Clonidine Pharmacokinetics in Pregnancy

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Number of Text Pages: 18
Number of Tables: 0
Number of Figures: 2
Number of References: 17
Number of Words:
  Abstract: 185
  Introduction: 127
  Discussion: 971

Non-standard Abbreviations:

CL/F = Apparent oral clearance; AUCτ = Area under the plasma concentration time curve over one dosing interval; CLrenal = Renal Clearance; Ae,τ = Amount of clonidine excreted unchanged in the urine over one dosing interval. CrCl = Creatinine clearance; fu = fraction unbound in plasma
ABSTRACT

The objective of this study was to determine the pharmacokinetic parameters of clonidine during pregnancy as compared to previously published data in non-pregnant subjects. Serial blood and urine samples were collected in 17 women during mid to late pregnancy over one steady-state dosing interval to determine clonidine noncompartmental pharmacokinetic parameters (n=17) and creatinine clearance. In six of these pregnant subjects, maternal and umbilical cord (venous and arterial) plasma samples were collected at the time of delivery for measurement of clonidine concentrations. Clonidine CL/F was found to be 440 ± 168 mL/min during pregnancy as compared to 245 ± 72 mL/min as previously reported in non-pregnant subjects (Cunningham et al., 1994, p < 0.0001). There was a strong correlation (r = 0.82, p < 0.001) between clonidine renal clearance, adjusted for variation in GFR, and urine pH. Umbilical cord to maternal plasma clonidine concentration ratios were 1.0 ± 0.1 (arterial) and 1.0 ± 0.1 (venous). In conclusion, clonidine is cleared more rapidly in pregnant women than in non-pregnant subjects. At the time of delivery, the fetus is exposed to similar plasma clonidine concentrations as the mother.
INTRODUCTION

Chronic hypertension during pregnancy is associated with complications, including fetal growth restriction, premature birth, preeclampsia, placental abruption, hypertensive crisis, and in some cases fetal demise. Very little information is available on the pharmacokinetic changes that occur for antihypertensive medications during pregnancy. Clonidine, a centrally acting, alpha-2 adrenergic agonist, has been used for the treatment of hypertension during pregnancy (Hartikainen-Sorri et al., 1987; Kobinger, 1978). In the non-pregnant population, approximately 60% of clonidine is eliminated unchanged by the kidneys (Davies et al., 1977). There are significant changes in kidney function during pregnancy, with creatinine clearance reported to increase by 50-60% (Davison and Noble, 1981). The pharmacokinetics of clonidine have not been studied during pregnancy. The objective of this study was to characterize the pharmacokinetics of clonidine during pregnancy.
METHODS

Following written informed consent, we examined steady-state pharmacokinetics of orally administered clonidine in the plasma of 17 pregnant women receiving clonidine during mid-pregnancy (22-26 weeks gestation) or late-pregnancy (34-38 weeks gestation). Six of these women participated in maternal and umbilical cord (arterial and venous) plasma sample collections at the time of delivery for measurement of clonidine concentrations. Gestational age was based on the last menstrual period or early ultrasound dating.

Subject Selection: This study was approved by the University of Washington Investigational Review Board. Subjects were eligible to participate if they were pregnant, ≥ 18 years of age, had a hematocrit ≥ 28%, and receiving oral clonidine for maternal hypertension.

Dosing Regimen: Oral clonidine therapy was not altered for study purposes. Dosages ranged from 0.15 to 0.30 mg per day, given in divided doses. The duration of the pharmacokinetic study was dependent on the subject’s dosage interval, which ranged from 6 to 12 hours. For three days prior to the study, oral clonidine tablets, from the same manufacturing lot, were provided. Subjects recorded the time of each dose on a study calendar. Subjects took their morning clonidine dose within 30 minutes of 8:00 a.m. Subsequent doses were taken within 30 minutes of scheduled administration times. On the morning of the pharmacokinetic study, subjects were asked to fast, with the exception of clear liquids, for five hours prior to clonidine administration until one hour after dosing.
Sample Collection: Serial plasma samples were collected at the following times: 0, 0.5, 1, 1.5, 2, 3, 4, 6, 8, 10 and 12 hours post dosing, or truncated to correspond to the subject’s dosing interval. Urine was collected over one dosing interval to determine clonidine renal clearance and creatinine clearance. Plasma samples, maternal and umbilical cord (arterial and venous), were collected at the time of delivery for quantifying clonidine concentrations. All samples were stored at -80°C until sample analysis.

