Hepatic Transporter Expression in Metabolic Syndrome: Phenotype, Serum Metabolic Hormones, and Transcription Factor Expression

Ajay C. Donepudi, Qiuqiong Cheng, Zhenqiang James Lu, Nathan J. Cherrington, and Angela L. Slitt

Department of Biomedical and Pharmaceutical Sciences, University of Rhode Island, Kingston, Rhode Island (A.C.D., Q.C., A.L.S); Arizona Statistical Consulting Laboratory, The Bio5 Institute (Z.J.L.) and Department of Pharmacology and Toxicology, College of Pharmacy (N.J.C.), University of Arizona, Tucson, Arizona

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ABSTRACT
Metabolic syndrome is a multifactorial disease associated with obesity, insulin resistance, diabetes, and the alteration of multiple metabolic hormones. Obesity rates have been rising worldwide, which increases our need to understand how this population will respond to drugs and exposure to other chemicals. The purpose of this study was to determine in lean and obese mice the ontogeny of metabolic hormones may play a role in regulating the genes involved in drug metabolism and transport.

Introduction
Obesity is a metabolic disease characterized by an increased body mass index (BMI $\geq 30$). It is a predominant risk factor for metabolic syndrome (MetS), which encompasses increases in body weight, adipose tissue mass, insulin resistance, and serum hormone levels (Grundy, 2004). One of the manifestations of metabolic syndrome is the development of hepatic lipid accumulation (e.g., steatosis) that represents nonalcoholic fatty liver disease (NAFLD) in conjunction with insulin resistance (Wanless and Lentz, 1990). There is a growing concern about absorption, distribution, metabolism, and excretion (ADME) in the obese population, as studies reveal altered ADME in people affected with obesity and metabolic diseases (Brill et al., 2012). Ob/ob mice have a mutation in the ob gene that encodes for leptin, resulting in a phenotype that has many characteristics common to MetS. Owing to lack of leptin, ob/ob mice exhibit hyperphagia, profound weight gain, hyperglycemia, glucose intolerance, elevated plasma insulin, and severe hepatic steatosis (Lindstrom, 2007). Most of these changes are observed predominantly in ob/ob mice after 3 weeks of age (Dubuc, 1976).

ADME of xenobiotics and several endogenous compounds are regulated by drug-metabolizing enzymes (DME) and drug transporters. Drug transporters are membrane-bound proteins that facilitate both uptake and efflux of xenobiotics, endogenous compounds, and their metabolites in various tissues, including liver. Hepatic transporter expression is an important determinant in maintaining systemic balance of endogenous compounds such as bile acids, hormones, and bilirubin (Lecureux et al., 2009). Multiple conditions can alter drug transporter expression in liver, such as obesity, oxidative stress, inflammation, drug-induced liver injury, and environmental toxicants (Geier et al., 2003; Aleksunes et al., 2008; Cheng et al., 2008). Previous studies document alterations in DME and drug transporter expression in obese and diabetic conditions (Cheng et al., 2008; More and Slitt, 2011).

In liver, transcription factors such as pregnane-X receptor (Pxr, Nr112), constitutive androstane receptor (Car, Nr113), farnesoid X receptor (Fxr, Nr1H4), and nuclear factor E2-related factor 2 (Nrf2, Fpargc1a, peroxisome proliferator-activated receptor gamma coactivator 1-alpha; Pxr (Nr112), pregnane-X receptor; WT, wild type.

