

Quantitative Analysis of Complex Drug-Drug Interactions between Cerivastatin and Metabolism/Transport Inhibitors Using Physiologically Based Pharmacokinetic Modeling[§]

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ABSTRACT

Cerivastatin (CER) was withdrawn from the world market because of lethal rhabdomyolysis. Coadministrations of CER and cyclosporine A (CsA) or gemfibrozil (GEM) have been reported to increase the CER blood concentration. CsA is an inhibitor of organic anion transporting polypeptide (OATP)1B1 and CYP3A4, and GEM and its glucuronide (GEM-glu) inhibit OATP1B1 and CYP2C8. The purpose of this study was to describe the transporter-/enzyme-mediated drug-drug interactions (DDIs) of CER with CsA or GEM based on unified physiologically based pharmacokinetic (PBPK) models and to investigate whether the DDIs can be quantitatively analyzed by a bottom-up approach. Initially, the PBPK models for CER and GEM/GEM-glu were constructed based on the previously reported standard protocols. Next, the drug-dependent parameters were optimized by Cluster Newton Method. Thus, described

concentration-time profiles for CER and GEM/GEM-glu agreed well with the clinically observed data. The DDIs were then simulated using the established PBPK models with previously obtained *in vitro* inhibition constants of CsA or GEM/GEM-glu against the OATP1B1 and cytochrome P450s. DDIs with the inhibitors were underestimated compared with observed data using the geometric means of reported values. To search for better described parameters within the range of *in vitro* values, sensitivity analyses were performed for DDIs of CER. Using the *in vitro* parameter sets selected by sensitivity analyses, these DDIs were well reproduced, indicating that the present PBPK models were able to describe adequately the clinical DDIs based on a bottom-up approach. The approaches in this study would be applicable to the prediction of other DDIs involving both transporters and metabolic enzymes.

Introduction

Cerivastatin (CER), a potent 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitor, was withdrawn from the world market because of the serious side effect of rhabdomyolysis (Wooltorton, 2001). In almost all lethal rhabdomyolysis cases, CER had been used with the lipid-lowering fibrate gemfibrozil (GEM) or

prescribed at the highest therapeutic dose of 0.8 mg/day (Furberg and Pitt, 2001). Coadministration of GEM was reported to increase the CER blood concentrations (Backman et al., 2002). It has been reported that the risk of myopathy during treatment with statins depends on their dose or concentrations (Brewer, 2003). Given this, it has been widely speculated that the fatal rhabdomyolysis might be due in part to the increase in systemic exposure to CER, leading to its increase in muscle exposure.

CER is actively transported into the liver by organic anion transporting polypeptide (OATP)1Bs and then metabolized by two different P450 isoforms, CYP3A4 and CYP2C8 (Mück, 2000; Shitara et al., 2003;

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ABBREVIATIONS: AUC, area under the blood concentration-time curve; β , ratio of hepatic and biliary excretion clearance to the sum of hepatic and biliary excretion clearance and passive diffusional clearance from the hepatocyte to the liver blood; CER, cerivastatin; C_h , drug concentrations in the liver; CL_{int} , sum of intrinsic clearance of biliary excretion, hepatic metabolism, and glucuronidation; $CL_{int,all}$, hepatic overall intrinsic clearance; $CL_{int,bile}$, intrinsic clearance of biliary excretion; CL_{glu} , intrinsic clearance of glucuronidation; $CL_{int,met}$, intrinsic clearance of hepatic metabolism; CL_r , renal clearance; CL_{tot} , total clearance; CNM, Cluster Newton Method; CsA, cyclosporine A; C_{tissue} , drug concentrations in tissue; DDIs, drug-drug interactions; DIN, a drug interaction number; EH, hepatic extracellular space; EHC, enterohepatic circulation; F_aF_g , intestinal availability; f_b , protein unbound fraction in blood; f_{bile} , contribution of biliary excretion to hepatic elimination; f_{glu} , contribution of glucuronidation to hepatic elimination; F_h , hepatic availability; f_n , hepatic protein unbound fraction; FLV, fluvastatin; $f_{m,CYP2C8}$, fraction metabolized by CYP2C8 of CER; f_p , protein unbound fraction in plasma; γ , ratio of influx intrinsic clearance by passive diffusion relative to efflux intrinsic clearance by passive diffusion through sinusoidal membrane; GEM, gemfibrozil; GEM-glu, gemfibrozil 1-O- β -glucuronide; HC, hepatocytes; HMG-CoA, 3-hydroxy-3-methylglutaryl coenzyme A; k_a , absorption rate constant; k_{bile} , biliary excretion rate constant; k_{deconj} , deconjugation rate constant of gemfibrozil 1-O- β -glucuronide; $k_{deg,CYP2C8}$, degradation rate constant of CYP2C8; k_f , fecal elimination rate constant; K_i , inhibition constant; $K_{i,app,CYP2C8}$, apparent dissociation constant between CYP2C8 and the inhibitor; k_{inact} , maximal inactivation rate constant; K_p , tissue to blood concentration ratio; $k_{transit}$, transit rate constant for intestinal compartment; OATP, organic anion transporting polypeptide; P450, cytochrome P450; PBPK, physiologically based pharmacokinetics; Pgp, prostaglandin P; PS_{act} , active uptake intrinsic clearance on sinusoidal membrane; $PS_{dif,eff}$, efflux intrinsic clearance by passive diffusion through sinusoidal membrane; PS_{inf} , sum of influx intrinsic clearance through sinusoidal membrane; $PS_{dif,inf}$, influx intrinsic clearance by passive diffusion through sinusoidal membrane; PTV, pitavastatin; $RE_{act,CYP2C8}$, ratio of the active enzyme relative to total CYP2C8; WSS, weighted sum of squares.

Shitara and Sugiyama, 2006). It was previously reported that CER concentrations were increased 2.5- to 5.0-fold by the concomitant use of GEM, as mentioned above, and by the treatment with the immunosuppressive agent cyclosporine A (CsA) in kidney transplant patients (Mück et al., 1999; Backman et al., 2002). CsA markedly inhibits OATP1B1-mediated hepatic uptake and, to a smaller extent, CYP3A4. GEM and its metabolite, GEM-glucuronide (GEM-glu), also moderately inhibit OATP1B1. Furthermore, GEM-glu irreversibly inhibits CYP2C8-mediated metabolism (Shitara et al., 2003; Ogilvie et al., 2006; Varma et al., 2015). Thus, the pharmacokinetic alteration under DDIs of CER is suggested to be caused by both transporter- and P450-mediated inhibition. Figure 1A shows the qualitative DDI mechanisms of CER with CsA or GEM in the liver.