Clonidine Concentrations: Plasma clonidine concentrations were quantified using a validated LC/MS assay. In brief, 250 µL plasma samples were mixed with 500 pg of internal standard (clonidine-d₄) in 500 µL of 0.1 M sodium carbonate (pH=9.3), and 3 mL of heptane:iso-amyl alcohol (98.5:1.5). Standards were prepared by spiking blank plasma with 0, 12.5, 25, 50, 100, 200, 250, 375, 500, and 750 pg of clonidine. Samples were shaken on a horizontal shaker for 15 minutes at room temperature. After centrifugation (1500 x g for 10 minutes), the samples were frozen on dry ice and the organic layer decanted to a clean culture tube. Sulfuric acid aliquots (2 mL of 0.05 M) were added to the organic layer and again shaken for 15 minutes. After centrifugation (1500 x g for 10 minutes), the samples were frozen on dry ice and the organic layer discarded. A 200 µL aliquot of 29.29% aqueous ammonium hydroxide was added to the aqueous layer along with 3 mL of heptane:iso-amyl alcohol (98.5:1.5) and shaken for 15 minutes. After centrifugation (1500 x g for 10 minutes), the samples were frozen on dry ice and the organic layer decanted to a clean culture tube. The organic layer was evaporated under a stream of nitrogen at 40°C. The residue was reconstituted with 75 µL of formic acid (12 mM):methanol (60:40), of which 10 µL were injected onto the LC/MS system.
Urine samples of 250 µL were mixed with 250 µL of 12 mM formic acid and 50 ng of internal standard (clonidine-d₄). Standards were prepared by spiking blank urine with 0, 1.25, 2.5, 6.25, 12.5, 18.75, and 25 ng of clonidine respectively. The samples were vortexed and 1 µL was injected onto the LC/MS.

The LC/MS system was an Agilent 1100 MSD (Agilent Technologies, Santa Clara, California, USA) consisting of binary pump, auto-sampler, column compartment and mass spectrometer. The HPLC column was a Phenomenex Synergi Polar RP80A, 250 mm x 2.0 mm, 4 µm. The column was maintained at 40° C. Mobile phase components were 12 mM formic acid (A) and methanol (B), with an initial flow rate of 0.25 mL/min. The initial mobile phase was maintained for 1 minute and consisted of 40% B for plasma samples and 35% B for urine samples. The percent of B was increased linearly over the next 4 minutes to 90% for plasma and 60% for urine and then maintained constant for 1 minute with a linear flow increase to 0.3 mL/minute for urine and plasma. For the return gradient, the percent of B was then decreased linearly over 1 minute to 40% (plasma) or 35% (urine), which was then maintained constant. The flow rate was maintained at 0.3 mL/minute for 1 minute and then linearly decreased over 1 minute to 0.25 mL/min and then maintained constant. The total run time was 12.5 minutes. For both plasma and urine assays, the mass spectrometer was operated in the positive electrospray mode and set for selective ion monitoring of m/z 230 (clonidine) and m/z 236 (clonidine-d₄) with dwell time of 289 msec for each ion.

**Pharmacokinetic Parameters:** Steady-state pharmacokinetic parameters were determined using standard noncompartmental techniques. The area under the plasma clonidine...
concentration time curve over one dosing interval (AUCτ) was calculated using the linear trapezoidal rule. Apparent oral clearance was calculated by CL/F = Dose/AUCτ. Clonidine renal clearance was calculated by CL_{renal} = Ae,τ/AUCτ, where Ae,τ = the amount of clonidine excreted unchanged in the urine over one dosing interval. Creatinine clearance was calculated by CrCl = (Volume_{Urine}•Creatinine_{Urine})/(Creatinine_{Serum}•τ). Clonidine renal clearance adjusted for GFR was calculated by CL_{renal adjusted for GFR} = CL_{renal} – fu•CrCl where the fraction unbound in plasma (fu) was assumed to be 0.80 (Hulter, 1979). In all our subjects, the relatively short dosing interval did not allow accurate estimation of clonidine half-life, which has previously been reported to be 11.1 ± 3.3 and 10.8 ± 2.2 hours in non-pregnant healthy adults (Cunningham et al., 1994; Fujimura et al., 1994).

Statistics: Our clonidine pharmacokinetic parameters in pregnancy were compared with previously published parameters in healthy, non-pregnant volunteers (CL/F, Cunningham et al., 1994 and Porchet et al., 1992; CL_{renal}, Fujimura et al., 1994). Linear regression was used to determine correlation between creatinine clearance and clonidine renal clearance. Unpaired Student’s T test was used to compare the pregnant and non-pregnant groups. A p value < 0.05 was considered significant. All results are reported as mean ± standard deviation.
RESULTS

Subjects: Seventeen subjects (10 White, 4 Black, 1 Hispanic/Latina, 1 Hispanic/Latina/Native American, and 1 Native American/White) completed the pregnancy phase of the study. The average age was $32.1 \pm 8.0$ years old, body weight $97.6 \pm 21.5$ kg (range 71.0 to 153.6 kg), and height $163.9 \pm 8.0$ cm. Five subjects were studied during mid-pregnancy (22-26 weeks gestation) and twelve subjects during late-pregnancy (34-38 weeks gestation). One subject completed both a mid- and late-pregnancy study day. The pharmacokinetic parameters were similar for each study day and thus, to eliminate bias, we only included her late-pregnancy study day results. Six of these pregnant women were studied at the time of delivery.