ABBREVIATIONS: ABC, ATP-binding cassette; ADME, absorption distribution metabolism and elimination; Bsep (Abcb11), bile salt export pump; P450, cytochrome P450; Car (Nr113), constitutive androstane receptor; DME, drug-metabolizing enzyme; Fxr (Nr1H4), farnesoid X receptor; GLP, glucagon-like peptide; Lxr (Nr1H3), liver X receptor; MetS, metabolic syndrome; NAFLD, nonalcoholic fatty liver disease; Nrf2 (Nfe2l2), nuclear factor E2-related factor 2; Ntcp (Slc20a1), Na+-taurocholate cotransporting polypeptide; Ppar-?, peroxisome proliferator-activated receptor alpha; Ppargc1?, peroxisome proliferator-activated receptor gamma coactivator 1-alpha; Pxr (Nr112), pregnane-X receptor; WT, wild type.
Novel biomarkers that could be associated with ADME changes in obesity. For example, Pxr and Car upregulate Cyp3a11 and Cyp2b10 expression, whereas Nrfl2 upregulates Nqo1 and Gste gene transcription and expression (Alekunne et al., 2006). With regard to transporters, hepatic Abcc2-4 induction by microsomal enzyme inducers is observed to be Nrfl2-dependent (Maher et al., 2005). Prototypical Pxr activators upregulate hepatic Abcc2, Abcc3, Na+-taurocholate cotransporting polypeptide (Ntcp), and solute-carrier organic anion transporter (Slco)1a4 expression (Cheng et al., 2005, 2007; Maher et al., 2005), whereas Car activators upregulate Abcc2-6 mRNA expression in liver (Cheng et al., 2005; Maher et al., 2005). An increase in mRNA expression of these transcription factors was observed in livers of 9-week-old ob/ob mice compared with WT mice (Xu et al., 2012), implicating coordinate regulation of drug-transporter and transcription-factor expression in steatosis. However, the correlation or coordinated expression of transcription factors and transporter expression during development of fatty liver disease has not been well described or documented.

Obesity alters levels of several metabolic hormones, such as resistin, glucagon, insulin, and incretins (Starke et al., 1984; Azuma et al., 2003; Reinehr et al., 2007), which may influence hepatic gene expression in obesity. Along with hormonal changes, diabetes and obesity causes insulin resistance accompanied by hyperglycemia, which are known to regulate expression of several hepatic genes (Kahn et al., 2006). Although increased incretin levels, such as glucagon-like peptide-1 (GLP-1), are observed in obese people, GLP-1 activity associated with insulin secretion is decreased compared with lean individuals (Lafererre et al., 2007). Several therapies that target these hormones have been identified for treatment of obesity and other metabolic diseases (Schmitz et al., 2007). Serum levels of such hormones as resistin, leptin, and insulin were correlated to metabolic disease such as obesity (Pantsulaiha et al., 2004), indicating obesity can alter levels of hormones involved in regulation of metabolism, or vice-versa. As more endocrine hormone–targeting drugs are being approved and developed to manage aspects of metabolic disease, there is a need to understand the influence of these hormones on processes that dictate hepatic gene expression involved in drug metabolism and disposition in both preclinical species and humans.

Leptin-deficient obese (ob/ob) mice are commonly used to model MetS and fatty liver disease (Lindstrom, 2007). This study aimed to correlate typical clinical endpoints such as metabolic hormones and physiologic factors with hepatic transcription factors, prototypical DME, and transporter-mRNA expression to better predict drug disposition. The findings of this study provide potential insight into possible measures and serum biomarkers that could be associated with ADME changes in obesity.

Materials and Methods

Animals and Husbands. Adult heterozygous Lep^{+/-} (OB, B6.V-Lep^{+/+}, stock no. 0000632) breeders were purchased from Jackson Laboratories (Bar Harbor, ME). Heterozygous breeders were mated and offspring were genotyped for sex and mutation of the leptin gene. The resulting wild-type or C57BL/6 mice (WT) used in this study were considered the WT controls and leptin-deficient homozygous mice were considered obese (ob/ob) mice. In ob/ob mice pronounced changes in physiologic factors have been observed from 3 weeks of age (Dubuc, 1976), so to identify correlation between physiologic factors and hepatic gene expression during development of obesity we chose mice of different ages, such as week 1 (pre-), week 3, week 4 (post-), and week 8 (adult) mice. Tissues from male and female WT and ob/ob were collected at 1, 3, 4, and 8 weeks (n = 4–5 per group). Blood was collected and serum was obtained after centrifugation at 5000 rpm for 5 minutes at 4°C. Livers were collected, snap frozen in liquid nitrogen, and stored at −80°C for future analysis. All animal experiments were approved by University of Rhode Island Institutional Animal Care and Use Committee.

RNA Extraction. Total RNA from livers was isolated by phenol-chloroform extraction method using RNA-Beet reagent (Tel-Test, Inc., Friendswood, TX), according to the manufacturer’s protocol. RNA concentration was quantified by absorbance at 260 nm using a Nanodrop ND1000 (Thermo Fisher Scientific, Waltham, MA) and the samples were diluted to 1 μg/μl. Formaldehyde–agarose gel electrophoresis followed by UV illumination was used to visualize 28S and 18S ribosomal RNA and confirm integrity.

