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Multidisciplinary Insights into the Structure–Function Relationship of the CYP2B6 Active Site^S

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ABSTRACT

Cytochrome P450 2B6 (CYP2B6) is a highly polymorphic human enzyme involved in the metabolism of many clinically relevant drugs, environmental toxins, and endogenous molecules with disparate structures. Over the last 20-plus years, *in silico* and *in vitro* studies of CYP2B6 using various ligands have provided foundational information regarding the substrate specificity and structure–function relationship of this enzyme. Approaches such as homology modeling, X-ray crystallography, molecular docking, and kinetic activity assays coupled with CYP2B6 mutagenesis have done much to characterize this originally neglected monooxygenase. However, a complete understanding of the structural details that make new chemical entities substrates of CYP2B6 is still lacking. Surprisingly little *in vitro* data has been obtained about the structure–function relationship of amino acids identified to be in the CYP2B6 active site. Since much attention has already been devoted to elucidating the function of CYP2B6 allelic variants, here we review the salient findings of *in silico* and *in vitro* studies of the CYP2B6 structure–function relationship with a deliberate focus on the active site. In addition to summarizing these complementary approaches to studying

structure–function relationships, we note gaps/challenges in existing data such as the need for more CYP2B6 crystal structures, molecular docking results with various ligands, and data coupling CYP2B6 active site mutagenesis with kinetic parameter measurement under standard expression conditions. Harnessing *in silico* and *in vitro* techniques in tandem to understand the CYP2B6 structure–function relationship will likely offer further insights into CYP2B6-mediated metabolism.

SIGNIFICANCE STATEMENT

The apparent importance of cytochrome P450 2B6 (CYP2B6) in the metabolism of various xenobiotics and endogenous molecules has grown since its discovery with many *in silico* and *in vitro* studies offering a partial description of its structure–function relationship. Determining the structure–function relationship of CYP2B6 is difficult but may be aided by thorough biochemical investigations of the CYP2B6 active site that provide a more complete pharmacological understanding of this important enzyme.

Introduction

The cytochromes P450 (P450) are a superfamily of heme-containing monooxygenases found in all kingdoms of life (Nelson, 2011). P450s participate in the metabolism of xenobiotics, such as clinically relevant drugs and environmental toxins (Guengerich, 2017), as well as endogenous compounds, including eicosanoids, vitamins, and steroid hormones (Nebert et al., 2013). As the only expressed member of the human 2B subfamily, cytochrome P450 2B6 (CYP2B6) has been the subject

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ABBREVIATIONS: BDE-47, 2,2',4,4'-tetrabromodiphenyl ether; 3-OH-BDE-47, 3-hydroxy-2,2',4,4'-tetrabromodiphenyl ether; 5-OH-BDE-47, 5-hydroxy-2,2',4,4'-tetrabromodiphenyl ether; 6-OH-BDE-47, 6-hydroxy-2,2',4,4'-tetrabromodiphenyl ether; 4'-OH-BDE-49, 4'-hydroxy-2,2',4,5'-tetrabromodiphenyl ether; BDE-99, 2,2',4,4',5-pentabromodiphenyl ether; BDE-100, 2,2',4,4',6-pentabromodiphenyl ether; 3-OH-BDE-100, 3-hydroxy-2,2',4,4',6-pentabromodiphenyl ether; 3'-OH-BDE-100, 3'-hydroxy-2,2',4,4',6-pentabromodiphenyl ether; 5'-OH-BDE-100, 5'-hydroxy-2,2',4,4',6-pentabromodiphenyl ether; 6'-OH-BDE-100, 6'-hydroxy-2,2',4,4',6-pentabromodiphenyl ether; BPA, 4-*tert*-butylphenylacetylene; 4-BP, 4-benzylpyridine; CB118, 2,3',4,4',5-pentachlorobiphenyl; 4-CPI, 4-(4-chlorophenyl)imidazole; CYP2B1, cytochrome P450 2B1; CYP2B4, cytochrome P450 2B4; CYP2B6, cytochrome P450 2B6; CYP2E1, cytochrome P450 2E1; Cyt b5, cytochrome b5; 7-EFC, 4-trifluoromethyl-7-ethoxycoumarin; 4-NBP, 4-(4-nitrobenzyl)pyridine; OCDD, 1,2,3,4,6,7,8,9-octachlorodibenzo-*p*-dioxin; P450, cytochrome P450; PBDE, polybrominated diphenyl ether; PCB, polychlorinated biphenyl; PCB-187, 2,2',3,4',5,5',6-heptachlorobiphenyl; PDB, Protein Data Bank; POA, phenoxyaniline; POR, P450 oxidoreductase; QSAR, quantitative structure–activity relationship; RP 73401 or Piclamilast, 3-cyclopentyl-*N*-(3,5-dichloro-4-pyridyl)-4-methoxybenzamide; SRS, substrate recognition site; WZ35, 1-(4-hydroxy-3-methoxyphenyl)-5-(2-nitrophenyl)penta-1,4-dien-3-1.

of experimental interest since its discovery in the late 1980s (Miles et al., 1988) due to its primary role in the metabolism of clinically important drugs (e.g., efavirenz, bupropion, methadone, ketamine, and cyclophosphamide) in addition to its extensive polymorphism (Zanger and Klein, 2013; Desta et al., 2021). As more CYP2B6 substrates were identified, there was a growing interest in characterizing *how* and *why* the active site structure of CYP2B6 functions in coordinated, substrate-specific oxidative catalysis with the eventual goal of computationally screening the metabolic profile of new chemical entities (Lewis, 1999; Lewis et al., 1999).

The computational power of predicting enzymatic structure–function relationships has grown within the last 20 years. Specifically, a surplus of *in silico* information concerning CYP2B6 such as its substrate/inhibitor characteristics (Wang and Halpert, 2002; Korhonen et al., 2007; Ekins et al., 2008; Wang et al., 2019b), 13 crystal structures (Table 1), active site amino acids (Supplemental Table 1 and Table 1), and plasticity (Shah et al., 2012; Wilderman and Halpert, 2012; Shah et al., 2018) has been elucidated. The techniques of quantitative structure–activity relationships (QSARs) (Wang and Halpert, 2002; Lewis et al., 2010; Dmitriev et al., 2021), homology modeling (Lewis et al., 1999; Bathelt et al., 2002; Lewis et al., 2002), X-ray crystallography (Gay et al., 2010; Halpert, 2011; Shah et al., 2018), and molecular docking (Niu et al., 2011; Radloff et al., 2013; Maldonado-Rojas et al., 2016) have been pivotal in this endeavor. For a full view of the significant *in silico* work done with CYP2B6, we have compiled a historical table of CYP2B6 homology models (Supplemental Table 1) and a modern table of CYP2B6 crystal structures/molecular docking results (Table 1) with predicted active site residues around various ligands noted. The *in silico* information collected through these techniques will be reviewed herein to exemplify advancements in the field of assessing the CYP2B6 structure–function relationship. However, *in vitro* activity data measuring the kinetics of CYP2B6 are also of utmost importance to these advancements.

Most studies investigating CYP2B6 kinetics have been more focused on identifying substrates of this enzyme than venturing deeper into its structure–function relationship. As a result, the *in vitro* insights reviewed here will underscore the subset of those exploring CYP2B6 mutagenesis (Domanski et al., 1999; Spatzenegger et al., 2003; Nguyen et al., 2008; Lin et al., 2016), substrate moiety modulation (Cox and Bumpus, 2014; Cox and Bumpus, 2016; Liu et al., 2016; Wang et al., 2019b), and substrate/stereo-regio-specificity (Roy et al., 1999; Totah et al., 2007; Erratico et al., 2015; Wang et al., 2018; Wang et al., 2019a). CYP2B6 mutagenesis has largely been performed to better understand the metabolic activity of allelic variants, which is thoroughly reviewed elsewhere (Zanger et al., 2007; Wang and Tompkins, 2008; Mo et al., 2009; Turpeinen and Zanger, 2012; Zanger and Klein, 2013; Naidoo et al., 2014; Langmia et al., 2021). Despite their pharmacological interest, these studies do not necessarily advance our understanding of the CYP2B6 active site and will not be explored here. Thus, reports performing mutagenesis to characterize CYP2B6 active site residues will be reviewed here for the first time. We have tabulated kinetic parameters for every published CYP2B6 active site mutant (Table 2) and the problematic expression system-dependence of CYP2B6 kinetic parameters that has permeated the *in vitro* work of the field (Table 3). Analyzing CYP2B6 substrates will be prioritized over inhibitors with the discussion of their K_m and k_{cat} (Table 2) to highlight meaningful biochemical insights into the structure–function relationship of the CYP2B6 active site.

Our current understanding of the CYP2B6 structure–function relationship has benefitted from *in silico* and *in vitro* data. Previous *in silico* work identifying six substrate recognition sites (SRSS) in P450 family 2 has been helpful to note where amino acids involved in CYP2B6 substrate binding are likely located (Gotoh, 1992), but this and further

information specific to CYP2B6 that has been elucidated *in silico* has not been thoroughly assessed *in vitro* (Table 2). Multidisciplinary studies utilizing *in silico* and *in vitro* techniques can provide hypothesis-generating information about the CYP2B6 active site and, by extension, shed further light on the CYP2B6 structure–function relationship.

In Silico Insights

Characterizing CYP2B6 Ligands. Early QSARs postulated that CYP2B6 ligands possess two (Wang and Halpert, 2002) to three (Ekins et al., 1999) hydrophobes and one (Ekins et al., 1999; Lewis, 2000; Wang and Halpert, 2002) to two (Lewis, 2000) hydrogen bond acceptor region(s). More general descriptors for CYP2B6 ligands have indicated the importance of nonplanarity, high volume, moderate molar mass, positive electrostatic potential, neutrality/weak basicity, hydrogen bond acceptor capacity, and, most importantly, high lipophilicity with a canonical “butterfly-wing conformation” (Ekins et al., 1999; Lewis, 2000; Lewis et al., 2001; Wang and Halpert, 2002; Appiah-Opong et al., 2008; Lewis et al., 2010). The hydrophobicity of CYP2B6 ligands is usually supplied by one or more aromatic rings (Lewis et al., 2002), their weak basicity coincides with the presence of protonatable nitrogen(s), and the number of hydrogen bond donors in the CYP2B6 active site is a limiting factor for the number of hydrogen bonds formed when binding ligands (Lewis et al., 2001; Lewis, 2003). The average molar mass of CYP2B6 ligands is about 250 to 300 g/mol, and, as more substrates were identified, each has about 2.5 hydrogen bond acceptors (Ekins et al., 2008). The nonplanarity of CYP2B6 ligands is evidenced by rotatable heterocyclic rings (Korhonen et al., 2007) and the ease of mapping substrates within a two-dimensional template to predict oxidation sites (Koyama and Yamazoe, 2011). While this description of CYP2B6 ligands is useful, many of these characterizations are generalizations due to the sensitivity and variability of the CYP2B6 structure–function relationship. For example, CYP2B6 has difficulty metabolizing planar molecules, yet planar dioxins have been predicted to readily bind and bioaccumulate within the enzyme (Maldonado-Rojas et al., 2016). This observation can be explained, at least in part, by the capacity of halogen atoms in dioxins, other persistent organic pollutants, and additional CYP2B6 substrates/inhibitors to promote tight ligand binding within CYP2B6 (Korhonen et al., 2007; Maldonado-Rojas et al., 2016; Wang et al., 2019b).

