Short Communication

Regio- and Stereo-Selective Oxidation of a Cardiovascular Drug, Metoprolol, Mediated by Cytochrome P450 2D and 3A Enzymes in Marmoset Livers

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ABSTRACT

A β-blocker, metoprolol, is one of the in vivo probes for human cytochrome P450 (P450) 2D6. Investigation of nonhuman primate P450 enzymes helps to improve the accuracy of the extrapolation of pharmacokinetic data from animals into humans. Common marmosets (Callithrix jaccus) are a potential primate model for preclinical research, but the detailed roles of marmoset P450 enzymes in metoprolol oxidation remain unknown. In this study, regio- and stereo-selectivity of metoprolol oxidations by a variety of P450 enzymes in marmoset and human livers were investigated in vitro. Although liver microsomes from cynomolgus monkeys and rats preferentially mediated S-metoprolol O-demethylation and R-metoprolol α-hydroxylation, respectively, those from humans, marmosets, minipigs, and dogs preferentially mediated R-metoprolol O-demethylation, in contrast to the slow rates of R- and S-metoprolol oxidation in mouse liver microsomes. R- and S-metoprolol O-demethylation activities in marmoset livers were strongly inhibited by quinidine and ketoconazole, and were significantly correlated with bufuralol 1'-hydroxylation and midazolam 1'-hydroxylation activities and also with P450 2D and 3A4 contents, which is different from the case in human livers that did not have any correlations with P450 3A-mediated midazolam 1'-hydroxylation. Recombinant human P450 2D6 and marmoset P450 2D6/3A4 enzymes effectively catalyzed R-metoprolol O-demethylation, comparable to the activities of human and marmoset liver microsomes, respectively. These results indicated that the major roles of P450 2D enzymes for the regio- and stereo-selectivity of metoprolol oxidation were similar between human and marmoset livers, but the minor roles of P450 3A enzymes were unique to marmosets.

Introduction

Species similarities and differences for metabolic clearance of drug candidates are important issues for drug development. For predicting human toxicokinetic and pharmacokinetic profiles of drug candidates, nonhuman primates have been used because of their physiologic and genetic similarity to those of humans (Orsi et al., 2011; Sasaki, 2015). Common marmosets (Callithrix jaccus), small New World Monkeys, are useful nonhuman primate species for preclinical testing due to the functional similarity of cytochrome P450 (P450) enzymes with those of humans (Uno et al., 2016). Recently, our studies demonstrated similar enzymatic properties of P450 3A and 2D enzymes (Uehara et al., 2015a; 2017b).

Metoprolol is a β-adrenergic blocking drug that is used in cardiovascular medicine. Racemic mixtures of metoprolol have been used for treating hypertension, angina pectoris, and arrhythmia, but the pharmacological effects reside in the S-enantiomer of metoprolol (Lennard et al., 1986). Metoprolol is metabolized through O-demethylation (65% of dose), α-hydroxylation (10% of dose), and N-dealkylation (10% of dose) in humans in vivo (Borg et al., 1975). The α-hydroxylation of metoprolol is catalyzed almost entirely, and O-demethylation of metoprolol is catalyzed partially through human P450 2D6 (Otton et al., 1988). Reportedly, human P450 2D6 preferentially catalyzed the R-enantiomer faster than the S-enantiomer (Mautz et al., 1995). In our recent studies, marmoset P450 2D6 and 2D8 enzymes catalyzed metoprolol O-demethylation (Uehara et al., 2015a), indicating that the roles of marmoset P450 enzymes in metoprolol metabolism need to be further elucidated for preclinical studies. In this study, the role of P450s for regio- and stereo-selective metoprolol oxidation was evaluated in marmosets and humans. We report herein the high affinity of marmoset P450 2D enzymes with R-metoprolol O-demethylation and the high capacity of marmoset P450 3A enzymes for R- and S-metoprolol O-demethylation.