Quantitative analyses of such complex DDIs, as in the case for CER, are challenging but essential for adequate assessment of the clinical use of drugs, particularly regarding safety and effectiveness. Regulatory agencies strongly recommend the evaluation of DDI potential with a mechanistic rationale for sponsors during drug development. In the regulatory guidelines/guidance on DDIs, the usefulness of the PBPK modeling approach is mentioned as a powerful tool to guide rationalized design of DDI studies and to quantitatively predict the potential of DDIs (<https://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm292362.pdf>; http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2012/07/WC500129606.pdf; <https://www.pmda.go.jp/files/000206158.pdf>). On the other hand, the regulatory authorities also document that the robustness of PBPK models should be evaluated by performing verification and modification since the choice of structural models and availability of drug-dependent parameters are key issues in these analyses (<https://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm531207.pdf>; http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2016/07/WC500211315.pdf).

Recently, our group proposed a standard scheme of PBPK analyses to predict quantitatively the DDIs primarily caused by the OATPs-mediated hepatic uptake (Yoshikado et al., 2016). In that study, similar inhibition constants (K_i) against OATP1Bs of inhibitors CsA or rifampicin were obtained regardless of the substrates, pitavastatin (PTV), or fluvastatin (FLV). In addition, the obtained *in vivo* K_i values of CsA against OATP1Bs were comparable with *in vitro* K_i (or IC_{50}) values in the presence of CsA preincubation. These results suggested the validity of the PBPK models of these drugs and the effectiveness of the drug-dependent parameter optimization scheme for predicting other DDIs involving these drugs.

The purpose of this study was to construct unified PBPK models to analyze transporter- and enzyme-mediated complex DDIs of CER with CsA or GEM based on the standard protocol. Because there are some unknown parameters whose initial values were difficult to set directly in the models because of limited information, the PK parameters of CER or GEM were estimated by a parameter estimation algorithm called the Cluster Newton Method (CNM) (Aoki et al., 2014). The CNM has the following advantages: it requires only the designation of relatively broad parameter ranges as an initial setting, whereas the conventional algorithm, such as the Levenberg-Marquardt method, requires identification of feasible initial parameters, and the results are obtained as multiple sets of optimized parameters. Previous studies have reported that CNM has been successfully applied in analyzing complex DDIs using PBPK models (Yoshida et al., 2013; Toshimoto et al., 2017).

Moreover, to investigate whether DDIs of CER can be quantitatively described by unified PBPK models based on a bottom-up approach, simulations for DDIs with CsA and GEM were carried out using *in vitro* values. It was reported that the *in vitro* inhibition-related values of the drugs for OATP1B1 or cytochromes P450s varied widely, possibly

affected by experimental conditions (Izumi et al., 2013, 2015). To evaluate the validity of these *in vitro* values and to select the well described parameter sets, therefore, sensitivity analyses for the values were carried out for DDIs of CER.

Materials and Methods

Construction of PBPK Models

The PBPK models were constructed to describe the blood concentration-time profiles of CER or GEM/GEM-glu according to their PK properties based on previously reported standard models with some modifications (Yoshikado et al., 2016). The model structures are shown in Fig. 1, B and D. The liver was divided into five compartments for hepatic extracellular and hepatocellular parts to reproduce the hepatic availability (F_h) of dispersion model. Rapid equilibrium in the muscle, skin, and adipose, whose distribution volume is not negligible, was also included. In addition, to represent the delay of the absorption process, an empirical transit compartment was added in connection with the intestinal compartment.

In the CER and GEM-glu case, to represent the enterohepatic circulation (EHC), three transit compartments between the compartments for hepatocytes and the intestine were incorporated. In the GEM/GEM-glu case, as a rapid equilibrium distribution of GEM in the liver could be assumed from the previous report, compartments for hepatic extracellular spaces and hepatocytes were unified into a single liver compartment (Kimoto et al., 2015). Since GEM-glu was reported to be a substrate for OATPs (1B1, 1B3, and 2B1), the GEM-glu was also assumed to be absorbed from the intestine (Kimoto et al., 2015). Moreover, glucuronidation and deconjugation were assumed to occur only in the liver and the intestine, respectively. The CsA PBPK model (Fig. 1C) and reported PK parameters were used in this study (Yoshikado et al., 2016).

Calculation of PK Parameters of CER, GEM, and GEM-glu

Based on the standard protocol that was previously proposed (Yoshikado et al., 2016), PK parameters of CER and GEM/GEM-glu were collected from the literature, *in silico* calculations, our experiments and calculation methods as shown in Supplemental Tables 1 and 3 (Davies and Morris, 1993; Mück et al., 1997; Shitara et al., 2003, 2004; Shitara and Sugiyama, 2006; Yang et al., 2008; Watanabe et al., 2009, 2011; Deguchi et al., 2011; Honkalampi et al., 2011; Kudo et al., 2013; Yoshikado et al., 2017). Some hybrid parameters ($CL_{int,all}$, β , γ , and f_{bile}) were used instead of intrinsic clearances (PS_{act} , $PS_{dif,inf}$, $PS_{dif,eff}$, $CL_{int,bile}$, and $CL_{int,met}$) in the same way as previously reported (Supplemental Tables 1 and 3; Tables 1 and 2) (Yoshikado et al., 2016). $K_{p,liver}$ of GEM was calculated based on the reported *in silico* method (Rodgers and Rowland, 2006) that uses $clogP$ and pK_a described in Scifinder Scholar (Chemical Abstracts Service, Columbus, OH). Hybrid parameters were defined as follows in eq. 1–5:

$$f_b \times CL_{int,all} = f_b \times (PS_{act} + PS_{dif,inf}) \times \frac{f_h \times CL_{int,met} + f_h \times CL_{int,bile}}{f_h \times PS_{dif,eff} + f_h \times CL_{int,met} + f_h \times CL_{int,bile}} \quad (1)$$

$$R_{dif} = \frac{PS_{dif,inf}}{PS_{act}} \quad (2)$$

$$\beta = \frac{f_h \times CL_{int,met} + f_h \times CL_{int,bile}}{f_h \times PS_{dif,eff} + f_h \times CL_{int,met} + f_h \times CL_{int,bile}} \quad (3)$$

$$\gamma = \frac{PS_{dif,inf}}{PS_{dif,eff}} \quad (4)$$

$$f_{bile} = \frac{CL_{int,bile}}{CL_{int,met} + CL_{int,bile}} \quad (5)$$

Initial ranges of unknown PK parameters for CNM optimization are shown in Tables 1 and 2. All drug-dependent parameters were calculated according to the methods reported by our group (Yoshikado et al., 2016; Kim et al., 2017; Toshimoto et al., 2017).

