



# Differences in Metformin and Thiamine Uptake between Human and Mouse Organic Cation Transporter 1: Structural Determinants and Potential Consequences for Intrahepatic Concentrations<sup>IS</sup>

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## ABSTRACT

The most commonly used oral antidiabetic drug, metformin, is a substrate of the hepatic uptake transporter OCT1 (gene name *SLC22A1*). However, OCT1 deficiency leads to more pronounced reductions of metformin concentrations in mouse than in human liver. Similarly, the effects of OCT1 deficiency on the pharmacokinetics of thiamine were reported to differ between human and mouse. Here, we compared the uptake characteristics of metformin and thiamine between human and mouse OCT1 using stably transfected human embryonic kidney 293 cells. The affinity for metformin was 4.9-fold lower in human than in mouse OCT1, resulting in a 6.5-fold lower intrinsic clearance. Therefore, the estimated liver-to-blood partition coefficient is only 3.34 in human compared with 14.4 in mouse and may contribute to higher intrahepatic concentrations in mice. Similarly, the affinity for thiamine was 9.5-fold lower in human than in mouse OCT1. Using human-mouse chimeric OCT1, we showed that simultaneous substitution of transmembrane helices TMH2 and TMH3 resulted in the reversal of affinity for metformin. Using homology modeling, we suggest several explanations, of which a different interaction of Leu155 (human

TMH2) compared with Val156 (mouse TMH2) with residues in TMH3 had the strongest experimental support. In conclusion, the contribution of human OCT1 to the cellular uptake of thiamine and especially of metformin may be much lower than that of mouse OCT1. This may lead to an overestimation of the effects of OCT1 on hepatic concentrations in humans when using mouse as a model. In addition, comparative analyses of human and mouse orthologs may help reveal mechanisms of OCT1 transport.

## SIGNIFICANCE STATEMENT

OCT1 is a major hepatic uptake transporter of metformin and thiamine, but this study reports strong differences in the affinity for both compounds between human and mouse OCT1. Consequently, intrahepatic metformin concentrations could be much higher in mice than in humans, impacting metformin actions and representing a strong limitation of using rodent animal models for predictions of OCT1-related pharmacokinetics and efficacy in humans. Furthermore, OCT1 transmembrane helices TMH2 and TMH3 were identified to confer the observed species-specific differences in metformin affinity.

## Introduction

Metformin is the most commonly prescribed oral antidiabetic drug. It reduces plasma glucose and has favorable effects on lipid metabolism.

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Metformin acts both in the liver and in the gut (Rena et al., 2017). In hepatocytes, metformin decreases glucose production and lipogenesis both by AMPK-dependent and AMPK-independent mechanisms.

At physiologic pH, metformin is almost entirely present as organic cation ( $pK_a$  of 12.04, estimated 99.998% positively charged molecules). Therefore, metformin depends highly on transporter proteins to enter hepatocytes. The organic cation transporter 1 (OCT1, gene name *SLC22A1*), which is strongly expressed in the sinusoidal membrane of hepatocytes, has been demonstrated to be the major hepatic uptake transporter of metformin (Wang et al., 2002; Shu et al., 2007). In humans, OCT1 is genetically highly variable. In total, 9% of Europeans and white Americans are carriers of two reduced function or loss-of-function OCT1 alleles and are so-called poor OCT1

**ABBREVIATIONS:** AMPK, AMP-activated protein kinase; DPBS, Dulbecco's phosphate-buffered saline; GLUT3, glucose 3 transporter; h, human; HBSS, Hanks' buffered salt solution; HEK, human embryonic kidney; IS, internal standard; IVIVE, in vitro to in vivo extrapolation;  $K_p$ , liver-to-blood partition coefficient; LC-MS/MS, liquid chromatography tandem mass spectrometry; m, mouse; MFS, major facilitator superfamily; OCT, organic cation transporter; TMH, transmembrane helix PET, positron emission tomography; DAPI, 4',6-diamidino-2-phenylindole; THTR, thiamine transporter; SLC, solute carrier.

transporters (Kerb et al., 2002; Shu et al., 2003; Tzvetkov et al., 2012; Seitz et al., 2015). In some specific populations, like Surui Indians, this percentage may increase up to 80% (Seitz et al., 2015). These genetic OCT1 variants are expected to affect the hepatic uptake and thus the efficacy of metformin.

Indeed, OCT1 knockout in mice reduced metformin concentrations in the liver by up to 30-fold (Wang et al., 2002, 2003) and abolished the glucose-lowering effects of metformin (Shu et al., 2007). OCT1 deficiency in mice has been suggested to lead to an absolute lack of metformin uptake into hepatocytes (Wang et al., 2002). Also in humans, genetic variants leading to decreased OCT1 activity were associated with reduced intrahepatic concentrations of metformin (Sundelin et al., 2017). However, despite some initial reports of reduced response to metformin in poor OCT1 transporters (Shu et al., 2007), larger studies and meta-analyses could not confirm the association of OCT1 genetic variants with reduced efficacy of metformin in humans (Zhou et al., 2009; Dujic et al., 2017).

Similarly, OCT1 was identified as a relevant transporter of thiamine (vitamin B1) in the mouse liver (Chen et al., 2014). OCT1 knockout in mice resulted in higher thiamine plasma levels, likely because of decreased hepatic extraction (Chen et al., 2014; Liang et al., 2018). However, a recent study in humans showed no differences in the plasma concentrations of thiamine or its metabolites in poor OCT1 transporters (Jensen et al., 2020). Taken together, this questions the suitability of mouse as a model for studying effects of OCT1 on metformin or thiamine-driven biologic processes without understanding the causes of the species differences.

One possible explanation for the variable results between human and mouse may be differences in the kinetics of OCT1-mediated uptake of metformin and thiamine. Although species differences in organ-specific OCT1 expression are well characterized (Gorboulev et al., 1997; Zhang et al., 1997; Green et al., 1999; Schmitt et al., 2003), there is only very limited data reporting differences in transport activity. The amino acid identity between the human and mouse OCT1 orthologs is 77%. Since the exact mechanism of substrate interaction with OCT1 is not known, it is difficult to predict to what extent the 23% different amino acids between the two orthologs can confer differences in uptake. Furthermore, metformin and thiamine were suggested to share common sites of ligand-transporter interaction within OCT1 (Chen et al., 2014). Therefore, similar differences in the uptake between human and mouse could be expected for these two substrates.

The aim of this study was to compare the uptake of metformin and thiamine by human and mouse OCT1 *in vitro* to explore underlying mechanisms causing differences in the hepatic concentrations in humans and mice. This should help to better interpret the data of mouse models and should improve the translation of mouse pharmacokinetic data to humans. Furthermore, we used the differences in uptake between human and mouse OCT1 as a tool to improve our understanding of the transport mechanism of OCT1.

## Materials and Methods

**Reagents.** Metformin hydrochloride, thiamine hydrochloride, ammoniumbicarbonate, dithiothreitol, and iodoacetamide were obtained from Sigma-Aldrich (Taufkirchen, Germany); buformin hydrochloride was obtained from Wako Chemicals (Neuss, Germany); and thiamine-d3 hydrochloride was obtained from Toronto Research Chemicals (North York, ON, Canada). All chemicals used in this study were purchased from commercial sources and had purities of 95% or higher. Dulbecco's modified Eagle's medium (DMEM), Hanks' buffered salt solution (HBSS), and additives for cell culturing were obtained from Life Technologies (Darmstadt, Germany). Dulbecco's phosphate-buffered saline (DPBS) was obtained from PAN-Biotech (Aidenbach, Germany). Poly-D-lysine (1–5 kDa), HEPES, bicinchoninic acid, and copper sulfate pentahydrate were

obtained from Sigma-Aldrich. Twelve-well plates were obtained from Nunc (Langensfeld, Germany), and tissue culture flasks were from Sarstedt (Nümbrecht, Germany). Acetonitrile, methanol, and formic acid in LC-MS/MS grade and sodium chloride were obtained from Merck (Darmstadt, Germany). SDS (ultrapure) was obtained from AppliChem (Darmstadt, Germany). Sequencing Grade Modified Trypsin and ProteaseMAX surfactant were obtained from Promega (Mannheim, Germany).

**Generation of OCT1 Constructs.** For overexpression of OCT1 in human embryonic kidney (HEK) 293 cells, pcDNA5/FRT expression vectors (Thermo Fisher Scientific, Darmstadt, Germany) containing wild-type, mutant, or chimeric OCT1 constructs were generated as follows or as described previously (Tzvetkov et al., 2012; Seitz et al., 2015). Human-mouse chimeric OCT1 was generated by restriction of human and mouse OCT1 genes with Bsu36I and BsaBI, separating the OCT1 gene into three fragments: from N terminus to large intracellular loop, from transmembrane helix (TMH) 7 to TMH9, and from TMH10 to C terminus (Fig. 3A). The fragments were ligated back together in the correct order but with different combinations of the species, and the resulting chimeric OCT1 genes were cloned into the pcDNA5/FRT vector after restriction of both gene and vector with HindIII and EcoRV. These constructs were then used for targeted chromosomal integration into HEK293 cells. Human-mouse chimeric OCT1 constructs with single TMH substitutions were generated using the overlap extension method (Horton et al., 1989) and primers listed in Supplemental Table 1. Point mutations in human and mouse OCT1 genes were introduced by site-directed mutagenesis in pcDNA5/FRT vectors containing human or mouse OCT1 wild-type genes, using primers listed in Supplemental Table 1. All generated constructs were validated by capillary sequencing of the complete open reading frame of OCT1 before transfection into HEK293 cells.

**Cell Lines and Cell Culturing.** HEK293 cells stably overexpressing human OCT1, mouse OCT1, rat OCT1, human-mouse chimeric OCT1, or human OCT2 were generated by targeted chromosomal integration using the Flp-In System (Life Technologies) as described previously (Tzvetkov et al., 2012; Seitz et al., 2015). Cells were cultured in Dulbecco's modified Eagle's medium supplemented with 10% FBS, 100 U/ml penicillin, and 100 µg/ml streptomycin at 37°C and 5% CO<sub>2</sub>.

**Transient Transfection of T-Rex-293 Cells for Cellular Uptake Experiments.** For transient transfection of OCT1 constructs into HEK293 cells for uptake experiments,  $5 \times 10^5$  T-Rex-293 cells (Life Technologies) were seeded per well of a 12-well plate precoated with poly-D-lysine. At 24 hours later, the cells were transfected with 100 µl of reaction mix per well, containing 2 µg pcDNA5/FRT vector with the OCT1 construct of interest, 0.5 µg pGFP-tpz vector, and 6.25 µl Lipofectamine 2000 (Thermo Fisher Scientific), according to the manufacturer's instructions. At 48 hours later, transfection efficacy was assessed microscopically by visualizing the GFP signal of the cotransfected GFP vector, and the cells were used for uptake experiments.

**Cellular Uptake Experiments.** At 48 hours prior to the experiment,  $6 \times 10^5$  cells were seeded per well of a 12-well plate. When using transiently transfected cells,  $5 \times 10^5$  T-Rex-293 cells were seeded per well of a 12-well plate 72 hours prior to the experiment, and they were transfected 24 hours later as described above. Twelve-well plates were precoated with poly-D-lysine.

Cellular uptake experiments were performed at 37°C and pH 7.4 using HBSS supplemented with 10 mM HEPES (in the following referred to as HBSS+). Cells were washed once with 1 ml of prewarmed (37°C) HBSS+, and uptake was started by adding 400 µl prewarmed HBSS+ containing the substrate. Uptake was stopped after 2 minutes by adding 2 ml ice-cold HBSS+. Cells were washed twice with 2 ml ice-cold HBSS+ and were lysed in 500 µl 80% acetonitrile supplemented with internal standard (Table 1). Intracellular substrate concentrations were measured by LC-MS/MS as described below and afterward were normalized to the total amount of protein in the sample as measured using the bicinchoninic acid assay (Smith et al., 1985).

**Quantification of Intracellular Substrate Concentration by LC-MS/MS.** For quantification of intracellular substrate concentrations, the cell debris was removed by centrifugation of the cell lysate at 16,000g for 15 minutes. In total, 350 µl of the supernatant was evaporated to dryness under nitrogen flow at 40°C. The sample was reconstituted in 200 µl 0.1% formic acid, and 5 or 15 µl was injected into the LC-MS/MS system for metformin and thiamine, respectively.

For LC-MS/MS quantification, an API 4000 tandem mass spectrometer (AB SCIEX, Darmstadt, Germany) was used. Samples were separated on a Brownlee SPP RP-Amide column (4.6 × 100 mm, 2.7 µm; PerkinElmer, Rodgau,

TABLE 1  
Parameters of quantitative LC-MS/MS analyses

Analyte	Quantifier Precursor Ion to Product Ion (m/z)	Retention Time (min)	IS	IS Precursor Ion to Product Ion (m/z)	Retention Time IS (min)	Mobile Phase (% Organic Solvent) <sup>a</sup>	Flow (μl/min)
Metformin	130.1 > 71	2.88	Buformin	158.1 > 60	4.0	3	300
Thiamine	265.3 > 122	2.47	Thiamine-d3	269.1 > 125	2.47	3	350

m/z, mass-to-charge ratio.

<sup>a</sup>Six parts acetonitrile + one part methanol.

Germany) using a mobile phase of 0.1% (v/v) formic acid and varying concentrations of organic solvent (parameters are listed in Table 1).

**Immunocytochemical Staining and Confocal Microscopy Analysis of OCT1-Overexpressing Cells.** For immunocytochemical staining of OCT1,  $6 \times 10^5$  HEK293 cells stably overexpressing human, mouse, or human-mouse chimeric OCT1 were seeded onto coverslips in 12-well plates 48 hours prior to the experiment. Coverslips were precoated with poly-D-lysine. Cells were washed twice with 1 ml DPBS for 10 minutes and were fixed with 100% ethanol for 20 minutes at  $-20^\circ\text{C}$ . After washing three times with DPBS for 5 minutes, cell membranes were permeabilized with DPBS/0.4% Tween 20 for 10 minutes. Cells were washed three times with DPBS for 5 minutes and blocked with blocking buffer (DPBS/5% FBS) for 1–3 hours. Cells were incubated with the primary antibodies diluted in blocking buffer (according to Supplemental Table 2) in a humid chamber overnight. The next day, after washing three times with DPBS for 5 minutes, the cells were incubated with the secondary antibodies diluted in blocking buffer (according to Supplemental Table 2) for 1 to 2 hours protected from light. After washing three times with DPBS for 5 minutes, coverslips were mounted with ROTI-Mount FluorCare DAPI (Carl Roth, Karlsruhe, Germany) onto microscope slides. The cells were analyzed using a laser scanning microscope (LSM780; Carl Zeiss, Jena, Germany), and the images were processed using the Fiji distribution of ImageJ2 (Schindelin et al., 2012; Rueden et al., 2017).