Crystallographic Advances with CYP2B6. For many years heterologous expression and purification of CYP2B6 was challenging due in part to the existence of the N-terminal anchor embedded in the cytoplasmic surface of the endoplasmic reticulum. Scott et al. first discovered that N-terminal truncation and mutation of CYP2B6 could facilitate increased expression and solubility in *E. coli* (Scott et al., 2001). Crystallization of CYP2B6 remained elusive until Bumpus et al. demonstrated the utility of the K262R polymorphic variant for increased solubility into cytosolic fractions (Bumpus et al., 2005). The Halpert Laboratory additionally found the Y226H mutation to further favor CYP2B6 crystallization and successfully solved the first CYP2B6 structure as an N-terminally truncated Y226H/K262R double mutant complexed with 4-(4-chlorophenyl)imidazole (4-CPI) (Gay et al., 2010). To date, 13 crystal structures of CYP2B6 in complex with 11 unique ligands above the heme prosthetic group have been solved through the utilization of N-terminal truncation and the Y226H/K262R double mutation (Table 1). Two of these structures also contain additional point mutations: I114V (Shah et al., 2017) and Y244W (Liu et al., 2016). Amino acids within 5 Å of complexed/docked ligands are canonically considered to be those pertinent for influencing the binding, orientation, and potential metabolism of the ligand (Lewis et al., 1999; Wang and Halpert, 2002; de Graaf et al., 2005; Mise et al., 2016; Hirakawa et al., 2018). Residues observed to

TABLE 1

CYP2B6 active site amino acids surrounding various ligands bound during crystallization or molecular docking experiments to results based on solved CYP2B6 crystal structures. Normal font entries correspond to results based on molecular docking utilizing the indicated CYP2B6 PDB ID. Residues **bolded** and underlined> entries correspond to results based on the bound ligand (proximal to the heme prosthetic group) were considered to be located in the active site, within 5 Å of the bound ligand.

Bound ligand PDB ID (reference)	K100	I101	V104	P106	F108	Y111	I114	F115	A116	N117	F206	I209	S210	S294	F297
Amlodipine 3UA5 (Shah et al., 2012)	X	X					X	X			X	X	X		X
4-benzylpyridine 3OOA (Shah et al., 2011)	X	X	X				X	X			X	X	X		X
Bornane 5UDA (Shah et al., 2017)	X	X			X		X	X			X	X	X	X	X
Bornyl bromide 5UAP (Shah et al., 2017)	X	X			X		X	X			X	X	X	X	X
(4)-3-sclareane 4RR1 (Shah et al., 2015)	X	X			X		X	X			X	X	X	X	X
4-(4-chlorophenyl)imidazole 3IBD (Gay et al., 2010)	X	X			X		X	X			X	X	X	X	X
Efavirenz 2- <u>4</u> -oxo-2-methyl-5 VB 6G (Shah et al., 2018)	X	X			X		X	X			X	X	X	X	X
(-)-mivrenyl bromide 5UEC (Shah et al., 2017)	X	X			X		X	X			X	X	X	X	X
(-)-mivrenyl bromide 5UFG (Shah et al., 2017)	X	X			X		X	X			X	X	X	X	X
4-(4-nitrobenzyl)pyridine 3QL8 (Shah et al., 2011)	X	X			X		X	X			X	X	X	X	X
(+)- α -pinene 4B1 (Wilderman et al., 2013)	X	X			X		X	X			X	X	X	X	X
(+)- α -pinene 4ZV8 (Liu et al., 2016)	X	X			X		X	X			X	X	X	X	X
Subinene 4RQL (Shah et al., 2015)	X	X			X		X	X			X	X	X	X	X
2,2',4,4'-tetraabromodiphenyl ether (BDE-47) 3IBD (Zhang et al., 2021)	X	X			X		X	X			X	X	X	X	X
BDE-47 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
3-hydroxy-2,2',4,4'-tetraabromodiphenyl ether (3-OH-BDE-47) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
5-hydroxy-2,2',4,4'-tetraabromodiphenyl ether (5-OH-BDE-47) 3UA5 (Maldonado-Rojas et al., 2016)	X	X	X		X		X	X			X	X	X	X	X
6-hydroxy-2,2',4,4'-tetraabromodiphenyl ether (6-OH-BDE-47) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
2,2',4,4',5'-pentaabromodiphenyl ether (BDE-99) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
2,2',4,4',6'-pentaabromodiphenyl ether (BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
3-hydroxy-2,2',4,4',6'-pentaabromodiphenyl ether (3-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
3'-hydroxy-2,2',4,4',6'-pentaabromodiphenyl ether (3'-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
5'-hydroxy-2,2',4,4',6'-pentaabromodiphenyl ether (5'-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
6'-hydroxy-2,2',4,4',6'-pentaabromodiphenyl ether (6'-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
Bromoketamine 3IBD (Wang et al., 2019b)	X	X			X		X	X			X	X	X	X	X
Bupropion 3IBD (Niu et al., 2011)	X	X			X		X	X			X	X	X	X	X
4- <i>tert</i> -butylphenylacetylene (BPA) 3IBD (Lin et al., 2011)	X	X			X		X	X			X	X	X	X	X
Cyclophosphamide 3UA5 (Lautier et al., 2016)	X	X			X		X	X			X	X	X	X	X
2,3',4',5'-penta-chlorobiphenyl (CB118) ^y 3IBD (Mise et al., 2016)	X	X			X		X	X			X	X	X	X	X
Deschloroketamine 3IBD (Wang et al., 2019b)	X	X			X		X	X			X	X	X	X	X
Elavirenz 3IBD (Niu et al., 2011)	X	X			X		X	X			X	X	X	X	X
Fluoroketamine 3IBD (Wang et al., 2019b)	X	X			X		X	X			X	X	X	X	X
Garcinol 5UFG (Bolla et al., 2021)	X	X			X		X	X			X	X	X	X	X
Herbacetin ^z 5UDA (Qian et al., 2022)	X	X			X		X	X			X	X	X	X	X
Ketamine 3IBD (Wang et al., 2019b)	X	X			X		X	X			X	X	X	X	X
Mephobarbital 3IBD (Niu et al., 2011)	X	X			X		X	X			X	X	X	X	X
1,2,3,4,6,7,8,9-octachlorodibenzo- <i>p</i> -dioxin (OCDD)	X	X			X		X	X			X	X	X	X	X
3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
2,2',3',4',5',6'-heptachlorobiphenyl (PCB-187) 3UA5 (Maldonado-Rojas et al., 2016)	X	X			X		X	X			X	X	X	X	X
Propofol 3IBD (Niu et al., 2011)	X	X			X		X	X			X	X	X	X	X
Propofol 2,6-di- <i>sec</i> -butyl phenol analog 3IBD (Niu et al., 2011)	X	X			X		X	X			X	X	X	X	X
Saquinone 3IBD (Gong et al., 2018)	X	X			X		X	X			X	X	X	X	X
Ticlopidine 3IBD (Gay et al., 2010)	X	X			X		X	X			X	X	X	X	X
Ticlopidine 5UFG (Bolla et al., 2021)	X	X			X		X	X			X	X	X	X	X
Ticlosoan 3IBD (Zhang et al., 2021)	X	X			X		X	X			X	X	X	X	X
1-(4-hydroxy-3-methoxyphenyl)-5-(2-nitrophenyl)pentan-1,4-dien-3-yl (WZ35) ^y 5UDA (Wang et al., 2020a)	X	X			X		X	X			X	X	X	X	X

TABLE 1 continued

Bound ligand PDB ID (reference)	Active site residues ^a															
	A298	G299	T300	E301	T302	L362	L363	G366	V367	P368	I370	D385	C475	V477	G478	K479
Amlodipine 3UA5 (Shah et al., 2012)	X	X	X	X	X	X	X	X	X	X			X	X		
4-benzopyridine 3OOA (Shah et al., 2011)	X				X	X	X	X	X	X			X	X		
Bornane 5UDA (Shah et al., 2017)	X				X	X	X	X	X	X			X	X		
Bornyl bromide 3UAF (Shah et al., 2017)	X				X	X	X	X	X	X			X	X		
[+]-3-carene 4RR1 (Shah et al., 2015)	X				X	X	X	X	X	X			X	X		
4-(4-chlorophenyl)imidazole 3HED (Gay et al., 2010)	X				X	X	X	X	X	X			X	X		
Efavirenz 2-tesoxo-2-methyl 5WBG (Shah et al., 2018)	X				X	X	X	X	X	X			X	X		
(-)-myrtenyl bromide 5UEC (Shah et al., 2017)	X				X	X	X	X	X	X			X	X		
(-)-myrtenyl bromide 5UFG ^b (Shah et al., 2017)	X				X	X	X	X	X	X			X	X		
4-(4-nitrobenzyl)pyridine 3OU8 (Shah et al., 2011)	X			X	X	X	X	X	X	X			X	X		
(+)-α-pinene 4I91 (Wilderman et al., 2013)	X				X	X	X	X	X	X			X	X		
(+)-α-pinene 4ZV8 ^c (Lin et al., 2016)	X				X	X	X	X	X	X			X	X		
Sabinene 3RQL (Shah et al., 2015)	X				X	X	X	X	X	X			X	X		
2,2',4,4'-tetrabromodiphenyl ether (BDE-47) 3IBD (Zhang et al., 2021)	X				X	X	X	X	X	X			X	X		
BDE-47 3UA5 (Maldonado-Rojas et al., 2016)	X				X	X	X	X	X	X			X	X		
3-hydroxy-2,2',4,4'-tetrabromodiphenyl ether (3-OH-BDE-47) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
5-hydroxy-2,2',4,4'-tetrabromodiphenyl ether (5-OH-BDE-47) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
6-hydroxy-2,2',4,4'-tetrabromodiphenyl ether (6-OH-BDE-47) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
2,2',4,4',5-pentabromodiphenyl ether (BDE-99) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
2,2',4,4',6-pentabromodiphenyl ether (BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
3-hydroxy-2,2',4,4',6-pentabromodiphenyl ether (3-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
3'-hydroxy-2,2',4,4',6-pentabromodiphenyl ether (3'-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
5-hydroxy-2,2',4,4',6-pentabromodiphenyl ether (5'-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
6-hydroxy-2,2',4,4',6-pentabromodiphenyl ether (6'-OH-BDE-100) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
Bromotamine 3IBD (Wang et al., 2019b)	X			X	X	X	X	X	X	X			X	X		
Bupropion 3IBD (Niu et al., 2011)	X			X	X	X	X	X	X	X			X	X		
4- <i>tert</i> -butylphenylacetyleane (BPA) 3IBD (Lin et al., 2011)	X			X	X	X	X	X	X	X			X	X		
Cyclophosphamide 3UA5 (Lauter et al., 2016)	X			X	X	X	X	X	X	X			X	X		
2,3',4,4',5-pentachlorobiphenyl (CB118) ^d 3IBD (Mise et al., 2016)	X			X	X	X	X	X	X	X			X	X		
Deschloroketamine 3IBD (Wang et al., 2019b)	X			X	X	X	X	X	X	X			X	X		
Efavirenz 3IBD (Niu et al., 2011)	X			X	X	X	X	X	X	X			X	X		
Fluoroketamine 3IBD (Wang et al., 2019b)	X			X	X	X	X	X	X	X			X	X		
Garcinol 5UFG ^e (Bolla et al., 2021)	X			X	X	X	X	X	X	X			X	X		
Herbacetin 5UDA (Qian et al., 2022)	X			X	X	X	X	X	X	X			X	X		
Ketamine 3IBD (Wang et al., 2019b)	X			X	X	X	X	X	X	X			X	X		
Morphobarbital 3IBD (Niu et al., 2011)	X			X	X	X	X	X	X	X			X	X		
1,2,3,4,6,7,8,9-octachlorodibenz-p-dioxin (OCDD)	X			X	X	X	X	X	X	X			X	X		
3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
2,2',3,4',5,5',6-heptachlorobiphenyl (PCB-187) 3UA5 (Maldonado-Rojas et al., 2016)	X			X	X	X	X	X	X	X			X	X		
Propofol 3IBD (Niu et al., 2011)	X			X	X	X	X	X	X	X			X	X		
Propofol 2,6-di- <i>sec</i> -butyl phenol analog 3IBD (Niu et al., 2011)	X			X	X	X	X	X	X	X			X	X		
Sauchinone 3IBD (Gong et al., 2018)	X			X	X	X	X	X	X	X			X	X		
Ticlopidine 3IBD (Gay et al., 2010)	X			X	X	X	X	X	X	X			X	X		
Ticlopidine 5UFG ^e (Bolla et al., 2021)	X			X	X	X	X	X	X	X			X	X		
Trileosan 3IBD (Zhang et al., 2021)	X			X	X	X	X	X	X	X			X	X		
1-(4-hydroxy-3-methoxyphenyl)-5-(2-nitrophenyl)pentan-4-dien-3-yl (WZ35) ^f 5UDA (Wang et al., 2020a)	X			X	X	X	X	X	X	X			X	X		

^a Amino acids are numbered according to full-length refSeq CYP2B6 (NX_P20813.1).