Materials and Methods

Detailed methods are shown separately in the Supplemental Material. Pooled liver microsomes from mice (CD-1, 8 males, 11 weeks of age), rats (Sprague-Dawley, 3 males), minipigs (Gottingen, 2 males, 7 months of age), dogs (10 males, >12 months of age), marmosets (5 males, sexually mature), and humans (74 males and 76 females, 18–82 years of age) were purchased from Corning Life Sciences (Woburn, MA). Pooled liver microsomes from cynomolgus monkeys (5 males, sexually mature) were purchased from Xenotech (Lenexa, KS). Liver microsomes from 17 individual human subjects were obtained from Corning Life Sciences. Individual liver microsomes were prepared from 23 marmosets (14 males and 9 females, >2 years of age) at the Central Institution for Experimental Animals (Kawasaki, Japan) in our laboratory, as described previously (Uehara et al., 2017a). This study was supported by a Japan Society for the Promotion of Science Grant-in-Aid for Young Scientists (B) [15K18934].

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ABBREVIATIONS: K_m, Michaelis constant; P450, cytochrome P450.
reviewed and approved by the Institutional Animal Care and Use Committee (Central Institution for Experimental Animals).

Activities of metoprolol O-demethylation and α-hydroxylation by liver microsomes and recombinant P450s were measured as described previously (Uehara et al., 2015a) with some minor modifications. A Pearson product-moment correlation was performed with GraphPad Prism (GraphPad, La Jolla, CA). Values of kinetic parameters \( V_{\text{max}} \), Michaelis constant \( (K_m) \), and \( V_{\text{max}}/K_m \) values were determined by fitting to an integrated form of the Michaelis-Menten equation using Kaleidograph (Synergy Software, Reading, PA).

Results and Discussion

R- and S-metoprolol oxidation activities by liver microsomes from humans, cynomolgus monkeys, marmosets, minipigs, dogs, rats, and mice were determined at a substrate concentration of 1.0 \( \mu \)M (Fig. 1). Rat liver microsomes preferentially catalyzed R- and S-metoprolol α-hydroxylation rather than O-demethylation, in contrast to the slow rates of R- and S-metoprolol oxidation in mouse liver microsomes. On the other hand, cynomolgus monkey liver microsomes effectively catalyzed R- and S-metoprolol O-demethylation rather than α-hydroxylation. Human, marmoset, and minipig liver microsomes showed roughly similar regio- and stereo-selective R- and S-metoprolol oxidation activities under the present conditions.

R- and S-metoprolol oxidation activities by human and marmoset recombinant P450 enzymes were determined at the substrate concentration of 10 \( \mu \)M (Supplemental Fig. 1). High catalytic activities of R-metoprolol O-demethylation by human and marmoset recombinant P450 2D6 enzymes were observed. Rates of R- and S-metoprolol O-demethylation by marmoset P450 3A4 and 3A90 enzymes were higher than those of human P450 3A4 and 3A5 enzymes. Human and marmoset P450 2D6 enzymes also catalyzed R- and S-metoprolol α-hydroxylation at slow rates. The roles of P450 enzymes in metoprolol O-demethylation by liver microsomes from humans and marmosets were investigated.

Correlations between R- and S-metoprolol O-demethylation activities and P450 probe oxidation activities in individual human and marmoset liver microsomes were determined (Supplemental Fig. 2). R- and S-metoprolol O-demethylation activities in human liver microsomes at substrate concentrations of 1.0 and 100 \( \mu \)M were significantly correlated to those of bufuralol 1'-hydroxylation \((r > 0.67)\). A high correlation coefficient \((r = 0.91)\) was observed for R-metoprolol O-demethylation activities at the substrate concentration of 1.0 \( \mu \)M. R- and S-metoprolol O-demethylation activities in marmoset liver microsomes at substrate concentrations of 1.0, 10, and 100 \( \mu \)M were also significantly correlated with bufuralol 1'-hydroxylation \((r > 0.77)\). High correlation coefficients \((r > 0.83)\) were observed for R- and S-metoprolol O-demethylation activities at low substrate concentrations of 1.0 or 10 \( \mu \)M. R- and S-metoprolol O-demethylation activities in marmoset liver microsomes at substrate concentrations of 1.0, 10, and 100 \( \mu \)M were also significantly correlated to those of midazolam 1'-hydroxylation \((r > 0.71)\), which is different from the case in human liver microsomes. In addition, R- and S-metoprolol O-demethylation activities in marmoset liver microsomes at the low substrate concentration of 1.0 \( \mu \)M were significantly correlated with P450 2D enzyme contents \((r > 0.63)\) and P450 3A4 enzyme contents \((r > 0.69)\) in marmoset liver microsomes (Supplemental Fig. 3).