**Quantification of OCT1 Protein Abundance by Targeted Proteomics.** Normal human liver tissue was obtained as excess material, which had to be removed for technical reasons during liver surgery. Patients had given their informed consent for research use of the tissues, and the procedures were approved by the ethics committee of the University Medicine Göttingen, Georg-August-Universität Göttingen (application number 26/01/17). Preparation of murine liver was carried out in compliance with the German laws on animal welfare (§ 4 Absatz 3 TierSchG), and all animals used were reported to the Landesveterinär- und Lebensmitteluntersuchungsamt Mecklenburg-Vorpommern.

Human ( $N = 12$ , eight female and four male) and murine [ $N = 18$ , each  $N = 6$  of C57BL/6N (two female and four male), C57BL/6J, and FVB mice (each three female and three male)] liver samples were mechanically crushed in a stainless-steel mortar system, precooled in liquid nitrogen. Approximately 100 mg tissue powder was used for isolation of native integral membrane using the ProteoExtract Native Membrane Extraction Kit according to the manufacturer's instructions (Merck). Tissues were additionally homogenized in a glass douncer during the step of cell lysis. Cell pellets of OCT1-overexpressing cells were directly used for extraction. Total protein content of the resulting membrane fraction was determined by bicinchoninic acid assay. If necessary, membrane fractions were adjusted to a maximum protein amount of  $2 \mu\text{g}/\mu\text{l}$ . Subsequently,  $100 \mu\text{l}$  of each membrane fraction was mixed with  $10 \mu\text{l}$  dithiothreitol (200 mM),  $40 \mu\text{l}$  ammonium bicarbonate buffer (50 mM, pH 7.8), and  $10 \mu\text{l}$  ProteaseMAX (1%, m/v) and incubated for 30 minutes at  $60^\circ\text{C}$  (denaturation). After cooling down,  $10 \mu\text{l}$  iodoacetamide (400 mM) was added, and the samples were incubated in a darkened water bath for 15 minutes at  $37^\circ\text{C}$  (alkylation). For protein digestion,  $10 \mu\text{l}$  trypsin (trypsin/protein ratio of 1:40) was added, and samples were incubated in a water bath for 16 hours at  $37^\circ\text{C}$ . Digestion was stopped by addition of  $20 \mu\text{l}$  formic acid (10%, v/v). All samples were stored at  $-80^\circ\text{C}$  until further processing. Finally,  $35 \mu\text{l}$  of the digested membrane fraction was mixed with  $35 \mu\text{l}$  isotope-labeled internal standard (IS) peptide mix ( $10 \text{ nM}$  of each IS; Thermo Fisher Scientific). All sample preparation and digestion steps were performed using Protein LoBind tubes (Eppendorf, Hamburg, Germany). Protein quantification was conducted on a 5500

QTRAP triple quadrupole mass spectrometer (AB Sciex) coupled to an Agilent Technologies 1260 Infinity system (Agilent Technologies) using validated LC-MS/MS methods as recently described (Drozdik et al., 2019). Transporter proteins and the respective proteospecific peptides and the stable isotope-labeled internal standard peptides considered in our analysis are given in Supplementary Table 3. Protein abundance of human OCT1 was determined by using three peptides, whereas mouse OCT1 and  $\text{Na}^+/\text{K}^+$ -ATPase were determined by using one peptide. For each peptide, two to three mass transitions have been monitored.

**IVIVE to Estimate the Liver Partition Coefficient of Metformin.** The uptake of metformin across the sinusoidal membrane into the liver was estimated based on in vitro uptake measurements in stably transfected HEK293 cells overexpressing human or mouse OCT1. In vitro clearance ( $CL_{in \text{ vitro}}$ ) was calculated as follows:

$$CL_{in \text{ vitro}} = \frac{v_{max}}{K_M},$$

where  $v_{max}$  is the maximum transport rate ( $\text{picomole} \times \text{minute}^{-1} \times \text{milligram protein}^{-1}$ ) and  $K_M$  is the Michaelis constant ( $\text{micromolar}$ ) determined in HEK293 cells. The obtained  $CL_{in \text{ vitro}}$  ( $\text{microliter} \times \text{minute}^{-1} \times \text{milligram protein}^{-1}$ ) was used for extrapolation toward total human or mouse liver clearance, which was mediated by active transport via human or mouse OCT1, respectively.

The active OCT1-mediated uptake into the liver ( $CL_{in,act}$ ) was calculated as follows:

$$CL_{in,act} = CL_{in \text{ vitro}} \times \frac{E_{in \text{ vivo}}}{E_{in \text{ vitro}}} \times LW \times \text{total protein per unit LW},$$

where  $E$  refers to the total OCT1 expression in human or mouse liver ( $E_{in \text{ vivo}}$ ) and in HEK293 cells overexpressing human or mouse OCT1 ( $E_{in \text{ vitro}}$ ) ( $\text{picomole} \times \text{milligram protein}^{-1}$ ). In this case, we assumed that 100% of the OCT1 protein is localized in the plasma membrane both in the liver and in HEK293 cells.  $LW$  refers to the liver weight in human and mouse (gram), respectively. The total protein amount per unit  $LW$  is given as ( $\text{milligram protein} \times \text{gram liver}^{-1}$ ) and was obtained from Sohlenius-Sternbeck (2006).

Passive diffusion into the liver [ $CL_{diff}$  ( $\text{microliter} \times \text{minute}^{-1} \times \text{milligram protein}^{-1}$ )] was estimated by using HEK293 cells transfected with the empty pcDNA5/FRT vector and was calculated as follows:

$$CL_{diff} = CL_{in \text{ vitro}} \times LW \times \text{total protein per unit LW}$$

The liver partition coefficient ( $K_p$ ) was calculated based on the extended clearance concept according to Guo et al. (2018):

$$K_p = \frac{CL_{in,act} + CL_{in, diff}}{CL_{ef,act} + CL_{ef,diff} + CL_{bile} + CL_{met}}$$

The following simplifications were made based on the findings that metformin is neither metabolized nor significantly excreted by transporters or secreted into the bile (Pentikäinen et al., 1979; Tucker et al., 1981): the clearances for transporter-mediated efflux ( $CL_{ef,act}$ ), for biliary excretion of unchanged drug ( $CL_{bile}$ ), and for metabolism ( $CL_{met}$ ) were set to zero. Furthermore, we assumed that the passive influx diffusion permeation ( $CL_{in, diff}$ ) is equal to the passive efflux diffusion permeation ( $CL_{ef, diff}$ ), here further designated simply as  $CL_{diff}$ .  $K_p$  was predicted using the following equation (Yabe et al., 2011; Shitara et al., 2013):

$$K_p = \frac{CL_{in,act} + CL_{diff}}{CL_{diff}}$$

As metformin had been shown to have negligible protein binding, the fraction unbound in plasma can be assumed as 1 (Tucker et al., 1981). The prediction of the liver partition coefficient for unbound drug concentration thereby was assumed as

$$K_{p,u} = K_p$$

**Computational Modeling.** Available templates for structural modeling were identified by using fold recognition methods offered by the pGenThreader server (available at <http://bioinf.cs.ucl.ac.uk/psipred/>) (Lobley et al., 2009). The human glucose 3 transporter (GLUT3; Protein Data Bank identifier (PDB ID) 4zw9) was selected as an optimal template for both human and mouse OCT1 modeling (Deng et al., 2015). This crystal structure of GLUT3 was selected based on multiple criteria, such as being resolved in high resolution (1.5 Å) and adopting an outward-occluded conformation, which is particularly useful for the investigation of substrate binding. Sequence-to-structure alignment between GLUT3 and human and mouse OCT1 sequence, respectively, was initially generated in PROMALS3D and subsequently revised and corrected by manual intervention (Pei et al., 2008). The large extracellular loop between TMH1 and TMH2 (88 residues) and one C-terminal intracellular loop (22 residues) were lacking structural templates and were therefore omitted for structural modeling purposes.

In total, 100 structural models were generated for both human and mouse OCT1 using Modeler 9.17 (Eswar et al., 2006). Energy minimization was performed to optimize the orientation of side chains. AMBER99SB-ILDN force field (Lindorff-Larsen et al., 2010) and GROMACS version 5.1.4 (Abraham et al., 2015) were used for steepest descent minimization. The convergence criterion was set to a maximum force <100.0 kJ/mol per nanometer. The final models for human and mouse OCT1 were selected on the basis of the MolProbity score ranking (<http://molprobity.biochem.duke.edu/>) and a proper orientation of the Asp474/475 residue, which is the main residue known to be implicated in ligand binding. Ramachandran outliers were visually inspected in Molecular Operating Environment 19 (Chemical Computing Group ULC, Montreal, QC, Canada). In silico models generated for human OCT1 and mouse OCT1 are available as supplement to this publication (Supplemental Files hOCT1.pdb and mOCT1.pdb).

Two independent algorithms—FTSite (Ngan et al., 2012) and SiteFinder in Molecular Operating Environment 19—were used to identify possible interaction sites in human and mouse OCT1 transporters. The FTSite program docks 16 small probe molecules to identify hot spots in the protein structure. Probe molecules are clustered, and the poses are ranked according to the empirical free energy function. The SiteFinder tool in MOE utilizes the alpha sphere method in which the protein cavities are explored by virtual spheres generated in the site. Every sphere can also differentiate with respect to potential hydrophobic or hydrophilic contacts. Predicted binding sites are ranked according to the number of alpha spheres located in every detected binding site. Standard settings of SiteFinder were applied. Hydrophobic interactions were analyzed by a helical wheel projection using DrawCoil 1.0 (available at <https://grigoryanlab.org/drawcoil/>).

**Data Analyses.** Kinetic parameters of metformin and thiamine transport ( $K_M$  and  $v_{max}$ ) were determined by nonlinear regression to the Michaelis-Menten equation using GraphPad Prism version 5.01 (GraphPad Software Inc., La Jolla, CA). The kinetic parameters or uptake values were compared between human and mouse OCT1, human-mouse chimeric OCT1, or mutant OCT1 using ANOVA followed by Tukey's honestly significant difference post hoc comparisons in SPSS Statistics version 25 (SPSS Inc., IBM, Chicago, IL).

## Results

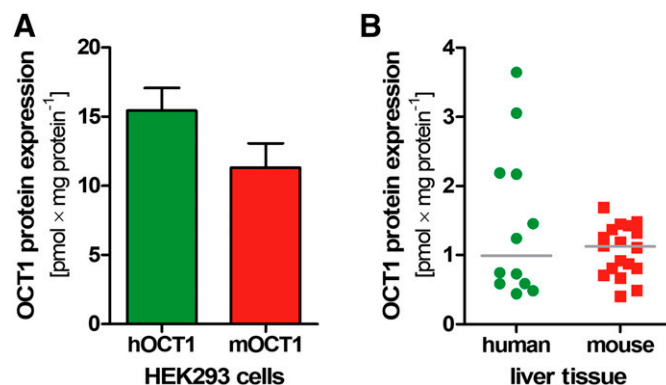
**Characterization of the Model System.** In this study, we used HEK293 cells stably transfected to overexpress human and mouse OCT1 by targeted chromosomal integration. As a first step, we characterized our model system with respect to the levels of OCT1 protein expression. We used targeted proteomics to quantify OCT1 expression in the stably transfected HEK293 cells and to compare it with OCT1 expression in human and mouse liver (Fig. 1). In HEK293 cells, OCT1 expression was 36% higher in the cells stably overexpressing human compared with the cells stably overexpressing mouse OCT1 (Fig. 1A). The native OCT1 expression in human and mouse liver was similar but more than 10-fold lower than in the

model cell lines (16-fold for human OCT1, 11-fold for mouse OCT1, Fig. 1B). We also compared the OCT1 expression in three mouse strains: C57BL/6 substrains J and N and FVB. C57BL/6J mice showed more than 30% higher expression than the other two strains (Supplemental Fig. 1).

**Differences in the Kinetics of Metformin Uptake between Human and Mouse OCT1.** More importantly, we compared the uptake of metformin between human and mouse OCT1. To this end, we performed concentration-dependent uptake measurements in stably transfected HEK293 cells. The maximal transport rates ( $v_{max}$ ) were 45% lower in human than in mouse OCT1 ( $v_{max}$  of 939 and 1353 pmol  $\times$  min<sup>-1</sup>  $\times$  pmol OCT1<sup>-1</sup>, respectively). The differences in the apparent affinity ( $K_M$ ) for metformin were much stronger (Fig. 2A). Mouse OCT1 showed a 4.9-fold higher affinity for metformin than human OCT1 ( $K_M$  of 491 and 2197  $\mu$ M, respectively,  $P < 0.0001$ ). This resulted in a 6.5-fold higher intrinsic clearance of mouse compared with human OCT1 (2.86 and 0.50  $\mu$ l  $\times$  min<sup>-1</sup>  $\times$  pmol OCT1<sup>-1</sup>, respectively). The strong differences in metformin kinetics were confirmed when using a transient transfection model (Supplemental Fig. 2; Supplementary Table 4). In transiently transfected HEK293 cells, we observed an 8.1-fold higher affinity of mouse OCT1 compared with human OCT1, resulting in a 12.5-fold difference in intrinsic clearance. Also, time-dependent analyses showed substantially higher uptake by mouse than by human OCT1 (Fig. 2B). The difference was strongest in the beginning of the incubation period (5.7-fold at 1 minute) but remained above 60% even after 30 minutes of incubation. There were no indications for different modes of transport between human and mouse OCT1 (Fig. 2C). After incubating with clinically relevant concentrations of 10  $\mu$ M metformin (Shu et al., 2007), we observed 4.4-fold higher intracellular concentrations in HEK293 cells overexpressing mouse than in those overexpressing human OCT1 (99.6 and 22.7  $\mu$ M, respectively, Fig. 2D).