QuantGene Multiplex Suspension Array. Mouse liver mRNA expression was determined using a QuantGene Plex 2.0 assay (Panomics/Affymetrix eBioscience, San Diego, CA) with a Bio-Plex System array reader with Luminex 100 xMAP technology, and data were acquired using Bio-Plex Software Data Manager (Bio-Rad, Hercules, CA). Assays were performed according to the manufacturer’s protocol. The optimal RNA input was determined prior to running the assay. Briefly, 500 ng of total RNA was incubated overnight at 53°C with xMAP beads containing oligonucleotide capture-probes, label extenders, and blockers. On the next day, beads and bound target RNA were washed and subsequently incubated with amplifier at 46°C for 1 hour. Samples were then washed and incubated with the label (biotin) at 46°C for 1 hour. Samples were washed with streptavidin-conjugated R-phycocerythrin, which binds biotinylated probes, and incubated at room temperature for 30 minutes. Streptavidin-conjugated R-phycocerythrin fluorescein was then detected for each analyte within each sample.

Branchen DNA Amplification Assay. Relative expression of bile salt-export pump (Abcb11, Bsep) and Na+-taurocholate cotransporting polypeptide (Slc10a1, Ntcp) mRNA was quantified using bDNA assay and previously described probe sets (Cheng et al., 2007). All reagents for analysis, including lysis buffer, amplifier/label probe diluted, and substrate solution, were supplied in the QuantGene 1.0 assay kit (Panomics/Affymetrix eBioscience). Briefly, the probe-set stocks containing capture extenders, label extenders, and blockers were diluted 1:100 in lysis buffer before use. On day 1, total RNA samples (10 μg) were added to wells containing 50 μl of capture hybridization buffer and 50 μl of diluted probe set. The RNA was allowed to hybridize overnight with the probe set at 53°C. On day 2, subsequent hybridization steps were followed as detailed in the manufacturer’s protocol, and fluorescence was measured with a GloRunner Microplate Lumimeter interfaced with GloRunner DXL Software (Turner BioSystems, Sunnyvale, CA). The fluorescence for each well was recorded as relative light units (RLU) (Donepudi et al., 2012).

Serum Metabolism-Related Hormone Levels. Serum metabolism-related hormones were quantified using a Millipore 10-plex kit (MMHMA-44K; EMD Millipore/Merck, Billerica, MA) on a Bioplex multiple array system. A custom Millipore-plex kit containing different targets such as insulin, glucagon, resistin, GLP-1, amylin and leptin was used and analyzed according to manufacturer’s protocol. Fluorescence was detected on a Bioplex multiple array reader system (Bio-Rad). Data were collected by Bioplex Software Manager 5.0 and plotted as average concentration (μg/ml). Serum glucose levels were analyzed using a Glucose Colorimetric Assay Kit (Cayman Chemical, Ann Arbor, MI).

Correlation Analysis. Correlations between the mRNA levels of genes related to drug metabolism and transcription factors were performed using either Statistica 9.1 software (StatSoft, Inc., Tulsa, OK) or canonical correlation analysis. Briefly, for canonical correlation analysis data generated was log-transformed and distributed in three blocks such that block 1 contained gene expression of drug transporters and phase 1 enzymes, block 2 contained transcription factor expression, and block 3 contained physiologic data—serum hormone and glucose levels and body and liver weights. Cross-block pairwise bivariate correlations were performed between each block, and heat maps were generated. Hierarchical clustering was performed using the same data with Pearson correlation. Data presented as heat maps or with r-value P ≤ 0.05 were considered a statistically significant correlation.

Statistical Analysis. The statistical significance between groups was determined by factorial analysis of variance (ANOVA) followed by a Duncan multiple-range post-hoc test, using Statistica 9.1 software (StatSoft, Inc.). Data are presented as mean ± S.E., with P ≤ 0.05 considered statistically significant.

