ABSTRACT

The cytosolic sulfotransferases (SULTs) are phase II conjugating enzymes that catalyze the transfer of a sulfonate group from the universal sulfate donor 3'-phosphoadenosine-5'-phosphosulfate to a nucleophilic group of their substrates to generate hydrophilic products. Sulfation has a major effect on the chemical and functional homeostasis of substrate chemicals. SULTs are widely expressed in metabolically active or hormonally responsive tissues, including the liver and many extrahepatic tissues. The expression of SULTs exhibits isoform-, tissue-, sex-, and development-specific regulations. SULTs display a broad range of substrates including xenobiotics and endobiotics. The expression of SULTs has been shown to be transcriptionally regulated by members of the nuclear receptor superfamily, such as the peroxisome proliferator-activated receptors, pregnane X receptor, constitutive androstane receptor, vitamin D receptor, liver X receptors, farnesoid X receptor, retinoid-related orphan receptors, estrogen-related receptors, and hepatocyte nuclear factor 4α. These nuclear receptors can be activated by numerous xenobiotics and endobiotics, such as fatty acids, bile acids, and oxysterols, many of which are substrates of SULTs. Due to their metabolism of xenobiotics and endobiotics, SULTs and their regulations are implicated in the pathogenesis of many diseases. This review is aimed to summarize the central role of major SULTs, including the SULT1 and SULT2 subfamilies, in the pathophysiology of liver and liver-related diseases.

SIGNIFICANCE STATEMENT

Sulfotransferases (SULTs) are indispensable in the homeostasis of xenobiotics and endobiotics. Knowing SULTs and their regulations are implicated in human diseases, it is hoped that genetic or pharmacological manipulations of the expression and/or activity of SULTs can be used to affect the clinical outcome of diseases.

Introduction

Sulfate conjugation (sulfation or sulfonation) is a major conjugating pathway responsible for the deactivation, detoxification, and excretion of xenobiotics and endogenous molecules (Falany, 1991). Sulfon conjugation was first recognized as an important metabolic pathway by Baumann (1876). At the chemical level, the cytosolic sulfotransferases (SULTs) catalyze the transfer of a negatively charged sulfonate group (SO3−) from the universal sulfate donor 3’-phosphoadenosine-5’-phosphosulfate (PAPS) onto a nucleophilic group of their substrates to generate hydrophilic products, which often promote the urinary excretion of the substrates. The high-energy sulfate donor PAPS can be generated by PAPS syntheses, including PAPSS1 and PAPSS2, as well as by ATP sulfurylase and two forms of adenosine 5’-phosphosulfate kinase (Mueller et al., 2018).

SULTs are widely expressed in the liver, as well as metabolically active or hormonally responsive extrahepatic tissues (Dooley et al., 2000; Gamage et al., 2006). This large family of enzymes is responsible for sulfating a variety of endogenous and exogenous molecules, including pharmaceuticals, procarcinogens, hormones, and neurotransmitters, as well as intermediates of endogenous metabolism (Dooley et al., 2000; Glatt et al., 2001; Negishi et al., 2001; Jancova et al., 2010). Sulfation often results in the inactivation of the substrates or reduced potency of ligands (Strott, 2002; Bjerregaard-Olesen et al., 2015) but with some exceptions. Their abundance in the liver and wide range of substrates suggest SULTs may act as important mediators for the development of liver diseases, such as hepatocellular carcinoma (Xie et al., 2017; Zou et al., 2017), liver fibrosis (Hardwick et al., 2013; Krattinger et al., 2016; Yetti et al., 2018), and drug-induced liver injury (Fang et al., 2016). Therefore, in this review we focus on the roles of the human and rodent SULTs in liver diseases.

ABBREVIATIONS: ABP, 4-aminoxyphenyl; APAP, acetaminophen; CF, cystic fibrosis; CFTR, cystic fibrosis transmembrane receptor; FXR, farnesoid X receptor; IGF-1, insulin-like growth factor 1; HCC, hepatocellular carcinoma; HNF4α, hepatocyte nuclear factor 4α; IKKβ, inhibitor of nuclear factor kappa-B kinase subunit β; I/R, ischemia-reperfusion; LXR, liver X receptor; NAFLD, nonalcoholic fatty liver disease; NASH, nonalcoholic steatohepatitis; PAPS, phosphoadenosine-5’-phosphosulfate; PXR, pregnane X receptor; SREBP, sterol regulatory element binding protein; SULT, sulfotransferase; WT, wild type.