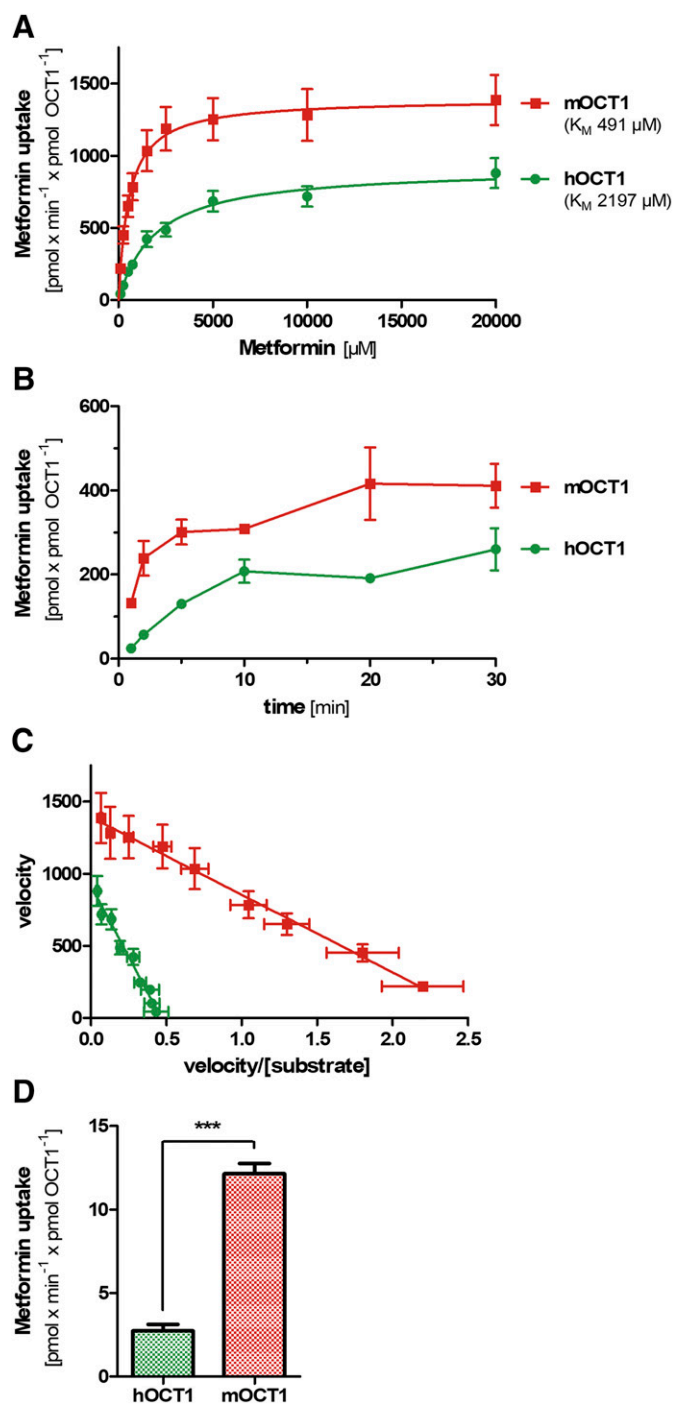
We also determined the transport kinetics of rat OCT1 (Supplemental Fig. 3). With a  $K_M$  of 422  $\mu$ M and intrinsic clearance of 30.5  $\mu$ l  $\times$  min<sup>-1</sup>  $\times$  mg protein<sup>-1</sup>, the rat ortholog did not differ significantly from mouse OCT1 ( $K_M$  of 491  $\mu$ M and intrinsic clearance of 37  $\mu$ l  $\times$  min<sup>-1</sup>  $\times$  mg protein<sup>-1</sup>) but differed strongly compared with human OCT1 ( $K_M$  of 2197  $\mu$ M and intrinsic clearance of 7.85  $\mu$ l  $\times$  min<sup>-1</sup>  $\times$  mg protein<sup>-1</sup>).

**Estimation of the Differences in Partition Coefficients of Metformin between Human and Mouse Livers Using IVIVE.** Assuming that OCT1 is the major determinant of metformin levels in the liver (Wang et al., 2002; Shu et al., 2007), it could be expected that



**Fig. 1.** OCT1 protein expression in (A) stably transfected HEK293 cells and (B) human and mouse liver. OCT1 expression in the membrane fraction of (A) HEK293 cells stably overexpressing human (green) or mouse (red) OCT1 or (B) human and mouse liver samples was measured by targeted LC-MS/MS. Please consider the difference in scaling of the y-axis. Shown are (A) means and S.E.M. of nine samples each and (B) concentrations of single samples (12 human and 20 mouse livers) and respective medians.





**Fig. 2.** Differences in metformin uptake between human and mouse OCT1. (A) Concentration-dependent uptake and (B) time-dependent uptake of metformin by human (green) and mouse (red) OCT1. OCT1-overexpressing HEK293 cells were incubated with (A) increasing concentrations of metformin for 2 minutes or (B) with 100  $\mu\text{M}$  metformin for up to 30 minutes. The uptake values were normalized to the amount of OCT1 protein in the respective HEK293 cells, as determined by targeted proteomics (see Fig. 1A). (C) Eadie-Hofstee transformation of the data in (A). (D) Intracellular metformin concentrations in HEK293 cells stably transfected with human or mouse OCT1 after incubation with 10  $\mu\text{M}$  metformin for 2 minutes. The intracellular concentrations were calculated assuming an intracellular volume of 1.2  $\mu\text{l}$  for  $1 \times 10^6$  HEK293 cells, following the estimations of Chien et al. (2016). All subfigures represent OCT1-mediated uptake that was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. Shown are means and S.E.M. of at least three independent experiments. \*\*\* $P < 0.001$  in a one-way ANOVA.

substantial differences in OCT1 clearance between human and mouse will result in substantial differences in the exposure to metformin in human and mouse liver. We used an IVIVE approach to estimate the  $K_p$  both in human and in mouse. Taking into account the differences in OCT1 expression (Fig. 1), we estimated the liver-to-blood  $K_p$  in human to be 3.34 compared with 14.4 in mouse (Table 2). Considering the estimated portal vein concentration of metformin in humans (Shu et al., 2007; Gormsen et al., 2016) and in mice (Wilcock and Bailey, 1994), we could expect 11-fold higher maximal intrahepatic concentrations in mice than in humans (66.9  $\mu\text{M}$  in human and 746  $\mu\text{M}$  in mouse, Table 2).

We compared the predicted values for mouse with the experimentally measured values (Wilcock and Bailey, 1994). The predicted  $K_p$  for mouse liver was almost 2-fold higher than the experimentally measured one (Table 2). In line with this, the predicted hepatic concentration in mouse was 98% higher. This suggests that our model overestimates the  $K_p$  in mice and that factors other than OCT1-mediated uptake may play a role.

To the best of our knowledge, there is no experimental data on hepatic concentrations of metformin in humans. However, using  $^{11}\text{C}$ -labeled metformin in PET analyses, Gormsen et al. (2016) estimated a hepatic  $K_p$  of 2.5 in humans, which is 34% lower than our estimation (Table 2) and supports the differences in hepatic metformin concentrations in vivo between human and mouse that were suggested by the IVIVE model.

**Identification of the Structural Causes for the Differences in Metformin Kinetics.** Next, we looked for structural differences between human and mouse OCT1 that confer the differences in their affinity for metformin. To this end, we generated chimeric constructs of human and mouse OCT1 (hmoOCT1 and mmhOCT1) and characterized their metformin uptake. We separated the protein into three parts: from N terminus to the large intracellular loop, from TMH7 to TMH9, and from TMH10 to C terminus (Fig. 3A). Concentration-dependent measurements pointed to the first six TMHs of OCT1 to confer the differences in affinity for metformin between human and mouse OCT1 (Fig. 3, B and C). Immunofluorescence staining demonstrated the correct membrane localization of the wild types and chimeras and suggested a reduced total expression of the mmhOCT1 chimera (Fig. 3D) that correlates with its reduced  $v_{\text{max}}$  (Fig. 3B).

To further narrow down the region within the first six TMHs conferring these differences, we generated chimeric constructs with single TMH substitutions between human and mouse OCT1. Uptake experiments at single concentrations pointed to TMH2 and TMH3 as being primarily involved. Substituting TMH2 or TMH3 in human OCT1 with TMH2 or TMH3 of mouse OCT1 resulted in the only significant increase of metformin uptake (Fig. 4A). In line with this, substituting TMH2 or TMH3 in mouse OCT1 with TMH2 or TMH3 of human OCT1 resulted in the strongest decrease of metformin uptake (by 77% and 55%, respectively; Fig. 4B). A single concentration of 100  $\mu\text{M}$  was chosen to optimally reflect the difference in the  $K_M$  based on the data from the wild-type constructs (Fig. 2). However, the effects (especially reduction of the uptake) may also be caused by a general reduction of activity (as observed for the mmhOCT1 chimera, Fig. 3B). To exclude this, we performed concentration-dependent measurements for TMH2- and TMH3-containing chimeras. We observed a strong increase in metformin affinity upon introduction of either mouse TMH2 or mouse TMH3 into human OCT1 (3-fold lower  $K_M$  compared with human OCT1, Fig. 4C). Vice versa, introduction of either human TMH2 or human TMH3 into mouse OCT1 did not significantly change affinity. However, simultaneous introduction of human TMH2 and TMH3 into mouse OCT1 resulted in a significantly decreased affinity (14-fold higher  $K_M$  compared with mouse OCT1, Fig. 4C), and the concentration-dependent uptake almost completely mimicked the uptake of human OCT1 (Fig. 4D). These experiments clearly

TABLE 2

Parameter of OCT1-mediated metformin pharmacokinetics in humans and mice measured experimentally or extrapolated using IVIVE

Parameter	Mouse					Human				
	Mean	n	S.D.	95% CI		Mean	n	S.D.	95% CI	
Maximal velocity, $v_{\max}$ ( $\text{pmol} \times \text{min}^{-1} \times \text{mg protein}^{-1}$ )	17,496	11	7097	12,727	22,265	14,703	11	4346	11,783	17,623
Affinity for metformin uptake, $K_M$ ( $\mu\text{M}$ )	491	11	155	387	595	2198	11	1154	1422	2973
Metformin in vitro clearance, $CL_{\text{in vitro}}$ ( $\mu\text{l} \times \text{min}^{-1} \times \text{mg protein}^{-1}$ )	37	11	16.1	26.2	47.9	7.85	11	3.9	5.23	10.5
Metformin passive diffusion, $CL_{\text{diff}}$ ( $\mu\text{l} \times \text{min}^{-1} \times \text{mg protein}^{-1}$ ) <sup>a</sup>	0.34	11	0.20	0.21	0.48	0.34	11	0.20	0.21	0.48
OCT1 expression in liver, $E_{\text{in vivo}}$ ( $\text{pmol} \times \text{mg protein}^{-1}$ )	1.27	20	0.72	0.93	1.61	1.44	12	1.09	0.75	2.13
OCT1 expression in vitro, $E_{\text{in vitro}}$ ( $\text{pmol} \times \text{mg protein}^{-1}$ )	11.3	9	5.23	7.3	15.3	15.4	9	4.92	11.7	19.2
Predicted metformin liver-to-blood partition coefficient, $K_{p,u}$ <sup>b</sup>	14.4	11	4.93	11.1	17.7	3.34	11	0.98	2.68	4.0
Predicted maximal hepatic metformin concentrations ( $\mu\text{M}$ ) <sup>c</sup>	746	11	254	575	659	66.9	11	19.6	53.7	80.0
Observed metformin $K_{p,u}$ <sup>d</sup>	6.8					2.5				
Observed maximal hepatic metformin concentrations ( $\mu\text{M}$ ) <sup>d</sup>	350									

S.D., standard deviation; n, number of independent measurements; CI, confidence interval.

<sup>a</sup>Passive diffusion was estimated based on the uptake in control HEK293 cells transfected with the empty vector pcDNA5 (Supplemental Fig. 2A). Therefore, the values do not differ between mouse and human.<sup>b</sup> $K_{p,u}$  was calculated as described in *Materials and Methods*. Liver weights used for the calculations were 1500 g for human and 2.5 g for mouse (Rogers and Dintzis, 2018). The total amount of protein was 90 and 115 mg  $\times$  g liver<sup>-1</sup> for human and mouse liver, respectively (Sohlenius-Sternbeck, 2006).<sup>c</sup>Hepatic concentrations were calculated assuming portal vein concentrations of 51.7  $\mu\text{M}$  for mouse (Wilcock and Bailey, 1994) and 20  $\mu\text{M}$  for human [double the  $C_{\max}$  observed in humans after 1 g of metformin (Shu et al., 2007; Gormsen et al., 2016)].<sup>d</sup>The experimental data of mouse  $k_p$  and hepatic concentrations were obtained from Wilcock and Bailey (1994) 30 min after an oral dose of 50 mg/kg metformin. The concentrations were calculated assuming 2.5 g average weight and 1.3 ml average volume of mouse liver. The human  $k_p$  was obtained from Gormsen et al. (2016). No experimental data of human liver concentrations were available.

identify TMH2 and TMH3 to confer the differences between human and mouse OCT1 in their affinity for metformin.