Results

Ontogeny of Serum Hormones in WT and Ob/Ob Mice. At week 1 of age, body weight was similar between all groups, and at weeks 4 and 8, ob/ob mice body weight had increased by at least 1.2- and 1.7-fold, respectively, compared with WT mice (Supplemental Data, Fig. 1A).

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Serum glucose levels increased from week 1 to 8 in both males and females (Supplemental Data, Fig. 1B). In ob/ob mice, serum glucose levels increased with age from week 3 in females and week 4 in males compared with week-1 ob/ob mice. Figure 1 illustrates serum metabolic hormone changes observed in both male and female WT and ob/ob mice from weeks 1, 3, 4, and 8. Interestingly, in both male and female WT mice such serum hormone levels as glucagon, resistin, and GLP-1 changed with age. However, in both male and female WT mice insulin and amylin levels were similar at all ages. In WT mice, serum glucagon, resistin, and GLP-1 levels were decreased by ~78% from week 3 compared with week 1. These age-dependent changes in serum glucagon and GLP-1 levels were not observed in either male and female ob/ob mice.

In both male and female mice, serum hormone levels were similar between WT and ob/ob mice at 1 week of age. However, in both male and female ob/ob mice, serum insulin and amylin increased with age after week 4 by 4-fold compared with week 1, whereas glucagon, resistin, and GLP-1 levels decreased by 57, 31, and 40% respectively. In both males and females, all serum hormone levels were significantly increased in ob/ob compared with 8-week-old WT mice.

Hepatic Drug Transporters and Prototypical Metabolizing-Enzyme Expression in Livers of WT and Ob/Ob Mice. Figure 2A illustrates hepatic efflux drug transporter expression observed in both male and female WT and ob/ob mice. In WT and ob/ob mice, ontogenic changes were observed in expression of Abc transporters such as Abcc1, 3, 4, 5, and Bsep levels in both males and females. At week 1, Abc transporter expression was similar between male and female WT and ob/ob mice. In male ob/ob mice at week 8, Abcc3, 4, and Abcg2 mRNA expression were higher than WT mice by 1.8-, 7-, and 2.3-fold, respectively. A similar increase was observed in female ob/ob mice compared with WT mice in higher than WT mice by 1.8-, 7-, and 2.3-fold, respectively. A similar male ob/ob mice at week 8, Abcc3, 4, and Abcg2 mRNA expression were expression was similar between male and female WT and ob/ob mice. In WT and ob/ob mice, ontogenic changes were observed in expression of hepatic uptake transporters such as Slco1a4, 1b2, and Slc10a1, although an increased expression of Slco1a4 was observed only at 3 and 4 weeks compared with respective week 1 counterparts.

Figure 2C illustrates hepatic DME expression in WT and ob/ob mice. In both male and female mice, Cyp3a11 mRNA expression increased with age, with no significant changes between ob/ob and WT counterparts at the same ages. In male and female ob/ob mice, Cyp2b10 and 4a14 mRNA expression increased from week 1 to 8, whereas no significant ontogenic changes were observed with age in WT mice. In males, Cyp2b10 and 4a14 expression in ob/ob mice were 4.3- and 2.2-fold higher than their WT counterparts, respectively. In females, Cyp4a14 mRNA expression in female ob/ob mice was 2- to 3-fold higher than WT counterparts at weeks 4 and 8. In female ob/ob mice significant increase in Cyp3a11 and Cyp2b10 gene expression by 1.3- and 1.8-fold was observed only at week 4 compared with respective WT counterparts. These changes in Cyp3a11 and Cyp2b10 expression in female ob/ob mice were not observed in week 8 mice.

Hepatic Transcription Factor Expression in WT and ob/ob Mice. In male and female, WT and ob/ob mice, there were no significant ontogenic changes in transcription factor mRNA expression with the exception of Pxr. Figure 3 illustrates the hepatic transcription factor mRNA expression levels in livers of male and female WT and ob/ob mice. In WT female mice, Pxr mRNA expression decreased with age by 51% compared with week 1, whereas ontogenic changes in Pxr expression were not observed in female ob/ob mice. In male ob/ob mice, Nrf2 mRNA expression was 2-fold higher at week 8 compared with week 1 counterparts. Male ob/ob mice have 2- to 3-fold higher mRNA levels of liver X receptor (Lxr), Fxr, and Nrf2 compared with their WT counterparts only at week 8. These changes in male ob/ob