The effects of P450 inhibitors for R-metoprolol O-demethylation live microsomes from humans and marmosets were determined at the substrate concentration of 10 \( \mu \)M (Supplemental Fig. 4). Dose-dependent suppression by quinidine (human P450 2D6 inhibitor, 2.0–20 \( \mu \)M) was observed for R-metoprolol O-demethylation in human liver microsomes (less than 23% of control activities) and for R- and S-metoprolol O-demethylation in marmoset liver microsomes (less than 10% of control activities). R- and S-metoprolol O-demethylation in marmoset liver microsomes was also dose-dependently suppressed by 1.0–10 \( \mu \)M ticlopidine (human P450 2C19 and 2D6 inhibitors; Ko et al., 2000) to ~26% of control activities. R- and S-metoprolol O-demethylation in marmoset liver microsomes were suppressed to less than 24% in the presence of 100 \( \mu \)M ketoconazole (human P450 3A inhibitor), in contrast to results showing that ketoconazole (100 \( \mu \)M) inhibited R- and S-metoprolol O-demethylation in human liver microsomes to half of control activities (Supplemental Fig. 4). Taken together, liver microsomal marmoset P450 2D6 effectively mediated R- and S-metoprolol O-demethylation in a manner similar to that in human liver microsomal P450 2D6, but the roles of P450 3A enzymes for R- and S-metoprolol O-demethylation in liver microsomes were partially different between human and marmoset livers.

Kinetic analyses for O-demethylation and α-hydroxylation of R- and S-metoprolol were performed using liver microsomes from humans and marmosets and recombinant P450 enzymes (Supplemental Fig. 5; Table 1). Liver microsomes from humans and marmosets showed low and high \( K_m \) values (humans, 15 and 190 \( \mu \)M; marmosets, 14 and 120 \( \mu \)M) for R-metoprolol O-demethylation activities with similar \( V_{\text{max}} \) values (humans, 0.15 and 0.48 nmol/min/mg protein; marmosets, 0.63 and 0.59 nmol/min/mg protein), respectively; those two component \( K_m \) values for R-metoprolol O-demethylation were lower than the \( K_m \) values (humans, 32 and 920 \( \mu \)M; marmosets, 21 and 150 \( \mu \)M) for S-metoprolol O-demethylation. Liver microsomes from humans and marmosets indicated low \( K_m \) values (37 and 27 \( \mu \)M) for R-metoprolol α-hydroxylation with low \( V_{\text{max}} \) values (0.054 and 0.047 nmol/min/mg protein) compared with those for S-metoprolol α-hydroxylation. Liver microsomes from humans and marmosets showed high \( V_{\text{max}}/K_m \) values (humans, 0.010 and 0.0025 ml/min/mg protein; marmosets, 0.045 and 0.0049 ml/min/mg protein) for R-metoprolol O-demethylation, in both the high- and low-affinity components. It has been suggested that the high affinity for R-metoprolol O-demethylation in

