IN VITRO-IN VIVO EXTRAPOLATION OF CYP2D6 INACTIVATION BY PAROXETINE: PREDICTION OF NONSTATIONARY PHARMACOKINETICS AND DRUG INTERACTION MAGNITUDE

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List of abbreviations: MBI, mechanism-based inactivation; DDI, drug-drug interaction; IVIVE, In vitro-in vivo extrapolation; f_{2D6}, fractional contribution of CYP2D6 to total oral clearance
ABSTRACT

Attempts at predicting drug-drug interactions perpetrated by paroxetine from *in vitro* data have utilized reversible enzyme inhibition models and have been unsuccessful to date, grossly underpredicting interaction magnitude. Recent data have provided evidence for mechanism-based inactivation of CYP2D6 by paroxetine. We have predicted the pharmacokinetic consequences of CYP2D6 inactivation by paroxetine from *in vitro* inactivation kinetics (\(k_{\text{inact}} \) 0.17 min\(^{-1}\), unbound \(K_i\) 0.315 \(\mu\)M), *in vivo* inhibitor concentrations, and an estimated CYP2D6 degradation half-life of 51 h, using a mathematical model of mechanism-based inhibition. The model-predicted accumulation ratio of paroxetine was 5 times that expected from single dose kinetics and in excellent agreement with the observed 5-6-fold greater accumulation. Magnitudes of interactions produced by paroxetine (20-30 mg/day) with desipramine, risperidone, perphenazine, atomoxetine, (S)-metoprolol and (R)-metoprolol were predicted, considering the contribution of CYP2D6 to their oral clearance. Predicted fold-increases in victim drug AUC were 5, 6, 5, 6, 4 and 6-fold, respectively, and are in reasonable agreement with observed values of 5, 6, >7, 7, 5 and 8-fold, respectively. Failure to consider microsomal binding *in vitro* adversely affected predictive accuracy. Simulation of the sensitivities of these predictions to model inputs suggests a 2-fold underprediction of interaction magnitude when a CYP2D6 degradation half-life of 14 h (reported for rat CYP3A) is used. In summary, the scaling model for mechanism-based inactivation successfully predicted the pharmacokinetic consequences of CYP2D6 inactivation by paroxetine from *in vitro* data.
INTRODUCTION

Paroxetine (Paxil®) is a widely used selective serotonin reuptake inhibitor (SSRI) antidepressant that is indicated for the treatment of major depressive disorder, obsessive compulsive disorder, panic disorder, generalized anxiety disorder, social anxiety disorder and post-traumatic stress disorder.

Paroxetine is an established perpetrator of drug-drug interactions (DDI’s) when co-administered with agents whose clearance is largely dependent on the activity of cytochrome P450 2D6 (CYP2D6). Examples of characterized clinical interactions with paroxetine include its effects on the kinetics of desipramine (Brøsen et al., 1993; Alderman et al., 1997), perphenazine (Özdemir et al., 1997), metoprolol (Hemeryck et al., 2000), risperidone (Spina et al., 2001) and atomoxetine (Belle et al., 2002) where the clearance of the victim drugs is impaired by 5-8-fold. In addition, paroxetine displays nonlinear accumulation kinetics with steady-state exposures exceeding projections from single dose kinetics by ~5-fold in CYP2D6 extensive metabolizers but not in CYP2D6 poor metabolizers (Kaye et al., 1989; Sindrup et al., 1992). This has been attributed to metabolic saturation following multiple dosing (Sindrup et al., 1992) although supporting evidence at the enzyme kinetic level is lacking. In vitro studies have reproducibly demonstrated potent inhibition of human liver microsomal CYP2D6 activity via an apparent competitive mechanism (von Moltke et al., 1995; Otton et al., 1996; Hemeryck et al., 2001). However, attempts at in vitro-in vivo extrapolation (IVIVE) of interaction magnitude under the assumption of reversible inhibition have been largely unsuccessful, even when nonspecific microsomal binding in vitro was considered (Hemeryck et al., 2001) or empirical approaches such as application of total plasma or even total intrahepatic concentrations of paroxetine were used in the predictions (von Moltke et al., 1995; Hemeryck et al., 2000).
Recent data have provided evidence for mechanism-based inactivation (MBI) of CYP2D6 by paroxetine (Bertelsen et al., 2003). Paroxetine produced a concentration and time-dependent inhibition of human liver microsomal CYP2D6 activity \textit{in vitro}, as measured by dextromethorphan O-demethylation rate. Kinetic analysis revealed that paroxetine produced a metabolism-dependent rapid loss of activity of the enzyme with a half-life of inactivation of 4 minutes ($k_{\text{inact}} = 0.17 \text{ min}^{-1}$), and biochemical evidence for metabolite intermediate complexation via a carbene-heme complex with CYP2D6 (Bertelsen et al., 2003).