Minireview

The Role of Sulfotransferases in Liver Diseases

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The Superfamily of Sulfotransferases

The SULTs superfamily contains 62 human SULT genes and 46 murine homologs as of 2016 (Mueller et al., 2015; Herrero et al., 2016). Among SULTs, the aryl-sulfotransferase (SULT1) and the hydroxysteroid sulfotransferase (SULT2) families are two principal subfamilies of SULTs that are the major contributors to the sulfonation of many xenobiotics, including pharmaceuticals and procarcinogens, and endobiotics, including steroids, thyroid, and neurotransmitters (Kauffman, 2004; Rein and Vermeulen, 2015).

The SULT1 family comprises five isoforms: phenol-sulfotransferases (SULT1A1/2) (Raftogianis et al., 1999); catecholamine phenol sulfotransferase (SULT1A3/4), which is only present in humans (Zou et al., 2017); thyroid hormone sulfotransferase (SULT1B1) (Saeki et al., 1998); iodothyronine sulfotransferase (SULT1C2/4) (Dubaisi et al., 2019); and estrogen sulfotransferase (EST/SULT1E1) (Zhang et al., 1998; Guo et al., 2015). The SULT2 family has two isoforms: alcohol/hydroxysteroid sulfotransferase (SULT2A1), which sulfonates hydroxysteroids (Mueller et al., 2018), such as androgens, estrogens at both the 3- and 17- positions (Ambadapadi et al., 2017), and bile acids (Huang et al., 2010), and SULT2B1b, which has a greater selectivity for 3-hydroxysteroids, such as cholesterol (Bi et al., 2018), but not for bile acids.

In addition to the SULT1 and SULT2 families of enzymes, the human genome contains two more sulfotransferase gene families, SULT4 and SULT6, encoding enzymes including SULT4A1, which is a brain-specific sulfotransferase associated with antipsychotic treatment response (Wang et al., 2014), and SULT6B1, whose physiologic function is largely unknown.

SULT1

SULT1A1/2

SULT1A1/2 in Liver Cancers. The human SULT1A1 and SULT1A2, also known as aryl/phenol or thermostable sulfotransferases, exhibit a broad substrate range with specificity for phenolic compounds (Raftogianis et al., 1997, 1999). SULT1A1 is widely distributed throughout the body with high abundance in the liver, lung, brain, skin, platelets, gastrointestinal tissues, and kidney (Hempel et al., 2005). The SULT1A2 gene is located on chromosome 16p11.2-12.1 in close proximity to its related isoform SULT1A1 (Gamage et al., 2006). Besides their classic role in facilitating the detoxification and excretion of their substrates and metabolites, SULT1A1 and SULT1A2 are also known to play a major role in the bioactivation of environmental mutagens and carcinogens, such as hydroxymethyl polycyclic aromatic hydrocarbons, N-hydroxy derivatives of arylamines, allylic alcohols, and heterocyclic amines, leading to mutagenicity and carcinogenesis through the binding of sulfonated metabolites to DNA (Falany, 1997; Weinshilboum et al., 1997; Hempel et al., 2005). Li et al. (2018) recently reported that the expression of liver SULT1A1/2 is highly associated with sex-dependent susceptibility of bladder and liver to the major human bladder carcinogen 4-aminobiphenyl (ABP). Both the parent ABP and its sulfonated metabolites are genotoxic (Chou et al., 1995). In this study, the authors observed that male bladders were more susceptible than female bladders to ABP. This was explained by the increased bladder exposure to ABP in male mice through androgen-dependent suppression of ABP sulfation in the liver, leading to increased bladder delivery of carcinogenic ABP (Li et al., 2018). The male preference in the bladder’s susceptibility to ABP was attenuated by knocking out the Sult1a1 gene. In contrast, female livers were more susceptible than male livers to ABP in mice, which was believed to be due to their higher liver exposure to carcinogens, including ABP and its toxic Sult1a1 metabolite, N-sulfate ester-ABP. Further studies revealed that Sult1a1 is positively associated with the increased formation of N-(deoxyguanosin-8-yl)-4-aminobiphenyl, a principal ABP-DNA adduct and the readout of tissue susceptibility to ABP, in hepatic cells. As summarized in Figure 1, androgen renders bladder more exposed to ABP in male mice by suppressing Sult-mediated ABP metabolism in liver, whereas the liver of female mice is more exposed to ABP and its carcinogenic metabolites because of the higher enzymatic activity of liver Sult1a1.