Parameter Optimization by the CNM. The PK parameters of CER and GEM/GEM-glu in the PBPK models were optimized by fitting to their blood concentration profiles using CNM (Mück et al., 1997; Honkalampi et al., 2011; Toshimoto et al., 2017). The optimization was carried out as described in the report by Toshimoto et al. (2017) with some modifications. Briefly, 1000 initial

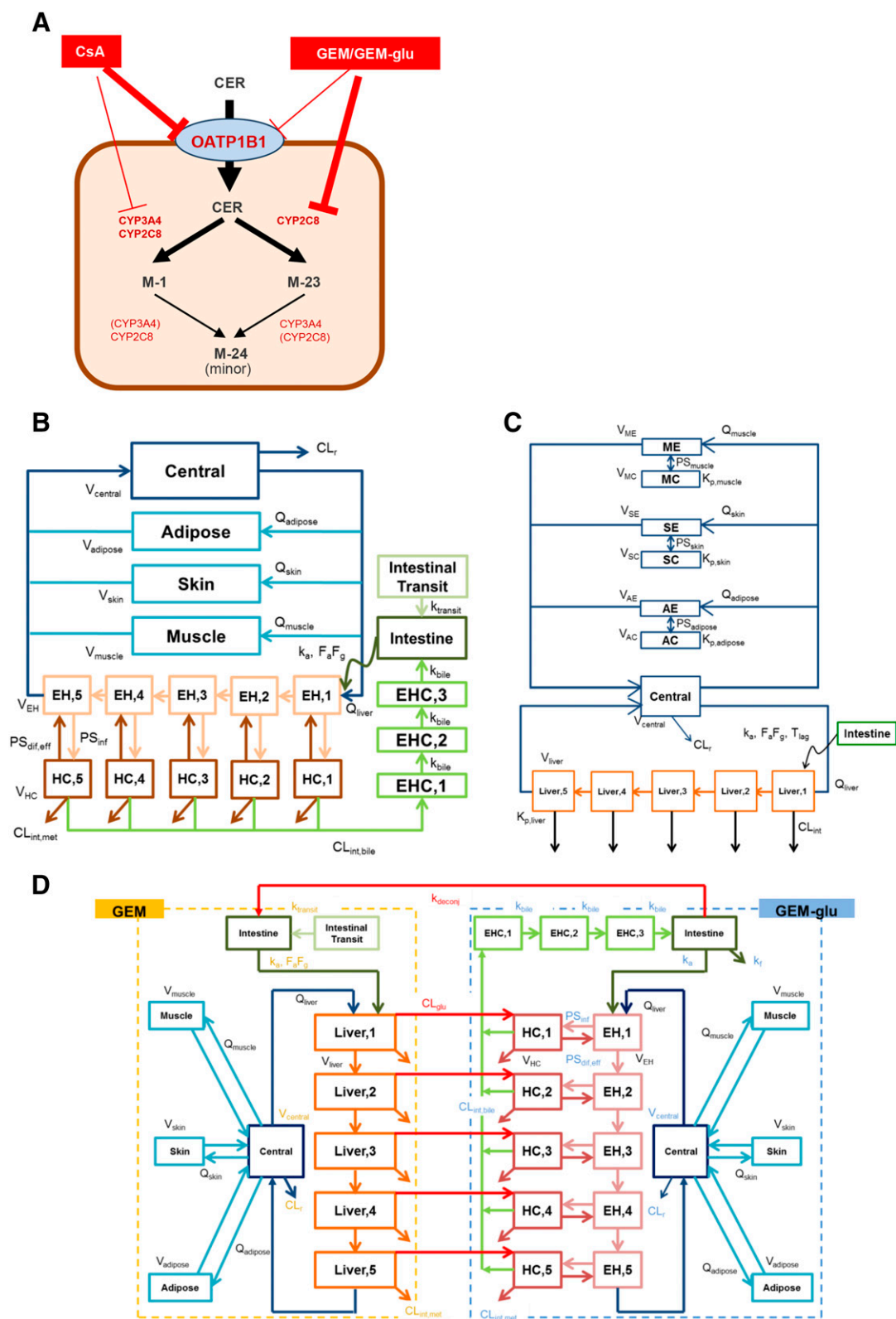


Fig. 1. Diagram of DDI in the liver (A) and the structures of PBPK models (B–D). (A) CER is transported into the liver by OATP1B1 and then metabolized by CYP2C8 and CYP3A4. Coadministration of CER and CsA, an inhibitor of OATP1B1 and CYP3A4 or GEM, inhibitors of OATP1B1 and CYP2C8 as GEM and its glucuronide (GEM-glu) increases the AUC and C_{max} of CER. (B–D) Structures of the PBPK models for CER (B), CsA (C), and GEM/GEM-glu (D). These models were based on previously reported models with some modifications (Yoshikado et al., 2016).

sets of parameters, whose ranges are described in Table 1 (CER) and Table 2 (GEM/GEM-glu), were first generated and optimized. The area under the blood concentration-time curve (AUC) value of CER after intravenous administration or

the values of GEM and GEM-glu after oral administration of GEM were used as the objective function for optimization. Procedures for parameter generation and optimization were repeated 50 times with 1000 different initial sets. Next, the

TABLE 1

Average, S.D., and coefficient of variant (CV%) of the 30 most optimized drug-dependent parameter sets of CER selected to minimize the WSS from the observed i.v. and p.o. data after the optimization of model parameters by CNM to explain the pharmacokinetic data under i.v. conditions

Parameters	Units	Initial Range	Average	S.D.	CV%
$V_{central}$	L/kg	0.075–0.75	0.092	0.039	42.5
k_a	/h	0.06–6	3.249	1.274	39.2
$k_{transit}$	/h	0.06–6	0.769	0.535	69.6
$1/\beta$		1.1–10	2.358	1.643	69.7
$f_b * CL_{int,all}$	L/h/kg	0.0372–3.72	0.638	0.033	5.2
k_{bile}	/h	0.06–6	4.029	1.082	26.9
$1/f_{bile}$		1.1–10	1.411	0.087	6.2

blood concentration-time profiles of CER and GEM/GEM-glu were simulated using the 50,000 optimized parameter sets. The weighted sum of squares (WSS) between the simulated and the observed profile was calculated as described in the following (eq. 6):

$$WSS = \sum_{i=1}^n \frac{(y_i - y'_i)^2}{y_i^2} \quad y_i: \text{ith observed value, } y'_i: \text{ith predicted value} \quad (6)$$

In the case of CER, the top 30 drug-dependent parameter sets were selected to minimize the WSS between the simulated and observed profiles after oral and intravenous administration, and the average of 30 selected parameter sets was used for subsequent DDI analyses. In the case of GEM/GEM-glu, some parameters were correlated to each other; thus, the top five drug-dependent parameter sets were selected to minimize the WSS between the simulated and observed profiles of GEM and GEM-glu after oral administration of GEM, and each parameter set was used for subsequent DDI analysis.

Parameter Optimization by Nonlinear Least-Squares Fitting

PK parameters of CER under control conditions and those of CsA in the DDI studies in the PBPK models were reoptimized by fitting to their blood concentration profiles (Mück et al., 1999; Backman et al., 2002) using the nonlinear least squares fitting software Napp (version 2.31) (Hisaka and Sugiyama, 1998). The weight for the calculation was set as 1 (the square root of the value).