Whereas \textit{in vitro-in vivo} scaling approaches for reversible inhibition of CYP enzymes have been extensively described over the last decade and applied in the prediction of drug-drug interactions, mathematical models for scaling interactions resulting from mechanism-based inhibition have only recently been described, with relatively fewer reported examples illustrating their predictive utility. Of notable mention are the reasonably successful predictions of the interactions perpetrated by fluoxetine, diltiazem, clarithromycin, verapamil and the HIV protease inhibitors with CYP3A4 substrates using a mathematical model of mechanism-based inhibition developed by Hall and colleagues (Mayhew et al., 2000; Wang et al., 2004; Ernest II et al., 2005). \textit{In vitro-In vivo} extrapolations using this model require a formal kinetic analysis of inactivation by the perpetrator agent to determine the potency of inactivation ($K_{i}$) and maximal inactivation rate ($k_{\text{inact}}$). An estimate of the enzyme-available concentration of the inhibitor is necessary (as for scaling models for reversible inhibition). In addition, an estimate of the \textit{in vivo} degradation rate ($k_{\text{degrad}}$) of the enzyme in humans is needed.

In this report, we describe the application of the scaling model for mechanism-based inhibition in predicting the pharmacokinetic consequences of CYP2D6 inactivation by paroxetine from \textit{in vitro} data. Specifically, we describe predictions of (a) the nonlinear
accumulation kinetics of paroxetine resulting from autoinhibition of clearance; and (b). the magnitude of drug-drug interactions of paroxetine with CYP2D6 substrate victim drugs. In addition, we have characterized the sensitivity of these predictions to estimates of model inputs that are nearly impossible to determine directly in humans, and therefore associated with the greatest uncertainty: *in vivo* enzyme degradation rate and the *in vivo* enzyme-available inhibitor concentration. The implications of these findings for assessment of clinical risk associated with mechanism-based inhibitory DDI’s are discussed.
METHODS

Scaling Model. The following mathematical model has been previously derived for IVIVE of drug interactions resulting from MBI (Mayhew et al., 2000):

\[
\frac{\text{CL}_{\text{int},I}}{\text{CL}_{\text{int},\text{control}}} = \frac{k_{\text{degrad}}}{k_{\text{degrad}} + \frac{1 \times k_{\text{inact}}}{1 + K_I}}
\]  

(1)

In Equation 1, \( \text{CL}_{\text{int},I} \) and \( \text{CL}_{\text{int},\text{control}} \) are the intrinsic clearance of the metabolic pathway of the victim drug in the presence and absence of the mechanism-based inactivator (concentration I). The inactivation rate constant \( k_{\text{inact}} \) (maximal inactivation rate at saturating concentration of the inactivator) and the potency \( K_I \) (inactivator concentration at which half-maximal inactivation rate is achieved), which are typically measured \textit{in vitro}, are the primary kinetic parameters describing the MBI process, and \( k_{\text{degrad}} \) represents the first-order rate constant for degradation (turnover) of the enzyme \textit{in vivo}. If the victim drug is dosed orally, completely absorbed, and cleared entirely by hepatic metabolism by the inactivated enzyme, the apparent oral clearance (\( \text{CL}_{\text{po}} \)) should equal \( \text{CL}_{\text{int}} \), and fractional decrement in oral clearance (FDCL) can thus be described as:

\[
\text{FDCL} = 1 - \frac{k_{\text{degrad}}}{k_{\text{degrad}} + \frac{1 \times k_{\text{inact}}}{1 + K_I}}
\]  

(2)

In the application of this model to scaling DDI’s perpetrated by paroxetine via MBI of CYP2D6, it was assumed that non-CYP2D6 clearance mechanisms of the victim drugs of interest (desipramine, risperidone, perphenazine, atomoxetine, (S)-metoprolol and (R)-metoprolol) were unaffected by paroxetine treatment. Thus, the fractional decrement of total oral clearance (\( \text{FDCL}_{\text{tot}} \)) was calculated as follows, where \( f_{2D6} \) represents the fractional contribution of CYP2D6 to the overall oral clearance of the specific victim drug:
The magnitude of the DDI (fold-increase in victim drug AUC) was predicted as follows:

\[
\frac{\text{AUC}_{\text{paroxetine phase}}}{\text{AUC}_{\text{control phase}}} = \frac{1}{1 - \text{FDCL}_{\text{tot}}}
\]

(4)

The above described approach was also used to assess the extent of nonlinear accumulation of paroxetine following multiple dose-administration, with the only conceptual difference being that paroxetine is considered as both the perpetrator and victim drug in the analysis (i.e. autoinactivation).

**Estimation of Model Inputs.** As prerequisites to the IVIVE exercise, the following model inputs were estimated: \(k_{\text{inact}}, K_I, k_{\text{degrad}}, I, \) and \(f_{2D6}\).

*In vitro* kinetic analysis of MBI of CYP2D6 by paroxetine has been previously described (Bertelsen et al., 2003) and the following kinetic parameters were reported: \(k_{\text{inact}}\) of 0.17 min\(^{-1}\) and \(K_I\) of 4.85 \(\mu\)M. A human liver microsomal protein concentration of 2.5 mg/mL was used in the preincubation with paroxetine in these studies of time and concentration-dependent effects on CYP2D6 activity. The estimated unbound fraction of paroxetine at the microsomal protein concentration used in the kinetic MBI studies is 0.065, extrapolated from the results of previously described human liver microsomal binding experiments (Hemeryck et al., 2001). Thus, the unbound \(K_I\) for paroxetine inactivation of CYP2D6 was determined as 0.315 \(\mu\)M. Both “total” and “unbound” \(K_I\) estimates (4.85 \(\mu\)M and 0.315 \(\mu\)M, respectively) were explored as inputs for scaling to assess the utility of nonspecific binding considerations in the estimation of inhibitor potency.