**Fig. 1.** Effect of age and leptin deficiency on phenotypic changes in WT and ob/ob mice. Metabolism-related hormone levels of WT and ob/ob mice at week-1, 3, 4 and 8. Metabolism-related hormone levels in WT and ob/ob mice were quantified in serum collected from WT and ob/ob mice at 1, 3, 4, and 8 weeks of age (n = 4-5 for each group/sex). Hormones were quantified using a Luminex-based assay. Data are presented as average concentration (µg/ml) ± S.E.M. (n = 4–5 mice per group). Asterisks (*) represent a statistical difference between wild type and ob/ob of same age group and hash mark (#) represent a statistical difference with respect to week-1 mice (P ≤ 0.05).
The role of sex and leptin in ontogenic changes in hepatic gene expression pattern is depicted pictorially using heat maps (Fig. 4). Gene expression values from male and female, and WT and ob/ob mice were log-transformed to generate heat maps for each group. These heat maps strongly indicate that leptin and sex regulate hepatic genes involved in metabolism and disposition. Differences in ontogenic pattern of hepatic gene expression between ob/ob mice and WT mice illustrate the influence of leptin on hepatic gene expression patterns. In WT males, hepatic transcription factors and efflux transporters, with the exception of Abcc2, showed high expression (red color) at week 1 with a gradual decrease in expression with age (low expression indicated with green color at week 8), whereas the same expression patterns were not observed in female ob/ob mice. In females only serum resistin and insulin levels correlated with all clinical markers, whereas in female mice, serum resistin and amylin correlated with most other physiologic factors during development of obesity. In both males and females, the correlations between physiologic factors suggest that changes in serum resistin levels may be associated with development of obesity, irrespective of sex.

Correlation Analysis between Metabolic Hormones and Phenotypic Changes. The role of sex and leptin in ontogenic changes in hepatic gene expression pattern is depicted pictorially using heat maps (Fig. 4). Gene expression values from male and female, and WT and ob/ob mice were log-transformed to generate heat maps for each group. These heat maps strongly indicate that leptin and sex regulate hepatic genes involved in metabolism and disposition. Differences in ontogenic pattern of hepatic gene expression between ob/ob mice and WT mice illustrate the influence of leptin on hepatic gene expression patterns. In WT males, hepatic transcription factors and efflux transporters, with the exception of Abcc2, 3, and Bsep, showed high expression (red color) at week 1 with a gradual decrease in expression with age (low expression indicated with green color at week 8), whereas the same expression patterns were not observed in male ob/ob mice. In females also only WT mice had high expression (red color) of efflux transporters, with the exception of Abcc2, 3, and Bsep, at week 1 and low expression (green color) at week 8. Only a few hepatic transcription factors such as peroxisome proliferator–activated receptor gamma coactivator 1-alpha (Ppargc1a), and Pxr showed similar patterns in both male and female WT mice. Table 1 illustrates correlations between physiologic factors, such as metabolic hormone levels, serum glucose, and body and liver weights, that were performed using canonical correlations. Canonical correlations were performed to determine how different metabolic hormones are correlated with each other during the development of obesity (Table 1).
In males, serum GLP-1 concentrations were correlated with only Bsep, Slco1a4, Cyp3a11, and 2e1 expression, whereas in females they were correlated with Abcc2, 3, Bsep, Slco1a1, 1a4, 1b2, Cyp3a11, 2b10, and 2e1 expression. In males, serum glucose levels were correlated to Abcc1, 3, 5, 6, Abcb1a, Bsep, Cyp3a11, and 2b20 expression, whereas they were correlated to Abcc3-5, Cyp3a11, and 2b10 in females. Both body and liver weights correlated to Abcc1, 3, 5, 6, Abcb1a, Bsep, Slco1a1, Cyp3a11, and 2b10 expression in males, whereas in females they correlated with Abcc1, 3–6, Cyp3a11, and 2b10 expression.