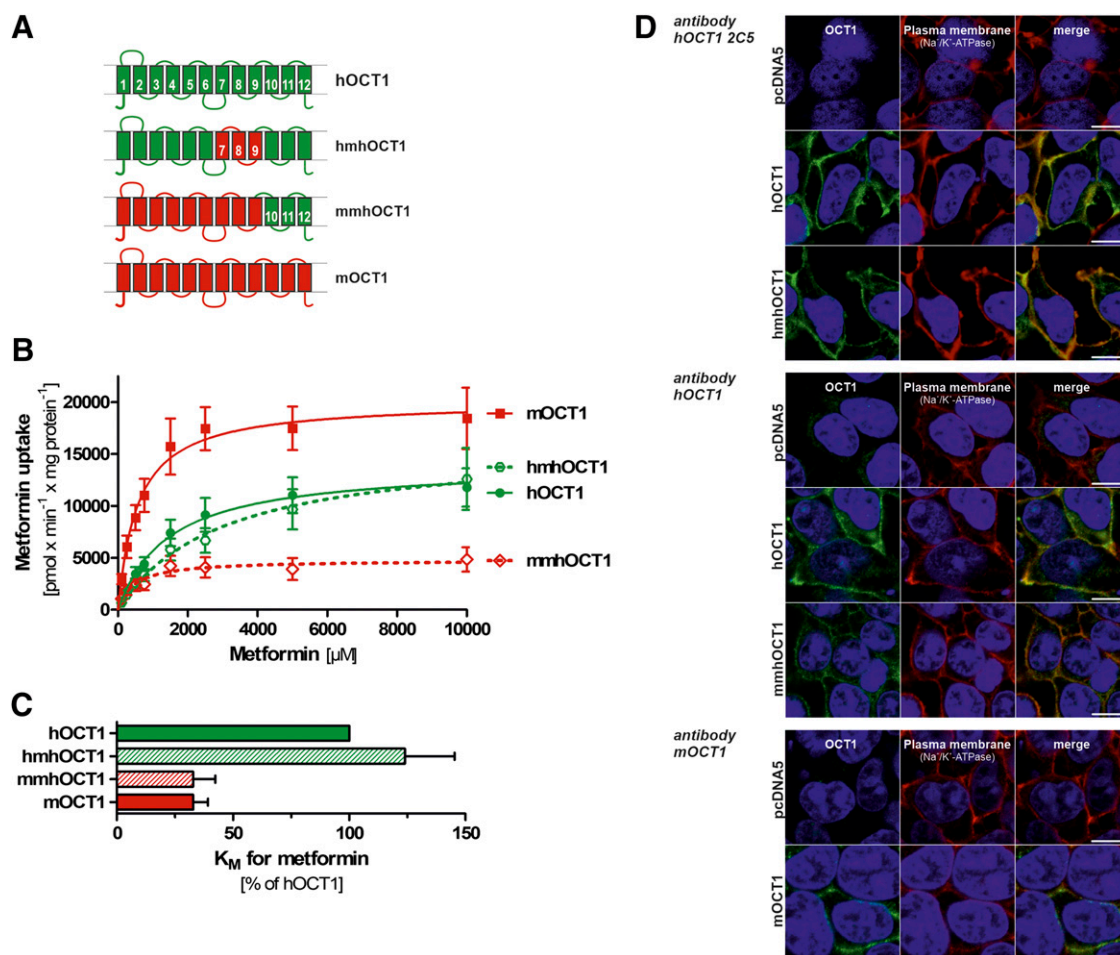
Next, we generated homology models of human and of mouse OCT1 to identify single amino acids within TMH2 and TMH3 that may confer the differences in metformin uptake. There were no major differences in the tertiary structure between human and mouse OCT1 as visible by superposition of the two models (Fig. 5A). Two major binding cavities with the involvement of TMH2 or TMH3 were identified using two distinct algorithms. One of the proposed binding cavities is located in the middle of the translocation pore and is a highly populated site with probe molecules (143 alpha spheres for human OCT1 and 120 alpha spheres for mouse OCT1). This “classical” binding site has been reported in several previous studies (Chen et al., 2017; Boxberger et al., 2018; Gorboulev et al., 2018). The binding site is enframed by TMH1, TMH4, TMH5, TMH7, TMH8, TMH9, TMH11, and (more importantly) TMH2. None of the five nonconserved amino acids in TMH2 (Fig. 5B; Supplemental Fig. 4) could be suggested to be directly involved in substrate binding. However, our structural models show that Leu155 in human (hLeu155) that corresponds to Val156 in mouse OCT1 (mVal156) in TMH2 can form a hydrophobic core packing with Ile35 located in TMH1 (Fig. 6, A and B) that may have an impact on tertiary structure stability (the mouse OCT1 protein is longer than the human one, resulting in a one-count shift in amino acid position after number 84). The stronger hydrophobic interaction between Leu155 and Ile35 in human OCT1 (compared with Val156 and Ile35 in mouse OCT1) might aggravate the entrance of substrates and conformational changes in this region of the transporter. These observations might explain a generally lower affinity for metformin uptake in human OCT1 compared with mouse OCT1.

Indeed, simultaneous introduction of mouse TMH3 and mutation of Leu155Val increased the affinity of human OCT1 by more than 70% (of the difference between human and mouse OCT1, Fig. 6C), and vice versa, simultaneous introduction of human TMH3 and mutation of Val156Leu decreased metformin affinity in mouse OCT1 by 55% (Fig. 6, C and D). Without the simultaneous introduction of TMH3, the mutation of Leu155Val in human showed only limited effects, and Val156Leu in mouse OCT1 showed no significant effects. This points to the importance of the interaction with TMH3 for the effects of hLeu155/mVal156.

An alternative explanation involving both TMH2 and TMH3 may be provided by the second predicted binding site. This binding cavity

(although with a lower score, as indicated by 51 alpha spheres for human OCT1 and 25 alpha spheres for mouse OCT1; Fig. 7, A and B, left panel) is a membrane-exposed pocket with involvement of residues from both TMH2 and TMH3. Interestingly, this “outer” cavity is framed by two nonconserved residues—one from each TMH—lying just opposite of each other. Whereas in mouse OCT1, these residues are valines (Val166 and Val182), in human OCT1, these residues are glycines (Gly165 and Gly181; Fig. 7, A and B). The glycines in human OCT1 could lead to a higher conformational flexibility of the helices in that region, whereas the valines in mouse OCT1 introduce hydrophobicity and potentially stronger interactions with ligands at this position. Indeed, mutation of valines 166 and 182 in mouse OCT1 to glycines (Val166Gly, Val182Gly), either alone or in combination, significantly decreased metformin uptake (Fig. 7C). However, the decrease was maximally 26%, and the reverse mutation of glycines in human OCT1 to valines (Gly165Val, Gly181Val) neither alone nor in combination affected metformin uptake. This suggests that the glycine-to-valine differences at positions 165/166 and 181/182 alone cannot explain the differences in metformin uptake between human and mouse OCT1.

As an alternative approach, we took advantage of the observation that the affinity of human OCT2 for metformin is rather similar to the affinity of mouse OCT1 than to the affinity of human OCT1 (Supplemental Fig. 5A). We mutated amino acids that are identical in mouse OCT1 and human OCT2 but are different in human OCT1 (Supplemental Fig. 5B) and analyzed the effects on metformin uptake. In addition to hLeu155/mVal156, which we already analyzed (Fig. 6C), this affected hPhe169/mIle170 in TMH2. Interestingly, this variation is the only difference between human and mouse OCT1 within the conserved A-motif (Fig. 5B), which is suggested to be important for both structure and function of MFS transporters by interacting with residues from surrounding TMHs and supporting conformational changes during the transport cycle (Henderson and Maiden, 1990; Pao et al., 1998; Quistgaard et al., 2016). Mutation of Ile170 in mouse OCT1 to the corresponding amino acid in human OCT1 (Ile170Phe) decreased metformin uptake by 28% ( $P = 4 \times 10^{-4}$ , Supplemental Fig. 5C). However, mutation of both Val156Leu and Ile170Phe in mouse OCT1 did not lead to a stronger decrease in metformin uptake than mutation of Val156Leu alone (43%, Supplemental Fig. 5C). Mutation of these amino acids in human OCT1 (Leu155Val and Phe169Ile) neither alone nor in combination had an effect on metformin uptake (Supplemental Fig. 5C). Thus, hPhe169/mIle170 neither alone nor



**Fig. 3.** Metformin uptake in human-mouse chimeric OCT1. (A) Schematic representation of human and mouse wild-type and human-mouse chimeric OCT1 constructs with numbering of the individual TMHs. Colors indicate the origin of the TMHs of either human (green) or mouse (red) OCT1. (B) Concentration-dependent uptake of metformin by human and mouse wild-type and human-mouse chimeric OCT1. OCT1-overexpressing HEK293 cells were incubated with increasing concentrations of metformin for 2 minutes. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. (C) Affinity for metformin ( $K_M$ ) of human and mouse wild-type OCT1 compared with human-mouse chimeric OCT1. Represented are  $K_M$  values of the data shown in (B) as percentage of human OCT1. Shown are means and S.E.M. of at least three independent experiments. (D) Membrane localization of OCT1 as assessed by immunofluorescence staining. To enable staining of human-mouse chimeric OCT1, two different antibodies against human OCT1 were used, binding in the large intracellular loop (2C5, top panel) or in the C terminus (middle panel). The antibody against mouse OCT1 binds in the C terminus and could therefore not be used for staining chimeric OCT1. Cells were costained with an antibody against Na<sup>+</sup>/K<sup>+</sup>-ATPase as a marker for the plasma membrane. Cell nuclei were stained with DAPI. Scale bar, 10 μm.

in combination with hLeu155/mVal156 can explain more than 30% of the observed differences in metformin affinity between the species. In addition, this suggests that independent mechanisms confer the higher affinity for metformin in mouse OCT1 and in human OCT2.

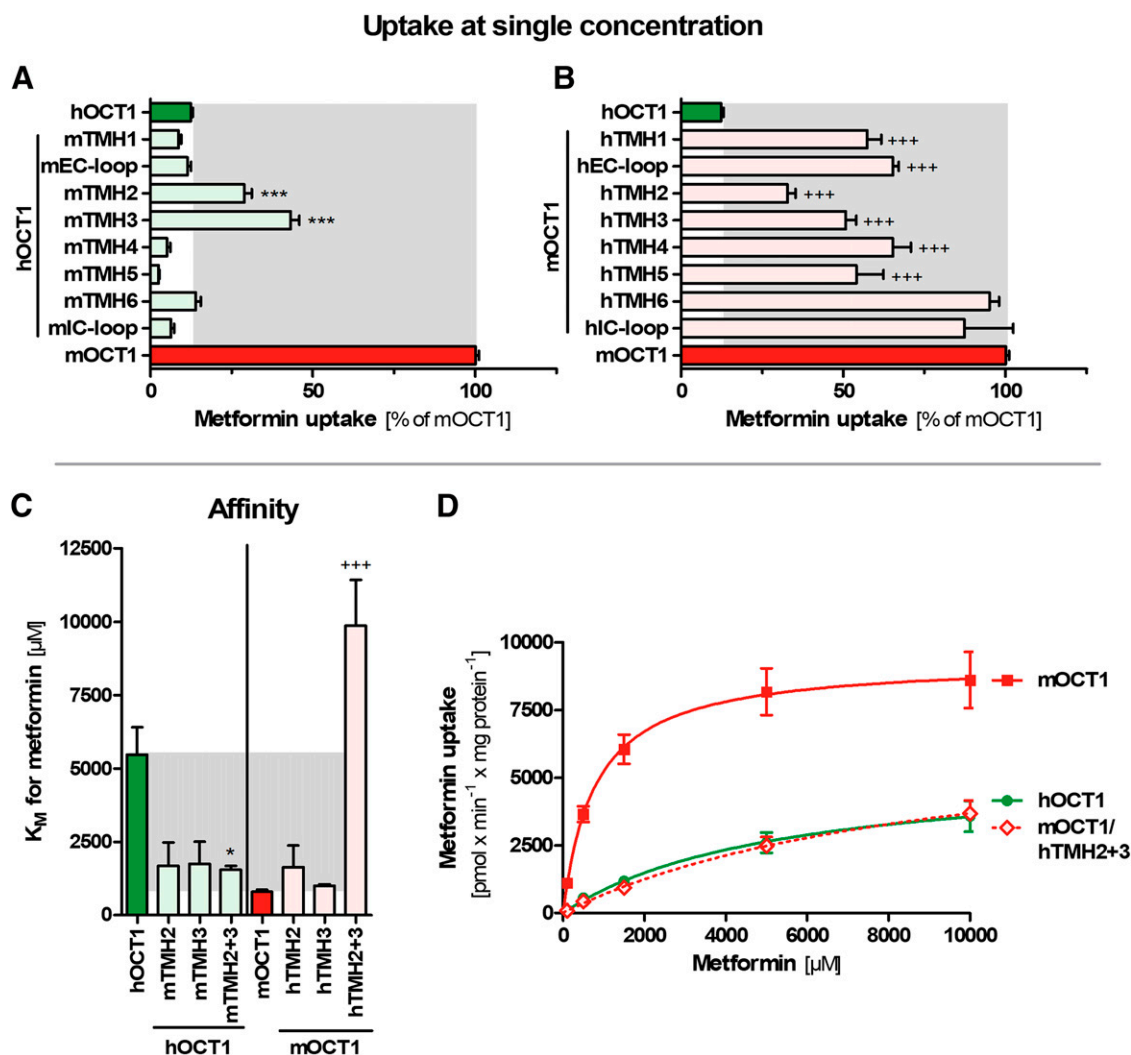
**Differences in the Kinetics of Thiamine Uptake between Human and Mouse OCT1.** Similar to metformin, we observed strong differences in the affinity for thiamine between human and mouse OCT1 (Fig. 8). Mouse OCT1 had a 9.5-fold higher apparent affinity for thiamine than human OCT1 ( $K_M$  of 143 and 1057 μM, respectively; Fig. 8A). In contrast to metformin, the lower affinity for thiamine of human OCT1 resulted in 80% higher maximal transport rates ( $v_{max}$  of 528 and 287 pmol × min<sup>-1</sup> × pmol OCT1<sup>-1</sup> for human and mouse OCT1, respectively). Nevertheless, this resulted in a 5.1-fold higher intrinsic clearance of thiamine by mouse OCT1 compared with human OCT1 (Fig. 8C).