In humans, it was suggested that the most common polymorphism in SULT1A1 (Arg213His) may have accounted for variations in interindividual susceptibility to hepatocellular carcinoma (HCC) (Boccia et al., 2015; Kim et al., 2015) because SULT1A1 activates environmental mutagens and carcinogens found in well done meat (Bellanri et al., 2018), food processing contaminant (Høie et al., 2016), and dietary flavonoids (Sak and Everaus, 2016). Interestingly, SULT1A1 deactivates carcinogens in cigarette smoke, and a significant interaction between SULT1A1 and smoking was found in a case-control study that included 221 patients with HCC and 290 control subjects (Boccia et al., 2015). Specifically, up to 36% of HCC cases occurred in smokers who carried the Arg213His allele. The high incidence of HCC among Arg213His-carrying smokers was explained by the low catalytic activity of this SULT1A1 variant in detoxifying cigarette smoke carcinogens (Boccia et al., 2015).

SULT1A1 in Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis. Nonalcoholic fatty liver disease (NAFLD) encompasses a spectrum of diseases ranging from simple steatosis to nonalcoholic...
SULT1A3/4 in Liver Cancers and NAFLD

SULT1A3/4, also known as catecholamine phenol sulfotransferase, plays a key role in maintaining the homeostasis of monoamine neurotransmitters, such as dopamine, through sulfation. Unlike SULT1A1/2, which are highly expressed in the hepatocytes, the level of SULT1A3/4 is low in an adult human liver, but they are highly expressed in the fetal liver (Dubaisi et al., 2019) and tumor tissues (Yamamoto et al., 2016). A recent report confirmed that the expression of SULT1A3/4 was increased in tumor tissues compared with adjacent normal tissues of patients with HCC, and the highly expressed SULT1A3/4 in liver tumor tissues was positively associated with its increased enzymatic activity in metabolizing dopamine and elevated metastatic capacity (Zou et al., 2017). Dopamine was a substrate for SULT1A3/4, as well as a regulator responsible for SULT1A3/4 transcriptional activation (Sidharthan et al., 2013). Functional studies showed that the induced expression of SULT1A3/4 in tumor tissues of patients with HCC was negatively correlated with dopamine concentration but positively associated with the epithelial-mesenchymal transition and cancer stem cell acquisition, which ultimately promotes tumor metastasis (Zou et al., 2017).

SULT1A3 activity was reported to be decreased in steatosis, diabetic cirrhosis, and alcoholic cirrhosis liver samples compared with nonfatty control livers (Yalcin et al., 2013), which may have led to reduced sulfation of acetaminophen and opioid drugs (Bairam et al., 2018).

SULTIB1 in Liver Diseases

SULTIB1 plays an important role in the metabolism of drugs, environmental toxins, and endogenous steroids, such as androstenediones, thryoxine, l-napthol, and thyroid hormones (Fujita et al., 1997; Wang et al., 1998; Gamage et al., 2006). However, few studies have shown the changes of SULTIB1 expression or activity under different liver disease conditions. According to the Gene Expression Omnibus data base, conflicting observations in SULTIB1 gene expression were reported among different species, the same disease models of the same species under different treatments, or even the same disease models of the same species with the same treatment. As shown in Table 1, there were no consistent changes of SULTIB1 gene expression under HCC or high-fat-diet–induced NASH/NAFLD. Moreover, the lack of study of the roles of SULTIB1 in liver diseases makes it even more challenging to interpret the pathophysiological significance of the altered expression of SULTIB1 in liver diseases.