![Fig. 1. R- and S-metoprolol oxidation activities by liver microsomes from humans, cynomolgus monkeys, marmosets, minipigs, dogs, rats, and mice.](image-url)
human liver microsomes and for R- and S-metoprolol O-demethylation in marmoset liver microsomes is accounted for by P450 2D6 enzymes because the corresponding recombinant P450 2D6 enzymes had a \( K_m \) value of 11 \( \mu \)M for R-metoprolol O-demethylation and \( K_m \) values of 62 and 61 \( \mu \)M for R- and S-metoprolol O-demethylation, respectively. Recombinant human and marmoset P450 2D6 enzymes showed high \( V_{\text{max}}/K_m \) values (3.3 and 0.48 ml/min/mmol) for R-metoprolol O-demethylation compared with S-metoprolol O-demethylation. Marmoset P450 2D8 enzymes showed low \( V_{\text{max}}/K_m \) values (0.028 and 0.024 ml/min/mmol) for R- and S-metoprolol O-demethylation compared with marmoset P450 2D6 enzymes. Marmoset P450 3A4A showed high \( K_m \) values (280 and 290 \( \mu \)M) and low \( V_{\text{max}} \) values (0.43 and 0.46 ml/min/mmol) for R- and S-metoprolol O-demethylation. In contrast, \( K_m \) values for R- and S-metoprolol O-demethylation in human P450 3A enzymes and other enzyme sources were high (over \( \sim 500 \) \( \mu \)M) under the present conditions.

Oral administration of metoprolol in cynomolgus monkeys and minipigs yielded plasma concentrations similar to their quantitative detection limits (Mogi et al., 2012; Shida, et al., 2015), suggesting rapid oxidative clearance of metoprolol in vivo in a manner similar to that of in vitro oxidation in cynomolgus monkeys (Fig. 1). At a high concentration of 100 \( \mu \)M (not at 1.0 \( \mu \)M, as in Fig. 1), metoprolol oxidation by minipig liver microsomal P450 2D25 enzymes was faster than that in human liver microsomal P450 2D6 enzymes (Mogi et al., 2012; Yamazaki, 2014), resulting in a low bioavailability of metoprolol after oral administration in minipigs. On the other hand, human plasma concentrations of P450 probes containing metoprolol reportedly can be extrapolated from the corresponding data in marmosets after oral administration using simplified physiologically based pharmacokinetic modeling with in vitro metabolic cleavage data (Utoh et al., 2016). In this study, liver microsomes from humans and marmosets preferentially mediated R-metoprolol O-demethylation rather than S-metoprolol O-demethylation (Fig. 1), which is different from cynomolgus monkeys. Stereo-selectivity for R-metoprolol O-demethylation by marmoset liver microsomes was efficiently mediated by liver microsomal P450 2D6, as shown in this study (Supplemental Fig. 1; Table 1), which is similar to reported human P450 2D-dependent R-metoprolol O-demethylation (Mautz et al., 1995). The calculated \( V_{\text{max}}/K_m \) values for R- and S-metoprolol O-demethylation in liver microsomes from humans and marmosets were comparable (Table 1), which showed stereo-selective metoprolol O-demethylation in a manner similar to that in recombinant human and marmoset P450 2D6 enzymes (Supplemental Fig. 1). The stereo-selectivity for R-metoprolol O-demethylation by marmoset liver microsomal P450 2D6 enzymes was not as predominant as that of human liver microsomal P450 2D6 enzymes in the correlation analyses (Supplemental Figs. 2 and 3) and inhibition assays with quinidine (Supplemental Fig. 4). On the other hand, R- and S-metoprolol O-demethylation by marmoset liver microsomes was efficiently mediated by liver microsomal P450 3A enzymes significantly correlated to midazolam 1'-hydroxylation and was strongly suppressed by ketoconazole (different from the cases of human liver microsomes). It was suggested that the roles of liver microsomal P450 3A enzymes for R- and S-metoprolol O-demethylation were different between humans and marmosets. In terms of R- and S-metoprolol O-demethylation activities by marmoset liver microsomes, the metabolite formation rate per milligram of protein was high compared with those of human liver microsomes (Fig. 1), possibly accounted for by marmoset P450 2D and 3A enzymes, which contributed equally at low substrate concentrations, as is evident from the correlation analysis (Supplemental Figs. 2 and 3). In this study, metabolite formation from R- and S-metoprolol in vitro elucidated P450 2D-dependent enantioselective metabolism in marmoset

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**TABLE 1**

Kinetic analyses for rates of O-demethylation and \( \alpha \)-hydroxylation of R- and S-metoprolol by liver microsomes and recombinant P450s.