Concentration-dependent measurements using human-mouse chimeric OCT1 also pointed to the first six TMHs of OCT1 to confer the differences in affinity for thiamine between human and mouse OCT1 (Fig. 9, A and B). Considering the observed key role of TMH2 and TMH3 in the transport of metformin, we analyzed thiamine uptake by human-mouse chimeric OCT1 carrying the simultaneous substitution of

both helices. Similar to metformin, simultaneous introduction of mouse TMH2 and TMH3 into human OCT1 resulted in a significant increase of affinity for thiamine ( $K_M$  of 456 μM compared with 1517 μM of human OCT1, Fig. 9, C and D). However, in contrast to metformin, introduction of human TMH2 and TMH3 into mouse OCT1 did not result in a significant decrease of affinity (Fig. 9C). Therefore, it could be concluded that TMH2 and TMH3 are also involved in the mechanisms conferring differences in thiamine uptake between human and mouse OCT1, but the mechanisms are not identical to the ones for metformin.

## Discussion

In this study, we report strong differences between human and mouse OCT1 in the transport of metformin and thiamine. The most pronounced difference was the substantially higher apparent affinity of mouse compared with human OCT1, which results in a much higher intrinsic uptake clearance. This was observed for both metformin and thiamine. As a consequence, higher concentrations of metformin may be reached in mouse than in human liver. Furthermore, differences in the affinity for metformin between human and mouse OCT1 could be attributed to



**Fig. 4.** Identification of TMH2 and TMH3 as major determinants of the differences in metformin uptake between human and mouse OCT1. (A and B) Human-mouse chimeric constructs with single TMH substitutions of each of the first six TMHs, the large extracellular (EC), or the large intracellular (IC) loop (human background, light green; mouse background, light red). Metformin uptake was measured at single concentrations of 100  $\mu M$  metformin and related to the uptake by human (green) and mouse (red) wild-type OCT1. (C) Effects on metformin affinity ( $K_M$ ) after substituting TMH2 and TMH3 alone or in combination between human and mouse OCT1. (D) Concentration-dependent metformin uptake of human (green) and mouse (red) wild-type OCT1 and mouse OCT1 with TMH2 and TMH3 of human OCT1 (red dotted line). In all cases, HEK293 cells transiently overexpressing OCT1 were incubated with metformin for 2 minutes. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. Shown are means and S.E.M. of at least three independent experiments. \* $P < 0.05$ ; \*\*\* $P < 0.001$  compared with (\*) human or (†) mouse OCT1 in a Tukey's post hoc analysis following one-way ANOVA.

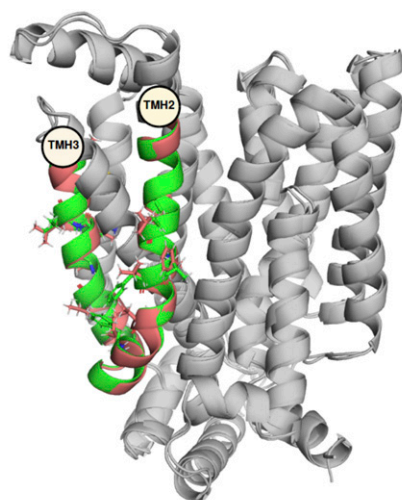
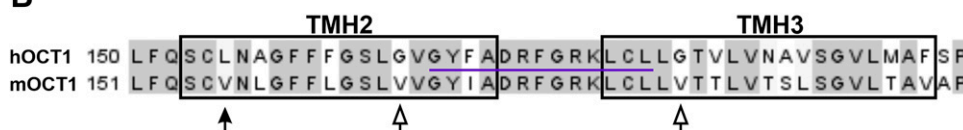
differences in TMH2 and 3 of the transporter, revealing new insights into the transport mechanism of metformin by OCT1.

We observed a 4.9-fold higher affinity for metformin by mouse than by human OCT1 and a 45% higher transport capacity (Fig. 2). The affinity of human OCT1 for metformin observed here ( $K_M$  of 2197  $\mu M$ , Fig. 2) is similar to previous reports (Shu et al., 2007; Umehara et al., 2007; Nies et al., 2009). Despite mouse being a commonly used model organism to study metformin pharmacokinetics and effects, to the best of our knowledge, this is the first study reporting metformin uptake kinetics via mouse OCT1. Moreover, we characterized metformin uptake by human and mouse OCT1 in parallel. Therefore, the obtained data are highly comparable, especially since we used a model system that is well characterized in terms of protein expression, enabling us to normalize uptake data to the amount of OCT1 protein. There are no previous reports on metformin uptake kinetics in mouse hepatocytes. However, comparison of human and rat hepatocytes showed a 27-fold higher metformin clearance in rats than in humans (Umehara et al., 2007), and

we observed highly similar uptake kinetics between rat and mouse OCT1 (Supplemental Fig. 3).

Based on our in vitro data on OCT1 affinity and on the differences in the portal vein concentrations, metformin concentrations can be expected to be about 11-fold higher in mouse than in human liver (Table 2). This could result in differences in the hepatic actions of metformin between human and mouse. Low metformin concentrations were suggested to activate AMPK (Zhou et al., 2001) and to suppress gluconeogenic gene expression and glucose production (Cao et al., 2014), whereas high metformin concentrations inhibit mitochondrial complex I (El-Mir et al., 2000) and lead to an AMPK-independent suppression of gluconeogenesis (Foretz et al., 2010). Especially as OCT1-overexpression has recently been shown to substantially increase the mitochondrial accumulation of the drug (Chien et al., 2016), mitochondrial effects of metformin may be more likely in mouse than in human liver. In general, our data warrants attention when using mouse data to extrapolate the hepatic effects of metformin in humans.



**A****B**

**Fig. 5.** Structural differences between human and mouse OCT1 with a focus on TMH2 and TMH3. (A) Superposition of human and mouse OCT1 structural models with TMH2 and TMH3 highlighted in green (human OCT1) and red (mouse OCT1). (B) Protein sequence alignment of human and mouse OCT1 using EMBOSS Needle (Madeira et al., 2019) with TMH2 and TMH3 highlighted and the conserved A-motif of MFS transporters underlined in violet. Coloring is based on amino acid identity. Arrows indicate the positions of amino acids hLeu155/mVal156 (closed arrow) and hGly165/mVal166 and hGly181/mGly182 (open arrows), which were of particular interest.

Because of the better uptake by mouse OCT1, the liver-mediated effects may be more pronounced in mice than in humans.

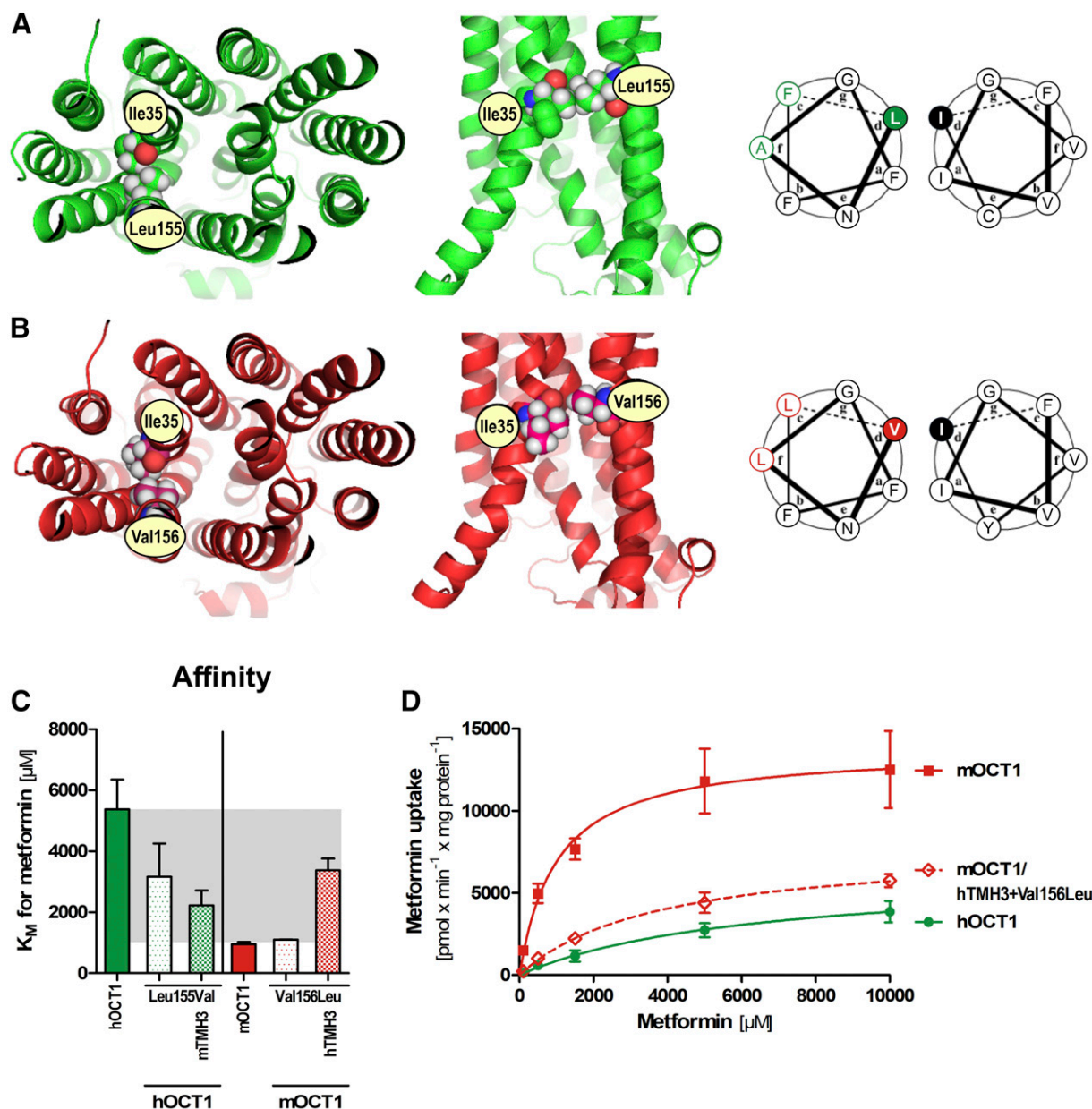
The effects of OCT1 deficiency are more pronounced in mice than in humans (Wang et al., 2002, 2003; Shu et al., 2007; Zhou et al., 2009; Dujic et al., 2017; Sundelin et al., 2017). A 30-fold decrease in hepatic metformin concentrations was shown in OCT1 knockout mice (Wang et al., 2002). Precise measurements in humans are difficult, but a study using PET imaging showed about 2-fold lower hepatic metformin concentrations in carriers of loss-of-function OCT1 variants (Sundelin et al., 2017). These numbers generally fit to the tendency observed here that OCT1-mediated uptake is about 14-fold higher compared with diffusion in mice and only 3-fold higher compared with diffusion in humans (Table 2). One explanation, supported by our data, is that because of the different efficacy of human and mouse OCT1 in transporting metformin, knockout in mice and loss-of-function genetic variants in humans do not have comparable effects on hepatic metformin concentrations. However, the fact that some of the human OCT1 genetic variants do not lead to a complete loss of metformin uptake should also be considered (Kerb et al., 2002; Shu et al., 2003; Seitz et al., 2015).

Our IVIVE calculations based on the uptake kinetics of mouse OCT1 overestimated the hepatic  $K_p$  and hepatic concentrations of metformin experimentally measured in mice (Table 2). The most probable reason for this is that the major reflection of the higher uptake clearance via mouse OCT1 is observed in the first minutes of the uptake (Fig. 2B), and the available experimental data are obtained after 30 minutes or more (Wilcock and Bailey, 1994). In the longer incubation, other factors, like reaching steady state of intracellular versus extracellular metformin concentrations, may play a role. Therefore, short-term differences in the concentrations of metformin between human and mouse liver may be more pronounced than the long-term differences. Nevertheless, accounting for the PET-based estimation of  $K_p$  of 2.5 in humans and a maximal portal vein concentration of 20  $\mu\text{M}$  (Shu et al., 2007; Gormsen et al., 2016), an intrahepatic concentration of about 50  $\mu\text{M}$  could be estimated for the human liver. This is 7-fold lower than the intrahepatic concentrations of metformin measured in mice (Wilcock and Bailey, 1994).

The involvement of alternative transporters like OCT3 that are not reflected in our IVIVE calculations is less probable. In the mouse liver,

OCT1 has much higher expression levels than OCT3 [OCT1-to-OCT3 mRNA ratio of about 30 (Chen et al., 2015)]. Consistently, up to 30-fold lower hepatic metformin concentrations were measured in OCT1 knockout mice (Wang et al., 2002), but there were no significant changes in OCT3 knockout mice (Chen et al., 2015; Lee et al., 2018). Also, in the human liver, OCT1 is expressed much more strongly than OCT3. The ratio of OCT1 to OCT3 in human liver is 22 based on protein quantification (Drozdziak et al., 2019) and 32 based on mRNA quantification (Nies et al., 2009). The intrinsic clearance of metformin can thus be estimated to be at least 11-fold lower by OCT3 than by OCT1, indicating that OCT1 is the predominant uptake transporter of metformin both in the mouse and in the human liver.

Similar to metformin, the affinity for thiamine was much higher by mouse than by human OCT1 (9.5-fold lower  $K_M$ , Fig. 8), which is supported by a previous study (Chen et al., 2014). Also, clear effects of OCT1 deficiency on thiamine levels were reported in mice (Chen et al., 2014; Liang et al., 2018) but not in humans (Jensen et al., 2020). One explanation may be that in humans, at “physiological” low concentrations, thiamine is predominantly transported by thiamine transporters THTR-1 and THTR-2, which have a substantially higher affinity ( $>1600$ -fold lower  $K_M$ ) but also a substantially lower capacity ( $>130$ -fold lower  $v_{\text{max}}$ ) than OCT1 (Jensen et al., 2020). In contrast, in mice, the differences in affinity and capacity of thiamine uptake between OCT1 and THTR-1 are much smaller (2.9-fold lower  $K_M$  and 7.6-fold lower  $v_{\text{max}}$ ), and mouse OCT1 showed more than 5-fold higher thiamine uptake than mouse THTR-1 at low concentrations [100 nM; (Chen et al., 2014)]. This, together with the much higher affinity of mouse compared with human OCT1, suggests that OCT1 may play a more important role in thiamine uptake at low concentrations in mice than in humans and thereby may contribute to the different effects of OCT1 deficiency on thiamine plasma levels between these species. Alternatively, the strong renal OCT1 expression in mouse but not in human (Gorboulev et al., 1997; Zhang et al., 1997; Green et al., 1999; Schmitt et al., 2003) may contribute to the different effects of OCT1 deficiency on systemic thiamine concentrations in the two species. However, compensatory effects of OCT2, which is strongly expressed in the kidney, are probable (Chen et al., 2014).