SULTIE1

SULTIE1 in Liver Cancer. Estrogen sulfotransferase (SULTIE1, also known as EST), widely expressed in human tissues such as the liver, lung, adipose tissue, and kidney (Barbosa et al., 2019), is best known for its function in the sulfoconjugation and deactivation of estrogen. This is because sulfonated estrogens fail to bind to the estrogen receptor and thus lose their hormonal activities (Song, 2001). As such, SULTIE1 has long been implicated in female sex hormone–related cancer, such as breast, endometrial, and ovarian cancers (Pasqualini, 2009; Mungenast et al., 2017; Sinreih et al., 2017; Xu et al., 2018). Interestingly, SULTIE1 is also reported to be associated with the occurrence of HCC in rats. In a study on inflammatory liver disease, upregulation of Sultlel in diethylnitrosamine-treated mouse livers was observed compared with the vehicle-treated livers using real-time reverse-transcription polymerase chain reaction (Lee et al., 2017). However, in the diethylnitrosamine-induced rat model of hepaticoma, the expression of SULTIE1 was decreased with the onset of hepatomas (Albrethsen et al., 2011). The causal relationship remains to be defined because the expression of SULTIE1 was increased in regenerating liver (Albrethsen et al., 2011). Moreover, the species difference and the human relevance of SULTIE1 in liver cancer need to be clarified.

SULTIE1 in NAFLD and NASH. Previous study in our laboratory found that loss of Sultle1/Est in female mice improved metabolic function with improved insulin sensitivity and reduced hepatic steatosis in ob/ob mice lacking Sultle1 (also known as the obe mice) compared with ob/ob mice (Gao et al., 2012). In an independent study, Sultle1 was suggested to play a role in sensitizing male mice to NAFLD/NASH by reducing the expression of Sultle1 in the liver of ob/ob mice lacking Sultle1/Est (Albrethsen et al., 2011). Functional studies also suggested that the hepatic expression of SULTIE1 was decreased with increasing severity of NAFLD compared with steatotic liver tissue. A limitation of this study is the absence of controls evaluated the effect of the Arg213His SULT1A1 gene polymorphism, which leads to a decreased enzymatic activity of SULT1A1, and the interaction of this polymorphism with lifestyle and dietary habits (Miele et al., 2014). Statistically significant interactions were reported for fruit intake and SULT1A1 gene, as the carriers of Arg213His SULT1A1 gene variant were at higher risk of developing NAFLD when taking high amounts of fruit. The presents of Arg213His SULT1A1 gene variant was also positively associated with grilled meat or fish intake in the development of NAFLD. In addition, many studies found that flavonoids in vegetables and fruits were inhibitors of multiple SULTs (Pai et al., 2001; Huang et al., 2009), which may have affected sensitivity to the development of NAFLD. However, there was no direct evidence linking dietary flavonoid–induced inhibition of SULT1A1 and NAFLD. On the other hand, NAFLD and its progression have a major effect on the expression and activity of multiple SULTs, such as SULT1A1, 1A3, 2A1, and 2B1 (Hardwick et al., 2013; Yalcin et al., 2013). Hardwick et al. (2013) reported an upregulation in protein and enzymatic activity of SULT1A1 in patients with simple steatosis but decreased expression of SULT1A1 in patients with NASH. However, these results are not without controversies. An independent study on normal subjects and patients with steatosis, diabetic cirrhosis, and alcoholic cirrhosis reported a significant decrease of SULT1A1 activity with increasing severity of liver disease from simple steatosis to cirrhosis (Yalcin et al., 2013). A follow-up study from the same group reported that in human liver tissues, sulfation of bisphenol A, an industrial chemical and endocrine disruptor, was substantially lower in livers from subjects with steatosis (23%), diabetic cirrhosis (16%), and cirrhosis (18%) relative to healthy livers (100%), resulting in a higher exposure of bisphenol A in patients with NAFLD (Yalcin et al., 2016). The discrepancies in reported SULT1A1 expression in steatosis might result from the differences in the stages of steatosis and from the gender and age of patients when the liver tissues were collected.

SULT1A1 in Acetaminophen-Induced Liver Injury. Acetaminophen (APAP), or Tylenol, is one of the most widely used drugs. Although the drug is safe at therapeutic doses, an overdose of APAP remains the leading cause of acute liver failure in the United States. It has been known that human SULT1A1 can catalyze APAP sulfation and facilitate its excretion from the urine. Consistent with their report that the expression and activity of SULT1A1 was increased in steatotic livers and decreased in NASH, Hardwick et al. (2013) showed that the formation of sulfate-APAP was increased in cytosolic fractions of human steatotic liver tissue compared with normal liver but reduced in NASH compared with steatotic liver tissue. A limitation of this study is that the results are largely associations. Future studies are necessary to determine whether SULT1A1 is necessary and sufficient to affect APAP hepatotoxicity.