^b The additionally mutated I114V CYP2B6 structure was used in these studies and position 114 was inconsistently observed to be within 5 Å of the ligands.

^c This study used the additionally mutated Y244W CYP2B6 structure and W244 was not observed in the active site. Notably, this mutation did not change the amino acids within 5 Å of the ligand; therefore, we grouped both crystal structures (4I91 and 4ZV8) into the same row of this table.

^d These entries reflect only the amino acids reported by the authors, although the existence of additional active site residues is likely.

TABLE 2 continued

Substrate [metabolic method]	Active site mutations ^b													References	
	V477E	V477F	V477I	V477T	V477W	V477Y	G478A	G478E	G478S	G478V	I114V/V477W	SRS 1 ^c	SRS 5 ^c		SRS 6 ^c
Androstenedione	—	—	—	—	—	—	—	—	—	—	—	nr	nr	nr	(Domanski et al., 1999)
[16 β -hydroxylation]	—	—	—	—	—	—	—	—	—	—	—	nc	nc	↑↑↑	(Radloff et al., 2013)
Bupropion	—	—	—	—	—	—	—	—	—	—	—	—	—	—	(Spatzenegger et al., 2003)
[4-hydroxylation]	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
7-butoxycoumarin	—	nr	—	—	—	—	—	—	—	—	—	—	—	—	
[Variant oxidation]	↑↑	↑↑	—	—	—	—	—	—	—	—	—	—	—	—	
Cyclophosphamide	nr	↓	↓	↑	↑	↑	nr	nr	nr	↓	↓	—	—	—	(Nguyen et al., 2008)
[4-hydroxylation]	↓↓↓	↑	↑	↓	↑	↓	↓↓↓	↓↓↓	↓↓↓	↓	↓	—	—	—	(Radloff et al., 2013)
Efavirenz	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
[8-hydroxylation]	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
3-cyclopentyl-oxo-N-(3,5-dichloro-4-pyridyl)-4-methoxybenzamide	—	—	nr	nr	—	—	—	—	—	—	—	—	—	—	(Domanski et al., 1999)
(RP 73401)	—	—	↓	↓	—	—	—	—	—	—	—	—	—	—	
[Cyclopentyl ring <i>trans</i> -hydroxylation]	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
4-trifluoromethyl-7-ethoxycoumarin	—	↑	nr	—	—	—	—	—	—	—	—	nr	nr	nr	(Domanski et al., 1999; Spatzenegger et al., 2003; Lin et al., 2016)
(7-EFC)	—	—	—	—	—	—	—	—	—	—	—	—	—	—	
[O-deethylation]	—	↑	↑	—	—	—	—	—	—	—	—	↓↓↓	↓↓↓	↑	

^b Amino acids are numbered according to full-length refseq CYP2B6 (NX_P20813.1).

^c These entries represent the mutations of SRS 1, 5, and 6 of CYP2B6 to the corresponding CYP2B1 sequences (Domanski et al., 1999).

^d The directional changes tabulated here are relative to the CYP2B6 wild-type. One arrow indicates a change between 1-fold and twofold; two arrows indicate a change between twofold and threefold; three arrows indicate a change greater than threefold; "nc" indicates no notable change; "nr" indicates no reported value; em-dashes indicate unexplored kinetics.

^e The observed effect on K_m and k_{cat} differed depending on the L363V CYP2B6 mutant being expressed in baculovirus-infected insect (Domanski et al., 1999)/*E. coli* JM109 cells (Spatzenegger et al., 2003).

TABLE 3
Expression system-dependence of kinetic parameters during the metabolism of select substrates by wild-type CYP2B6.^a

SUBSTRATE [METABOLIC METHOD]	EXPRESSION SYSTEMS										
	Human B-lymphoblastoid cell microsomes	293FT cell microsomes	COS-1 cell microsomes	COS-7 cell microsomes	Supernomes	Baculovirus- infected insect cell microsomes	<i>Saccharomyces cerevisiae</i> W(R) cell microsomes	<i>Escherichia coli</i> Topp 3 cells	<i>Escherichia coli</i> C41 DE3 cells	<i>Escherichia coli</i> DH5a cells	<i>Escherichia coli</i> JM109 cells
Bupropion [4-hydroxylation]	K_m : 107.5 μM k_{cat} : NR ^b (Bkins and Wrighton, 1999)	— ^c	K_m : 87 μM k_{cat} : 7.9 min^{-1} (Radloff et al., 2013)	—	—	K_m : 65 μM^d k_{cat} : 2.73 min^{-1} (Lang et al., 2004) K_m : 153 μM^d k_{cat} : 2.46 min^{-1} (Klein et al., 2005)	—	K_m : 8.8 μM k_{cat} : 2.6 min^{-1} (Bumpus et al., 2005)	K_m : 95 μM k_{cat} : 6.8 min^{-1} (Zhang et al., 2011)	K_m : 0.5 μM k_{cat} : 47 min^{-1} (Lee et al., 2018)	—
Cyclophosphamide [4-hydroxylation]	—	—	—	—	K_m : 1.400 μM k_{cat} : 338 min^{-1} (Xie et al., 2003)	K_m : 4.900 μM k_{cat} : 62.5 min^{-1} (Nguyen et al., 2008)	—	K_m : 3.600 μM k_{cat} : 75.4 min^{-1} (Raccor et al., 2012)	—	—	
Elavirenz [8-hydroxylation]	—	K_m : 4.09 μM k_{cat} : 0.35 min^{-1} (Watanabe et al., 2018)	K_m : 2.07 μM k_{cat} : 1.1 min^{-1} (Radloff et al., 2013)	—	K_m : 2.680 μM k_{cat} : 301 min^{-1} (Ariyoshi et al., 2011)	K_m : 4.030 μM k_{cat} : 36 min^{-1} (Lauter et al., 2016)	—	K_m : 14.3 μM k_{cat} : 4.3 min^{-1} (Bumpus et al., 2006)	K_m : 7.3 μM k_{cat} : 4.0 min^{-1} (Zhang et al., 2011)	—	
4-trifluoromethyl-7- ethoxycoumarin (7-EFC) [O-deethylaton]	K_m : 1.7 μM k_{cat} : NR (Bkins and Wrighton, 1999)	K_m : 7.58 μM k_{cat} : 2.11 min^{-1} (Watanabe et al., 2018)	K_m : 5.50 μM k_{cat} : 3.93 min^{-1} (Jimno et al., 2003)	K_m : 7.0 μM^e k_{cat} : 21.1 min^{-1} (Watanabe et al., 2010)	K_m : 9.0 μM k_{cat} : 1.4 min^{-1} (Ogburn et al., 2010)	K_m : NR k_{cat} : 3.74 min^{-1} (Domanski et al., 1999)	K_m : 3.5 μM k_{cat} : 17 min^{-1} (Lauter et al., 2016)	K_m : NR k_{cat} : 6.8 min^{-1} (Bumpus and Hollenberg, 2008)	K_m : 4.8 μM k_{cat} : 1.2 min^{-1} (Zhang et al., 2011)	K_m : 8.5 μM k_{cat} : 14 min^{-1} (Spatzenegger et al., 2003)	

^aAccording to our literature search, this is only a sample of kinetic parameters differing for the same CYP2B6 isoform, substrate, and metabolic method within various expression systems. Expression system-dependence of CYP2B6 kinetics can also be observed for polymorphic and nonpolymorphic CYP2B6 mutants (Domanski et al., 2003; Spatzenegger et al., 1999; Spatzenegger et al., 2003; Gadel et al., 2013; Zanger and Klein, 2013; Calinski et al., 2015; Gadel et al., 2015; Imaishi and Goto, 2018; Wang et al., 2020b).

^bNR^c indicates values that were not reported.

^cEm-dashes indicate unexplored kinetics.

^dWhile CYP2B6 expression and thus k_{cat} determination was more reliable within insect cell microsomes, the authors only reported K_m values via CYP2B6 bupropion hydroxylase activity from COS-1 cell expression.

^eThese values are the average of multiple replicate averages for CYP2B6.1 K_m and k_{cat} determination reported by the authors.

site (Bathelt et al., 2002) were visualized via CYP2B6 crystal-based molecular docking with cyclophosphamide (Lautier et al., 2016). The oxazaphosphorin ring of cyclophosphamide was surrounded by F206, A298, T302, and L363 (pocket A) with one chloroethyl side chain located near I114, F115, S294, and F297 (pocket B); the second chloroethyl side chain was bound by the roof of the binding cavity, composed of residues I209, V367, and V477 (pocket C) (Bathelt et al., 2002; Lautier et al., 2016).

The importance of hydrogen bonding for ligand binding/orientation should not be underestimated. S294 and A298 contributed hydrogen bonds to the molecularly docked 1-(4-hydroxy-3-methoxyphenyl)-5-(2-nitrophenyl)penta-1,4-dien-3-1 (WZ35) (Wang et al., 2020a). F297, A298, and K479 also mediate hydrogen bonds for the stabilization of herbacetin above the CYP2B6 heme (Qian et al., 2022). Mephobarbital is stabilized through hydrogen bonds from E301, T302, and K479 along with many of the hydrophobic contacts already discussed (Niu et al., 2011). E301 exhibits an interesting behavior within the CYP2B6 crystal complexed with 4-CPI as it flexes out of the active site to hydrogen bond with a nearby water molecule rather than the ligand, although this allows 4-CPI to interact hydrophobically with F206 (Gay et al., 2010). This same phenomenon is observed for crystallized CYP2B6 complexed with 4-BP and 4-(4-nitrobenzyl)pyridine (4-NBP) where E301 interacts directly with Q172 and indirectly with S304 in a P450 subfamily 2B-conserved hydrogen bond network stabilizing the E and I helices (Shah et al., 2011). However, the side chain of E301 protrudes into the cavity where 4-CPI would be in the CYP2B6-amlodipine crystal to interact with T305 and R308 (Shah et al., 2012; Chen et al., 2018). The plasticity of E301 is further noted through molecular docking as bupropion induces its movement to the edge of the CYP2B6 active site, which is more centralized within when propofol and efavirenz are docked (Niu et al., 2011). E301 also forms a hydrogen bond with the CYP2B6 covalent modification site, T302, that is broken upon covalent binding of BPA to cause deleterious effects on active site conformation, plasticity, and catalysis (Lin et al., 2011; Watanabe et al., 2018).

Many compounds are stabilized within CYP2B6 by a combination of forces such as hydrogen bonding between E301 and PBDEs in addition to hydrophobic interactions between L363 and compounds like 2,2',4,4',5-pentabromodiphenyl ether (BDE-99), PCB-187, and OCDD (Maldonado-Rojas et al., 2016). L363 has long been defined as a residue critical for the CYP2B6 structure–function relationship with its further creation of hydrophobic contacts with 3-cyclopentylloxy-*N*-(3,5-dichloro-4-pyridyl)-4-methoxybenzamide (RP 73401 or Piclamilast), 7-butoxycoumarin, the terminal methyl of the 7-ethoxy group of 4-trifluoromethyl-7-ethoxycoumarin (7-EFC), and the *N*-methyl group of diazepam (Domanski et al., 1999; Lewis et al., 1999; Domanski and Halpert, 2001; Spatzenegger et al., 2003; Niu et al., 2011; Wang et al., 2019b). However, L363 was found to be confoundingly insignificant compared with other residues in the binding of 4-CPI, efavirenz, propofol, and the propofol 2,6-di-*sec*-butyl phenol analog (Gay et al., 2010; Niu et al., 2011). Relative to other hydrophobic CYP2B6 active site residues, L363 and V477 weakly interact with efavirenz and the benzyl groups of propofol and the propofol 2,6-di-*sec*-butyl phenol analog, which differs from their strong hydrophobic interactions with bupropion and mephobarbital (Niu et al., 2011). The local hydrophobicity and flexibility of V477 in the CYP2B6 active site nonetheless affects the metabolism of many of the same substrates that L363 is pertinent for such as RP 73401 and 7-butoxycoumarin (Domanski et al., 1999; Spatzenegger et al., 2003; Gay et al., 2010; Shah et al., 2012). Like many phenylalanine residues, L363 and T302 have also been implicated in conjugated π systems upon binding saquinone and herbacetin, respectively (Gong et al., 2018; Qian et al., 2022).