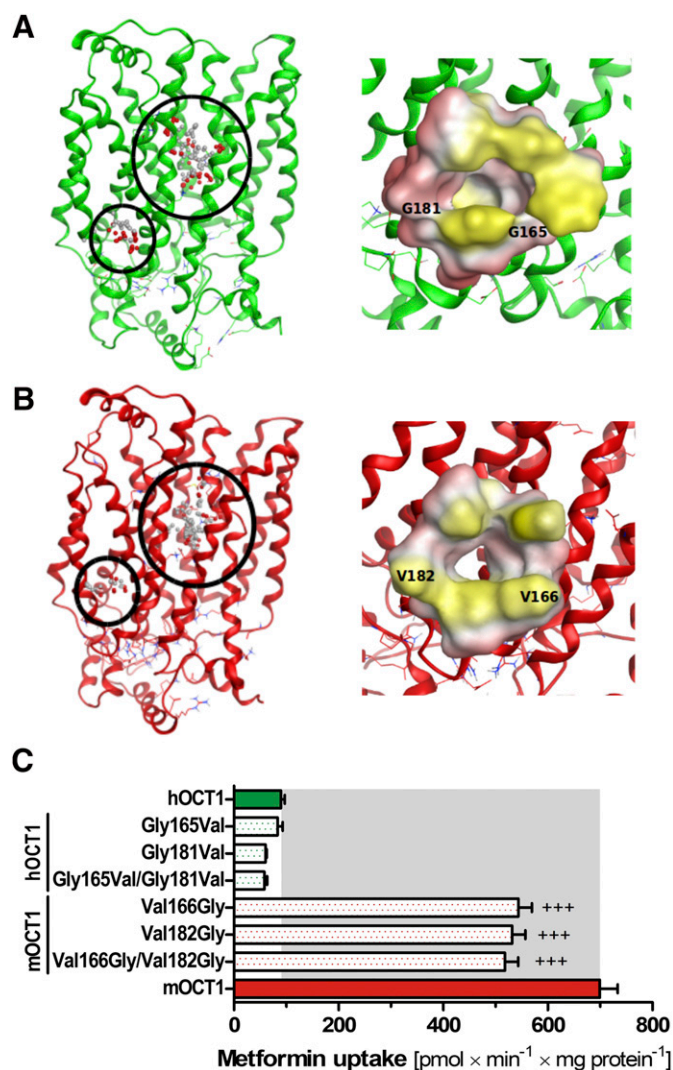


**Fig. 6.** Potential involvement of hLeu155/mVal156 in TMH2 of human/mouse OCT1 in conferring the differences in metformin affinity. Hydrophobic interactions between hLeu155/mVal156 and Ile35 in (A) human and (B) mouse OCT1 in (left panel) top view and (middle panel) side view. (A and B, right panel) Helical wheel projection of TMH2 and TMH1 showing the positioning of hLeu155/mVal156 (TMH2) and Ile35 (TMH1) in the viewing “d” of the helical wheels, respectively. Nonconserved amino acids are highlighted in color, and Ile35 is highlighted in black. (C) Effects on metformin affinity ( $K_M$ ) after simultaneous substitution of Leu155Val and mouse TMH3 in human OCT1 and Val156Leu and human TMH3 in mouse OCT1. (D) Concentration-dependent metformin uptake of human (green) and mouse (red) wild-type OCT1 and mouse OCT1 with Val156Leu mutation and human TMH3 (red dotted line). In all cases, HEK293 cells transiently overexpressing OCT1 were incubated with metformin for 2 minutes. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. Shown are means and S.E.M. of four independent experiments. \* $P < 0.05$ , \*\*\*/\* $P < 0.001$  compared with (\*) human or (+) mouse OCT1 in a Tukey’s post hoc analysis following one-way ANOVA.

Transmembrane helices TMH2 and TMH3 were experimentally identified to confer the differences in metformin affinity between human and mouse OCT1. Three independent hypotheses were generated to identify the single amino acids responsible: 1) differences in the hydrophobic interaction of hLeu155/mVal156 (TMH2) with Ile35 (TMH1), 2) higher flexibility by Gly165 and Gly181 in human as opposed to higher hydrophobicity and ligand interaction by Val166 and Val182 in mouse OCT1, and 3) similar affinities of mouse OCT1 and human OCT2 caused by Val156 and/or Ile170 that differ in human OCT1 (Ile170 being located within the conserved

A-motif of the MFS transporters). Experimentally, the strongest effects were observed by mutating Val156 to Leu together with exchanging TMH3 (Fig. 6), supporting the first hypothesis the most. The mechanisms may be expected to be similar for rat OCT1, since the potentially involved amino acids are identical between rat and mouse OCT1 (Supplemental Fig. 3D).

Another interesting observation is that the substitution of a single TMH (either 2 or 3) is sufficient to increase the affinity of human OCT1, but both human TMHs are needed to decrease the affinity of mouse OCT1 (Fig. 4C). This may suggest that a decreased affinity requires an



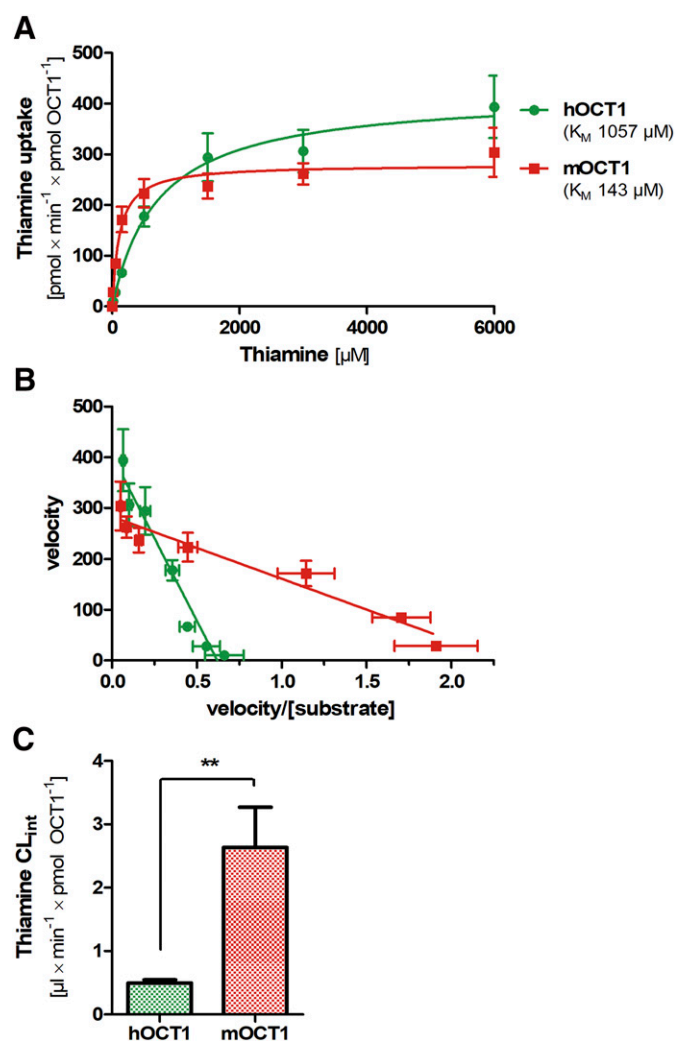
**Fig. 7.** Potential involvement of TMH2 and TMH3 in conferring the differences in metformin affinity between human and mouse OCT1. (A and B, left panel) Two predicted binding cavities within the TMH2-TMH3 region in (A) human and (B) mouse OCT1 structural models. (A and B, right panel) Top view of the “inner” binding cavity with protein surface colored according to lipophilicity (red, hydrophilic; yellow, hydrophobic; white, neutral surface) with (A) glycine residues 165 and 181 in human OCT1 and (B) valine residues 166 and 182 in mouse OCT1 highlighted. (C) Effect of mutations of Gly165Val and Gly181Val in human and Val166Gly and Val182Gly in mouse OCT1 on metformin uptake. HEK293 cells transiently overexpressing OCT1 were incubated with 100  $\mu\text{M}$  metformin for 2 minutes. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. Shown are means and S.E.M. of two to four independent experiments performed in duplicates.  $+++P < 0.001$  compared with mouse OCT1 in a Tukey’s post hoc analysis following one-way ANOVA.

interaction between the two TMHs and removing one of the TMHs is enough to destroy this interaction.

From the structural perspective, the “knob-into-hole” motif of the hydrophobic interaction between hLeu155/mVal156 in TMH2 with Ile35 in TMH1 (Fig. 6) is somewhat analogous to coiled-coil structures (Liu et al., 2006). Interestingly, hLeu155/mVal156 are located at position “d” when depicting the helix as a helical wheel projection (Fig. 6). Since this position is suggested to be more vulnerable to amino acid substitution (Zhu et al., 1993), hLeu155/mVal156 may have a huge impact on tertiary structure stability. The stronger hydrophobic interaction between Leu155 and Ile35 in human OCT1 (compared with Val156 and Ile35 in mouse OCT1) might obstruct substrate entry and conformational

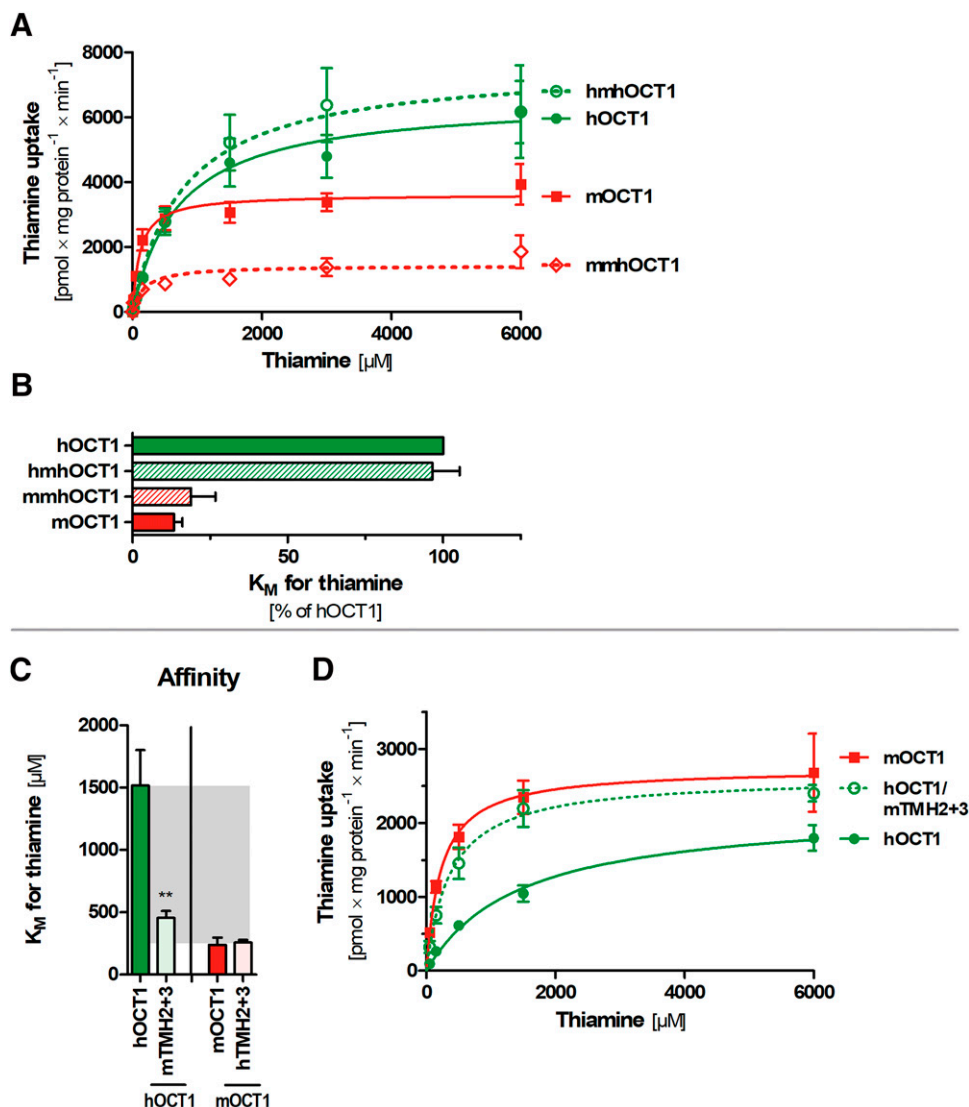
changes in this region, thereby potentially explaining the lower affinity for metformin.

Another aspect to be considered is the substrate-specific effects of OCT1. Recently, Morse et al. (2020) reported substantial differences in the uptake kinetics between human and mouse primary hepatocytes for ondansetron and tropisetron but not for sumatriptan and fenoterol. In our study, the differences in the affinity between human and mouse OCT1 were comparable between metformin and thiamine (Figs. 2 and 8). This is consistent with previous reports suggesting similar binding sites of metformin and thiamine in OCT1 (Chen et al., 2014). However, our data show that the affinities for these two compounds are conferred by similar, but not identical, structures in OCT1 (Figs. 4 and 9). This underlines the polyspecificity of OCT1 and points out that structure-to-function relations of OCT1 need to be established separately for each substrate and, based on the present results, also for each species.



**Fig. 8.** Differences in thiamine uptake between human and mouse OCT1. (A) Concentration-dependent uptake of thiamine by human (green) and mouse (red) OCT1. OCT1-overexpressing HEK293 cells were incubated with increasing concentrations of thiamine for 2 minutes. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. Background thiamine levels were subtracted from all values to exclude influence of endogenous thiamine on the measurement. The uptake values were normalized to the amount of OCT1 protein in the respective HEK293 cells, as determined by targeted proteomics (see Fig. 1A). (B) Eadie-Hofstee transformation of the data in (A). (C) Comparison of the intrinsic clearance ( $\text{CL}_{\text{int}}$ ) between human and mouse OCT1 calculated using  $v_{\text{max}}$  and  $K_{\text{M}}$  of the data in (A). Shown are means and S.E.M. of at least three independent experiments.  $**P < 0.01$  in a one-way ANOVA.