Simulation of CER Concentration Profiles under DDI Conditions

In this study the competitive inhibition of CsA against OATP1B1 and CYP3A4 and that of GEM/GEM-glu against OATP1B1, as well as mechanism-based inhibition of GEM-glu against CYP2C8, were incorporated into the analyses. CER is metabolized by CYP3A4 and CYP2C8 (Mück, 2000; Shitara et al., 2003; Shitara and Sugiyama, 2006), so the sum of f_{m_CYP2C8} and f_{m_CYP3A4} was assumed to be 1. The blood concentration-time profile of CER under DDI

conditions with CsA was simulated with consideration to the inhibition of a hepatic uptake and competitive inhibition of CYP3A4 expressed by eq. 7 and 8:

Inhibition of OATP1B1-Mediated Hepatic Uptake.

$$PS_{act}(+CsA) = \frac{PS_{act}(control)}{1 + \frac{f_{b_CsA} \times C_{h_CsA}}{K_{i_OATP1B1}(CsA)}} \quad (7)$$

Inhibition of P450-Mediated Metabolism.

$$CL_{int,met}(+CsA) = CL_{int,met}(control) \times \left(f_{m_CYP2C8} + \frac{1 - f_{m_CYP2C8}}{1 + \frac{f_{b_CsA} \times C_{h_CsA}}{K_{i_CYP3A4}(CsA)}} \right) \quad (8)$$

where PS_{act} represents the intrinsic clearance for hepatic active uptake of CER from blood; $K_{p,liver_CsA}$ represents liver-to-blood concentration ratio of CsA; $CL_{int,met}$ represents the intrinsic clearance for hepatic metabolism mediated by CYP2C8 and CYP3A4 of CER; f_{b_CsA} and f_{h_CsA} represent the unbound fraction of CsA in the blood and the liver compartment, respectively; C_{h_CsA} represents the concentration of CsA in the liver compartment; f_{m_CYP2C8} represents the fraction metabolized by CYP2C8 of CER; and $K_{i_OATP1B1}(CsA)$ and $K_{i_CYP3A4}(CsA)$ represent the inhibition constant of CsA for OATP1B1-mediated hepatic uptake and CYP3A4-mediated hepatic metabolism, respectively.

Similarly, the blood concentration-time profile of CER under DDI conditions with GEM was simulated considering the inhibition of hepatic uptake by GEM and GEM-glu and a mechanism-based inhibition of CYP2C8 by GEM-glu expressed by eq. 9 and 10:

Inhibition of OATP1B1-Mediated Hepatic Uptake.

$$PS_{act}(+GEM) = \frac{PS_{act}(control)}{1 + \frac{f_{b_GEM} \times C_{h_GEM}}{K_{i_OATP1B1}(GEM)} + \frac{f_{b_glu} \times C_{EH_glu}}{K_{i_OATP1B1}(glu)}} \quad (9)$$

Inhibition of P450-Mediated Metabolism.

$$CL_{int,met}(+GEM) = CL_{int,met}(control) \times (f_{m_CYP2C8} \times RE_{act-CYP2C8} + (1 - f_{m_CYP2C8})), \quad (10)$$

where f_{b_GEM} and f_{b_glu} represent the unbound fraction in the blood of GEM and GEM-glu, respectively; $K_{p,liver_GEM}$ represents liver-to-blood concentration ratio of GEM; C_{h_GEM} represents the concentration of GEM in the liver compartment; C_{EH_glu} represents the concentration of GEM-glu in the hepatic extracellular space compartment; $RE_{act-CYP2C8}$ represents the ratio of the active enzyme relative to total CYP2C8; and $K_{i_OATP1B1}(GEM)$ and $K_{i_OATP1B1}(glu)$ represents the inhibition constant for OATP1B1-mediated hepatic uptake of GEM and GEM-glu, respectively.

TABLE 2

Optimized top five drug-dependent parameter sets of GEM/GEM-glu selected to minimize the weighted sum of squares (WSS) from the observed clinical data by Cluster Newton Method (CNM)

	Parameters	Units	Initial Range	1st	2nd	3rd	4th	5th
GEM	$V_{central}$	L/kg	0.075–0.75	0.077	0.075	0.077	0.089	0.076
	k_a	/h	0.06–6	1.463	2.574	1.611	1.275	4.733
	$k_{transit}$	/h	0.06–6	3.645	2.872	2.378	6.585	1.488
	$f_b * CL_{int,all}$	L/hr/kg	0.0105–1.05	0.157	0.173	0.165	0.154	0.199
	$1/f_{glu}$		1.1–10	1.210	1.118	1.263	1.105	1.273
GEM-glu	$V_{central}$	L/kg	0.075–0.75	0.076	0.076	0.087	0.150	0.088
	k_a	/h	0.06–6	0.258	0.109	0.106	0.155	0.142
	R_{dif}		0.0275–0.11	0.051	0.103	0.064	0.079	0.096
	$1/\beta$		1.1–10	5.727	2.119	6.318	14.039	2.779
	$f_b * CL_{int,all}$	L/hr/kg	0.0156–1.56	0.245	0.159	0.293	0.314	0.184
	k_{bile}	/h	0.06–6	0.205	0.209	2.235	0.095	3.807
	k_{deconj}	/h	0.06–6	0.856	1.179	0.073	0.189	0.824
	k_f	/h	0.06–6	0.938	1.414	0.906	0.067	2.466
WSS				1.202	1.319	1.514	1.521	1.586

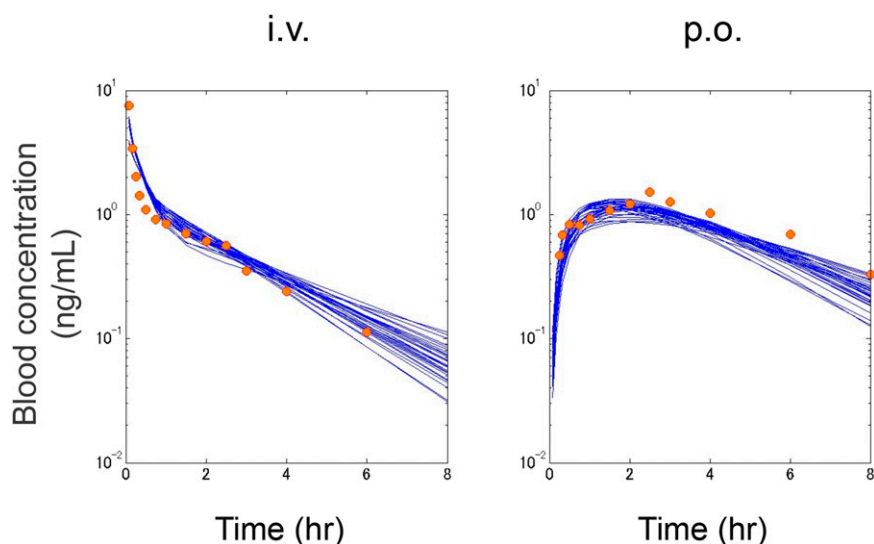


Fig. 2. Blood concentration-time profile of CER after single i.v. (0.1 mg) or p.o. (0.2 mg) administration. Fitting analysis was performed by CNM. Solid lines represent the fitted time course results using the 30 most optimized drug-dependent parameter sets selected to minimize the WSS from the observed intravenous and oral data after optimization of model parameters to explain the PK data under intravenous conditions. The closed circles represent the observed values (Mück et al., 1997). Optimized parameters are described in Table 1. Fixed pharmacokinetic parameters are described in Supplemental Table 1.