An indirect approach was used to determine \(k_{\text{degrad}}\) for CYP2D6 from the time course of return of CYP2D6 activity (measured as the dextromethorphan/dextrorphan urinary metabolic...
ratio) to baseline following discontinuation of paroxetine administration (20 mg/day for 10 days) in healthy volunteers (Liston et al., 2002). In the aforementioned study, CYP2D6 activity returned to baseline post-cessation of paroxetine administration, with a half-life of 69.6 hours. This half-life of return of CYP2D6 activity to baseline should reflect two processes: CYP2D6 turnover half-life and paroxetine steady-state half-life (18.3 hours following dosing at 20 mg/day (Kaye et al., 1989)). The CYP2D6 turnover half-life was thus determined using a noncompartmental deconvolution (component analysis) approach. Deconvolution of the mean residence time of paroxetine at steady-state of 26.4 hours (18.3/0.693) from the mean time of return of CYP2D6 activity to baseline of 100.4 hours (69.6/0.693) afforded a mean CYP2D6 degradation time of 74 hours (100.4-26.4), translating to a CYP2D6 degradation half-life of 51 hours (0.693 × 74) and a first-order degradation rate constant $k_{\text{degrad}}$ of 0.0136 h$^{-1}$ for CYP2D6.

The inhibitor concentration ($I$) was estimated as the unbound steady-state average concentration of paroxetine from total systemic exposures that were either measured in the specific drug-drug interaction studies or reported in previously published literature on the clinical pharmacokinetics of paroxetine (Kaye et al., 1989), and an unbound fraction ($f_u$) of 0.05 in human plasma (Kaye et al., 1989). Other estimates of inhibitor concentration including the total systemic concentration and the steady-state maximum unbound concentration at the inlet to the liver (estimated using Equation 5 which includes the steady-state systemic paroxetine $C_{\text{max}}$ and an additive portal venous “absorption” component as described by Kanamitsu et al., 2000) were also evaluated as alternative inputs to the model.

$$I_{\text{inlet,max},u} = f_u \times I_{\text{inlet,max}} = f_u \times \left(C_{\text{max}} + \frac{k_a \times Dose \times f_a}{Q_h}\right)$$

(5)

For application of Equation 5, the absorption rate constant ($k_a$) for paroxetine was estimated as 0.01 min$^{-1}$ using Equation 6, based on a mean $T_{\text{max}}$ of 5 hours and a half-life of 23 hours.
(elimination rate constant $k_{el}$ of 0.03 hr$^{-1}$). Complete absorption ($f_a =1$) of paroxetine was assumed based on the human absorption/disposition of paroxetine (Kaye et al., 1989) and a hepatic blood flow ($Q_h$) of 1500 mL/min was assumed (Davies and Morris, 1993).

$$T_{\text{max}} = \frac{\ln \left( \frac{k_a}{k_{el}} \right)}{k_a - k_{el}}$$

(6)

The fractional contribution of CYP2D6 to the overall oral clearance of each victim drug ($f_{2D6}$) was estimated from its clinical pharmacokinetics in CYP2D6 extensive metabolizers and poor metabolizers (Table 1) as follows, where $CL_{\text{po,CYP2D6 PM}}$ (or $AUC_{\text{CYP2D6 PM}}$) and $CL_{\text{po,CYP2D6 EM}}$ (or $AUC_{\text{CYP2D6 EM}}$) are literature values of the respective apparent oral clearance values (or area under the plasma concentration-time curve following an oral dose) of the victim drug (atomoxetine (Sauer et al., 2003), desipramine (Brøsen et al., 1993), R-metoprolol (Lennard et al., 1983), S-metoprolol (Lennard et al., 1983), paroxetine (Sindrup et al., 1992), perphenazine (Dahl-Puustinen et al., 1989), or risperidone (Huang et al., 1993)) in the poor metabolizer and extensive metabolizer subpopulations:

$$f_{2D6} = \left( 1 - \frac{CL_{\text{po,CYP2D6 PM}}}{CL_{\text{po,CYP2D6 EM}}} \right) = \left( 1 - \frac{AUC_{\text{CYP2D6 EM}}}{AUC_{\text{CYP2D6 PM}}} \right)$$

(7)

**Sensitivity Analysis.** The sensitivities of the model-derived predictions of drug interaction magnitude (fold-increase in victim drug AUC) to uncertainties in estimates of CYP2D6 degradation half-life and enzyme-available inhibitor concentration ($I$) were simulated over a wide range of relevant values for these input parameters (CYP2D6 half-life values of 10-100 hours, and enzyme-available paroxetine concentrations of 1-1000 nM), for victim drugs with
varying relative contributions of CYP2D6 to overall oral clearance ($f_{2D6}$ ranging from 0.5-1.0), using Equations 2-4. An unbound $K_I$ of 0.315 $\mu$M was used for these simulations.
RESULTS

Prediction of Nonstationary Pharmacokinetics of Paroxetine. Steady-state average plasma concentrations of 35-42.5 ng/mL are achieved following dosing with paroxetine at 20-30 mg QD, translating to unbound systemic exposures of 5.3-6.5 nM (Kaye et al., 1989; Sindrup et al., 1992). Application of the scaling model with the previously described input parameters (k_{degrad} of 0.0136 h^{-1}, k_{inact} of 0.17 min^{-1}, and an unbound K_{I} for CYP2D6 inactivation of 0.315 µM) suggests that in patients receiving paroxetine at 20-30 mg QD doses, 93% of CYP2D6 will be inactivated. Upon consideration of paroxetine’s f_{D6} of 0.86, the net decrement in paroxetine oral clearance at steady-state as a consequence of autoinactivation of CYP2D6-mediated hepatic metabolism is predicted to be 80%, translating to a predicted accumulation ratio that is 5 times that expected if the kinetics were time-independent. This prediction is in excellent agreement with the observed 4.9-5.7-fold greater than expected accumulation following multiple dose administration of paroxetine at 20-30 mg QD in clinical pharmacokinetic studies (Kaye et al., 1989; Sindrup et al., 1992).