Together these analyses indicate that correlation between physiologic factors and hepatic gene expression differs between males and females during development of obesity. In males, during development of obesity serum resistin, glucagon, and glucose levels correlated with most of hepatic Abc transporter expression compared with serum amylin, insulin, and GLP-1 levels. In females, serum glucose showed a higher correlation with hepatic Abc transporter expression than serum metabolism-related hormones. In females, during development of obesity, hepatic uptake transporter mRNA expression correlated with serum levels of resistin, glucagon, and GLP-1 (Fig. 5 and (Supplemental Table 2). Apart from serum metabolism-related hormones, serum glucose levels and body and liver weight stood as a markers for change in gene expression and physiologic factors during development of obesity (Table 1, Fig. 5, (Supplemental Fig. 2, and (Supplemental Table 2).

Figure 6 illustrates the correlation between transcription factors with drug transporters and phase-I enzyme expression in male and female mice. Correlation between transcription factors, drug transporters, and DME expression was performed using canonical correlations, and the data generated was presented in the form of heat maps. In males, during development of obesity and MetS, expression patterns of all Abc transporters with the exception of Bsep were correlated with all transcription factors analyzed, with minor exceptions. In male mice, Abcc3 expression did not strongly correlate with Pxr, Car, Ppar-α, and Ppargc1α expression, whereas Abcc5 was not correlated with Fxr. Correlation between Abc transporters and transcription factors showed sex-dependent changes during development of obesity. Interestingly in female mice only, Abcc2, 6, and Abcb1a expression correlated with all transcription factors analyzed, whereas Abcg2 expression was correlated with all transcription factors with the exception of Car during development of obesity. In female mice, Abcc3 expression correlated to Car, Lxr, Fxr, and Nrf2 expression, whereas Abcc4 expression was correlated to Pxr, Lxr, Fxr, and Nrf2 expression. In both male and female mice, uptake transporter Slco1b2 expression correlated to all transcription factor expressions, whereas Slco1a4 expression was correlated with Car, Lxr, Fxr, and Nrf2, and Ppargc1α expression. Expression of DMEs such as Cyp2e1 and 4a14 was correlated with all transcription factor expressions in both male and female mice. In male mice, Cyp2b10 expression was correlated to Lxr, Fxr, and Nrf2 expression, whereas in females Cyp3a11 and 2b10 were correlated with Car, Lxr, Fxr, and Nrf2 expression.

Discussion

Obesity and ensuing MetS is a major health concern in the United States (Grundy, 2004). Ob/ob mice are often used to model MetS and NAFLD because they have elevated multiple serum biomarkers, hepatic steatosis, and hepatic gene-expression changes that are similar to humans with uncontrolled MetS (Lindstrom, 2007). Drug transporter and DME expression were altered during NAFLD and could have resulted in altered drug elimination (Lickteig et al., 2007; Cheng et al., 2008; Barshop et al., 2011; Hardwick et al., 2012). Moreover, changes in hepatic uptake and efflux transporter expression in ob/ob mice are similar to those observed in livers of diet-induced obese mice or humans with steatosis (Cheng et al., 2008; More and Slitt, 2011).
Abc transporters enable compounds to pass from hepatocytes into bile or blood (Faber et al., 2003). In both WT and ob/ob mice, Abcc1 and 5 were expressed highly at week 1 and decreased with age. This pattern in expression of Abcc1—high expression at initial stages and decrease at later age—is also observed in liver regeneration after 90% hepatectomy, indicating that Abcc1 has a lesser role in adult liver (Kimura et al., 2012). Abcc3 and 4 are basolateral efflux transporters with relatively low expression in liver; they are induced by microsomal inducers during several disease conditions (Slitt et al., 2003; Mennone et al., 2006; More and Slitt, 2011). In both obese males and females Abcc3 and 4 mRNA levels increased with age, and also compared with their WT littermates, whereas no ontogenic changes were observed in Abcc3 and 4 expression with age in WT mice. Apical efflux transporters such as Abcc2, Abcg2, Abcb1a, and Bsep play a major role in excretion of xenobiotics and endogenous substances from liver to bile. In both male and female mice, Abcc2 and Abcb1a mRNA levels are unaltered with age and obesity. Previous studies indicated no change in mRNA levels or an increase in protein levels of Abcc2 in ob/ob mice, which is consistent with our results (Cheng et al., 2008). Abcc2 expression during obesity is species-specific; in obese Zucker rats Abcc2 expression decreases, whereas it increases in ob/ob mice, but in humans there is no change with obesity (Pizarro et al., 2004; Cheng et al., 2008; More and Slitt, 2011). In male ob/ob mice, Bsep expression was similar to respective WT littermates, whereas in females Bsep expression decreased with progression of obesity. The sex-specific difference in Bsep expression was may be attributable to a sex-specific difference in growth hormone responsiveness in obesity (Cocchi et al., 1993), which is known to regulate Bsep expression (Cheng et al., 2007).