<table>
<thead>
<tr>
<th>Enzymes</th>
<th>Reaction</th>
<th>R-Metoprolol</th>
<th>S-Metoprolol</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>( K_m )</td>
<td>( V_{\text{max}} )</td>
<td>( V_{\text{max}}/K_m )</td>
</tr>
<tr>
<td>Liver microsomes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Human</td>
<td>O-Demethylation</td>
<td>15 ± 7 (( K_m ))</td>
<td>0.15 ± 0.06 (( V_{\text{max}} ))</td>
</tr>
<tr>
<td></td>
<td></td>
<td>190 ± 70 (( K_m ))</td>
<td>0.48 ± 0.04 (( V_{\text{max}} ))</td>
</tr>
<tr>
<td>Marmoset</td>
<td>( \alpha )-Hydroxylation</td>
<td>37 ± 3</td>
<td>0.054 ± 0.001</td>
</tr>
<tr>
<td></td>
<td>O-Demethylation</td>
<td>14 ± 6 (( K_m ))</td>
<td>0.63 ± 0.28 (( V_{\text{max}} ))</td>
</tr>
<tr>
<td></td>
<td></td>
<td>120 ± 99 (( K_m ))</td>
<td>0.59 ± 0.24 (( V_{\text{max}} ))</td>
</tr>
</tbody>
</table>

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\( R \) and S-metoprolol (1.0–500 \( \mu \)M) were incubated with pooled liver microsomes (0.20 mg/ml) at 37°C for 15 minutes. Kinetic parameters were calculated from a fitted curve by nonlinear regression (mean ± S.E.) with Michaelis-Menten equations: \( v = \frac{V_{\text{max}} [S]}{K_m + [S]} \). Units of enzyme activities for liver microsomes and recombinant P450 proteins are nmol/min/mg protein and nmol/min/mmol P450, respectively. Units of \( V_{\text{max}}/K_m \) for tissue microsomes and recombinant P450 proteins are ml/min/mg protein and ml/min/mmol P450, respectively. NA, not available.

*\( \alpha \)-Catalytic activity at a substrate concentration of 500 \( \mu \)M.
metabolites separately in marmosets in vivo would be of great interest.

Similarly, regio- and stereo-selective oxidation of a classic \( \beta \)-blocker, propranolol, in human, cynomolgus monkey, and marmoset livers has been investigated (Narimatsu et al., 2011). Propranolol enantiomers were \( S \)-enantiomer stereo-selectively oxidized by human P450 2D6, cynomolgus monkey P450 2D6 (2D17), and marmoset P450 2D6 enzymes mainly into 4-hydroxypropranolol, followed by 5-hydroxypropranolol and \( N \)-desisopropylpropranolol, but cytochromes P450 2D6 (2D17) enzymes had stronger high \( N \)-desisopropylpropranolol formation activity among these three prime P450 2D enzymes. These results, together with the results in the present study, suggest similar regio- and stereo-selectivity of human and marmoset P450 2D6 enzymes.

Metoprolol pharmacokinetics in humans has been influenced by human P450 2D6 genotype groups (Blake et al., 2013). There was a significant difference with nearly 40% lower \( R \)- than \( S \)-metoprolol concentrations in plasma between ultra-rapid and extensive metabolizers having two active P450 2D6 alleles and two P450 2D6-null alleles (Blake et al., 2013). Marmoset P450 enzymes show high sequence homology to their human counterparts (Uno et al., 2016), and genetic polymorphisms have recently been found in the marmoset P450 2C19 gene (Uehara et al., 2015b, 2016). However, we have not found marmoset P450 2D6 non-synonymous variants even after screening the genomes of nearly 80 individual marmosets by direct sequence analysis (S. Uehara, unpublished data). If marmoset P450 2D6 variants might change the properties of enzymes like marmoset P450 2C19 (Uehara et al., 2015b, 2016), marmosets harboring genetic variants would be suitable models for enantioreselective drug metabolism associated with the polymorphic P450 2D enzyme in livers.