Phenylalanine residues in the putative CYP2B6 active site are uniquely involved in hydrophobic interactions, π - π stacking, and halogen- π

interactions. The bulky F108, F115, F206, F292, and F297 residues were found to be most involved in individual small movements culminating into large changes in the CYP2B6 active site volume, which ranges from 279 Å³ to 755 Å³ depending on the bound ligand (Halpert, 2011; Shah et al., 2011; Shah et al., 2012; Jang et al., 2014). F108 and F115 exhibit concerted movement in the binding of 4-CPI such that F108 shifts toward the G helix to accommodate the flexion of F115 out of the CYP2B6 active site (Gay et al., 2010). F108 is a residue not usually within 5 Å of CYP2B6 ligands, but the nitro moiety of 4-NBP points directly toward it and forces its accommodating flexion (Shah et al., 2011). Without the nitro moiety, the benzyl group of 4-BP points toward F115 instead (Shah et al., 2011). F108 also forms an important bromine- π bond for the stabilization of bornyl bromide in the CYP2B6 active site (Shah et al., 2017). (-)-Myrtenyl bromide was crystallized in a wild-type CYP2B6 active site and formed a bromine- π bond with F297; (-)-myrtenyl bromide was also crystallized in an I114V mutant CYP2B6 active site, allowing it to form bromine- π bonds with F108, F115, and F297 (Shah et al., 2017). The efavirenz 2-desoxo-2-methyl analog crystallized within CYP2B6 was found to form a chlorine- π bond with F108 that was allowed by a corresponding flexion of F206 and F297 to expand the active site volume (Shah et al., 2018). F206 also moves to occupy the space created by the flexion of E301 in the CYP2B6-4-CPI crystal while entering the active sites of the CYP2B6-4-BP and CYP2B6-4-NBP crystals as F297 rotates out (Gay et al., 2010; Halpert, 2011; Shah et al., 2011). The same concerted movement of F206 and F297 occurs to a more exaggerated extent to shrink the active site around (+)- α -pinene (Wilderman et al., 2013).

It has been hypothesized that there are two binding modes within the CYP2B6 active site, termed “A” and “B”; the former is more common and characterized by F206 moving into and F297 moving out of the active site while the latter involves the inverse action (Shah et al., 2015). Regardless, π - π stacking between F206 and selegiline along with herbacetin has also been predicted to be important for their binding/orientation/metabolism (Salonen et al., 2003; Qian et al., 2022). F297 similarly mediates π - π stacking with the benzyl groups of propofol, the propofol 2,6-di-*sec*-butyl phenol analog, and mephobarbital (Niu et al., 2011). F206 forms a sort of π stacking interaction with the -NH group of amlodipine’s pyridine moiety while F297 forms a chlorine- π bond with the chlorophenyl moiety of 4-CPI to stabilize these ligands (Shah et al., 2015). F297 was deemed exceptionally important for facilitating π - π stacking with ketamine, bromoketamine, fluoroketamine, and deschloroketamine (Wang et al., 2019b). Phenylalanine residues are also capable of forming hydrogen bonds with ligands, as seen between F297, the propofol 2,6-di-*sec*-butyl phenol analog, and herbacetin (Niu et al., 2011; Qian et al., 2022). The F-G cassette (Fig. 1) contains many phenylalanine residues and forms the “active site roof” in ligand-bound structures, is embedded within the lipid bilayer in ligand-unbound structures, and is thought of as the most plastic region of P450 subfamily 2B isoforms (Halpert, 2011; Wilderman and Halpert, 2012; Li et al., 2020). The hydrophobic F-G cassette helps create a lipophilic polarization within the CYP2B6 active site such that larger hydrophobic and smaller hydrophilic pockets contribute to substrate specificity; the identity of residues within those pockets delineate the form of chemical contribution (e.g., hydrogen bonding, hydrophobic interactions, π - π stacking) while also sterically contributing to such specificity (Mise et al., 2016).

In Silico Challenges with CYP2B6. In silico methods may be useful for explaining in vitro results, but they are not always 100% accurate on their own (Salonen et al., 2003; Radloff et al., 2013; Dmitriev et al., 2021) as some favorable ligand binding events visualized with X-ray crystallography or molecular docking can be unproductive in vitro (Talakad et al., 2011; Kobayashi et al., 2014; Maldonado-Rojas et al.,

2016). A complete *in silico* characterization of the CYP2B6 structure–function relationship is limited due to many variables influencing the CYP2B6–ligand binding/metabolism event (de Graaf et al., 2005). For example, CYP2B6 has been shown to have no regioselectivity for triclosan monohydroxylation (Zhang et al., 2021), catalyze testosterone 16 α - and 16 β -hydroxylation (Ekins and Wrighton, 1999), and strictly perform efavirenz 8-hydroxylation (Ward et al., 2003). While computerized methods are quicker/cheaper than *in vitro* methods for the pharmacological screening of new chemical entities, the measuring of *in vitro* catalytic data remains necessary to establish their full metabolic profile. The *in vitro* data providing insights into the structure–function relationship of the CYP2B6 active site will be reviewed next.

In Vitro Insights

CYP2B6 Substrate, Stereo-, and Regioselectivity. Recent work has been dedicated to understanding two important considerations for CYP2B6 substrate specificity: the substrate stereochemistry (Roy et al., 1999; Totah et al., 2007; Wang et al., 2018; Wang et al., 2019a) and, as detailed in the previous section, the various moieties composing the substrate (Ekins et al., 2008; Lewis et al., 2010; Cox and Bumpus, 2014; Cox and Bumpus, 2016; Liu et al., 2016; Wang et al., 2019b). Generally, CYP2B6 has a lower K_m for the (*S*)-enantiomers but a higher k_{cat} for the (*R*)-enantiomers of substrates (Niwa et al., 2011). CYP2B6 and CYP3A4 also tend to differ in their enantioselectivity as seen with ifosfamide since the former preferentially metabolizes (*S*)-ifosfamide while the latter prefers (*R*)-ifosfamide (Roy et al., 1999; Niwa et al., 2011). Some have attributed this observation to the potential for an enantiomeric/mirror arrangement of the CYP2B6 and CYP3A4 active sites (Lu et al., 2006). (*S*)-methadone has a lower K_m for *N*-demethylation by CYP2B6, which is also the principal metabolizer of racemic methadone (Gerber et al., 2004; Crettol et al., 2005; Zanger et al., 2007; Totah et al., 2008). CYP2B6 preferentially hydroxylates (*S*)-bupropion to (*S,S*)-hydroxybupropion (Coles and Kharasch, 2008; Sager et al., 2016; Wang et al., 2020b) and is solely responsible for the *N*-demethylation of (*S*)-mephobarbital (Kobayashi et al., 1999; Wang and Tompkins, 2008). An intriguing feature of CYP2B6-mediated bupropion metabolism is the selective formation of (*R,R*)- and (*S,S*)-hydroxybupropion but neither (*R,S*)- nor (*S,R*)-hydroxybupropion when using (*R*)- and (*S*)-bupropion as substrates, respectively (Coles and Kharasch, 2008; Bamfo et al., 2022). Explanations for these observations remain elusive but may manifest with deeper exploration of the structure–function relationship of the CYP2B6 active site. Contrary to its general (*S*)-enantioselectivity, CYP2B6 had a 77% lower K_m value for (*R*)-citalopram than (*S*)-citalopram (Niwa et al., 2011). The CYP2B6 K_m values for (*R*)-fluoxetine (Niwa et al., 2011), (*S*)-ketamine (Niwa et al., 2011; Wang et al., 2018), (*R*)-norketamine (Desta et al., 2012), and (*S*)-nicotine (Bloom et al., 2019) were all lower than their opposite enantiomers. The enantioselectivity of CYP2B6 for (*S*)-efavirenz was found to be the greatest with a higher active site affinity and 10-fold greater k_{cat} than (*R*)-efavirenz (Wang et al., 2019a).

The k_{cat} values are larger for (*S*)-ifosfamide 4-hydroxylation (Roy et al., 1999; Niwa et al., 2011), (*R*)-methadone *N*-demethylation (Gerber et al., 2004; Gadel et al., 2015), (*S*)-bupropion 4-hydroxylation (Coles and Kharasch, 2008; Wang et al., 2020b), and (*S*)-fluoxetine *N*-demethylation (Niwa et al., 2011) than their opposite enantiomers. The k_{cat} values for ketamine (Niwa et al., 2011; Li et al., 2013) and norketamine (Desta et al., 2012) are approximately equivalent between the (*R*)- and (*S*)-enantiomers. The k_{cat} in a recent CYP2B6–ketamine stereospecificity study, however, was higher for (*S*)-ketamine (Wang et al., 2018). This selection of example substrates seems to contradict the generalization made by Niwa et al. regarding K_m and k_{cat} , but there are confounding variations

in kinetic parameters for the metabolism of the same substrates by CYP2B6 in the same or different expression systems (Table 3). For example, a lower CYP2B6 K_m for (*S*)-ketamine was found in an insect cell expression system but not within COS-1 cells (Li et al., 2013). The issue of lacking continuity in CYP2B6 kinetic parameters based on the expression system will be discussed further later. Beyond enantioselectivity, there are various moieties of substrates that seem to specifically promote CYP2B6-mediated metabolism (Cox and Bumpus, 2014; Wang et al., 2019b). Interesting observations have been made in this regard but remain unexplained from the perspective of the CYP2B6 structure–function relationship. For example, deuterating ifosfamide on its alpha and alpha' carbons reduced its K_m for CYP2B6 by over 50% (Calinski et al., 2015). Previous work has also tested the CYP2B6 structure–function relationship through novel modulations in the efavirenz chemical scaffold (Cox and Bumpus, 2014; Cox and Bumpus, 2016).

Most notably, efavirenz analogs lacking an intact oxazinone ring were not readily oxidized by CYP2B6 (Cox and Bumpus, 2014). The benzoxazine analog had an increased catalytic efficiency, revealing the efavirenz carbonyl group to abrogate CYP2B6-mediated metabolism due to possible steric clashes with L363 and V367 (Niu et al., 2011; Cox and Bumpus, 2016) or overstabilization in the active site via hydrogen bonds donated from E301 and T302 (Cox and Bumpus, 2014). Opening the cyclopropyl group increased K_m by over twofold as the greater rotational degrees of freedom may cause steric clashes (Cox and Bumpus, 2014). Efavirenz analogs with intact oxazinone rings and heteroatom substitutions remained as CYP2B6 substrates but had altered K_m values (Cox and Bumpus, 2016). Replacing the carbonyl with a methyl group increased the affinity for the CYP2B6 active site by over twofold, corroborating previous findings (Cox and Bumpus, 2014; Cox and Bumpus, 2016). An angularly projected methyl group off the oxazinone ring could avoid steric clashes and form favorable hydrophobic interactions with L363 and V367 (Cox and Bumpus, 2016). However, replacing the hydrogen bond-accepting ether of efavirenz with a secondary amine or saturated carbon increased the CYP2B6 K_m by over threefold (Cox and Bumpus, 2016). Liu et al. performed a similar study with 4-substituted-7-alkoxycoumarin derivatives where 7-EFC displayed the highest catalytic efficiency (Liu et al., 2016). Increasing the 7-alkoxy chain length from an ethyl to a propyl and butyl group caused a proportional decrease in CYP2B6 K_m and k_{cat} (Liu et al., 2016). A similar decrease occurred when increasing the bulk of 7-ethoxycoumarin derivatives by placing a methyl group at position 4, but turnover was increased when that bulky 4-substituent was halogenated (Liu et al., 2016). The identity of substrate halogen atoms generally increases affinity for CYP2B6 such that dibromo > dichloro for phenoxyaniline (POA) analogs (Chen et al., 2018) and Br > Cl > F for ketamine analogs (Wang et al., 2019b). The effect of halogenating CYP2B6 substrates is substrate-dependent but generally promotes high CYP2B6 binding affinity by increasing substrate hydrophobicity (Chen et al., 2018; Wang et al., 2019b).