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kappa-B kinase subunit β (IKKβ) IKKβ deficiency in both genders but aggrivated in male but not female IkbkbΔhep mice with hepatocyte IKKβ deficiency. Microarray analysis on liver tissues from male and female wild-type (WT) and IkbkbΔhep mice fed high cholesterol and saturated fat diet showed a significant upregulation of Sult1e1 gene in IkbkbΔhep mice compared with WT mice. Among the four groups, the male IkbkbΔhep mice expressed the highest level of Sult1e1 gene, which was associated with decreased plasma estradiol levels. The authors’ mechanism study indicated that the enrichment of liver X receptor (LXR) α to LXRε on the Sult1e1 gene promoter of male IkbkbΔhep mice might be responsible for their heightened sensitivity to NASH. These results suggest hepatocyte IKKβ is protective in males due at least in part to its ability to repress LXR-responsive induction of Sult1e1 (Matsushita et al., 2017).

**SULT1E1 in Cholestasis.** Few studies have focused on the role of SULT1E1 in cholestasis, but a recent study suggested that cholestasis-induced farnesoid X receptor (FXR) activation can lead to the suppression of SULT1E1 and hence impede hepatic deactivation of estrogens (Liu et al., 2018). In this study, the authors reported the positive correlation between elevated bile acids levels and estradiol levels in patients with obstructive cholestasis or primary biliary cirrhosis. By using female WT and Fxr–/– mice, the authors went on to show that cholestasis-induced accumulation of estradiol was associated with a downregulation of liver SULT1E1 expression in an FXR-dependent manner. Mechanistic studies using human hepatoma Huh7 cells showed that bile acid activation of FXR repressed SULT1E1 by inhibiting the positive regulation of SULT1E1 by hepatocyte nuclear factor 4α (HNF4α) (Liu et al., 2018).

**SULT1E1 in Liver Injury Induced by Sepsis and Ischemia/Reperfusion.** SULT1E1 is relatively thoroughly studied in inflammation-based conditions. Sepsis, resulting from the host’s deleterious systemic inflammatory response to microbial infections, is a major cause of mortality in the intensive care unit. Although sepsis and its associated inflammation are known to decrease the expression and activity of many drug-metabolizing enzymes, we observed a major induction of Sult1e1 and compromised estrogen activity in the liver of mice subjected to the bacterial lipopolysaccharide or cecum ligation and puncture models of sepsis. The inflammatory induction of Sult1e1 gene by sepsis was mediated by nuclear factor κB. Reciprocally, the expression and activity of Sult1e1 can impact the clinical outcome of sepsis. Specifically, ablation of the Sult1e1 gene or pharmacological inhibition of the Sult1e1 enzyme by Triclosan sensitized mice to sepsis-induced death in an estrogen-dependent manner. The increased sepsis-induced lethality in Sult1e1 knockout mice was explained to be due to increased estrogen activity and the resultant attenuated sepsis-induced prosurvival inflammatory response (Chai et al., 2015c).

Hepatic ischemia-reperfusion (I/R) injury is a major cause of liver damage. The pathogenesis of hepatic I/R injury is a dynamic process including the deprivation of blood and oxygen supply during the ischemic phase, followed by their restoration during the reperfusion phase, which is associated with oxidative stress and inflammation. An induction of Sult1e1 in rats subjected to I/R was reported as early as 2006 but without a clearly defined mechanism or understanding of the biologic significance (Svetlov et al., 2006). More recently, we reported a systemic analysis of the effect of liver I/R on the expression of Sult1e1. We showed that the expression of Sult1e1 was markedly induced by I/R in the mouse liver. The ablation of Sult1e1 protected female mice from the injury in an estrogen-dependent manner but heightened liver injury in male mice in an androgen-dependent manner. Further mechanism study established Sult1e1 as a direct transcriptional target of nuclear factor erythroid 2–related factor, a key transcriptional factor responsible for the activation of an array of genes to adapt the cells to hypoxic or oxidative damages. Based on these results, we proposed that inhibition of SULT1E1, at least in females, may represent an effective approach to manage hepatic I/R injury (Guo et al., 2015).