Pharmacokinetics: Figure 1 is a representative clonidine plasma concentration-time curve. The average maternal peak clonidine concentration was $574 \pm 279$ pg/mL. Mean apparent oral clearance of clonidine was significantly higher in pregnant women ($440 \pm 168$ mL/min) compared with the literature reported non-pregnant population ($245 \pm 72$ mL/min, $p < 0.0001$; $233 \pm 38$ mL/min, $p < 0.0001$) (Cunningham et al., 1994 {male n=4, female n=6, 20-42 years, radioimmunoassay}; Porchet et al., 1992 {healthy volunteers n=10, 23-34 years, radioimmunoassay}). Creatinine clearance was $166 \pm 75$ mL/min in the pregnant women. Interestingly, clonidine renal clearance in pregnancy ($153 \pm 67$ mL/min) was not significantly different from that seen in the non-pregnant population ($183 \pm 55$ mL/min, $p = 0.3$) (Fujimura et al., 1994 {male n=8, 26-38 years, 61-81 kg, GC/MS and GC/electron capture detection assays}). Only $36 \pm 11\%$ of the clonidine was excreted unchanged in the urine in pregnancy compared with $59 \pm 18\%$ in the non-pregnant population ($p < 0.0001$) (Arndts, 1983{n=19, radioimmunoassay}). There was a poor correlation ($r = 0.26$) between clonidine renal clearance
and creatinine clearance (data not shown). However, a strong correlation (r = 0.82, p < 0.001) existed between clonidine renal clearance, corrected for GFR, and urine pH (range 5.8-7.5, Figure 2).

At parturition, the mean umbilical cord to maternal plasma clonidine concentration ratios were 1.0 ± 0.1 (venous) and 1.0 ± 0.1 (arterial), which remained fairly constant between 4 and 10 hours post last maternal clonidine dose (data not shown). The average maternal clonidine plasma concentration at the time of delivery was 404 ± 164 pg/mL (range 164-603 pg/mL). The average venous and arterial umbilical cord concentrations at the time of delivery were 394 ± 142 pg/mL (range 191-586 pg/mL) and 402 ± 147 pg/mL (range 204-614 pg/mL) respectively.
DISCUSSION

The obstetricians at the University of Washington have frequently observed a diminished response to clonidine in pregnant patients near the end of a standard 12-hour dosing interval. As a result, they often shorten the dosing interval to 6 – 8 hours (i.e., prescribe more frequent dosing) when treating maternal hypertension. Since kidney function, as measured by creatinine clearance, is known to increase during pregnancy and clonidine is predominantly eliminated by the kidneys in non-pregnant subjects, we hypothesized that pregnancy may alter the pharmacokinetics of clonidine. Indeed, the pregnant subjects in our study have a significantly higher apparent oral clearance of clonidine than previously reported in non-pregnant healthy male and female subjects with similar age ranges. It is possible that the difference may in part reflect clonidine assay specificity in that we utilized an LC/MS assay and the previously published studies utilized radioimmunoassays. However, both radioimmunoassays demonstrate cross-reactivity with only one of clonidine’s metabolites. The cross-reacting metabolite is present in very low concentrations in human plasma making it very unlikely to be the explanation for the differences seen in this study. (Darda et al., 1978) The question then becomes what is the mechanism underlying the increase in apparent oral clearance of clonidine. Possible mechanisms include changes in renal clearance, metabolic clearance, plasma protein binding, and/or bioavailability.

Despite the increase in creatinine clearance seen during pregnancy, clonidine renal clearance was not significantly different than previously reported in healthy male volunteers. The situation may be complicated by the dependence of net secretion and/or reabsorption of clonidine upon urine pH. Clonidine is a basic drug with a pKa reported to be 8.05 (Dollery, 1991a). At low
urine pH, clonidine is ionized (protonated) resulting in net renal secretion, whereas at high urine pH, net renal reabsorption occurs. Thus, the rather large variation in urine pH observed in this study might have masked the impact of increased renal filtration seen during pregnancy resulting in no apparent change in clonidine renal clearance. In any case, clonidine renal clearance appears not to be the principal reason for the increase in clonidine apparent oral clearance.