In conclusion, metoprolol oxidation activities were evaluated with respect to regio- and stereo-selective \( R \)- and \( S \)-metoprolol \( O \)-demethylation in humans and marmosets. The high affinities of human and marmoset P450 2D6 enzymes with respect to regio- and stereo-selective metoprolol oxidation were similar to those of human P450 2D6, but the minor roles of P450 2D3 enzymes were unique to marmosets. Therefore, marmosets may be a good model for P450 2D-dependent regio- and enantio-selective drug metabolism in preclinical studies.

**Authorship Contributions**

**Participated in research design:** Uehara, Uno, and Yamazaki.

**Conducted experiments:** Uehara and Ishii.

**Contributed new reagents or analytic tools:** Inoue and Sasaki.

**Performed data analysis:** Uehara, Ishii, and Yamazaki.

**Wrote or contributed to the writing of the manuscript:** Uehara, Uno, and Yamazaki.

**References**


Supplemental Data

Regio- and stereo-selective oxidation of a cardiovascular drug metoprolol mediated by cytochrome P450 2D and 3A enzymes in marmoset livers

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Drug Metabolism and Disposition

Supplemental Methods

Chemicals and enzymes

*R* and *S*-metoprolol and *α*-hydroxymetoprolol were purchased from Santa Cruz Biotechnology (Santa Cruz, CA); bufuralol and *O*-desmethylmetoprolol from Toronto Research Chemicals (Toronto, ON, Canada); and ketoconazole, ticlopidine, quinidine, and N,N,N′-triethylenetriphosphoramide (ThioTEPA) from Sigma-Aldrich (St. Louis, MO). Montelukast and *α*-naphthoflavone were purchased from Tokyo Kasei (Tokyo, Japan) and Wako Pure Chemicals (Osaka, Japan), respectively. Recombinant marmoset P450s were prepared by heterologous expression systems in *Escherichia coli* as described previously (Uehara et al., 2016a; Uehara et al., 2016b; Uehara et al., 2017). Recombinant human P450s expressed in baculovirus-infected insect cells were purchased from Corning Life Sciences. All other reagents used were the highest quality commercially available.

Analysis of *R*- and *S*-metoprolol oxidation

Activities of metoprolol *O*-demethylation and *α*-hydroxylation by liver microsomes and recombinant P450s were measured as described previously (Uehara et al., 2015a) with some minor
modifications. Briefly, a typical incubation mixture (0.25 mL) contained recombinant P450 protein (20 pmol equivalents/mL) or liver microsomes (0.20 mg/mL), an NADPH-generating system (0.25 mM NADP⁺, 2.5 mM glucose 6-phosphate, and 0.25 U/mL dehydrogenase), and 1.0-500 μM R- and S-metoprolol in 100 mM potassium phosphate buffer (pH7.4). Effects of α-naphthoflavone (human P450 1A inhibitor), ThioTEPA (human P450 2B6 inhibitor), montelukast (human P450 2C8 inhibitor), ticlopidine (human P450 2C19 inhibitor), quinidine (human P450 2D6 inhibitor), and ketoconazole (human P450 3A inhibitor) were investigated at a substrate concentration of 10 μM metoprolol. The reaction was started by adding the NADPH generating system, incubated at 37°C for 15 minutes, and terminated by adding 25 μL of 60% HClO₄ (w/v). Reaction mixtures were centrifuged at 10,000g for 5 minutes; supernatants were analyzed directly by a Prominence-I LC-2030C high-performance liquid chromatography system with fluorescence (Shimadzu, Kyoto, Japan) at an excitation wavelength of 222 nm and an emission wavelength of 300 nm. Samples were injected into a reversed-phase C₁₈ column (Mightysil RP-18 GP, 5 μm, 250 × 4.6 mm; Kanto Chemical, Tokyo, Japan) with elution by 20% acetonitrile in 20 mM NaClO₄ (pH2.5) at 1.0 mL/minute. The retention times of α-hydroxymetoprolol, O-desmethyldemetoprolol, and metoprolol were 5.1, 5.5, and 17.1 min, respectively, under the present conditions. Bufuralol 1'-hydroxylation and midazolam 1'-hydroxylation activities by liver microsomes were measured as described previously (Uehara et al., 2015b). The quantitation of liver microsomal P450 2D and 3A proteins was performed by immunoblotting in a previous study (Uehara et al., 2017).