**SULT1E1 in Cystic Fibrosis.** Cystic fibrosis (CF), characterized by mutations of both copies of the cystic fibrosis transmembrane receptor (CFTR) gene, is an inherited disorder that causes progressive and eventually fatal damage to the lungs, digestive system, and other organs in the body. Liver is an organ that can be affected by CF. A series of papers published from the laboratory of Charles Falany suggested that the hepatic SULT1E1 may play a role in the progression of liver damage in patients with CF (Li and Falany, 2007; He et al., 2008; Falany et al., 2009). Specifically, elevated hepatic SULT1E1 activity was observed in mouse models of CF (Li and Falany, 2007) and HepG2 cells cocultured with human MMNK-1 cholangiocytes with repressed CFTR (He et al., 2008). The induction of SULT1E1 in CFTR-deficient MMNK-1 cells/ HepG2 cells coculture system was found to be dependent on the activation of LXR, and SULT1E1 induction led to alterations in the expression of estrogen responsive genes, including insulin-like growth factor 1 (IGF-1), glutathione S-transferase P1 (GST-P1), and carbonic anhydrase II, due to decreased estradiol levels. These results suggest that the induction of hepatic SULT1E1 in patients with CF may have facilitated the development of CF liver disease (Falany et al., 2009).

**SULT2**

**SULT2A1**

**SULT2A1 in Cholestasis.** Cholestasis is an accumulation of bile acids in the liver as a result of increased bile acid production in the liver.
and/or insufficient detoxication and elimination of bile acids from the liver. SULT2A1, highly expressed in the liver and adrenal gland, mainly catalyzes the sulfation of hydroxysteroids, including bile acids (Rado- 
minska et al., 1990), hydroxysteroid dehydrogenase and its role in the detoxification of bile acids and thereby preventing cholestasis via its enzymatic activity in bile acid sulfation. The expression of SULT2A1 is transcriptionally regulated by several nuclear receptors, such as pregnane X receptor (PXR; Sonoda et al., 2002), constitutive androstane receptor (Saini et al., 2004), LXRα (Uppal et al., 2007; Ou et al., 2014), HNF4α (Fang et al., 2007), and FXR (Song et al., 2001).

The activation of SULT2A1 by PXR, constitutive androstane receptor, and LXRα was believed to play critical roles in the anticholestatic activity of these receptors in rodent models of cholestasis (Sonoda et al., 2002; Saini et al., 2004; Ou et al., 2014). In humans, it was reported that liver SULT2A1 expression is decreased in human obstructive cholestasis due to gallstone biliary obstruction (Chai et al., 2015b). In a more recent clinical study, Wunsch et al. (2015) showed that liver SULT2A1 expression is decreased in patients with primary sclerosing cholangitis, which was proposed to be due to microRNA-378a-5p-mediated inhibition of the PXR/SULT2A1 axis. Table 2 summarizes several studies suggesting that activation of SULT2A1 expression through drug treatment protects against the development of cholestasis.

**SULT2A1 in Other Liver Diseases.** Downregulations of SULT2A1 gene were found in human HCC (Huang et al., 2005) and alcoholic liver disease (Yang et al., 2019b), but not in NASH (Suga et al., 2019). An increased liver Sult2a1 gene expression was observed in choline-deficient, L-amino-acid–defined, high-fat-diet–induced mouse model of NASH (Naga et al., 2019).

**SULT2B1b**

**SULT2B1b in Hepatocellular Carcinoma.** The cholesterol sulfotransferase SULT2B1b catalyzes the sulfation of cholesterol to synthesize cholesterol sulfate. Yang et al. (2013) found that SULT2B1b expression promotes the proliferation of HCC cells, which may have contributed to the progression of HCC. Specifically, the expression of SULT2B1b was found to be higher in HCC tumor tissues than their adjacent normal tissues. Moreover, overexpression of SULT2B1b promoted the growth of the mouse HCC Hepa1-6 cells, whereas knockdown of SULT2B1b inhibited the cell growth with induced cell-cycle arrest and apoptosis via upregulating the expression of fatty acid synthase, downregulating the expression of CyclinB1, BCL2, and MYC in vitro and in vivo. A follow-up mechanistic study from the same group reported that SULT2B1b promotes the proliferation of HCC cells via the activation of the β-catenin/matrix metalloproteinase 7 pathway in hepatocyes and therefore enhances epithelial–mesenchymal transition (Yang et al., 2019a).