Differential equation (eq. 11) for RE_{act_CYP2C8} is expressed as follows considering that the mechanism-based inhibition by GEM-glu

$$\frac{dRE_{act_CYP2C8}}{dt} = -\frac{k_{inact_CYP2C8(glu)} \times RE_{act_CYP2C8} \times f_{h_glu} \times C_{HC_glu}}{K_{i,app_CYP2C8(glu)} + f_{h_glu} \times C_{HC_glu}} + k_{deg_CYP2C8} \times (1 - RE_{act_CYP2C8}), \quad (11)$$

where $k_{inact_CYP2C8(glu)}$ represents the maximum inactivation rate constant by GEM-glu; $K_{i,app_CYP2C8(glu)}$ represents the apparent dissociation constant between CYP2C8 and GEM-glu; C_{HC_glu} represents the concentration of GEM-glu in the hepatocytes compartment; f_{h_glu} represents the unbound fraction of GEM-glu in the hepatocytes compartment; and k_{deg_CYP2C8} represents the degradation rate constant of CYP2C8.

All in vitro inhibition constant (K_i , $K_{i,app}$, or IC_{50}) values were derived from the University of Washington's Metabolism and Transport Drug Interaction Database (<http://www.druginteractioninfo.org>) for each inhibitor against the specific enzyme/transporter in various experimental conditions (Supplemental Table 5) (Kim et al., 2017). The differential equations used in these analyses are shown in Supplemental Text.

Results

Analysis of the Blood Concentration-Time Profile of CER. To obtain the optimized drug-dependent parameters of CER in the PBPK model (Fig. 1B), its blood concentration-time profile was first fitted by CNM to the clinically observed data after single intravenous administration of CER (Mück et al., 1997). Fixed parameters used in the analysis are summarized in Supplemental Table 1. The 30 most optimized parameter sets were then selected to minimize the WSS from the observed PK data of single intravenous and oral administration of CER (Mück et al., 1997). The averages of the 30 most-optimized parameter sets are described in Table 1. As shown in Fig. 2, the reproduced concentration profiles after single intravenous and oral administration of CER agreed well with the clinically observed data (Mück et al., 1997).

Analysis of the Blood Concentration-Time Profile of CsA. To perform the DDI analysis of CER with CsA, the PK parameters, except k_a , $f_h * CL_{int}$ and dose (there is no information about dose in the literature), of CsA were reoptimized based on the previously reported PBPK model (Fig. 1C) (Yoshikado et al., 2016) with the clinically observed data of the kidney transplant patients receiving stable CsA treatment (Mück et al., 1999) using Napp. All parameters, except k_a , $f_h * CL_{int}$, and dose, were the same as previously reported (Yoshikado et al., 2016). In that study,

CsA was stably orally administered with steady state defined as unaltered CsA dosing and blood concentration of 90–120 ng/ml according to the local regimen at stable transplant function for at least 1 month, assessed by at least three blood-level measurements. Thus, the fitting analysis was performed with twice-daily administration of CsA for 6 days (under steady-state conditions). Optimized parameters and fitted concentration-time profiles are shown in Supplemental Table 2 and Supplemental Fig. 1, respectively. Estimated $f_h * CL_{int}$ was similar to initial values, within 1.5 times (Supplemental Table 2), and the reproduced concentration profile of CsA agreed well with the clinically observed data (Mück et al., 1999).

Fitting Analysis of the Blood Concentration-Time Profile of GEM/GEM-glu. To perform the DDI analysis of CER with GEM, the blood concentration-time profiles of GEM and GEM-glu were fitted by CNM based on the PBPK model (Fig. 1D) to the clinically observed PK data after single oral administration of GEM (Honkalammi et al., 2011). Fixed PK parameters used in the analysis are summarized in Supplemental Table 3. Since there are some correlated parameters and the average value cannot be used in this case, the five most-optimized drug-dependent parameter sets were selected to minimize the WSS from the observed PK data of GEM and GEM-glu (Honkalammi et al., 2011) to perform DDI analyses using each five-parameter set. Optimized parameter sets are described in Table 2. As shown in Fig. 3, the reproduced concentration profiles of GEM and GEM-glu after oral administration of GEM agreed well with the clinically observed data (Honkalammi et al., 2011). Only the best described result of GEM/GEM-glu was used in initial DDI simulation and sensitivity analyses, although final DDI simulation using the five best described drug-dependent parameter sets after sensitivity analyses were performed in each five case.

Simulation of the Blood Concentration-Time Profile of CER under DDI Conditions with CsA or GEM. To perform the DDI analysis of CER with CsA or GEM, the blood concentration-time profiles of CER under control conditions were first reproduced by reoptimization using Napp software of the values of k_a , $f_b * CL_{int,all}$, and k_{bile} based on the PBPK model with the average of the obtained 30 most optimized drug-dependent parameter sets as initial values (Fig. 1B). Optimized parameters and fitted concentration-time profiles of CER are shown in Supplemental Table 4 and Supplemental Fig. 2, respectively. Estimated values of $f_b * CL_{int,all}$ in both cases were similar to