As aforementioned, the regioselectivity of CYP2B6 can obfuscate its structure–function relationship by being strict or unspecified depending on the substrate. Uniquely, CYP2B6 can mediate the *O*-demethylation and *ortho*-hydroxylation of methoxychlor (Hodgson and Rose, 2007; Wang and Tompkins, 2008). PBDEs such as BDE-47 can also form up to nine different metabolites when oxidized by CYP2B6 (Erratico et al., 2015). The K_m and k_{cat} values were very similar for the CYP2B6-mediated formation of 4'-hydroxy-2,2',4,5'-tetrabromodiphenyl ether (4'-OH-BDE-49), 5-hydroxy-2,2',4,4'-tetrabromodiphenyl ether (5-OH-BDE-47), and 6-hydroxy-2,2',4,4'-tetrabromodiphenyl ether (6-OH-BDE-47), implying that CYP2B6 showed no preference for *ortho*-, *meta*-, or *para*-hydroxylation (Erratico et al., 2015). Interestingly, 2,2',4,4',6-pentabromodiphenyl ether (BDE-100) differs from the BDE-47 structure only by a third *ortho*-

substituted bromine atom, which significantly lowers its ability to be metabolized by CYP2B6 (Gross et al., 2015). The variety of metabolites formed from CYP2B6-mediated BDE-47 metabolism suggests that it can bind within the active site in a variety of metabolically active orientations (Erratico et al., 2015). CB118 must also either bind with positional variance within the CYP2B6 active site or its single binding mode places its carbons 3 and 5 in close enough proximity to the heme for monooxygenation (Mise et al., 2016). The former case is probably more likely considering efavirenz hydroxylation on carbons 7 and 8 but the strict 8-hydroxyefavirenz formation by CYP2B6 (Ward et al., 2003) and 7-hydroxyefavirenz formation by CYP2A6 (Ogbum et al., 2010). CYP2B6-mediated metabolism of 8-hydroxyefavirenz to 7,8-dihydroxyefavirenz and 8,14-dihydroxyefavirenz has also been observed, further exemplifying the existence of more than one productive binding mode and flexible regioselectivity (Avery et al., 2013). Without any further research into the structure–function relationships between CYP2B6 and these substrates, it is difficult to identify the reason for these observations.

Site-directed mutagenesis of CYP2B6 followed by metabolic characterization with various substrates is the “gold-standard” for elucidating *in vitro* insights into the structure–function relationship. Common substrates that have been used in these efforts include bupropion (Lang et al., 2004; Klein et al., 2005; Radloff et al., 2013; Wang et al., 2020b), cyclophosphamide (Nguyen et al., 2008; Ariyoshi et al., 2011; Raccor et al., 2012), 7-EFC (Domanski et al., 1999; Jinno et al., 2003; Spatzenegger et al., 2003; Kumar et al., 2007; Liu et al., 2016; Watanabe et al., 2018), efavirenz (Bumpus et al., 2006; Zhang et al., 2011; Watanabe et al., 2018; Wang et al., 2019a), and ketamine (Li et al., 2013; Wang et al., 2018) among others. Bupropion 4-hydroxylation (Desta et al., 2007) and (*S*)-efavirenz 8-hydroxylation (Wang et al., 2019a) are excellent probes for CYP2B6 activity in complex expression systems like human liver microsomes as such reactions are catalyzed nearly exclusively by CYP2B6. Although they are not clinical pharmaceuticals, monoterpenes have also been suggested as ideal *in vitro* CYP2B6 active site probes (Wang et al., 2019b). There are several CYP2B6 substrates that, to our knowledge, have not been kinetically parameterized with CYP2B6 mutants (Supplemental Table 2). The only substrates that have been used to kinetically characterize CYP2B6 active site mutants are androstenedione, bupropion, 7-butoxycoumarin, cyclophosphamide, efavirenz, RP 73401, and 7-EFC (Table 2).

Functionality of CYP2B6 Active Site Mutants. Substantial mutagenesis research has been done on clinically relevant CYP2B6 polymorphs, which are largely outside the active site and have been extensively reviewed by others (Zanger et al., 2007; Wang and Tompkins, 2008; Mo et al., 2009; Turpeinen and Zanger, 2012; Zanger and Klein, 2013; Naidoo et al., 2014; Langmia et al., 2021). A thorough summary of structure–function findings with CYP2B6 polymorphs outside of the active site is deserving of an independent review due to the large number of variants, and thus we will not explore them here. The sparse number of publications investigating the CYP2B6 active site through *in vitro* structure–function studies within the last 6 years (Table 2) reveals the potential to reignite such interests within our field. Originally, L363 within the K-K' loop of CYP2B6 (Fig. 1) was of particular interest as it is a unique residue compared with other P450 family 2 enzymes that have valine at that position (Domanski et al., 1999; Spatzenegger et al., 2003). An L363V mutation within CYP2B6 was found to increase the k_{cat} for androstenedione 16 β -hydroxylation (Domanski et al., 1999) but decreased the k_{cat} for 7-butoxycoumarin oxidation (Spatzenegger et al., 2003) and the catalytic efficiency for RP 73401 hydroxylation (Domanski et al., 1999). More rigid and planar substrates like androstenedione may favor the larger active site volume that V363 allows. L363V had no significant effect on the k_{cat} of 7-EFC *O*-deethylation in baculovirus-infected insect cells

(Domanski et al., 1999) yet decreased that catalytic efficiency in *E. coli* (Spatzenegger et al., 2003). Nonetheless, L363 in addition to F107 of the CYP2B6 B' helix (Fig. 1) were implicated in substrate-specific pertinence for RP 73401 metabolism (Domanski et al., 1999) while L363 also conferred a higher binding affinity for POA congeners that was decreased with a V363 substitution (Chen et al., 2018). There may be something necessary in the bulkier presence of F107 and L363 for allowing binding and oxidative metabolism of larger substrates with more rotational degrees of freedom like RP 73401 and 7-butoxycoumarin. F107 seems to possess substrate-specific importance as its mutation to valine or isoleucine had no significant effect on the catalytic efficiency of cyclophosphamide 4-hydroxylation (Nguyen et al., 2008) and 7-EFC *O*-deethylation (Domanski et al., 1999), respectively.

Single amino acid changes have even been shown to augment the cooperativity of CYP2B6 catalysis with L363V showing positive cooperativity and V367L showing negative cooperativity upon 7-EFC metabolism (Spatzenegger et al., 2003). A V367L single mutation within the K-K' loop of CYP2B6 (Fig. 1) also caused an elevated k_{cat} for cyclophosphamide 4-hydroxylation, which was an observation that was not made for the V367F/S/T/H mutations (Nguyen et al., 2008) nor when V367L CYP2B6 had 7-EFC or 7-butoxycoumarin as substrates (Spatzenegger et al., 2003). It is notable, though, that a V367A mutation decreased the k_{cat} for 7-EFC *O*-deethylation (Lin et al., 2016). V367 thereby affects CYP2B6-mediated metabolism on a substrate-specific basis with some substrates favoring a larger hydrophobic residue while others favor a smaller hydrophobic residue at position 367. Although CYP2B6.1 natively prefers 16 β -hydroxylation of testosterone and androstenedione, placing the less bulky alanine within the F helix and K-K' loop at positions 206 and 367, respectively (Fig. 1), seems to increase the enzymatic preference for 16 α -hydroxylation, which may be due to D-ring reorientation upon substrate binding that forces the 16 α -carbon closer to the heme iron (Lin et al., 2016). The only polymorphic residue within the CYP2B6 active site belongs to the CYP2B6.35 haplotype, which had no catalytic activity with 7-EFC and efavirenz as substrates (Watanabe et al., 2018). This may be attributed to the deleterious presence of apoenzyme and location of G110V and I114T in SRS 1 (Gotoh, 1992; Watanabe et al., 2018); however, the other polymorphisms included within the CYP2B6.35 haplotype (E148D, M198T, and A279P) could also be responsible for the observed lack of enzymatic activity (Honda et al., 2011; Radloff et al., 2013; Watanabe et al., 2018; Desta et al., 2021). An I114T single mutant also displayed a remarkably reduced catalytic efficiency with bupropion and efavirenz as substrates (Radloff et al., 2013), suggesting that the small size of G110 and the hydrophobicity of I114 promotes wild-type levels of CYP2B6 catalysis.

Singularly testing CYP2B6 mutants from complex haplotypes is useful for determining the functional effects of discrete changes to CYP2B6, as was done for I114T despite its presence within CYP2B6.35 (Radloff et al., 2013). However, multiple mutations within one CYP2B6 isoform can have interactive effects on catalysis. The I114V/V477W CYP2B6 mutant remarkably enhanced the catalytic efficiency of cyclophosphamide 4-hydroxylation (Nguyen et al., 2008). The I114V substitution in the B'-C loop (Fig. 1) independently decreased the K_m of this reaction, perhaps by increasing accessibility for cyclophosphamide through the previously identified substrate access channel 2 (Cojocaru et al., 2007; Nguyen et al., 2008; Shah et al., 2012; Jang et al., 2014; Lautier et al., 2016), while the V477I, V477F, and V477W substitutions with expanding size and hydrophobicity increased the catalytic efficiency in that order (Nguyen et al., 2008). Altering position 477 to have polarity with V477S/T/Y/N/D/E mutations largely decreased cyclophosphamide metabolism (Nguyen et al., 2008). Another study has also suggested that replacing V477 with a larger hydrophobic residue like

phenylalanine may encourage a more catalytically active orientation of 7-EFC, 7-ethoxycoumarin, and 7-butoxycoumarin in the CYP2B6 active site (Spatzenegger et al., 2003). Further substantiating the substrate specificity of active site residue importance, wild-type V477 is apparently beneficial for the cyclopentyl ring *trans*-hydroxylation of RP 73401 (Domanski et al., 1999). Mutagenesis within CYP2B6 SRSs also

displays substrate specificity as discretely mutating CYP2B6 SRS 1 and 5 to the corresponding CYP2B1 sequences maintained wild-type levels of androstenedione 16 β -hydroxylation but diminished 7-EFC *O*-deethylation (Domanski et al., 1999). The same was done for CYP2B6 SRS 2 and 6 where the former decreased the k_{cat} while the latter increased the k_{cat} with androstenedione and 7-EFC as substrates (Domanski et al., 1999).

The hydrophobicity of position 478 is also important for wild-type CYP2B6 activity as G478S and G478E single mutants were essentially inactive with cyclophosphamide as a substrate (Nguyen et al., 2008). Interestingly, a G478V single mutant had a similar catalytic efficiency to the wild-type for cyclophosphamide 4-hydroxylation while G478A significantly decreased that catalytic efficiency (Nguyen et al., 2008). Position 478 may require a hydrophobic residue that is either small (like glycine) or bulkier (like valine) for normal CYP2B6 functionality. The moderate bulk of A478 could potentially allow unfavorable steric clashes to occur within its seemingly hydrophobic pocket. Maintaining I209 and S294 as active site residues within the F helix and I helix, respectively (Fig. 1), is also necessary as the I209L and S294D CYP2B6 single mutants significantly diminished the k_{cat} with 7-butoxycoumarin and 7-EFC as substrates (Spatzenegger et al., 2003). In this case, the steric positioning of I209 appears advantageous to the CYP2B6 active site, and its mere isomerization is deleterious, even though the opposite was true with the L313I mutation (Watanabe et al., 2018). The localized polarity/hydrogen bonding capacity created by S294 near the heme may be important for CYP2B6-substrate interactions and/or topological interactions within CYP2B6. Furthermore, L199M increased, S207A did not affect, and K236N decreased, the CYP2B6 catalytic efficiency with cyclophosphamide as a substrate (Nguyen et al., 2008), which may be due to the localization of these residues in the F helix and G helix (Fig. 1), causing intricate modulations in the coordinated plasticity of the F-G cassette. An F206A mutation within CYP2B6 resulted in a slight decrease in k_{cat} with 7-EFC (Lin et al., 2016), suggesting that both the hydrophobicity and bulk at position 206 may be important for substrate binding/metabolism similar to the wild-type.