**Fig. 9.** Thiamine uptake in human-mouse chimeric OCT1 constructs. (A) Concentration-dependent uptake of thiamine by human and mouse wild-type and human-mouse chimeric OCT1. For detailed description of chimeric constructs see figure 3A. (B) Affinity for thiamine ( $K_M$ ) of human and mouse wild-type compared with human-mouse chimeric OCT1. Represented are  $K_M$  values of the data shown in (A) as percentage of human OCT1. (C) Affinity for thiamine ( $K_M$ ) of human and mouse wild-type OCT1 compared with human-mouse chimeric OCT1 carrying the simultaneous substitution of TMH2 and TMH3. (D) Concentration-dependent thiamine uptake of human (green) and mouse (red) wild-type OCT1 and human OCT1 with TMH2 and TMH3 of mouse OCT1 (green dotted line). HEK293 cells (A and B) stably or (C and D) transiently overexpressing OCT1 were incubated with increasing concentrations of thiamine for 2 minutes. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. Background thiamine levels were subtracted from all values to exclude the influence of endogenous thiamine on the measurement. Shown are means and S.E.M. of at least three independent experiments.  $**P < 0.01$  in a Tukey's post hoc analysis following one-way ANOVA.

In conclusion, mouse OCT1 has a much higher affinity for metformin and thiamine than human OCT1. This may be an important factor contributing to the substantially higher metformin concentrations measured in the mouse than in the human liver in vivo and should be considered when interpreting findings about the hepatic mechanism of action of metformin that are obtained in mouse models. The determinants of the differences in metformin affinity between human and mouse OCT1 are clearly located in TMH2 and TMH3 and comprise hLeu155/mVal156 (TMH2) and amino acid(s) in TMH3. The underlying mechanism is probably complex, and the identification of the precise amino acids in TMH3 and additional protein structures in that region involved needs further investigation.

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#### Authorship Contributions

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Conducted experiments: Meyer, Tuerkova, Wenzel.

Contributed new reagents or analytic tools: Tuerkova, Römer, Seitz, Gaedcke, Zdravil.

Performed data analysis: Meyer, Tuerkova, Römer, Wenzel, Oswald, Zdravil, Tzvetkov.

Wrote or contributed to the writing of the manuscript: Meyer, Tuerkova, Römer, Oswald, Brockmöller, Zdravil, Tzvetkov.

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## **Supplemental Material**

# **Differences in metformin and thiamine uptake between human and mouse organic cation transporter OCT1: structural determinants and potential consequences for intrahepatic concentrations**

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## Supplementary Tables

Supplementary Table S1. Primers used for the generation of OCT1 constructs

Primer	Sequence (5' → 3')	TMH/amino acid substitution
m/rOCT1_F	GTTGGTGAGGAAGCTTACCCAGCCATGCCACCGTGGA	
human1_for	CGTTTAAACTTAAGCTTGCATGCT	TMH1
hm_1_rev	CCAGGACTCTGGCAGTGGTGGTC	
hm_EC_for	GACCACCACTGCCAGAGTCCTGG	
hm_EC_rev	GGACTGAAAGAGGTCCAGCTTCCAGG	EC-loop
hm_2_for	CCTGGAAGCTGGACCTCTTTCAGTCC	
hm_2_rev	AGCTTACGGCCAAACCTGTCTGCAA	TMH2
hm_3_for	TTGCAGACAGGTTTGGCCGTAAGCT	
hm_3_rev	TGCAGCAGGCGGAAGAGCAGCATGGA	TMH3
hm_4_for	TCCATGCTGCTCTTCCGCCTGCTGCA	
hm_4_rev	CGTTCTTCTGGAGCCCGAGCC	TMH4
hm_5_for	GGCTCGGGCTCCAGAAGAACG	
hm_5_rev	GCCAGCTGCAGCCAGCGCCAGT	TMH5
hm_6_for	ACTGGCGCTGGCTGCAGCTGGC	
hm_6_rev	AACAGCCACCGAGGGGA	TMH6
hm_IC_for	TCCCCTCGGTGGCTGTT	
hm_IC_rev	CAGGATGAAGGTGCGCTTCCTCAG	IC-loop
L155V_hfor	GACCTCTTTCAGTCCTGT <b>GT</b> GAAATGCGGGCTTCTTCTTT	Leu155Val in hOCT1
L155V_hrev	AAAGAAGAAGCCCGCATT <b>CAC</b> ACAGGACTGAAAGAGGTC	
V156L_mfor	GACCTTTTTTCAGTCCTGT <b>TT</b> GAACTTGGGCTTCTTCCTG	Val156Leu in mOCT1
V156L_mrev	CAGGAAGAAGCCCAAGTT <b>CAA</b> ACAGGACTGAAAAAGGTC	
G165V_hfor	TTCTTCTTTGGCTCTCT <b>CGTT</b> GTTGGCTACTTTGCAGAC	Gly165Val in hOCT1
G165V_hrev	GTCTGCAAAGTAGCCA <b>ACA</b> ACGAGAGAGCCAAAGAAGAA	
G181V_hfor	CGTAAGCTGTGTCTCCTG <b>GTG</b> ACTGTGCTGGTCAACGCG	Gly181Val in hOCT1
G181V_hrev	CGCGTTGACCAGCACAGT <b>CACC</b> AGGAGACACAGCTTACG	
V166G_mfor	TTCTTCTTGGGCTCCCTG <b>GGT</b> GTTGGGTTACATTGCAGAC	Val166Gly in mOCT1
V166G_mfor	GTCTGCAATGTAACCCAC <b>ACCC</b> AGGGAGCCCAGGAAGAA	
V182G_mfor	CGTAAGCTCTGCCTCCTG <b>GGG</b> AACCACTCTGGTCACCTC	Val182Gly in mOCT1
V182G_mrev	GAGGTGACCAGAGTGGT <b>TCCC</b> CAGGAGGCAGAGCTTACG	

Affected codons are underlined, changed bases are boldfaced; TMH, transmembrane helix; EC-loop, large extracellular loop; IC-loop, large intracellular loop; hOCT1, human OCT1; mOCT1, mouse OCT1



**Supplementary Table S2. Antibodies and dilutions used for immunocytochemical staining of OCT1-overexpressing cells**

	Antibody	Source	Dilution	Applied to cell line			
				hOCT1	hnh	mmh	mOCT1
Primary antibody	Monoclonal mouse anti-hOCT1 2C5 (NBP1-51684)	Novus Biologicals, Abingdon, UK	1:400	X	X	-	-
	Monoclonal rabbit anti-hOCT1 (Orb32029)	Biorbyt Ltd. Cambridge, UK	1:50	X	-	X	-
	Polyclonal rabbit anti-mOCT1	Hermann Koepsell, University of Würzburg (Meyer-Wentrup et al., 1998)	1:50	-	-	-	X
	Monoclonal rabbit anti-Na <sup>+</sup> /K <sup>+</sup> -ATPase (EP1845Y)	Abcam, Cambridge, UK	1:200	X	X	-	-
	Monoclonal mouse anti-Na <sup>+</sup> /K <sup>+</sup> -ATPase (sc-48345)	Santa Cruz Biotechnology, Heidelberg, Germany	1:100	X	-	X	X
Secondary antibody	Alexa Fluor® 488 goat anti - mouse IgG (H+L), polyclonal	Thermo Fisher Scientific, Darmstadt, Germany	1:400	X	X	-	-
			1:200	X	-	X	X
	Alexa Fluor® 546 goat anti - rabbit IgG (H+L), polyclonal	Thermo Fisher Scientific, Darmstadt, Germany	1:400	X	X	-	-
			1:100	X	-	X	X

**Supplementary Table S3. Proteospecific peptides for quantitative LC-MS/MS analyses of OCT1 protein expression**

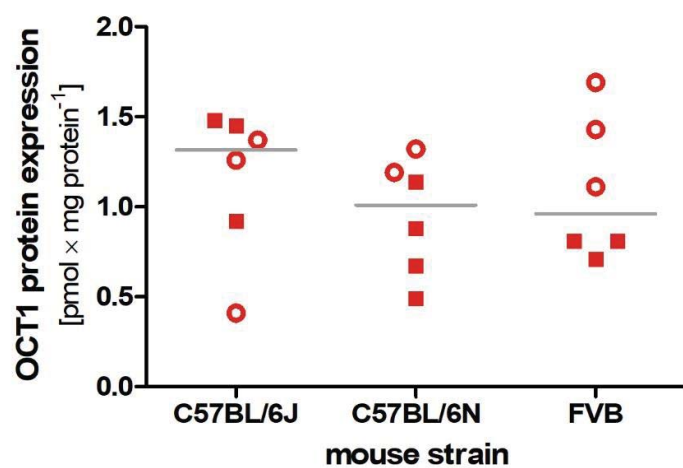
Protein	Peptide	Q1	Q3.1	Q3.2	Q3.3
Human OCT1	ENTIYLK	441	423.5	536.5	-
	ENTIYLK*	445.2	431.5	544.6	-
Human OCT1	LPPADLK	377.3	543.5	211.2	-
	LPPADLK*	381.4	551.5	211.2	-
Human OCT1	DAENLGR	388.1	459.3	588.4	-
	DAENLGR*	393.4	469.3	598.4	-
Mouse OCT1	GVALPETIEEAENLGR	850.1	680	228.2	341.2
	GVALPETIEEAENLGR*	855.2	684.7	228.2	341.3
Na <sup>+</sup> /K <sup>+</sup> -	LSLDELHR	328.	435.2	391.7	669.3
ATPase	LSLDELHR*	331.5	440.3	396.8	679.4

\*stable isotope-labeled internal standard peptide

Supplementary Table S4. Comparison of the parameters of transport kinetics between stably and transiently transfected HEK293 cells

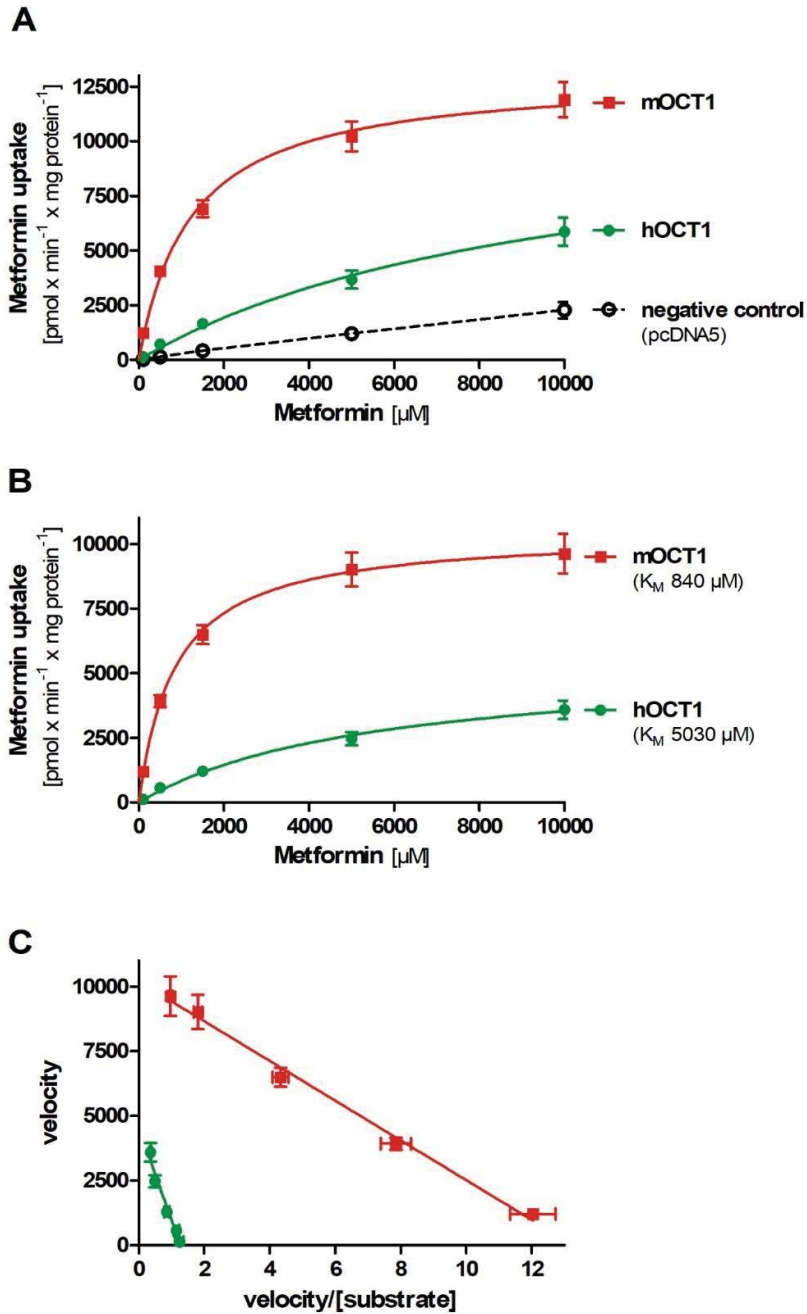
Parameter	Mouse					Human				
	Mean	n	SD	95% CI		Mean	n	SD	95% CI	
Metformin (stably transfected cells)										
Affinity for metformin uptake, $K_M$ [ $\mu M$ ]	491	11	155	387	595	2197	11	1154	1422	2973
Maximal velocity, $v_{max}$ [ $pmol \times min^{-1} \times mg \text{ protein}^{-1}$ ]	17496	11	7097	12727	22265	14703	11	4346	11783	17623
Maximal velocity, $v_{max}$ [ $pmol \times min^{-1} \times pmol \text{ OCT1}^{-1}$ ]	1353	11	549	985	1722	939	11	278	753	1126
Metformin $CL_{in vitro}$ [ $\mu L \times min^{-1} \times mg \text{ protein}^{-1}$ ]	37	11	16.1	26.2	47.9	7.85	11	3.9	5.23	10.5
Metformin (transiently transfected cells)										
Affinity for metformin uptake, $K_M$ [ $\mu M$ ]	840	14	225	710	970	5030	14	2101	3817	6243
Maximal velocity, $v_{max}$ [ $pmol \times min^{-1} \times mg \text{ protein}^{-1}$ ]	10187	14	3109	8392	11982	5169	14	2203	3897	6441
Metformin $CL_{in vitro}$ [ $\mu L \times min^{-1} \times mg \text{ protein}^{-1}$ ]	12.4	14	2.69	10.8	13.9	1.09	14	0.36	0.88	1.3
Thiamine (stably transfected cells)										
Affinity for thiamine uptake, $K_M$ [ $\mu M$ ]	143	5	96.3	22.9	262	1057	5	341	634	1480
Maximal velocity, $v_{max}$ [ $pmol \times min^{-1} \times mg \text{ protein}^{-1}$ ]	3712	5	1031	2432	4992	8261	5	3720	3642	12880
Maximal velocity, $v_{max}$ [ $pmol \times min^{-1} \times pmol \text{ OCT1}^{-1}$ ]	287	5	79.7	188	386	528	5	238	233	823
Thiamine $CL_{in vitro}$ [ $\mu L \times min^{-1} \times mg \text{ protein}^{-1}$ ]	34.1	5	18.3	11.3	56.8	7.71	5	1.75	5.54	9.88
Thiamine (transiently transfected cells)										
Affinity for thiamine uptake, $K_M$ [ $\mu M$ ]	238	3	103	-19	495	1517	3	491	298	2736
Maximal velocity, $v_{max}$ [ $pmol \times min^{-1} \times mg \text{ protein}^{-1}$ ]	2772	3	823	728	4816	2226	3	404	1222	3230
Thiamine $CL_{in vitro}$ [ $\mu L \times min^{-1} \times mg \text{ protein}^{-1}$ ]	12.1	3	1.56	8.20	15.9	1.54	3	0.37	0.62	2.45