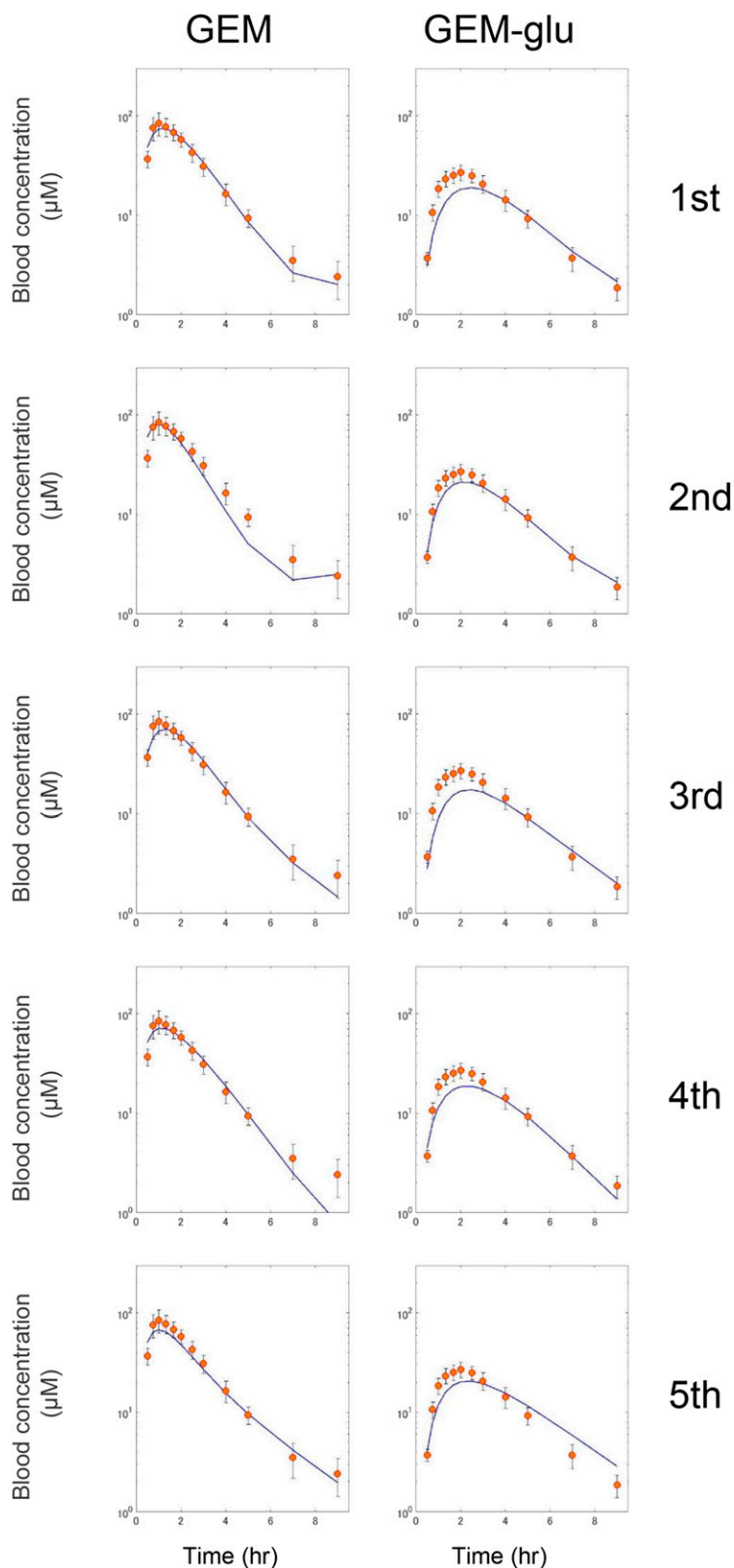


Fig. 3. Blood concentration-time profile of GEM/GEM-glu after single oral administration of GEM (600 mg). Fitting analysis was performed by the CNM. Solid lines represent the fitted time course results using the first to fifth most optimized drug-dependent parameter sets, which were selected to minimize the WSS from the observed PK data of GEM/GEM-glu. Closed circles represent the observed values (Honkalampi et al., 2011). Optimized parameters are described in Table 2. Fixed PK parameters are described in Supplemental Table 3.

initial values, within 1.2 times (Supplemental Table 4), and the obtained concentration profiles of CER under control conditions agreed well with the clinically observed data (Mück et al., 1999; Backman et al., 2002). Subsequently, the CER concentration profiles under DDI conditions with CsA or GEM were simulated, as shown in Supplemental Fig. 3,

based on the PBPK models (Figs. 1B–D) using the geometric mean of *in vitro* inhibition constants (Supplemental Table 5) (Kim et al., 2017), together with the drug-dependent parameters estimated in the present study described in Tables 1 and 2, and Supplemental Tables 2 and 4. In the case of DDI with CsA, the *in vivo* K_i value of CsA against

OATP1B1-mediated transport of PTV or FLV was reported to be comparable with in vitro K_i (or IC_{50}) values in the presence of CsA preincubation (Li et al., 2014; Yoshikado et al., 2016). In this study, therefore, in vitro K_i (or IC_{50}) values of CsA with preincubation against OATP1B1 were used for DDI analysis. Furthermore, in the case of DDI with GEM the ratio of $K_{i_OATP1B1(glu)}$ to $K_{i_OATP1B1(GEM)}$ was fixed at the ratio of the geometric means of the reported in vitro value (0.371) (Kim et al., 2017) to simplify the analysis, using the same technique as Kudo et al. (2013). As shown in Supplemental Fig. 3A, simulated C_{max} of the CER concentration profile was lower than the observed data in DDI with CsA. Similarly, as shown in Supplemental Fig. 3B, the C_{max} and $t_{1/2}$ were underestimated in DDI with GEM.

Sensitivity Analyses of the Parameters of CER and CsA Under DDI Conditions with CsA. To search for better described DDI-related parameters of CER with CsA from reported in vitro values, sensitivity analyses were carried out for $K_{i_OATP1B1}$ and K_{i_CYP3A4} of CsA under DDI condition with CsA (Supplemental Figs. 4A and B). Upper and lower limits of the range of inhibition constants were set by maximum and minimum value of the respective in vitro constant (Supplemental Table 5) (Kim et al., 2017). As shown in Supplemental Figs. 4A and B, the simulated CER concentration profile was changed by alteration of $K_{i_OATP1B1}$ but was hardly changed by K_{i_CYP3A4} . To examine a relationship between the blood/liver concentration of CsA and the in vitro K_i value, unbound blood and intrahepatic concentration-time profiles of CsA were simulated (Supplemental Fig. 4E) using CsA parameters that have been previously reported and in Supplemental Table 2. The simulated unbound CsA blood concentration was comparable to the in vitro $K_{i_OATP1B1}$, whereas the intrahepatic unbound concentration was much lower than the in vitro K_{i_CYP3A4} .

It is reported that CER is a substrate of P-glycoprotein (Pgp) (Kivistö et al., 2004) and CsA is an inhibitor of Pgp (Yoshida et al., 2012), indicating that CsA might also inhibit intestinal Pgp and that this inhibition might increase the magnitude of DDI. Yet, in these DDI analyses, intestinal inhibition of Pgp or CYP3A4 are not incorporated. Accordingly, sensitivity analyses were performed to take account of intestinal Pgp and CYP3A4 inhibition by changing $F_a F_g$ and/or k_a (in the case that $F_a F_g$ is assumed to be 1) values only in the DDI condition (Supplemental Figs. 4C and D). Results showed that the alteration of these parameters greatly affected the simulated concentration profile of CER.

Sensitivity Analyses of the Parameters of CER and GEM/GEM-glu under DDI Conditions with GEM. To search for better described DDI-related parameters of CER with GEM from reported in vitro values, sensitivity analyses were carried out for $K_{i_OATP1B1}$ of GEM/GEM-glu, $K_{i_app_CYP2C8}$, and k_{inact_CYP2C8} of GEM-glu under DDI conditions with GEM (Supplemental Figs. 5A–C). Upper and lower limits of the range of the in vitro parameters in Supplemental Figs. 5A–C were set by maximum and minimum value of the respective in vitro parameters. As shown in Supplemental Fig. 5A, the simulated CER concentration profile was changed by alteration of $K_{i_OATP1B1}$. On the other hand, CER concentration was not affected by changing the parameters related to the mechanism-based inhibition, $K_{i_app_CYP2C8}$ and k_{inact_CYP2C8} , of GEM-glu (Supplemental Figs. 5B and C). To examine a relationship between the blood concentrations of GEM/GEM-glu and the in vitro K_i value, unbound blood concentration-time profiles of GEM (black line) and GEM-glu (blue line) were simulated (Supplemental Fig. 5E) by using their parameters described in Table 2 and Supplemental Table 3. As shown in this graph, the unbound blood concentration of GEM-glu was comparable to in vitro $K_{i_OATP1B1}$. Supplemental Fig. 5F shows the fraction of active CYP2C8 simulated using the previously reported geometric mean of in vitro values ($K_{i_app_CYP2C8}$ and k_{inact_CYP2C8} of GEM-glu, and k_{deg_CYP2C8}) (Supplemental Tables 3 and 5), indicating

that the fraction of active CYP2C8 after multiple administration of GEM was quite low, less than 0.05.