References


Supplemental Fig. 1.

R- and S-metoprolol O-demethylation (A, B) and α-hydroxylation (C, D) activities by recombinant P450s. R- (closed) and S- (open) metoprolol (at substrate concentrations of 10 μM) were incubated with human recombinant P450s (expressed in insect cell microsomes; A, C) and marmoset P450s (expressed in E. coli membranes; B, D) in triplicate determinations.
Supplemental Fig. 2.

Relationship between $R$- and $S$-metoprolol $O$-demethylation activities and P450 2D/3A probe activities in individual liver microsomes from humans (A, C) and marmosets (B, D). $R$- (closed, solid line) and $S$- (open, dashed line) metoprolol (at substrate concentrations of 1.0 μM, circles; 10 μM, triangles; and 100 μM, squares) were incubated with liver microsomes from 17 individual humans and 23 individual marmosets. Bufuralol and midazolam hydroxylation activities were determined at substrate concentrations of 1.0 and 5.0 μM, respectively. Pearson’s product moment correlation coefficients were evaluated by $t$-test; *** and **, $p < 0.001$ and $p < 0.01$, respectively.
Supplemental Fig. 3.

Relationship between $R$- and $S$-metoprolol $O$-demethylation activities and P450 2D (A) and 3A (B) contents in individual liver microsomes from marmosets. $R$- (closed, solid line) and $S$- (open, dashed line) metoprolol (at substrate concentrations of 1.0 $\mu$M, circles; and 100 $\mu$M, squares) were incubated with liver microsomes from 23 individual marmosets. Pearson’s product moment correlation coefficients were evaluated by $t$-test; ***, **, and *, $p < 0.001$, $p < 0.01$, and $p < 0.05$, respectively.
Supplemental Fig. 4.

Effects of P450 effectors on R- and S-metoprolol O-demethylation activities in liver microsomes from humans (A, C) and marmosets (B, D). Pooled liver microsomes (0.20 mg protein/mL) were incubated with metoprolol (10 μM) in the absence (control) or presence of P450 effectors. R- and S-metoprolol O-demethylation activities in pooled liver microsomes from humans (74 males and 76 females) without effectors (control) were 0.039 and 0.024 nmol/min/mg protein, respectively, whereas R- and S-metoprolol O-demethylation activities in pooled liver microsomes from marmosets (14 males and 9 females) without effectors (control) was 0.14 and 0.12 nmol/min/mg protein, respectively. The numbers (1, 2, 10, and 20) indicate P450 effector concentrations co-incubated with the substrate for 15 min. Each bar represents the mean in triplicate determinations.
Supplemental Fig. 5.

Enzyme kinetics for R- and S-metoprolol O-demethylation by recombinant marmoset and human P450 2D/3A enzymes and liver microsomes from marmosets and humans. Kinetic analyses were performed for R- (closed) and S- (open) metoprolol O-demethylation by pooled liver microsomes from humans (A) and marmosets (B) and recombinant human P450 2D6 (C), marmoset P450 2D enzymes (D; P450 2D6, circles; P450 2D8, triangles), human P450 3A enzymes (E; P450 3A4, circles; P450 3A5, triangles), and marmoset P450 3A enzymes (F; P450 3A4, circles; P450 3A90, triangles). Each data represents the mean of duplicate determinations.