M365I and C475I CYP2B6 single mutants within the K-K' loop and β 4-1 strand, respectively (Fig. 1), notably decreased k_{cat} with cyclophosphamide (Nguyen et al., 2008) but not when M365I and C475S had RP 73401 and 7-EFC as substrates (Domanski et al., 1999). Even though methionine and isoleucine have a comparable capacity for steric clashes, the effect of amino acid chemistry at position 365 is apparently substrate-specific. The nucleophilic nature of residue 475 seems important for CYP2B6-mediated catalysis, and the functionality of C475S makes it clear that there is no disulfide bridge formed by C475 that is pertinent for the structural/catalytic integrity of CYP2B6. Furthermore, M103V, R109K, N291S, and T292L CYP2B6 single mutants decreased the k_{cat} for RP 73401 and 7-EFC oxidation, but the S284H, N289M, T292A, and I370R mutations had less significant effects on k_{cat} (Domanski et al., 1999). It is interesting to consider T292 in the I helix of CYP2B6 (Fig. 1) and, therefore, its proximity to the active site. The preservation of CYP2B6 catalytic activity with the nonconservative T292A mutation versus the loss of function with the bulkier yet still hydrophobic leucine at position 292 suggests a unique and unpredictable mutational sensitivity within the I helix. Such speculation is corroborated by the aforementioned effects of the S294D and L313I mutations (Spatzenegger et al.,

2003; Watanabe et al., 2018). The apparent sensitivity and elusiveness of the CYP2B6 structure–function relationship can be explained as a challenging product of its relatively uncharacterized active site (Table 2) and rampant expression system dependence of kinetic parameters (Table 3).

In Vitro Challenges with CYP2B6. The prominent in vitro challenges of substrate-dependent and expression system-dependent changes in catalysis remain unsolved (Domanski et al., 1999; Jinno et al., 2003; Bumpus et al., 2005; Klein et al., 2005; Wang et al., 2006; Zhao and Halpert, 2007; Wang and Tompkins, 2008; Honda et al., 2011; Zhang et al., 2011; Gadel et al., 2013). The former problem is expected and by nature connected with a thorough understanding of the CYP2B6 structure–function relationship, but the latter problem may obscure such an understanding. Though many kinetic data are compared internally within a single study, compiling these data and comparing across studies/expression systems reveals distinct inconsistencies that challenge the elucidation of the CYP2B6 structure–function relationship (Table 3). This is especially apparent when the kinetic parameters of K_m and k_{cat} should theoretically be constants for identical enzymatic isoforms catalyzing the same reactions on the same substrates. Therefore, it may be that the significant challenge of detailing the structure–function relationship of CYP2B6 is related to the discontinuity of catalytic data between the same isoforms with the same substrates in expression systems that are as different as 293FT cell microsomes (Watanabe et al., 2018) and *E. coli* (Zhang et al., 2011), as similar as COS-1 (Jinno et al., 2003) and COS-7 cell microsomes (Watanabe et al., 2010) or even identical (Gerber et al., 2004; Gadel et al., 2015). It is likely that standardizing in vitro experimental conditions for heterologous expression of CYP2B6.1 and its mutants when determining their kinetic parameters would provide the necessary framework for cross-study comparisons. It may be possible to optimize the CYP2B6 expression system, P450 oxidoreductase (POR):P450 ratio (El-Serafi et al., 2015; Wang et al., 2018; Wang et al., 2020b), and presence/absence/concentration of cytochrome b5 (cyt b5) (Shebley and Hollenberg, 2007; Bumpus and Hollenberg, 2008; Calinski et al., 2015; Yoo et al., 2019) for accessibility, affordability, and, most importantly, experimental accuracy and precision for laboratories in the field to use.

As the electron transferring redox partners of P450s that may have polymorph- and expression system-dependent levels of expression/activity, POR and cyt b5 can influence CYP2B6 substrate binding/metabolism in a complex and relatively unpredictable fashion (El-Serafi et al., 2015; Chen et al., 2018; Yoo et al., 2019). Moreover, the impact of membrane interactions involving the N-terminal domain of CYP2B6 on active site architecture also remains unknown. Until the heterologous expression of CYP2B6 is better optimized for cross-study comparisons, kinetic parameters, and structure–function findings elucidated from studies employing the heterologous expression of CYP2B6, particularly N-terminally truncated CYP2B6, should be interpreted with caution. Other enigmatic themes controverting common biochemical intuition are abundant within the CYP2B6 structure–function relationship. CYP2B6 has often been compared with CYP2B1 in an effort to extrapolate conclusions about the former enzyme's elusive behavior, but these reports should be handled carefully as their metabolic profiles are not always identical (Bumpus et al., 2005; Wang and Tompkins, 2008; Lin et al., 2016). Moreover, whole SRS mutations cannot necessarily confer the substrate specificity of CYP2B1 to CYP2B6 (Domanski et al., 1999), and the same is true for single residue mutations with F477V CYP2E1 obtaining CYP2B6-like activity with 7-EFC but V477F CYP2B6 not gaining activity with the CYP2E1-specific substrate, *p*-nitrophenol (Spatzenegger et al., 2003). CYP2B6 structure–function investigators have hypothesized that mutagenesis within active site residues is only useful for augmenting the function of certain P450 isoforms to behave more like other P450 isoforms if the substrates they natively metabolize are very similar (Spatzenegger

et al., 2003; Zhao and Halpert, 2007). Otherwise, changes in the highly conserved P450 backbone are necessary to alter the productive metabolism of substrates in a directed fashion (Spatzenegger et al., 2003; Zhao and Halpert, 2007; Fasan et al., 2019). The malleable CYP2B6 active site mold may be limited in its ability to metabolize various substrates of other P450s by the upper limits of its backbone flexibility. CYP2B6 was aptly dubbed an unusual P450 isoform due to the inexplicable preclusion of generalizations about catalytic consequences upon its mutagenesis (Wang et al., 2020b). However, what may seem inexplicable now may still be explained in the future.

Fertile Grounds for Future Insights

The ongoing optimization of current and advent of new *in silico* technologies will undoubtedly continue bolstering the accuracy of protein structure–function predictions. As such, new CYP2B6 crystals must continue to be solved with clinically relevant ligands or those that have been studied extensively *in vitro*. These may include bupropion, cyclophosphamide, 7-EFC, and ketamine in their native structures or as tight-binding analogs similar to the most recent CYP2B6-efavirenz analog crystal (Shah et al., 2018). As a remarkably specific (Stevens et al., 1997; Domanski et al., 1999) and recently neglected CYP2B6 substrate, RP 73401 may prove useful within X-ray crystallography or molecular docking studies for elucidating more about the putative CYP2B6 active site. A ligand-free CYP2B6 crystal structure is also still lacking. Few *in silico* studies have explored the enantioselectivity of CYP2B6 other than with the purported two-planar template model of its active site (Koyama and Yamazoe, 2011). Therefore, the chirality of substrates should be considered when performing X-ray crystallography or molecular docking for further insights into the stereochemical mold of the CYP2B6 active site. While crystallographic data are likely preferred over molecular docking results, the latter are still useful (Domanski et al., 1999; Bathelt et al., 2002; Niu et al., 2011; Wang et al., 2019b) and need continued performance with additional substrates for analyzing the CYP2B6 structure–function relationship, especially with lower affinity substrates that are difficult to crystallize within CYP2B6. The application of NMR-based tools (Estrada et al., 2013; Estrada et al., 2016; Bart and Scott, 2017), isothermal titration calorimetry (Zhao and Halpert, 2007), and deuterium exchange mass spectrometry (Halpert, 2011) for *in silico* CYP2B6 characterization will likely be advantageous for better understanding its structure–function relationship as well (Wilderman and Halpert, 2012). Above all else, more *in silico* work needs to be done with characterizing the functions of CYP2B6 active site residues.

As previously mentioned, Lewis and others pioneered many of the CYP2B6 structure–function studies with the use of homology models (Supplemental Table 1). Now that we have access to 13 CYP2B6 crystal structures (Table 1), however, there is a significant amount of increasingly accurate *in silico* findings and reasonable speculations to be made for the function of CYP2B6 active site residues in binding ligands and interacting with one another. These findings may enhance our understanding of the CYP2B6 pharmacophore for calculating QSARs, perhaps allowing an easier differentiation between CYP2B6 substrates/inhibitors and explaining why select substrates also function as inhibitors (Bumpus et al., 2005; Bumpus et al., 2006; Korhonen et al., 2007; Ekins et al., 2008; Lewis et al., 2010; Lee et al., 2018). This may also allow a more rational mutagenesis of CYP2B6 to alter its functionality in directed ways. Harnessing in tandem the power of *in silico* tools such as X-ray crystallography and molecular docking with *in vitro* techniques like site-directed mutagenesis and kinetic parameter measurement may be the key to rationalizing CYP2B6 substrate specificity, stereospecificity, and regio-specificity (Kumar et al., 2003; Kumar et al., 2007; Zhao and Halpert, 2007; Nguyen et al., 2008; Koyama and Yamazoe, 2011; Radloff et al.,

2013; Watanabe et al., 2018; Wang et al., 2019b). It may be possible for polymorphic mutations surrounding the active site to translate into it and impact metabolism through modulations in protein flexibility (Zhao and Halpert, 2007; Shah et al., 2012; Wilderman and Halpert, 2012; Kobayashi et al., 2014; Shah et al., 2015; Shah et al., 2016), solubility/stability (Bumpus et al., 2005; Kumar et al., 2007; Hofmann et al., 2008; Watanabe et al., 2010; Kobayashi et al., 2014; Watanabe et al., 2018), and substrate channel access (Cojocaru et al., 2007; Shah et al., 2012; Jang et al., 2014; Lautier et al., 2016) mediated by energetically stabilizing/destabilizing and sterically favoring/hindering residue contacts (Gay et al., 2010; Watanabe et al., 2010; Halpert, 2011; Shah et al., 2011; Shah et al., 2012; Wilderman et al., 2013; Kobayashi et al., 2014; Shah et al., 2015; Shah et al., 2018; Watanabe et al., 2018). However, understanding the structure–function relationship of the CYP2B6 active site at least as well as we currently understand the structure–function relationship of CYP2B6 polymorphs may be of immense pharmaceutical, medical, environmental, and biochemical usefulness.

A multidisciplinary approach combining *in silico* and *in vitro* tools to study CYP2B6 has been successfully employed before (Domanski et al., 1999; Spatzenegger et al., 2003; Kent et al., 2006; Korhonen et al., 2007; Nguyen et al., 2008; Niu et al., 2011; Radloff et al., 2013; Gong et al., 2018; Watanabe et al., 2018; Wang et al., 2019b; Zhang et al., 2021). These studies set the precedent that strongly urges future CYP2B6 studies to use a multidisciplinary method. It may prove beneficial to systematically perform *in vitro* site-saturation mutagenesis of CYP2B6 active site residues and measure the corresponding catalytic data with various clinically relevant substrates or persistent organic pollutants. The changes that these mutations induce within the enzyme structure, interactions governing substrate binding/metabolism, and potentially the identity of metabolites can then be visualized and/or predicted *in silico* to advance and reinforce the speculations to be made about the CYP2B6 structure–function relationship. Elucidating how slight changes above the CYP2B6 heme affect substrate metabolism may even eventually allow an engineering of CYP2B6 or other P450 subfamily 2B enzymes to perform coordinated oxidative reactions that are humanly valuable (Kumar et al., 2003; Zhao and Halpert, 2007), as was accomplished with a bacterial P450 by the Arnold Laboratory (Fasan et al., 2019). More may be known about other P450 subfamily 2B enzymes like rat CYP2B1 and rabbit CYP2B4 (Zhao and Halpert, 2007) or even the CYP2B6.6 polymorph (Jinno et al., 2003; Xie et al., 2003; Crane et al., 2012; Xu et al., 2012; Li et al., 2013; Zanger and Klein, 2013; Court et al., 2014) than the CYP2B6 active site, making it evident that a similar level of knowledge about the CYP2B6 active site can be attained if the effort is dedicated toward it.