The f_{m_CYP2C8} value of CER was previously estimated to be 0.61 in in vitro studies using human liver microsomes (Shitara et al., 2003, 2004). In contrast, it was already reported that the activities of metabolism by human liver microsomes greatly depended on the buffer components and their concentration and thus the in vitro f_m value is suggested to be greatly altered by buffer condition (Kudo et al., 2016). We therefore conducted a sensitivity analysis for f_{m_CYP2C8} (Supplemental Fig. 5D). An increase in $t_{1/2}$ was observed with an increase in the value of f_{m_CYP2C8} , and the simulated CER curves came closer to the clinical data with an increase in f_{m_CYP2C8} value.

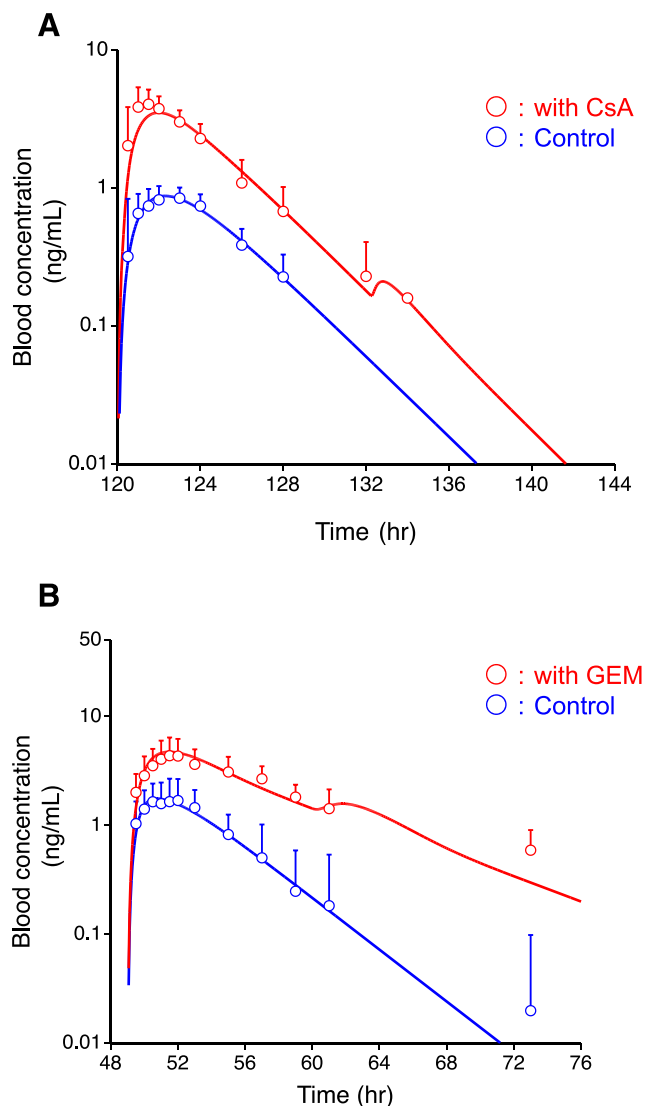


Fig. 4. Simulated blood concentration-time profile of CER under control and DDI conditions using the best described in vitro parameter sets (Table 3) after the sensitivity analyses (Supplemental Figs. 4 and 5) of model parameters to explain PK data under DDI conditions. (A) Control and DDI with CsA after single oral administration of 0.2 mg CER (111 mg of CsA was orally administered twice daily for 6 days). The solid blue and red lines represent the simulated time course under control and DDI conditions, respectively. The open circles represent the observed values (Mück et al., 1999). (B) Control and DDI with GEM after single oral administration of 0.3 mg CER (600 mg of GEM was orally administered twice daily for 3 days). The solid blue and red lines represent the simulated time course under control and DDI conditions, respectively. The open circles represent the observed values (Backman et al., 2002).

Simulation of the Blood Concentration-Time Profile of CER under Control and DDI Conditions Using the Best Described In Vitro Parameter Sets after the Sensitivity Analyses. Finally, the CER concentration profiles under control and DDI conditions with CsA (Fig. 4A) or GEM (Fig. 4B) were simulated using the best described in vitro parameter sets (Table 3) after the sensitivity analyses together with the parameters estimated in the present study described in Tables 1 and 2 and Supplemental Tables 2 and 4. As observed in these graphs, the obtained concentration profiles of CER under control and DDI conditions with CsA or GEM agreed well with the clinically observed data (Mück et al., 1999; Backman et al., 2002). In this study, sensitivity analyses for DDI between CER and GEM were performed using the most optimized parameter set of GEM/GEM-glu only, but blood concentration-time profiles of CER under control and DDI conditions with GEM were simulated using the best described in vitro parameter sets after sensitivity analyses (Table 3) with the second to fifth most-optimized parameter sets (Supplemental Fig. 6).

Discussion

CER pharmacokinetics were reported to be altered by DDI with CsA or GEM (Mück et al., 1999; Backman et al., 2002). Coadministration of CsA increased the AUC but not the elimination half-life ($t_{1/2}$) of CER, whereas GEM increased both AUC and $t_{1/2}$. In this study, we attempted to describe the transporter-/enzyme-mediated such complex DDIs of CER based on unified PBPK models as well as to investigate whether the DDIs can be quantitatively analyzed by a bottom-up approach.

After separate optimization, the PBPK models for CER and inhibitors were linked, taking into consideration the appropriate inhibitions of OATP1B1 and P450s. As shown in Supplemental Fig. 3, using the geometric means of in vitro values, simulated CER concentration profiles under DDI conditions were lower than the observed data. In contrast, after sensitivity analyses, the reproduced concentration-time profiles of CER obtained using the best described drug-dependent parameter sets (Table 3) under DDI conditions agreed well with the clinically observed data as shown in Fig. 4. These results indicate that it is quite possible to describe quantitatively the clinical DDIs based on a bottom-up approach, taking into consideration the plausible ranges of inhibition potency (e.g., from minimum to maximum of in vitro values).