However, when nonspecific microsomal binding of paroxetine in vitro was not considered in the scaling process (that is, a “total” K_{I} value of 4.85 µM was used as input to the model), the predicted percent loss of CYP2D6 activity following multiple dose administration of paroxetine at 20-30 mg QD was only 45-50%, translating to an approximately 40% decrement in paroxetine steady-state oral clearance, and accumulation that is 1.7-fold in excess of that expected from the single dose kinetic profile. Failure to consider nonspecific microsomal binding in vitro therefore resulted in an approximately 3-fold underprediction of the extent of nonlinearity in accumulation of paroxetine.
Application of total systemic steady-state average concentrations of paroxetine (106-130 nM following dosing with paroxetine at 20-30 mg QD (Kaye et al., 1989; Sindrup et al., 1992)) or estimated steady-state maximum unbound concentrations of paroxetine at the inlet to the liver (~27-40 nM following dosing with paroxetine at 20-30 mg QD, calculated using Equation 5) as inputs to the scaling model did not increase predictive accuracy (a 6-7-fold greater than expected accumulation was predicted, compared to a 5-fold prediction using steady-state average systemic unbound concentrations).

The performance of the scaling model in predicting the steady-state kinetics of paroxetine from the single dose kinetic profile is illustrated in Figure 1. For purposes of this exercise, previously published data on the single dose kinetics of paroxetine (Sindrup et al., 1992) were fitted to a one compartment pharmacokinetic model and the following parameters estimated: first-order absorption rate constant (k_a) 0.908 hr^{-1}; apparent volume of distribution (V/F) 3087 L; and elimination rate constant (k_e) 0.063 hr^{-1}. The expected steady-state profile following administration of 14 daily doses of 30 mg was traced by superposition of the fitted single dose kinetic profile (Winnonlin Enterprise version 3.2 software), assuming linear accumulation kinetics. These profiles for the first and fourteenth dose are represented in Figure 1 by solid lines. Assuming a hepatic blood flow of 21 mL/min/kg (Davies and Morris, 1993), the impact of autoinactivation on the systemic clearance and oral bioavailability of paroxetine at steady-state were estimated from the model-predicted fractional decrement in apparent oral clearance (intrinsic clearance), in the context of a well-stirred pharmacokinetic model (Wilkinson and Shand, 1975). Assuming that autoinactivation does not alter volume of distribution (but would alter the apparent volume of distribution (V/F) due to alteration of the bioavailability F) or absorption kinetics, and that changes in systemic clearance will be reflected as changes in
elimination half-life, the following pharmacokinetic parameters for paroxetine post-CYP2D6 inactivation were derived: apparent volume of distribution (V/F) 1397 L; and elimination rate constant (k_{el}) 0.0282 hr⁻¹. The corresponding superpositioned steady-state profile is represented in Figure 1 by a dashed line profile and is is reasonably good agreement with the observed steady-state pharmacokinetic data. In contrast, when nonspecific microsomal binding is not considered in the scaling, the resulting pharmacokinetic parameter estimates (apparent volume of distribution (V/F) 2262 L; and elimination rate constant (k_{el}) 0.0525 hr⁻¹) predict a steady-state profile (represented by the dotted line) that substantially underpredicts the observed extent of accumulation.

**Prediction of Drug Interaction Magnitudes.** The results of *in vitro-in vivo* extrapolation of drug interaction magnitude for CYP2D6 substrate victim drugs upon co-administration of paroxetine are detailed in Table 2. The scaling model predicts that administration of 20-30 mg daily doses of paroxetine results in a >90% loss of CYP2D6 activity (FDCL_{2D6} 0.9-0.95). Across all victim drugs and drug interaction studies, the mean accuracy of prediction of the fold-increase in victim drug AUC by paroxetine was 89% (range: 76%-104%) when nonspecific microsomal binding was considered.

However, failure to consider nonspecific microsomal binding led to a greater than 3-fold underprediction of interaction magnitude (mean accuracy of 29%, range: 22%-35%). When alternative estimates of inhibitor concentration (total systemic concentration or unbound maximal concentration at the inlet to the liver estimated using Equation 5) were used as inputs to the scaling model (with consideration of nonspecific microsomal binding), the overall predictive accuracy did not improve to favor their selection as the estimate of [I]. The mean accuracy was
117% (range: 90%-140%) when the estimated unbound maximal concentration at the inlet to the liver was used, and was 127% (range: 97%-140%) when the total systemic concentration was used.