In conclusion, the fertile grounds for cultivating future CYP2B6 structure–function insights exist as a two-step plan. We advise that a multidisciplinary approach be used that takes advantage of the well-substantiated predictive power of *in silico* tools and the *in vitro* utility of site-directed/site-saturation mutagenesis paired with kinetic parameterization. Furthermore, work should focus on CYP2B6 active site residues and could even extend to those involved in POR/cyt b5 binding and electron transfer to better understand the biochemistry of CYP2B6. It is also important to consider standardizing *in vitro* experimental conditions for cross-study comparisons of wild-type and mutant CYP2B6 catalytic constants. The surplus of information that is still unknown about the CYP2B6 structure–function relationship could begin to be revealed by the multidisciplinary symbiosis of *in silico* and *in vitro* data to allow for more profound structural and functional speculations about CYP2B6.

Authorship Contributions

Wrote the manuscript: Angle and Cox.

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**Multidisciplinary Insights into the Structure-Function Relationship of the
CYP2B6 Active Site
(Supplemental Tables)**

Drug Metabolism and Disposition

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Supplemental Table 1. Historical *in silico* insights of predicted active site amino acids and their topological locations around various ligands within P450 homology modeled CYP2B6.

Homology Model of CYP2B6	PDB ID	Ligand Bound/Docked	Predicted Active Site Amino Acids^{a,b}	Topological Location in CYP2B6	Reference
CYP102	Not Reported	Tamoxifen	<u>F206,</u> <u>S210,</u> <u>F297,</u> Q473	Helix F, F-F' loop, Helix I, C-terminal coil	(Lewis and Lake, 1997)
	1FAG	4-trifluoromethyl-7-ethoxycoumarin	T205, <u>F206,</u> <u>S294,</u> <u>T302,</u> <u>L363,</u> Q473, <u>C475</u>	Helix F, Helix F, Helix I, Helix I, K-K' loop, C-terminal coil, β 4-1 strand	(Lewis et al., 1999)
	1FAG	(S)-mephenytoin	R109, <u>I114,</u> <u>F206,</u> <u>T302,</u> Q473, E474, <u>C475</u>	B'-C loop, B'-C loop, Helix F, Helix I, C-terminal coil, β 4-1 strand, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Bupropion	<u>I114,</u> T205, <u>F206,</u> <u>L363,</u> Q473, E474	B'-C loop, Helix F, Helix F, K-K' loop, C-terminal coil, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Testosterone	<u>I101,</u> <u>I114,</u> T205, <u>F206,</u> <u>I209,</u> <u>T302,</u> <u>L363,</u> E474, <u>C475</u>	B-B' loop, B'-C loop, Helix F, Helix F, Helix F, Helix I, K-K' loop, β 4-1 strand, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Antipyrine	<u>I114,</u> <u>F206,</u> <u>T302,</u> <u>C475</u>	B'-C loop, Helix F, Helix I, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Diazepam	<u>I114,</u> <u>F206,</u> <u>T302,</u> <u>L363,</u> Q473, E474, <u>C475</u>	B'-C loop, Helix F, Helix I, K-K' loop, C-terminal coil, β 4-1 strand, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Methoxychlor	<u>I114,</u> T205, <u>F206,</u> <u>T302,</u> Q473,	B'-C loop, Helix F, Helix F, Helix I, C-terminal coil,	(Lewis et al., 1999)

			<u>C475</u>	β 4-1 strand	
	1FAG	Cyclophosphamide	<u>I114,</u> <u>S294,</u> <u>T302,</u> <u>L363,</u> Q473, <u>C475</u>	B'-C loop, Helix I, Helix I, K-K' loop, C-terminal coil, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Ifosfamide	<u>S294,</u> <u>A298,</u> <u>T302,</u> <u>L363,</u> E474 <u>C475</u>	Helix I, Helix I, Helix I, K-K' loop, β 4-1 strand, β 4-1 strand	(Lewis et al., 1999)
	1FAG	7-pentoxyresorufin	<u>I114,</u> T205, <u>F206,</u> <u>T302,</u> <u>L363,</u> M365, <u>C475</u>	B'-C loop, Helix F, Helix F, Helix I, K-K' loop, K-K' loop, β 4-1 strand	(Lewis et al., 1999)
	1FAG	7-benzyloxyresorufin	<u>I101,</u> <u>I114,</u> T205, <u>F206,</u> <u>T302,</u> <u>L363,</u> M365, <u>C475</u>	B-B' loop, B'-C loop, Helix F, Helix F, Helix I, K-K' loop, K-K' loop, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Orphenadrine	<u>I114,</u> <u>F115,</u> <u>F297,</u> <u>T302,</u> <u>L363,</u> E474	B'-C loop, B'-C loop, Helix I, K-K' loop, Helix I, β 4-1 strand	(Lewis et al., 1999)
	1FAG	Selegiline	<u>I114,</u> <u>F115,</u> <u>F206,</u> <u>A298,</u> <u>T302,</u> <u>L363</u>	B'-C loop, B'-C loop, Helix F, Helix I, K-K' loop, Helix I	(Salonen et al., 2003)
CYP2C5	1DT6	4-trifluoromethyl-7-ethoxycoumarin	<u>I114,</u> <u>F115,</u> T205, <u>F206,</u> <u>S294,</u> <u>F297,</u> <u>T302,</u> <u>L363,</u> <u>V477</u>	B'-C loop, B'-C loop, Helix F, Helix F, Helix I, Helix I, Helix I, K-K' loop, C-terminal coil	(Lewis et al., 2002)
	1DT6	4-trifluoromethyl-7-ethoxycoumarin (orientation 1)	<u>F206,</u> <u>A298,</u> <u>E301,</u> <u>T302,</u> T305, T306,	Helix F, Helix I, Helix I, Helix I, Helix I, Helix I,	(Wang and Halpert, 2002)

			<u>L362</u> , <u>L363</u> , <u>V367</u> , <u>V477</u> , <u>G478</u> , <u>K479</u>	K-K' loop, K-K' loop, K-K' loop, C-terminal coil, C-terminal coil, β4-2 strand	
1DT6	4-trifluoromethyl-7-ethoxycoumarin (orientation 2)		<u>K100</u> , M103, <u>I114</u> , <u>F115</u> , <u>F206</u> , <u>F297</u> , <u>A298</u> , <u>T302</u> , <u>V367</u> , <u>V477</u>	B-B' loop, B-B' loop, B'-C loop, B'-C loop, Helix F, Helix I, Helix I, Helix I, K-K' loop, C-terminal coil	(Wang and Halpert, 2002)
1DT6	7-benzyloxyresorufin		<u>I114</u> , <u>F115</u> , <u>F206</u> , <u>F297</u> , <u>A298</u> , <u>E301</u> , <u>T302</u> , <u>L363</u> , <u>V367</u> , <u>V477</u> , <u>G478</u>	B'-C loop, B'-C loop, Helix F, Helix I, Helix I, Helix I, Helix I, K-K' loop, K-K' loop, C-terminal coil, C-terminal coil	(Wang and Halpert, 2002)
1DT6	Cyclophosphamide		<u>I114</u> , <u>F115</u> , <u>F206</u> , <u>I209</u> , L216, <u>S294</u> , <u>F297</u> , <u>A298</u> , <u>T302</u> , <u>L363</u> , <u>V367</u> , <u>V477</u>	B'-C loop, B'-C loop, Helix F, Helix F, Helix F', Helix I, Helix I, Helix I, Helix I, K-K' loop, K-K' loop, C-terminal coil	(Bathelt et al., 2002)
1DT6	Ifosfamide		<u>I114</u> , <u>F115</u> , <u>F206</u> , <u>I209</u> , L216, <u>S294</u> , <u>F297</u> , <u>A298</u> , <u>T302</u> , <u>L363</u> , <u>V367</u> , <u>V477</u>	B'-C loop, B'-C loop, Helix F, Helix F, Helix F', Helix I, Helix I, Helix I, Helix I, K-K' loop, K-K' loop, C-terminal coil	(Bathelt et al., 2002)
1DT6	7-ethoxycoumarin		M103, <u>I209</u> , <u>L363</u> , <u>V367</u> , <u>V477</u>	B-B' loop, Helix F, K-K' loop, K-K' loop, C-terminal coil	(Spatzenegger et al., 2003)

	1DT6	4-trifluoromethyl-7-ethoxycoumarin	<u>S294,</u> <u>L363,</u> <u>V367,</u> <u>V477</u>	Helix I, K-K' loop, K-K' loop, C-terminal coil	(Spatzenegger et al., 2003)
	1DT6	7-pentoxyresorufin	<u>I114,</u> <u>F206,</u> S207, <u>T302,</u> <u>V367</u>	B'-C loop, Helix F, Helix F, Helix I, K-K' loop	(Lewis et al., 2004)
	1N6B	4-trifluoromethyl-7-ethoxycoumarin	<u>K100,</u> <u>I114,</u> <u>F115,</u> T205, <u>F206,</u> <u>T302,</u> <u>L363</u>	B-B' loop, B'-C loop, B'-C loop, Helix F, Helix F, Helix I, K-K' loop,	(Lewis et al., 2006)
CYP2B1 ^c	N/A	3-cyclopentyloxy-N-(3,5-dichloro-4-pyridyl)-4-methoxybenzamide (RP 73401)	M103, R109, <u>L363,</u> M365, <u>V477</u>	B-B' loop, B'-C loop, K-K' loop, K-K' loop, C-terminal coil	(Domanski et al., 1999)
CYP2B4	Not Reported	Bergamottin	<u>A298,</u> <u>T302,</u> <u>L363</u>	Helix I, Helix I, K-K' loop	(Kent et al., 2006)
	1SUO	4-trifluoromethyl-7-ethoxycoumarin	<u>K100,</u> <u>I114,</u> <u>F115,</u> T205, <u>F206,</u> <u>T302,</u> <u>L363</u>	B-B' loop, B'-C loop, B'-C loop, Helix F, Helix F, Helix I, K-K' loop,	(Lewis et al., 2006)
	1SUO	Cyclophosphamide	<u>V104,</u> I110, <u>I114,</u> <u>F115,</u> <u>F206,</u> F208, <u>I209,</u> <u>F297,</u> <u>A298,</u> <u>E301,</u> <u>T302,</u> S320, <u>L363,</u> <u>V367,</u> <u>V477,</u> <u>G478</u>	B-B' loop, B'-C loop, B'-C loop, B'-C loop, Helix F, Helix F, Helix F, Helix I, Helix I, Helix I, Helix I, Helix I, Helix J, K-K' loop, K-K' loop, C-terminal coil, C-terminal coil	(Nguyen et al., 2008)
	1SUO	4-(4-chlorophenyl)imidazole	<u>S210,</u> <u>F297,</u> <u>T302</u>	F-F' loop, Helix I, Helix I	(Lewis et al., 2010)
^a Amino acids are numbered according to full-length refseq CYP2B6 (NX_P20813-1). ^b Bolded and underlined amino acids indicate those found in the CYP2B6 active site according to the CYP2B6 crystal structures that have been solved and molecular docking results that have been yielded to date (Table 1). ^c The CYP2B1 crystal structure utilized within Domanski et al., 1999 was homology modeled from a consensus of P450 BM-3, P450cam, and P450terp (Szklarz et al., 1995) and then used as a homology model for CYP2B6.					