**SULT2B1b in NAFLD and Metabolic Liver Disease.** SULT2B1b is also reported to play an important role in NAFLD. Overexpression of SULT2B1b decreased serum and hepatic lipids in mouse models of NAFLD via the suppression of LXR agonist 25-hydroxycholesterol, significantly increased the formation of sulfated metabolite of 25-hydroxycholesterol in the liver tissue and decreased serum and hepatic lipid levels, including triglycerides, total cholesterol, free cholesterol, and free fatty acids, as compared with the control group both in WT and Ldlr<sup>−/−</sup> mice. SULT2B1b may have inhibited steatosis by sulfonation and deactivation of oxysterols, the endogenous agonists for the lipogenic nuclear receptor LXR. Indeed, gene expression analysis showed that overexpression of SULT2B1b was accompanied by the reduced expression of LXR target lipogenic genes, such as SREBP-1c, SREBP-2, acetyl-CoA carboxylase-1, and fatty acid synthase (Bai et al., 2012). The inhibition of NAFLD was also observed in transgenic mice that overexpress SULT2B1b in the liver (Shi et al., 2014).

In addition to inhibiting lipogenesis, SULT2B1b and its metabolic byproduct cholesterol sulfate exhibit major activity in inhibiting hepatic gluconeogenesis and relieving metabolic liver disease (Shi et al., 2014; Bi et al., 2018). Initially, we reported that SULT2B1b can inhibit hepatic gluconeogenesis by suppressing the gluconeogenic activity of HNF4α (Shi et al., 2014). In this study, we found that the treatment with cholesterol sulfate or transgenic overexpression of SULT2B1b in the liver inhibited hepatic gluconeogenesis and attenuated metabolic abnormalities in both the high-fat-diet–induced obesity and ob/ob mice. At the mechanistic level, treatment with cholesterol sulfate or overexpression of SULT2B1b inhibited gluconeogenesis via the suppression of acetyl-CoA synthetase expression, leading to decreased acetylation and nuclear exclusion of HNF4α. In a subsequent study, we reported that SULT2B1b can be transcriptionally and positively regulated by HNF4α (Bi et al., 2018), an observation that led to our hypothesis that the transactivation of SULT2B1b by HNF4α represents a negative feedback

### TABLE 2

SULT2A1 expression in cholestasis

<table>
<thead>
<tr>
<th>Compounds</th>
<th>Disease models</th>
<th>Species</th>
<th>Expression of Sult2a1</th>
<th>Mediators</th>
<th>Clinical outcome</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yangonin</td>
<td>Estrogen-induced cholestasis</td>
<td>Mouse</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Gao et al., 2018; Dong et al., 2019</td>
</tr>
<tr>
<td>Aurapetine</td>
<td>17α-Ethinylestradiol–induced cholestasis</td>
<td>Mouse</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Wang et al., 2019</td>
</tr>
<tr>
<td>Tamoxifen</td>
<td>Cholestasis</td>
<td>Human</td>
<td>/</td>
<td>FXR</td>
<td>/ Enhancement</td>
<td>Bansal and Lau, 2019</td>
</tr>
<tr>
<td>Calculus bovis sativus</td>
<td>Estrogen-induced cholestasis</td>
<td>Rat</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Xiang et al., 2019</td>
</tr>
<tr>
<td>N-3 polyunsaturated fatty acids</td>
<td>/</td>
<td>Human cell</td>
<td>/</td>
<td>FXR</td>
<td>/ Induce BA</td>
<td>Cieslak et al., 2018</td>
</tr>
<tr>
<td>Corilagin</td>
<td>ANIT-induced cholestasis</td>
<td>Rat</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Yang et al., 2018</td>
</tr>
<tr>
<td>Doxorubicin</td>
<td>Cholic acid-induced cholestasis</td>
<td>Mouse</td>
<td>/</td>
<td>p53</td>
<td>Attenuation</td>
<td>Chen et al., 2017</td>
</tr>
<tr>
<td>Gemjepsend</td>
<td>ANIT-induced cholestasis</td>
<td>Rat</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Wang et al., 2017</td>
</tr>
<tr>
<td>Alisol B 23-acetate</td>
<td>Estrogen-induced cholestasis</td>
<td>Mouse</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Meng et al., 2015a</td>
</tr>
<tr>
<td>Alisol B 23-acetate</td>
<td>ANIT-induced cholestasis</td>
<td>Mouse</td>
<td>/</td>
<td>FXR</td>
<td>Attenuation</td>
<td>Meng et al., 2015b</td>
</tr>
<tr>
<td>Oleandric acid</td>
<td>Bile duct ligation–induced cholestasis</td>
<td>Rat</td>
<td>/</td>
<td>/</td>
<td>Attenuation</td>
<td>Chai et al., 2015a</td>
</tr>
<tr>
<td>Oleandric acid</td>
<td>Lithocholic acid–induced cholestasis</td>
<td>Mouse</td>
<td>/</td>
<td>ERR2</td>
<td>Attenuation</td>
<td>Chen et al., 2014</td>
</tr>
</tbody>
</table>