The performance of the scaling model is illustrated in Figure 2 for the interaction of paroxetine with desipramine (Alderman et al., 1997). For purposes of this exercise, the steady-state pharmacokinetic profile of desipramine (50 mg QD) in the control phase (Alderman et al., 1997) was fitted to a one compartment pharmacokinetic model with an absorption lag time and the following parameters estimated: first-order absorption rate constant (k_a) 0.381 hr⁻¹; absorption lag time (t_{lag}) 0.79 hr; apparent volume of distribution (V/F) 1762 L; and elimination rate constant (k_e) 0.0408 hr⁻¹. Assuming a hepatic blood flow of 21 mL/min/kg (Davies and Morris, 1993), the impact of CYP2D6 inactivation by paroxetine on the systemic clearance and oral bioavailability of desipramine were estimated from the model-predicted fractional decrement in apparent oral clearance (FDCL_{tot} 0.81 at a paroxetine dosage of 30 mg QD; Table 2), in the context of a well-stirred pharmacokinetic model (Wilkinson and Shand, 1975). Assuming that the interaction does not alter volume of distribution (but would alter the apparent volume of distribution (V/F) due to alteration of F) or absorption kinetics, and that changes in systemic clearance will be reflected as changes in desipramine’s elimination half-life, the following pharmacokinetic parameters for desipramine post-CYP2D6 inactivation were derived: apparent volume of distribution (V/F) 1122 L; and elimination rate constant (k_e) 0.0122 hr⁻¹. The corresponding superpositioned steady-state profile is represented in Figure 2 by a dashed line profile and is reasonably good agreement with the observed steady-state pharmacokinetic profile (Alderman et al., 1997) of desipramine (50 mg QD) with co-administered paroxetine (30 mg QD to steady-state). In contrast, when nonspecific microsomal binding is not considered in
the scaling (FDCL_{tot} 0.473 at a paroxetine dosage of 30 mg QD; Table 2), the resulting pharmacokinetic parameter estimates (apparent volume of distribution (V/F) 1387 L; and elimination rate constant (k_{el}) 0.0273 hr^{-1}) predict a steady-state profile (represented by the dotted line in Figure 2) that substantially underpredicts the observed interaction, illustrating the importance of nonspecific microsomal binding considerations in quantification of inactivator potency.

Sensitivity Analyses. Simulations of the sensitivity of the model-predicted fold-increase in victim drug AUC to the estimated enzyme-available inhibitor concentration and CYP2D6 degradation half-life are shown in Figures 3 and 4, respectively. Sensitivities of the predicted interaction magnitude to both these input parameters were greatest when the victim drug was exclusively cleared by CYP2D6 (f_{2D6} ~1.0). For the victim drugs investigated here (f_{2D6} \leq 0.9), use of steady-state average total plasma paroxetine concentration following 10 mg BID or 20-30 mg QD dosing in the various reported paroxetine DDI studies (I_{ss} in the 80-160 nM range) or corresponding estimates of steady-state maximum portal venous unbound concentration (I_{inlet,max,u} 17-42 nM using Equation 5) instead of steady-state average unbound concentration (I_{ss,u} in the 4-8 nM range) did not significantly alter the predicted magnitude of the drug interaction, since this range of inhibitor exposures (4-160 nM) is outside of the dynamic range of the sensitivity curve (Figure 3). Figure 4 suggests that the use of a 14 hr half-life for CYP2D6 (as used for CYP3A4 (Mayhew et al., 2000) based on the half-life of rat CYP3A) instead of the 51 hr estimate of human CYP2D6 half-life would result in a 2-fold underprediction of interaction magnitude.
DISCUSSION

The predictive utility of a previously described mathematical model for IVIVE of DDIs resulting from MBI (Mayhew et al., 2000) has been demonstrated in this investigation for CYP2D6-inhibitory DDIs involving the SSRI antidepressant paroxetine. Across substrates, the accuracy of the model in predicting the magnitude of DDIs was approximately 90%. Thus, it can be inferred that previously published reports of underpredictions of the magnitude of DDIs perpetrated by paroxetine are explained by application of a competitive inhibition model for a mechanism-based enzyme inactivator. Nonlinear accumulation of paroxetine was also predicted by the model with an accuracy greater than 90%, suggesting that autoinactivation of CYP2D6 rather than reversible saturation of the enzyme may largely explain this pharmacokinetic phenomenon.

Nonspecific microsomal binding has been shown to significantly impact the determination of metabolic intrinsic clearance and enzyme kinetic parameters, with its consideration in the prediction of human clearance from in vitro metabolism data reproducibly improving predictive accuracy (Obach, 1997; Obach, 1999; Obach, 2000; Venkatakrishnan et al., 2000). More recently, it has been demonstrated that nonspecific microsomal binding can account for underestimation of inhibitor potency (i.e. overestimation of IC₅₀ or Kᵢ values) when dealing with lipophilic basic perpetrator drugs (Tran et al., 2002; Margolis and Obach, 2003), with the potential implication being erroneous underestimation of drug interaction risk. The risks of not considering nonspecific microsomal binding in estimating inhibitory potency may be greater when dealing with mechanism-based inactivators, due to the relatively high microsomal concentrations that are typically used during preincubation of the inactivator in these experiments. In this investigation, application of a “total” Kᵢ based on nominal inhibitor
concentrations added in vitro resulted in a gross underestimation of the magnitude of drug-drug interactions with several CYP2D6 substrates, in contrast to a 90% accurate prediction on average when an “unbound” $K_i$ estimate was used. When nonspecific binding was not considered, despite application of a MBI model that accounts for time-dependent loss of enzyme activity, a <2-fold increase in victim drug AUC is predicted upon paroxetine co-administration, significantly underestimating the observed 5-8-fold magnitudes of clinical interactions of paroxetine with CYP2D6 substrates.