Supplemental Table 2. Kinetic parameters for the CYP2B6.1-mediated metabolism of substrates for which there are no reported catalytic constants of CYP2B6 mutants.					
Substrate	Metabolic Method	Expression System^a	K_m (μM)^a	k_{cat} (min⁻¹)^a	Reference
Amitriptyline	<i>N</i> -demethylation	Human B-lymphoblastoid cell microsomes	144.4	NR	(Ekins and Wrighton, 1999)
Antipyrine	4-hydroxylation	Human B-lymphoblastoid cell microsomes	17,700	NR	(Ekins and Wrighton, 1999)
Arteether	<i>O</i> -deethylation	Human B-lymphoblastoid cell microsomes	28	NR	(Ekins and Wrighton, 1999)
Azinphos-methyl	<i>S</i> -oxidation	Supersomes	2.2	2.8	(Buratti et al., 2002)
2,2',4,4'-tetrabromodiphenyl ether (BDE-47)	3-hydroxylation	Supersomes	6.4	0.0106	(Feo et al., 2013)
	4-hydroxylation		2.7	0.150	(Erratico et al., 2013)
	4'-hydroxylation		1.2	0.270	(Erratico et al., 2013)
	5-hydroxylation		3.8	0.948	(Feo et al., 2013)
			5.8	0.300	(Erratico et al., 2013)
	6-hydroxylation		4.2	0.202	(Feo et al., 2013)
2,2',4,4',5-pentabromodiphenyl ether (BDE-99)	1'-hydroxylation	Supersomes	0.1	0.2116	(Erratico et al., 2012)
	2-hydroxylation		0.3	0.0068	(Erratico et al., 2012)
	4-hydroxylation		0.3	0.0276	(Erratico et al., 2012)
	4'-hydroxylation		0.9	0.2467	(Erratico et al., 2012)
	5'-hydroxylation		2.4	0.4573	(Erratico et al., 2012)
	6'-hydroxylation		0.3	0.0141	(Erratico et al., 2012)
2,2',4,4',6-pentabromodiphenyl ether (BDE-100)	3-hydroxylation	Supersomes	5.2	0.0099	(Gross et al., 2015)
	4'-hydroxylation		5.5	0.066	(Gross et al., 2015)
	5'-hydroxylation		4.9	0.166	(Gross et al., 2015)
	6'-hydroxylation		7.0	0.043	(Gross et al., 2015)
(<i>R</i>)-1,3-Benzodioxolyl- <i>N</i> -methylbutanamine (MBDB)	<i>N</i> -demethylation	Supersomes	138	2.5	(Niwa et al., 2011)
(<i>S</i>)-1,3-Benzodioxolyl- <i>N</i> -methylbutanamine (MBDB)			119	0.7	(Niwa et al., 2011)
(<i>R</i>)-1,3-Benzodioxolyl- <i>N</i> -methylbutanamine (MBDB)	Demethylation		83	3.3	(Niwa et al., 2011)

(S)-1,3-Benzodioxolyl-N-methylbutanamine (MBDB)			56	0.2	(Niwa et al., 2011)
7-benzyloxyresorufin	N-demethylation	Human B-lymphoblastoid cell microsomes	1.28	NR	(Ekins and Wrighton, 1999)
7-benzyloxyquinoline	O-debenzylation	Baculovirus-infected insect cell microsomes	NR	0.55	(Renwick et al., 2001)
Bromoketamine	N-demethylation	Baculovirus-infected insect cell microsomes	10	94	(Wang et al., 2019)
4-chloromethyl-7-ethoxycoumarin	O-deethylation	Human B-lymphoblastoid cell microsomes	33.7	NR	(Ekins and Wrighton, 1999)
Carbaryl	Methyl hydroxylation	Supersomes	45	15.54	(Tang et al., 2002)
	4-hydroxylation		11	0.80	(Tang et al., 2002)
	5-hydroxylation		110	0.29	(Tang et al., 2002)
Cinnarizine	p-hydroxylation	Human B-lymphoblastoid cell microsomes	17.2	NR	(Ekins and Wrighton, 1999)
Clobazam	N-demethylation	Supersomes	289	5.70	(Giraud et al., 2004)
3-cyano-7-ethoxycoumarin	O-deethylation	Human B-lymphoblastoid cell microsomes	71.3	NR	(Ekins and Wrighton, 1999)
Deschloroketamine	N-demethylation	Baculovirus-infected insect cell microsomes	184	83	(Wang et al., 2019)
Dextromethorphan	N-demethylation	Human B-lymphoblastoid cell microsomes	350	NR	(Ekins and Wrighton, 1999)
Diazepam	N-demethylation	Human B-lymphoblastoid cell microsomes	181	NR	(Ekins and Wrighton, 1999)
Diazinon	S-oxidation	Supersomes	14.83	5.44	(Ellison et al., 2012)
	Dearylation		13.94	2.60	(Ellison et al., 2012)
1,2-dibromoethane	2-bromo acetaldehyde formation	Human B-lymphoblastoid cell microsomes	9,700	NR	(Ekins and Wrighton, 1999)
N,N-diethyl-m-toluamide (DEET)	Ring methyl hydroxylation	Supersomes	40.2	22.3	(Usmani et al., 2002)
			46.2	34.7	(Edwards et al., 2005)
Disulfoton	S-oxidation	Supersomes	11.2	62.2	(Usmani et al., 2004)
Efavirenz <i>trans</i> -alkene analog	7- or 8-hydroxylation	Supersomes	0.23	0.13	(Cox and Bumpus, 2014)

Efavirenz 13-propyl analog	5- or 7-hydroxylation and 8-hydroxylation		1.10	0.97	(Cox and Bumpus, 2014)
Efavirenz benzoxazine analog	7- or 8-hydroxylation		0.22	3.2	(Cox and Bumpus, 2014)
Efavirenz methyl benzoxazine analog	Methyl hydroxylation		1.4	NR	(Cox and Bumpus, 2016)
	5-, 7-, or 8-hydroxylation		1.0	NR	(Cox and Bumpus, 2016)
Efavirenz quinazolinone analog	Benzene ring hydroxylation		11	NR	(Cox and Bumpus, 2016)
Efavirenz <i>trans</i> -alkene quinazolinone analog	5-, 7-, or 8-hydroxylation		13	NR	(Cox and Bumpus, 2016)
Efavirenz quinolinone analog	Benzene ring hydroxylation		14	NR	(Cox and Bumpus, 2016)
Efavirenz benzo[d][1,3]dioxin-2-one analog	Unknown hydroxylation		5.5	NR	(Cox and Bumpus, 2016)
Endosulfan- α	<i>S</i> -oxidation	Supersomes	16.2	11.4	(Casabar et al., 2006)
			5.58	10.31	(Lee et al., 2006)
(-)-fenchone	6-exo-hydroxylation	Supersomes	150	12.9	(Miyazawa and Gyoubu, 2007)
	6-endo-hydroxylation		260	5.33	(Miyazawa and Gyoubu, 2007)
	10-hydroxylation		200	10.66	(Miyazawa and Gyoubu, 2007)
Fluoroketamine	<i>N</i> -demethylation	Baculovirus-infected insect cell microsomes	40	103	(Wang et al., 2019)
(<i>R</i>)-fluoxetine	<i>N</i> -demethylation	Supersomes	126	1.19	(Niwa et al., 2011)
(<i>S</i>)-fluoxetine			546	2.55	(Niwa et al., 2011)
7-hydroxyefavirenz	14-hydroxylation	Supersomes	62.4	1.3	(Ogburn et al., 2010)
8-hydroxyefavirenz	14-hydroxylation	Supersomes	2.12	8.5	(Ward et al., 2003)
			23.2	4.21	(Ogburn et al., 2010)
Ifosfamide	<i>N</i> -dechloro-ethylation	<i>Escherichia coli</i> C41 DE3	2,000	0.4	(Calinski et al., 2015)
	4-hydroxylation		4,600	3.8	(Calinski et al., 2015)
Deuterated ifosfamide ^c	<i>N</i> -dechloro-ethylation		2,000	0.2	(Calinski et al., 2015)
	4-hydroxylation		2,000	4.1	(Calinski et al., 2015)
(<i>R</i>)-ifosfamide	<i>N</i> -dechloro-ethylation	Supersomes	1,900	8.2	(Roy et al., 1999)
(<i>S</i>)-ifosfamide			900	11.8	(Roy et al., 1999)
(<i>R</i>)-ifosfamide	4-hydroxylation		NR	<0.4	(Roy et al., 1999)

			2,240	1.98	(Niwa et al., 2011)
(S)-ifosfamide			NR	2.08	(Roy et al., 1999)
			1,270	17.1	(Niwa et al., 2011)
Imipramine	N-demethylation	<i>Saccharomyces cerevisiae</i> microsomes	383	NR	(Ekins and Wrighton, 1999)
β -ionone	4-hydroxylation	Supersomes	5.6	572.8	(Marumoto et al., 2017)
Lidocaine	N-deethylation	Human B-lymphoblastoid cell microsomes	537.6	NR	(Ekins and Wrighton, 1999)
Loperamide	N-demethylation	Supersomes	65.6	0.008	(Kim et al., 2004)
Malathion	S-oxidation	Supersomes	3.09	3.6	(Buratti et al., 2005)
(S)-mephenytoin	N-demethylation	Human B-lymphoblastoid cell microsomes	564	NR	(Ekins and Wrighton, 1999)
(S)-mephobarbital	N-demethylation	Human B-lymphoblastoid cell microsomes	264	0.008	(Kobayashi et al., 1999a)
Methiocarb	S-oxidation	Supersomes	11.0	28.0	(Usmani et al., 2004)
Methyl parathion	S-oxidation	Supersomes	1.25	10.39	(Ellison et al., 2012)
	Dearylation		141	1.42	(Ellison et al., 2012)
Midazolam	1'-hydroxylation	Human B-lymphoblastoid cell microsomes	46.1	NR	(Ekins and Wrighton, 1999)
Naphthalene	1-hydroxylation	Supersomes	58.6	20.2	(Cho et al., 2006)
	2-hydroxylation		93.8	0.8	(Cho et al., 2006)
	1,2-dihydroxylation		49.5	2.2	(Cho et al., 2006)
Nevirapine	3-hydroxylation	Human B-lymphoblastoid cell microsomes	834	NR	(Erickson et al., 1999)
Nonane	2-hydroxylation	Supersomes	79.0	76.0	(Edwards et al., 2005)
2-nonanol	C-oxidation	Supersomes	35.0	72.8	(Edwards et al., 2005)
(R)-norketamine	5-hydroxylation	Supersomes	21.6 ^b	16.72 ^b	(Desta et al., 2012)
	C-oxidation		286.5	13.0	(Desta et al., 2012)
(S)-norketamine	5-hydroxylation	Supersomes	36.1 ^b	16.3 ^b	(Desta et al., 2012)
	C-oxidation		146.8	7.61	(Desta et al., 2012)

Parathion	S-oxidation	Supersomes	0.8	1.7	(Buratti et al., 2002)
		NR	0.61	4.827	(Foxenberg et al., 2007)
	Dearylation		0.74	1.804	(Foxenberg et al., 2007)
Pethidine	N-demethylation	Supersomes	262	82	(Murray et al., 2020)
Phorate	S-oxidation	Supersomes	32.1	70.8	(Usmani et al., 2004)
Propofol	4-hydroxylation	Human B-lymphoblastoid cell microsomes	10	21	(Court et al., 2001)
(-)- <i>cis</i> -rose oxide	9-hydroxylation	Supersomes	159.2	60.67	(Nakahashi et al., 2015)
(-)- <i>trans</i> -rose oxide			73.80	154.0	(Nakahashi et al., 2015)
Sertraline	N-demethylation	Human B-lymphoblastoid cell microsomes	30.7	2.04	(Kobayashi et al., 1999b)
(+)- <i>cis</i> -3,5-dimethyl-2-(3-pyridyl) thiazolidin-4-one hydrochloride (SM-12502)	S-oxidation	Human B-lymphoblastoid cell microsomes	1,767	NR	(Ekins and Wrighton, 1999)
Styrene	Glycol formation	Human B-lymphoblastoid cell microsomes	180	NR	(Ekins and Wrighton, 1999)
Verapamil	O-demethylation	Human B-lymphoblastoid cell microsomes	137.4	NR	(Ekins and Wrighton, 1999)

^a NR indicates data/values that were not reported.

^b These authors analyzed the diastereomeric metabolites formed by CYP2B6-mediated 5-hydroxylation of (*R,S*)-norketamine; CYP2B6 showed the highest intrinsic clearance with its production of (*2S,5S;2R,5R*)-hydroxynorketamine, the K_m and k_{cat} of which is reported.

^c This ifosfamide was deuterated on its alpha and alpha' carbons.

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