In this analysis, the best described $K_{i_OATP1B1}$, with a minimum in vitro value of $0.014 \mu\text{M}$, is nearly equal to the in vivo K_i for PTV ($0.0118 \mu\text{M}$) and *FLV* ($0.00958 \mu\text{M}$), suggesting the validity of the value (Yoshikado et al., 2016). Furthermore, the CER blood concentration profile under DDI was substantially changed by taking account of

the intestinal inhibition of Pgp and CYP3A4, and the simulated CER curves came to fit better with an alteration of these values, suggesting that CsA has a high inhibitory potential against Pgp-mediated efflux and/or CYP3A4-mediated metabolism of CER in the intestine (Supplemental Figs. 4C and D). Tachibana et al. (2009) reported that a drug interaction number (DIN, defined as the ratio of inhibitor dose to inhibition constant) is useful in classifying the risk of intestinal DDI involving Pgp and CYP3A4. In that report, Pgp inhibitors with a DIN for Pgp greater than 27.91 and CYP3A4 inhibitors with a DIN for CYP3A4 greater than 9.41 were reported to have a high risk. For this CsA case, calculated DIN_{Pgp} and $\text{DIN}_{\text{CYP3A4}}$ were 54.291 ($111 \text{ mg}/1.7 \mu\text{M}$) (Yoshida et al., 2013) and 24.881 ($111 \text{ mg}/3.71 \mu\text{M}$) (Kim et al., 2017), indicating that the intestinal Pgp and CYP3A4 were strongly inhibited by CsA. Although further study using a more mechanistic model with consideration given to intestinal inhibition, such as a segregated flow model (Chow and Pang, 2013), ACAT model (Agoram et al., 2001) or ADAM model (Jamei et al., 2009a,b), is needed, these results suggest that the clinical DDI between CER and CsA was caused by inhibition of both hepatic OATP1B1 and intestinal Pgp and/or CYP3A4 by CsA.

According to Supplemental Fig. 5E, the simulated unbound blood concentration of GEM-glu was comparable to the range of in vitro $K_{i_OATP1B1(\text{glu})}$. In addition, the best described $K_{i_OATP1B1(\text{GEM})}$ and $K_{i_OATP1B1(\text{glu})}$ were minimum in vitro values of 4.00 and $1.48 \mu\text{M}$, respectively. These values closely match the in vivo K_i for repaglinide (14.2 and $5.48 \mu\text{M}$), suggesting the validity of these values (Kudo et al., 2013). The fraction of active CYP2C8 after multiple administration of GEM was less than 0.05. Furthermore, the simulated CER curves came to fit better with an increase in f_{m_CYP2C8} (Supplemental Fig. 5D). These results suggest that the in vivo DDI between CER and GEM was due mainly to inhibition of CYP2C8-mediated metabolism by GEM-glu and partly to OATP1B1-mediated uptake by GEM/GEM-glu. Moreover, they also suggest the importance of properly estimating the contributions of each P450 to total metabolism to quantitatively describe the DDI.

Our group already reported that five tanks in series model (five-liver model) was useful for the hepatic elimination of high-clearance drugs (Watanabe et al., 2009); however, it was unclear whether the five-liver model is also robust for the drugs with relatively high F_h , such as CER (Mück et al., 1997). Therefore, by using a one- or three-liver model, the concentration-time profile of CER was refitted to the clinically observed data. Then, DDI simulations between CER and CsA in one- and three-liver models were performed in the same way as the five-liver model (Supplemental Fig. 3) to examine the impact of the number of liver compartments (Supplemental Fig. 7). Optimized parameters are shown in Supplemental Tables 6–8. As shown in the figure, there is little

TABLE 3
In vitro parameter sets after sensitivity analyses

Parameters	Units	Values	Sources
DDI conditions with CsA	μM	0.014	Supplemental Fig. 4A
$K_{i_OATP1B1}$ of CsA			
K_{i_CYP3A4} of CsA	μM	3.71	Supplemental Fig. 4B
F_g of CER		0.791 (control)/1 (DDI)	Supplemental Fig. 4C
k_a of CER	/h	0.789 (control)/2.5 (DDI)	Supplemental Fig. 4D
f_{m_CYP2C8} of CER		0.85	Supplemental Fig. 5D
DDI conditions with GEM			
$K_{i_OATP1B1}$ of GEM	μM	4.00	Supplemental Fig. 5A
$K_{i_OATP1B1(\text{glu})}/K_{i_OATP1B1(\text{GEM})}$		0.371	Ratio of geometric mean of in vitro K_i (Supplemental Table 5)
$K_{i_OATP1B1}$ of GEM-glu	μM	1.48	
$K_{i_app_CYP2C8}$ of GEM-glu	μM	26.7	Supplemental Fig. 5B
k_{inact_CYP2C8} of GEM-glu	/h	4.05	Supplemental Fig. 5C
f_{m_CYP2C8} of CER		0.85	Supplemental Fig. 5D

CsA, cyclosporine A; DDI, drug-drug interactions; GEM, gemfibrozil.

difference in reproducibility among all liver models, indicating the robustness of the five-liver model.

Although the existence of the biliary excretion and/or metabolism of CER in the mouse, rat, and dog was reported previously (Boberg et al., 1998), the typical second peak was not reported in the clinical data (Mück et al., 1997, 1999; Backman et al., 2002). Thus, fitting analyses to the control data (intravenous/oral) and subsequently the DDI simulation with CsA were performed using the model without EHC in the same way as with EHC to examine whether EHC is needed (Supplemental Fig. 8). Optimized parameters are shown in Supplemental Tables 9 and 10. In these analyses, the same scaling factor for K_p values of muscle, skin, and adipose were set and optimized to agree with the distribution phase of intravenous data. As shown in Supplemental Fig. 8, fitted concentration-time profiles of the intravenous/oral data (Supplemental Fig. 8A) and control data in the DDI study (Supplemental Figs. 8B–E; blue lines) also agreed well with the clinically observed data. On the other hand, the simulated concentration-time profile under DDI conditions obtained using the best described parameter sets in the non-EHC model (Supplemental Fig. 8F) was lower than the simulated concentration-time profile obtained in the EHC model (Fig. 4A) and the observed data even after sensitivity analyses for $K_{IOATP1B1}$ and $K_{ICYP3A4}$ of CsA (Supplemental Figs. 8B and C) and $F_a F_g$ and/or k_a of CER (Supplemental Figs. 8D and E). These results indicate that the clinically observed data were not perfectly reproduced using the model without EHC, suggesting that EHC of cerivastatin in humans exists to some extent. Although further study is needed to validate the adequate number of EHC compartments in cerivastatin cases, we selected the three EHCs model in these analyses for the reason that three compartments were thought to be enough to describe the delay in transit between the hepatocyte and intestine and for consistency with our previous study.

In conclusion, the results of this study revealed that the degree of complex DDIs between CER and the inhibitors CsA and GEM, involving both transporters and metabolic enzymes, could be quantitatively described based on a bottom-up approach and some sensitivity analyses, although variabilities of reported in vitro K_i values should be always kept in mind. The PBPK models and approaches in this study would be applicable to the prediction of other DDIs involving both transporters and metabolic enzymes.

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Authorship Contributions

Participated in research design: Yao, Toshimoto, Kim, Yoshikado, Sugiyama.

Performed data analysis: Yao, Toshimoto, Kim.

Wrote or contributed to the writing of the manuscript: Yao, Toshimoto, Kim, Yoshikado, Sugiyama.

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