A key assumption of the scaling model used here is that enzyme inactivation is the sole mechanism of these DDIs, since the reversible component of CYP2D6 inhibition by paroxetine is not considered. Further, it is also assumed that the effect of paroxetine is substrate-independent. This is based on the assumption that the MBI completely prevents substrate binding in a productive orientation at the enzyme active site(s), such that metabolic capacity towards all substrates would be equally affected. Atypical kinetics of dextromethorphan metabolism by CYP2D6 have been described (Yu et al., 2001), suggesting that multiple binding orientations are possible. Thus the theoretical possibility remains that the assumption of substrate independence of $K_i$ and $k_{\text{inact}}$ of mechanism-based inactivation of CYP2D6 by paroxetine may not be true, if the MBI results in an orientation-specific (and hence substrate-dependent) loss of substrate binding at the enzyme active site. Nevertheless, paroxetine reproducibly impairs the clearance of structurally diverse CYP2D6 substrates, which would suggest that the likelihood of paroxetine producing substrate-specific CYP2D6 inhibition is small and of limited clinical relevance.

The lack of information on the in vivo degradation rates of many human drug metabolizing CYPs represents a significant limitation that increases the uncertainty in risk
assessment, even when the desired in vitro data are available. In the present investigation, we have utilized an indirect approach of pharmacokinetic deconvolution to estimate the half-life of CYP2D6 in humans as ~51 hours. A comparable mean half-life estimate of 50 hours (range 25-80 hours) has been reported for human CYP2E1 by measuring the time-course of return of chlorzoxazone 6-hydroxylation clearance to baseline following administration of the rapidly cleared CYP2E1 inactivator disulfiram (Emery et al., 1999). Thus, the estimated CYP2D6 half-life of 51 hours is in excellent agreement with the degradation half-life estimated for another human CYP isoform within the same family, using a similar indirect approach. When information on enzyme half-life is not available, compound-specific analysis of the sensitivity of the IVIVE model to $k_{\text{degrad}}$ represents a useful approach to estimating the level of confidence in the predicted DDI magnitude. The same is applicable when there is uncertainty in the estimated enzyme-available inhibitor levels due to possible active hepatic uptake (von Moltke et al., 1998) or transient high concentrations during first-pass through the liver (Ito et al., 1998; Kanamitsu et al., 2000).

In the current retrospective analysis, the possibility of concentrative hepatic uptake of paroxetine has not been considered due to the lack of data on the extent of hepatocellular uptake of paroxetine, or identified biochemical mechanisms of active hepatic uptake transport of this drug. Paroxetine displays extensive hepatic distribution, with a liver/plasma total concentration ratio of ~26 (von Moltke et al., 1995). However, it is unknown whether this extent of distribution is significantly in excess of that expected from the relative extents of plasma and hepatic tissue binding. The choice of inhibitor concentration (systemic unbound vs. systemic total vs. estimated maximum unbound concentration at the inlet to the liver) did not significantly influence the predicted drug interaction magnitudes in the current retrospective analysis because
the limits of the range of these alternative exposure estimates following clinical doses of paroxetine were outside the dynamic range of the sensitivity curve for these victim drugs. Thus, a true test of the applicability of Iss,u of paroxetine as a surrogate of enzyme-available concentrations will require DDI studies employing subtherapeutic doses that yield exposures in the dynamic range of the sensitivity curve. The successful prediction of paroxetine DDIs in the current analysis therefore does not necessarily support the general applicability of unbound systemic concentrations of an inhibitor in the prediction of interaction magnitude. While the application of unbound systemic exposures is theoretically appealing and has been successful for many structurally distinct mechanism-based inactivators (Mayhew et al., 2000; Wang et al., 2004; Ernest II et al., 2005), there are many reversible inhibitors for which there is evidence to the contrary. A recent large retrospective analysis points to examples of underprediction of several interactions when this metric of inhibitor concentration was used in the context of a reversible inhibition model, although nonspecific microsomal binding in vitro was not considered in this analysis (Ito et al., 2004). Even after correction for in vitro microsomal binding and ruling out MBI, Levy and colleagues have shown that the in vivo potencies of fluvoxamine as an inhibitor of CYPs 1A2 and 2C19 are 10-40-fold greater than in vitro estimates (Yao et al., 2001; Yao et al., 2003). Thus, the level of certainty in enzyme-available inhibitor concentrations is a major determinant of the confidence in prospective prediction of DDI magnitude. It is thus recommended that when quantitative predictions of interaction magnitude are made from in vitro data, an analysis of sensitivity to the uncertainty in enzyme-available inhibitor concentrations over the clinical dose/exposure range be performed. When the sensitivity is large, it may be best to express the predicted interaction magnitude as a range derived from a range of estimated [I], weighted by their likelihood inferred from available
experimental evidence for the specific perpetrator drug (e.g. information on uptake transport, portal/systemic gradients, etc.).