Clonidine is a low hepatic extraction ratio drug. (Davies et al. 1977) Change in hepatic blood flow is not a likely reason for the increased clonidine apparent oral clearance. For orally administered medications, changes in hepatic blood flow should not change the oral AUC for either high or low extraction ratio drugs. Although we expect that intestinal blood flow may be increased as a result of the increased cardiac output during pregnancy, this would likely result in increased bioavailability and higher plasma concentrations rather than the lower concentrations seen during pregnancy. Plasma protein binding is unlikely to be the reason for the 1.8-fold increase in clonidine apparent oral clearance because it is only 20% bound in the plasma. Even if the percent bound for clonidine decreased to zero during pregnancy, it would not explain the changes seen in clonidine apparent oral clearance. Although not expected for clonidine, a drug with presumably good oral absorption, it is possible that the extent of absorption could be decreased in pregnancy.

Accordingly, alteration in hepatic and/or intestinal intrinsic clearances are the most likely reasons for the higher apparent oral clearance of clonidine during pregnancy. A lower recovery of unchanged drug in the urine was observed in our pregnant women compared to data reported for non-pregnant subjects (36% versus 59%), which is entirely consistent with the hypothesis
that pregnancy shifts the major route of elimination for clonidine from the kidneys to organs involved in drug metabolism (Arndts, 1983, Davies et al., 1977). The enzymes involved in clonidine metabolism are currently unknown; however, pregnancy does increase the activities of CYP3A, CYP2D6 and CYP2C9 (Hebert et al., 2008; Yerby et al., 1990; Tracy, 2005). Clonidine is metabolized to 5 metabolites including: p-hydroxy-clonidine; dichlorophenylguanidine; 1-(2,6-dichloro-4-hydroxyphenyl)-guanidine; 2-[(2,6-dichlorophenyl)-imino-a1-imidazolidine-4-one and 2-[2,6-dichloro-4-hydroxyphenyl)-imino]-imidazolidine-4-one. (Darda et al., 1978) Clonidine’s primary metabolite (p-hydroxy-clonidine) is < 10% of unchanged drug in the urine from non-pregnant subjects. (Darda et al., 1978, Arndts et al., 1979) The metabolites of clonidine have been reported to be inactive. (Lowenthal DT, 1980) Further research is underway to characterize which drug-metabolizing enzymes are involved in the non-renal clearance of clonidine.

In our study, clonidine plasma concentrations in the umbilical cord were approximately the same as in the maternal samples. This is similar to data reported by others (Hartikainen-Sorri et al., 1987; Boutroy et al., 1988). Boutroy et. al. reported umbilical to maternal concentration ratio to be 0.87 ± 0.14 ng/mL and three out of five newborns exposed in utero to clonidine (maternal dosage 0.3-0.45 mg/day) experienced hypertension during the first 48 hours of life. Hypertension in these infants may in part be related to genetic predisposition, although it could also be attributed to a paradoxical neonatal reaction to clonidine. Rebound hypertension can be ruled out since clonidine concentrations in the infants were similar at the time of birth and at 24 hours of age.
In conclusion, the clinical observation of decreased clonidine effect at the end of a 12-hour dosing interval probably reflects the increased apparent oral clearance during pregnancy, which may necessitate higher or more frequent clonidine dosing. The increase in apparent oral clearance of clonidine during pregnancy is attributable to a change in its non-renal clearance. Renal clearance of clonidine did not show any change despite the increase in creatinine clearance. The apparent lack of change in clonidine renal clearance may be due to the high variability in urine pH, subsequently altering tubular reabsorption of clonidine. Non-renal clearance plays a much more significant role in overall clonidine clearance during pregnancy than in the non-pregnant population. Further research is needed to characterize clonidine pharmacodynamics and metabolism during pregnancy. Moreover, clonidine does cross the placenta and similar concentrations are measured in maternal and umbilical cord plasma. Fetal and neonatal safety requires further investigation.
REFERENCES


FOOTNOTES

This work was supported in part by: NIH/NICHD through support of the Obstetric-Fetal Pharmacology Research Unit Network grant #5U10HD047892; UW Multidisciplinary Predoctoral Research Training Program – a Roadmap Initiative from the National Institutes of Health/National Center for Research Resources (T32 RR023256), P. Mitchell & S. Marshall, Program Directors; NIH grant #M01RR00037 and NIH/NCRR grant #RR023256.

LEGENDS FOR FIGURES

Figure 1. Representative steady-state oral clonidine plasma concentration time profile during late pregnancy (34.1 weeks gestation).

Figure 2. Correlation between clonidine renal clearance, corrected for GFR, and urine pH during mid-pregnancy (22-26 weeks gestation, n=5) and late-pregnancy (34-38 weeks gestation, n=12).
Figure 1

Clonidine Subject #8 Late Pregnancy

Clonidine Plasma Concentration (pg/mL) vs. Time Post-Dosing (Hr)
Figure 2

Clonidine renal clearance corrected for GFR vs. Urine pH

\[ y = -145x + 982 \quad r = 0.82, \quad P < 0.001 \]