**TABLE 1**

The correlation matrices for variables in phenotypical factors during development of obesity in male and female mice

<table>
<thead>
<tr>
<th></th>
<th>Amylin</th>
<th>Resistin</th>
<th>Insulin</th>
<th>Glucagon</th>
<th>GLP-1</th>
<th>Glucose</th>
<th>Body weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male mice</td>
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<td></td>
<td></td>
<td></td>
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<tr>
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<td>0.4*</td>
<td>0.02</td>
<td>−0.05</td>
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<td></td>
</tr>
<tr>
<td>Body weight</td>
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<td>0.35*</td>
<td>0.44</td>
<td>−0.13</td>
<td>0.94*</td>
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<tr>
<td>Liver weight</td>
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<td>−0.52*</td>
<td>0.36*</td>
<td>−0.12</td>
<td>−0.15</td>
<td>0.93*</td>
<td>0.99*</td>
</tr>
<tr>
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<td></td>
<td></td>
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<tr>
<td>Resistin</td>
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<td>0.57*</td>
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<tr>
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Fig. 4. Heat maps for gene expression analysis performed in both male and female WT and ob/ob mice. Gene expression values are log-transformed and heat maps were generated using R language.
Previous studies showed that Abc transporter expression is regulated by several transcription factors (Maher et al., 2005). Interestingly, significant correlations were observed between hepatic Abc transporters and transcription factor expression during development of obesity. These correlations during development of obesity between transcription factors and hepatic efflux transporters showed sex-dependent changes, and are much more pronounced in week-8 mice (Supplemental Table 1). In both male and females, hepatic uptake transporter Slco1a1 mRNA levels increased with age only in WT mice. In both male and female ob/ob mice Slco1a1 mRNA levels decreased significantly at week 8. Previous studies showed that Slco1a1 mRNA expression is androgen-dependent and negatively regulated by microsomal enzyme inducers that activate transcription factors Pxr, Car, Ppar-α, and Nrf2 (Lu et al., 1996; Cheng et al., 2005). Ob/ob mice are known to have decreased androgen levels (Swerdloff et al., 1976) and increased transcription factor expression (Xu et al., 2012). Obesity did not affect the ontogeny of Slco1a4, 1b2, and Ntcp in both males and females.

The mRNA expression of phase-I biotransformation enzymes, such as Cyp3a11 and 2e1, was similar between WT and ob/ob mice in this study. Cyp2e1 expression during obesity and diabetes appears to be species-specific. In humans, Cyp2e1 in liver is increased during obesity and diabetes, whereas in mice it appears to be unchanged or decreased (Enriquez et al., 1999; Wang et al., 2003; Cheng et al., 2008). In males, Cyp2b10 mRNA levels increased with development of obesity compared with WT mice, whereas in female Cyp2b10 mRNA levels remained unchanged. These sex-specific changes in Cyp2b10 expression were observed with phenobarbital treatment, which is a known inducer of the Cyp2b family (Larsen et al., 1994; Cheng et al., 2008). Insulin treatment altered Cyp3a, 2e1, and 4a expression in hepatocytes (Kim et al., 2003), indicating that metabolism-related hormones can regulate phase-I enzyme expression. Our results indicate that in female mice, P450 enzyme mRNA expression during development of obesity correlates most closely with resistin and GLP-1, whereas in males these correlations are less well associated. Sexually dimorphic regulation of P450 enzyme expression might be responsible for sex-specific differences in expression and correlations with metabolism-related hormones (Hernandez et al., 2009).