In summary, the results of the present investigation suggest several important implications for clinical DDI risk assessment from in vitro data when dealing with enzyme inactivators. Mechanism-based inactivation is characterized not only by concentration-dependent, but also time-dependent enzyme inhibition. As a result, risk assessment cannot be performed by a simple comparison of inactivator potency (Ki) against its in vivo exposure. In the case of paroxetine, I/Ki ratio at the 30 mg QD dosing regimen is 0.02 -- an extremely low value that would typically be suggestive of minimal risk in the context of reversible inhibition. If total plasma concentrations are used, the I/Ki ratio is still only 0.4, a relatively low value that would only be suggestive of medium DDI risk (applying considerations that hold for reversible inhibition (von Moltke et al., 1998; Tucker et al., 2001; Bjornsson et al., 2003; Obach, 2003; Venkatakrishnan et al., 2003)). Paroxetine however produces 5-8-fold increases in the exposure of CYP2D6 substrates -- observations that are reconciled well by the MBI model, as shown in this investigation. In fact, for a substrate that relies entirely on CYP2D6 (f2D6 = 1), the model predicts that paroxetine (30 mg QD) will produce a 14-fold increase in victim drug AUC. Thus, when MBI is suspect, risk assessment is not straightforward. It requires thorough kinetic analysis and IVIVE using the appropriate scaling model (Mayhew et al., 2000), with consideration of factors such as microsomal binding in vitro, to enable unbiased estimation of inhibitor potency, to ultimately increase confidence in clinical DDI risk assessment from in vitro data.
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REFERENCES


Footnotes

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LEGENDS FOR FIGURES

**Figure 1.** Prediction of nonstationary kinetics of paroxetine by the MBI scaling model.
Symbols represent the observed plasma concentration-time data on Days 1 and 14 of paroxetine administration at 30 mg QD, from Sindrup et al. (1992). Solid lines are the best fit pharmacokinetic curve for the Day 1 data, and the expected Day 14 profile derived by superpositioning. The dashed and dotted lines represent the scaling model-predicted Day 14 profile from *in vitro* MBI kinetic parameters and the Day 1 profile, with and without consideration of *in vitro* nonspecific microsomal binding of paroxetine, respectively. Note the underprediction of nonlinearity in accumulation kinetics of paroxetine when *in vitro* microsomal binding is ignored during scaling.

**Figure 2.** Prediction of the desipramine-paroxetine interaction by the MBI scaling model.
Symbols represent the observed desipramine steady-state plasma concentration-time data prior to (open circles) and following (filled circles) steady-state co-administration of paroxetine (Alderman et al., 1997). The solid line is the best fit curve for the control desipramine steady-state pharmacokinetic data. The dashed and dotted lines represent the scaling model-predicted profiles following steady-state paroxetine co-administration from *in vitro* MBI kinetic parameters and the baseline pharmacokinetic parameters of desipramine, with and without consideration of *in vitro* nonspecific microsomal binding of paroxetine, respectively. Note the underprediction of drug interaction magnitude when *in vitro* microsomal binding is ignored during scaling.
Figure 3. Analysis of the sensitivity of the MBI scaling model-predicted drug-drug interaction magnitude (fold-increase in victim drug AUC) to the estimated enzyme-available inhibitor concentration (I) for victim drugs with $f_{2D6}$ values ranging from 0.5-1.0, using Equations 2-4. Note that the sensitivity increases at higher values of $f_{2D6}$ and is relatively small when $f_{2D6} \leq 0.90$, as is the case for all the victim drugs examined in this investigation.

Figure 4. Analysis of the sensitivity of the MBI scaling model-predicted drug-drug interaction magnitude (fold-increase in victim drug AUC) to the estimated CYP2D6 degradation half-life for victim drugs with $f_{2D6}$ values ranging from 0.5-1.0, using Equations 2-4. Note that the use of a 14 hour estimate of CYP2D6 half-life (as opposed to the 51 hour estimate) results in an approximately 2-fold underestimation of interaction magnitude for victim drugs examined in this investigation (mean $f_{2D6} \sim 0.87$).
Table 1. Estimation of the fractional contribution of CYP2D6 to the overall oral clearance of drug interaction victims of interest.

<table>
<thead>
<tr>
<th>Victim Drug</th>
<th>CYP2D6 EM/PM Pharmacokinetics</th>
<th>f_{2D6}</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atomoxetine</td>
<td>(CL_{po} \text{ (L/h/kg): 0.373 / 0.0357})</td>
<td>0.90</td>
<td>Sauer et al., 2003</td>
</tr>
<tr>
<td>Desipramine</td>
<td>(CL_{po} \text{ (L/h): 102 / 15})</td>
<td>0.85</td>
<td>Brøsen et al., 1993</td>
</tr>
<tr>
<td>R-Metoprolol</td>
<td>(AUC \text{ (ng·h/mL): 408 / 3802})</td>
<td>0.89</td>
<td>Lennard et al., 1983</td>
</tr>
<tr>
<td>S-Metoprolol</td>
<td>(AUC \text{ (ng·h/mL): 679 / 3431})</td>
<td>0.80</td>
<td>Lennard et al., 1983</td>
</tr>
<tr>
<td>Paroxetine(^a)</td>
<td>(AUC \text{ (nmol·h/L): 550 / 3910})</td>
<td>0.86</td>
<td>Sindrup et al., 1992</td>
</tr>
<tr>
<td>Perphenazine</td>
<td>(AUC \text{ (ng·h/mL): 2.18 / 16.8})</td>
<td>0.87</td>
<td>Dahl-Puustinen et al., 1989</td>
</tr>
<tr>
<td>Risperidone</td>
<td>(AUC \text{ (ng·h/mL): 32 / 278})</td>
<td>0.89</td>
<td>Huang et al., 1993</td>
</tr>
</tbody>
</table>