## Supplementary Figures

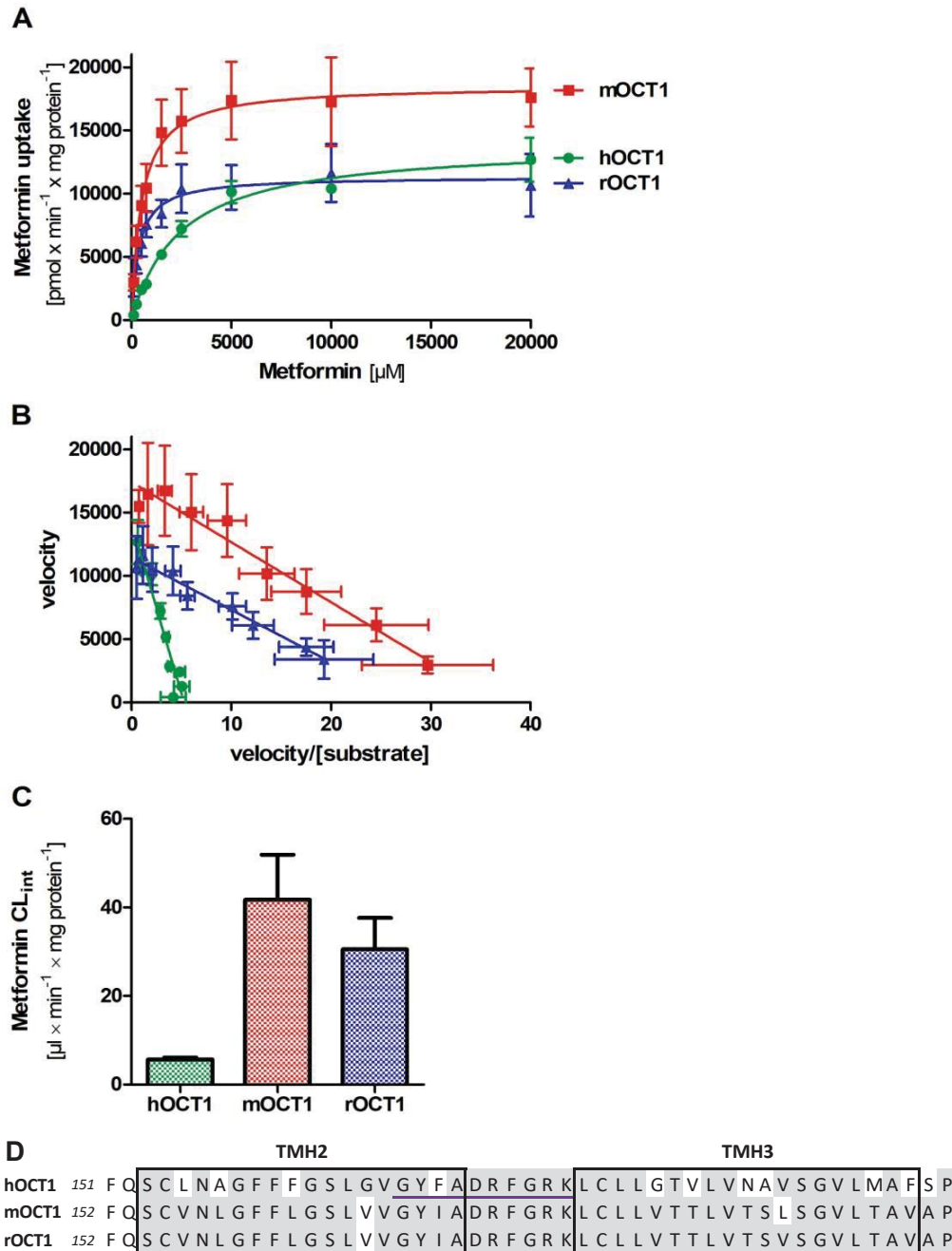


**Supplementary Figure S1. Comparison of OCT1 protein expression between different mouse strains** OCT1 protein expression in the membrane fraction of mouse liver samples was measured using targeted proteomics. Boxes represent male, open circles represent female animals. Shown are the medians of six animals per mouse strain.

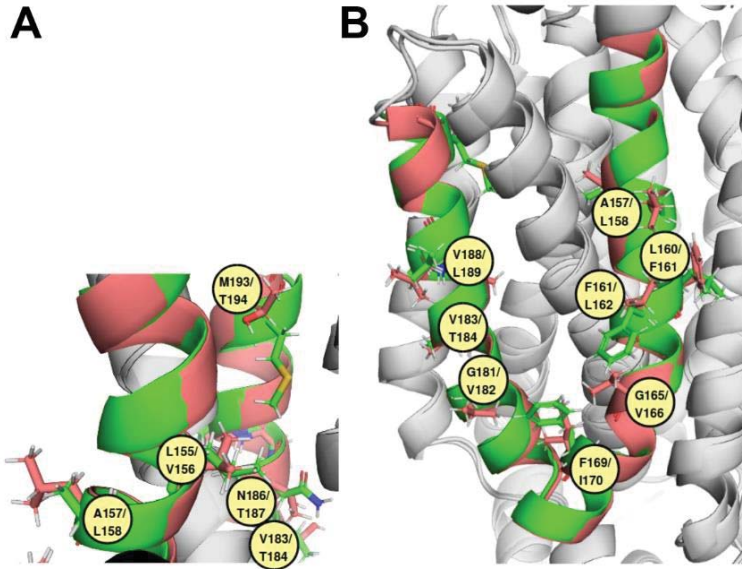




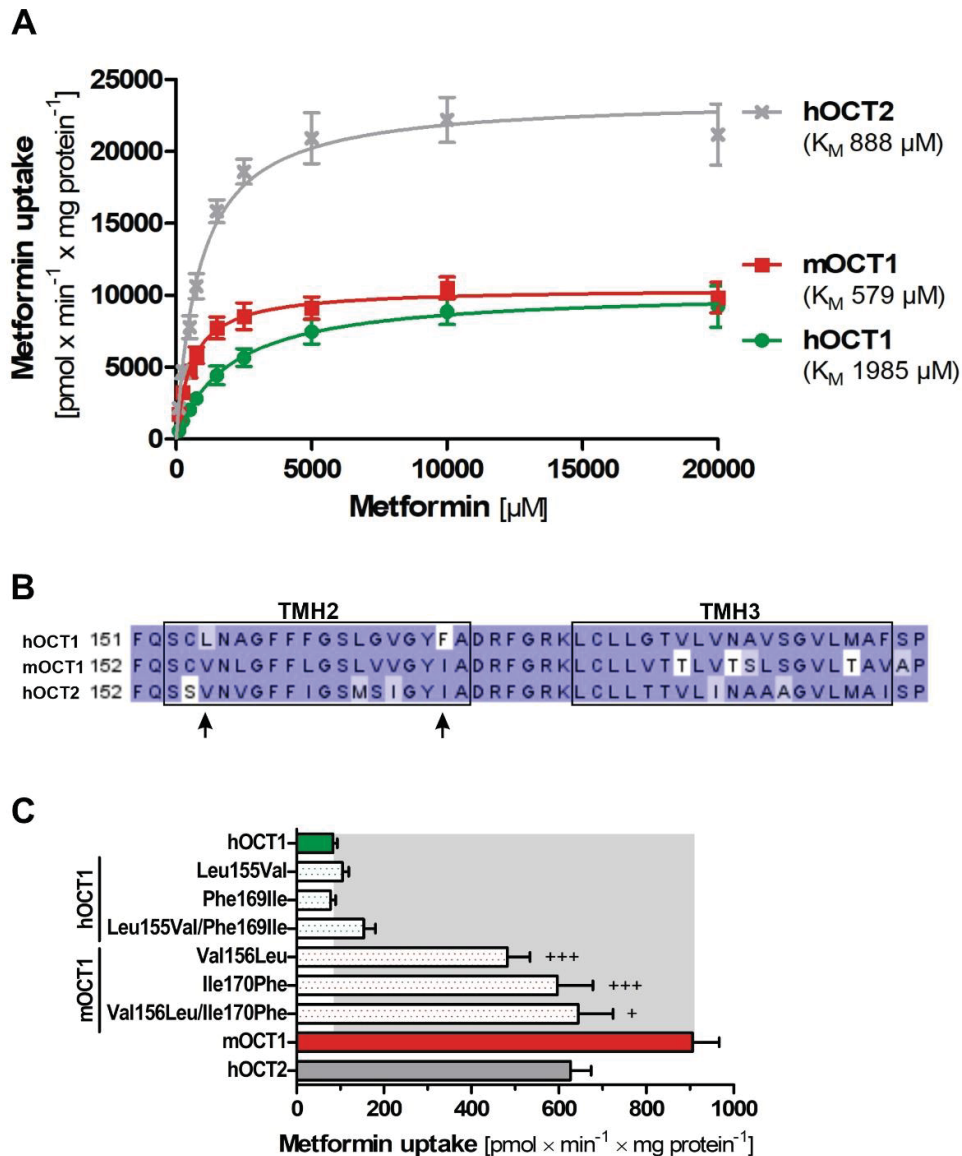
**Supplementary Figure S2. Differences in metformin uptake between human and mouse OCT1 using transiently transfected HEK293 cells** (A) Concentration-dependent uptake of metformin by human OCT1 (green), mouse OCT1 (red) and the negative control pcDNA5 (empty vector). (B) Concentration-dependent uptake of metformin by human and mouse OCT1. HEK293 cells transiently overexpressing OCT1 were incubated with increasing concentrations of metformin for 2 min. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. (C) Eadie-Hofstee transformation of the data in B. Shown are means and standard errors of the means of 14 independent experiments.



**Supplementary Figure S3. Differences in metformin uptake between human, mouse, and rat OCT1** (A) Concentration-dependent uptake of metformin by human (green), mouse (red), and rat (blue) OCT1. OCT1-overexpressing HEK293 cells were incubated with increasing concentrations of metformin for 2 min. OCT1-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing OCT1. (B) Eadie-Hofstee transformation of the data in A. (C) Comparison of the intrinsic clearance ( $CL_{int}$ ) between human, mouse, and rat OCT1 calculated using  $v_{max}$  and  $K_M$  of the data in A. Shown are only measurements of all three OCT1 orthologs in parallel, therefore, data may differ slightly from data shown in Fig. 2A Shown are means and standard errors of the means of four independent experiments. (D) Protein sequence alignment of TMH2 and TMH3 of human, mouse, and rat OCT1 with the conserved A-motif of the MFS underlined in violet. Coloring based on amino acid identity.



**Supplementary Figure S4. Structural overview of the TMH2/THM3 region in human and mouse OCT1 *in silico* models** Superposition of human and mouse OCT1 structural models with focus on TMH2 and TMH3 and non-conserved residues highlighted and labeled. (A) Inside view and (B) outside view on TMH2 and TMH3.



**Supplementary Figure S5. Differences in metformin uptake between human OCT1, mouse OCT1, and human OCT2** (A) Concentration-dependent uptake of metformin by human OCT1 (green), mouse OCT1 (red) and human OCT2 (grey). OCT1 or OCT2-overexpressing HEK293 cells were incubated with increasing concentrations of metformin for 2 min. OCT1 or OCT2-mediated uptake was calculated by subtracting the uptake of control cells (pcDNA5) from the uptake of cells overexpressing human or mouse OCT1 or human OCT2. Shown are only measurements of all three OCT paralogues in parallel, therefore, data may differ slightly from data shown in Fig. 2A. (B) Protein sequence alignment of TMH2 and TMH3 of human OCT1, mouse OCT1, and human OCT2 using the CLUSTAL O(1.2.4) multiple sequence alignment tool (Madeira et al., 2019). Coloring based on the BLOSUM62 matrix. Arrows indicate amino acids mutated in C. (C) Effect of mutations of hLeu155/mVal156 and hPhe169/mIle170 in human/mouse OCT1, respectively, on metformin uptake. HEK293 cells transiently overexpressing OCT1 were incubated with 100  $\mu$ M metformin for 2 min. Shown are means and standard errors of the means of three independent experiments. +++  $P < 0.001$  compared to mouse OCT1 in a Tukey's post hoc analysis following one-way ANOVA.