ANT, alpha-naphthylisothiocyanate; BA, bile acid; ERR2, estrogen receptor–related receptor 2.
to limit the gluconeogenic activity of HNF4α. Consistent with our hypothesis we showed that downregulation of Sult2B1b promoted the gluconeogenic activity of HNF4α as a result of decreased expression of the HNF4α deacetylase Sirt1 and increased HNF4α acetylation. Sult2b1b expression was also induced by HNF4α upon fasting. Ablation of Sult2B1b in mice led to increased gluconeogenic gene expression and an elevated fasting glucose level, suggesting that SULT2B1b plays a restrictive role in HNF4α-mediated fasting responsive gluconeogenesis. In the same study, we also designed and synthesized thiocholesterol, a hydrolysis-resistant derivative of cholesterol sulfate, which showed more superior activity than the native cholesterol sulfate in inhibiting gluconeogenesis and improving insulin sensitivity in high-fat-diet–induced diabetic mice. Based on these results, we conclude that the HNF4α–SULT2B1b–cholesterol sulfate axis represents a key endogenous mechanism to prevent uncontrolled gluconeogenesis.

**SULT2B1b in APAP-Induced Liver Injury.** In a recent study, we uncovered an unexpected role for SULT2B1b in APAP-induced liver injury. Hepatic overexpression of SULT2B1b enhanced the sensitivity of mice to APAP-induced liver injury, whereas ablation of the Sult2B1b in mice conferred resistance to the APAP hepatotoxicity (An et al., 2019). This is a somewhat surprising result considering that sulfation is generally considered to be a metabolic pathway that detoxifies APAP. Consistent with our previous finding that SULT2B1b is a transcriptional target of HNF4α, Hnf4α overexpression also sensitized mice to APAP-induced hepatotoxicity in a Sult2b1b-dependent manner, indicating that the HNF4α–SULT2B1b axis plays a unique role in APAP-induced hepatotoxicity and that SULT2B1b induction might be a risk factor for APAP toxicity (An et al., 2019).

As summarized in Figure 2, SULT2B1b plays important roles in the development of liver cancer, NAFLD, and sensitivity to APAP-induced liver injury. Accumulating evidence shows altered expression and/or regulation of SULTs in liver diseases can reciprocally alter clinical consequences of liver diseases. Sex-, tissue-, and development-specific regulations of SULTs, such as SULT1A1/2 and SULT1E1, appeared to be important factors affecting the pathogenesis of liver diseases. In most cases, the effect of the SULTs on liver diseases can be explained by the sulfonation of xenobiotics and endobiotics, which has a major impact on the chemical and functional homeostasis of these chemicals.

Despite the progress in our understanding of the role of SULTs in liver diseases, there are a number of remaining questions and challenges: 1) the identification of previously unknown pathophysiologic functions of SULT isoforms and understanding the molecular mechanisms underlying the roles of SULT beyond regulating the sulfation and deactivation of a variety of endogenous and exogenous molecules; 2) exploring SULTs and SULT-mediated signaling pathways as therapeutic targets for disease conditions such as NAFLD/NASH, sepsis- or drug-induced liver injury, and even cancers; 3) investigating the species specificity of the SULTs function, as the majority of the in vivo and mechanistic studies have been focused on rodents; human studies are necessary to determine whether the rodent results are translatable to humans; and 4) the humanization of cytochrome P450 enzymes, such as CYP3A4 (Granvil et al., 2003; Cheung et al., 2006) and 2D6 (Corchero et al., 2001) have been reported, but the humanization of SULTs is a largely unexplored territory.

**Authorship Contributions**

Wrote or contributed to the writing of the manuscript: Y. Xie, W. Xie.

**References**


regulatory element binding protein-1c signaling pathway and reduces serum and hepatic lipids in mouse models of nonalcoholic fatty liver disease. Metabolism 66:836–845.


