistry and physiology A, Comparative physiology* **77**:655-662.
- Thompson EM, Price YE, and Wright NA (1990) Kinetics of enteroendocrine cells with implications for their origin: a study of the cholecystokinin and gastrin subpopulations combining tritiated thymidine labelling with immunocytochemistry in the mouse. *Gut* **31**:406-411.
- Thomson AB, Cheeseman CI, Keelan M, Fedorak R, and Clandinin MT (1994) Crypt cell production rate, enterocyte turnover time and appearance of transport along the jejunal villus of the rat. *Biochimica et biophysica acta* **1191**:197-204.
- Walker BE and Leblond CP (1958) Sites of nucleic acid synthesis in the mouse visualized by radioautography after administration of C14-labelled adenine and thymidine. *Experimental cell research* **14**:510-531.
- Weinstein WM (1974) Epithelial cell renewal of the small intestinal mucosa. *The Medical clinics of North America* **58**:1375-1386.
- Wright NA, Britton DC, Bone G, and Appleton DR (1977) An in vivo stathmokinetic study of cell proliferation in human gastric carcinoma and gastric mucosa. *Cell and tissue kinetics* **10**:429-436.