\(^a\) Estimated following a single dose of paroxetine
**Table 2.** Model-predicted magnitudes of drug-drug interactions of paroxetine with CYP2D6 substrates, without and with consideration of nonspecific microsomal binding. FDCL\textsubscript{2D6} and FDCL\textsubscript{tot} refer to the fractional decrement in oral clearance via the CYP2D6 component of net clearance, and the fractional decrement in total oral clearance, respectively; \(f_{2D6}\) refers to the fractional contribution of CYP2D6 to net apparent oral clearance; and \([PX]\) is the steady-state average plasma unbound concentration of paroxetine (PX).

<table>
<thead>
<tr>
<th>Victim Drug</th>
<th>(f_{2D6})</th>
<th>PX mg/d</th>
<th>[PX] nM</th>
<th>Predicted FDCL\textsubscript{2D6}</th>
<th>Predicted FDCL\textsubscript{tot}</th>
<th>Predicted AUC Ratio</th>
<th>Actual AUC Ratio</th>
<th>Percent Accuracy</th>
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<td></td>
<td>Microsomal binding considered?</td>
<td>Microsomal binding considered?</td>
<td>Microsomal binding considered?</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>NO</td>
<td>YES</td>
<td>NO</td>
<td>YES</td>
<td>NO</td>
</tr>
<tr>
<td>Atomoxetine</td>
<td>0.90</td>
<td>20</td>
<td>4.6\textsuperscript{a}</td>
<td>0.413</td>
<td>0.915</td>
<td>0.374</td>
<td>0.827</td>
<td>1.6</td>
</tr>
<tr>
<td>Desipramine</td>
<td>0.85</td>
<td>20</td>
<td>5.0\textsuperscript{a}</td>
<td>0.437</td>
<td>0.922</td>
<td>0.373</td>
<td>0.786</td>
<td>1.6</td>
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<tr>
<td></td>
<td></td>
<td>30</td>
<td>8.1\textsuperscript{a}</td>
<td>0.554</td>
<td>0.949</td>
<td>0.473</td>
<td>0.810</td>
<td>1.9</td>
</tr>
<tr>
<td></td>
<td></td>
<td>20</td>
<td>4.0\textsuperscript{a}</td>
<td>0.383</td>
<td>0.904</td>
<td>0.327</td>
<td>0.771</td>
<td>1.5</td>
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<tr>
<td>R-Metoprolol</td>
<td>0.89</td>
<td>20</td>
<td>5.9\textsuperscript{b}</td>
<td>0.478</td>
<td>0.933</td>
<td>0.427</td>
<td>0.833</td>
<td>1.7</td>
</tr>
<tr>
<td>S-Metoprolol</td>
<td>0.80</td>
<td>20</td>
<td>5.9\textsuperscript{b}</td>
<td>0.478</td>
<td>0.933</td>
<td>0.383</td>
<td>0.748</td>
<td>1.6</td>
</tr>
<tr>
<td>Perphenazine</td>
<td>0.87</td>
<td>20</td>
<td>5.9\textsuperscript{b}</td>
<td>0.478</td>
<td>0.933</td>
<td>0.416</td>
<td>0.811</td>
<td>1.7</td>
</tr>
<tr>
<td>Risperidone</td>
<td>0.89</td>
<td>20</td>
<td>5.9\textsuperscript{b}</td>
<td>0.478</td>
<td>0.933</td>
<td>0.423</td>
<td>0.825</td>
<td>1.7</td>
</tr>
</tbody>
</table>

\(\textsuperscript{a}\) Paroxetine steady-state plasma unbound concentration from the exposure measured in this study

\(\textsuperscript{b}\) Paroxetine steady-state plasma unbound concentration following 20 mg QD dosing (Kaye et al., 1989)

\(\textsuperscript{c}\) The 7-fold increase is a lower bound since half-life was prolonged but only AUC\textsubscript{0-8h} was measured; Predicted accuracy is thus not calculated (NC) for the perphenazine-paroxetine interaction.
FIGURE 1

Dose 1

Dose 14

Plasma Paroxetine Concentration (ng/mL)

Time (hours) post-first dose

- Observed Concentration-time Profiles
- Expected by Linear Accumulation
- MBI Model-predicted Profile
- MBI Model-predicted Profile (Nonspecific binding not considered)
FIGURE 2

Time post-dose (hours)

Desipramine Plasma Concentration (ng/mL)

- Steady-State Data in Control Phase
- Steady-State Data in Paroxetine (30 mg/d) Phase
- Fit of control phase dataset
- Predicted Kinetics in Paroxetine Phase Using MBI Model
- Predicted Kinetics in Paroxetine Phase Using MBI Model (Nonspecific binding not considered)
FIGURE 4

Estimated CYP2D6 Degradation Half-Life (hours)

Model-Predicted Fold-Increase in AUC

14 hr  51 hr

f_{2D6}

1.0

0.95

0.90

0.85

0.5

Estimated CYP2D6 Degradation Half-Life (hours)