Incretins, such as amylin and GLP-1, play a key role in insulin secretion and maintenance of blood glucose levels. In both male and female ob/ob mice, GLP-1 and amylin levels were increased compared with WT. Amylin is secreted along with insulin from pancreas and is involved in maintaining serum glucose levels (Moreno et al., 2011). Changes in amylin and insulin levels in our study were observed at the same time. Moreover, changes in amylin and insulin levels, along with resistin, showed significant correlation with other physiologic
parameters analyzed in females during development of obesity. Hyperglycemia was identified as one of the important factors causing several metabolic abnormalities during obesity and MetS (Reinehr et al., 2007). Drug therapies targeting incretins, such as GLP-1 agonists or DPP4 inhibitors, were developed as potential treatments for MetS (Schmitz et al., 2004). In humans, GLP-1 mimetic administration was shown to have drug-drug interaction by altering area under curve and Cmax of drugs such as acetaminophen, lovastatin, digoxin, ethinyl estradiol, and lisinopril (Hurren and Pinelli, 2012). Although these changes were thought to be the result of delayed gastric emptying by GLP-1 mimetics, more detailed studies are needed to explain the identified changes in drug disposition. Correlation analyses in our study indicate that incretins such as GLP-1 and amylin may regulate DME and hepatic drug transporter expression, and changes in these incretin levels could potentially alter ADME of a drug via altered expression of DME and drug transporters.

Several studies have shown correlations between alteration in serum levels of various hormones during metabolic disorders. Studies also shown that hormones such as estrogen, progesterone, and androgens could alter hepatic uptake and efflux transporter expression (Geier et al., 2003; Kalabis et al., 2007). The role of metabolic hormone regulation of genes involved in drug metabolism is less well known. Acute or chronic administration of metabolism-related hormones, such as glucagon and GLP-2, can alter transporter expression and function. For example, GLP-2, an incretin secreted along with GLP-1, was shown to regulate Abcc2 expression in intestine (Villanueva et al., 2010). In human and rat hepatocytes, short term glucagon treatment altered Abcc3 function (Chandra et al., 2005). Our laboratory also showed glucagon treatment induces ABCC2-4 mRNA expression in human hepatocytes and that fasting can increase biliary excretion of Dibromsulphathalein (DBSP), an Abcc2 substrate (Kulkarni et al., 2014). Not only transporters but also phase-II DME expression were altered with glucagon and insulin treatment (Kim et al., 2003), indicating metabolism-related hormones have a potential to alter drug metabolism and related gene expression. Correlations observed in our study indicate that not only sex-related hormones, such as estrogen and testosterone (Lu et al., 1996; Geier et al., 2003), but also hormones involved in energy metabolism, such as resistin, glucagon, and GLP-1, are potential regulators of hepatic uptake transporter expression.

In summary, expression patterns in ob/ob mice were similar to WT littermates at week 1. In both males and females, as obesity developed and metabolic hormone changes occurred, more robust expression changes in liver were observed after weeks 4 and 8. Drug transporter and DME expression were associated with metabolism-related hormones. Correlations revealed that the metabolism-related hormones, such as resistin, GLP-1, amylin, and glucagon were either coregulated or involved in regulation of hepatic gene expression, which needs further investigations. We recognize that species differences and use of mRNA data must be taken into consideration when extrapolating from our findings. This study was an initial step in exploring how clinical and serum measures can aid in the prediction of altered ADME processes and how metabolic hormone levels might regulate hepatic transporter expression during metabolic stress. Although this study did not identify a single biomarker for altered expression in hepatic drug transporters and DME expression during development of obesity, it did indicate that metabolic hormones may play role in regulating gene expression for processes that dictate drug metabolism and disposition. Identifying the roles of serum metabolic hormones, such as resistin, GLP-1, amylin, and glucagon, in regulating genes involved in drug metabolism is at present an unexplored field of study. Correlations observed in this study were obtained using mRNA levels, and future studies need to confirm these correlations at a protein and activity level. To summarize, this study provided insight into potential relationships between metabolic hormones and transporter expression, as well as insight into predictive markers for altered transporter expression.

**Authorship Contributions**

- **Performed data analysis**: Donepudi, Slitt.
- **Wrote or contributed to the writing of the manuscript**: Donepudi, Lu, Cherrington, Slitt.

**References**


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Address correspondence to: Dr. Angela L. Slitt, Department of Biomedical and Pharmaceutical Sciences, University of Rhode Island, 7 Greenhouse Road, Kingston, RI 02881. E-mail: aslitt@uri.edu