Identification of Human UGT2B7 As the Major Isoform Involved In the O-Glucuronidation of Chloramphenicol

Mei Chen, Barbara LeDuc, Stephen Kerr, David Howe and David A. Williams

Massachusetts College of Pharmacy and Health Sciences, Boston, MA (MC, BL, SK, DAW) and Vertex Pharmaceuticals Incorporated, Cambridge, MA (DH)
O-Glucuronidation of Chloramphenicol

Address correspondence to:

Dr. David A Williams,
Massachusetts College Pharmacy and Health Sciences,
Department of Pharmaceutical Sciences,
179 Longwood Ave,
Boston, MA 02115 USA.
Voice tel. 617 732 2934
Fax : 617-732-2228
E-mail: david.williams@mcphs.edu

Abbreviations: LC-MS/MS, liquid chromatography-tandem mass spectrometry; MRM, Multiple Reaction Monitoring; DMSO, dimethyl sulfoxide; HLM, pooled human liver microsomes; UDPGA, UDP-glucuronic acid; UGT, UDP-glucuronosyltransferase; AZT, azidothymidine; CP, chloramphenicol.
ABSTRACT:

Chloramphenicol (CP), a broad spectrum antibiotic, is eliminated in humans by glucuronidation. The primary UGT enzymes responsible for CP O-glucuronidation remain unidentified. We have previously identified the 3-O-CP (major) and 1-O-CP (minor) glucuronides by β-glucuronidase hydrolysis, LC/MS/MS and 1D/2D-H-NMR. Reaction phenotyping for the glucuronidation of CP with 12 expressed human liver UGT isoforms has identified UGT2B7 as having the highest activity for 3-O- and 1-O-CP glucuronidation with minor contributions from UGT1A6 and UGT1A9. The kinetics of CP 3-O-glucuronidation by pooled HLM exhibited biphasic Michaelis-Menten kinetics with the apparent high affinity $K_m^1$ and low affinity $K_m^2$ values of 46.0 and 1027 µM, whereas expressed UGT2B7 exhibited Michaelis-Menten kinetics with the apparent $K_m$ values 109.1 µM. The formation of 1-O-CP glucuronide by pooled HLM and expressed UGT2B7 exhibited substrate inhibition kinetics with apparent $K_m$ values of 408.2 µM and 115.0 µM, respectively. AZT and hyodeoxycholic acid (substrates of UGT2B7) inhibited 3-O- and 1-O-CP glucuronidation in pooled HLMs. In 10 donor HLM preparations, both CP 3-O- and CP 1-O-glucuronidation showed a significant correlation with AZT glucuronidation (UGT2B7) ($r_s = 0.85$ and $r_s = 0.83$, respectively) at 30 µM CP while no significant correlation was observed between CP 3-O-glucuronidation and serotonin glucuronidation (UGT1A6) or propofol glucuronidation (UGT1A9) at this CP concentration. These results suggest that UGT2B7 is the primary human hepatic UGT isoform catalyzing 3-O- and 1-O-CP glucuronidation with minor contributions from UGT1A6 and UGT1A9.
Introduction

CP (Fig. 1) is a broad-spectrum antibiotic which has been used to treat serious infections in humans, often when other antibiotics have failed. However, the use of this antibiotic is limited because of its potentially life-threatening side effects, such as aplastic anemia in adults or Grey Baby syndrome in premature and newborn infants (Suhrland and Weisberger, 1963). Despite these dangers, CP is still widely used in developing countries due to the increased cost and decreased availability of newer antibiotics (Brook 2004).

Although CP has a complex metabolic pattern, glucuronidation has been recognized as the major pathway for adult human elimination with up to 85% of CP appearing in the urine as its glucuronide (Glazko et al. 1950; Glazko 1966). Nonetheless, the UGT isoforms responsible for this glucuronidation remain unidentified. CP is rapidly absorbed having an oral bioavailability of 80-90%, indicating little pre-systemic glucuronidation and with 5-15% of CP doses excreted unchanged in the urine. The drug is distributed into all tissues including the brain and spinal cord. Its elimination half-life of 4-6 hours suggests no enterohepatic cycling occurs.

The incidence of toxic effects of CP in neonates was found to be strongly related to age, as nine out of ten babies who showed toxic effects were less than 9 days old (Kaufmann et al., 1981). Subsequent pharmacokinetic studies in these neonates confirmed an elevation of plasma CP levels suggesting the parent drug was responsible for the toxic effects of the Grey Baby syndrome (Weiss et al., 1960; Kaufmann et al., 1981). These clinical findings illustrate the
importance of understanding the impact of age-dependent developmental differences on the
activity of drug-metabolizing liver enzymes and drug clearance which can markedly alter the
pharmacokinetics of a drug in infants as compared with adults (Kaufmann et al., 1981; Kearns et
al., 2003). Because the mechanisms underlying these age-dependent developmental differences
are largely unknown, the identification of the main human liver UGT isoforms involved in drug
clearance is crucial to understanding the age-related variations in plasma drug concentration that
lead to altered drug efficacy and/or toxicity in infants, which may necessitate an age-based dose
adjustment.

Glucuronidation is inefficient at birth (Kaufmann et al., 1981; Kearns et al., 2003). As a
result of low UGT isoform activity at birth, the clearance half-life for CP in newborns < 3 months
postnatal is ~10 hours, at 6 months 4-6 hours, and at 4 years of age, 4-6 hours (Miles, 1983). The
reduced glucuronidation and thus slow elimination of CP has been cited as being responsible for
the delayed systemic clearance and subsequent toxicity of this drug in newborns (Lambdin et al.,
1960; Suhrland et al., 1963; Weiss et al., 1960).

Despite the widespread clinical use of CP for over five decades, the role of the liver in
human CP O-glucuronidation appears not to have been characterized. Furthermore, the UGT
isoforms contributing to CP O-glucuronide formation have yet to be identified. We have
previously identified the CP 3-O- (major) and CP 1-O-glucuronide (minor) conjugates (Chen et
al., 2007). The determination of the specific human UGT isoforms responsible for the O-
glucuronidation of CP could provide evidence into the reasons for the newborn toxicity of CP.
Therefore, in this study we sought to gain insight into the in vivo hepatic CP clearance by 1) phenotyping the specific UGT isoforms involved in the in vitro CP 3-O- and 1-O-glucuronidation by expressed human UGTs, 2) determining the kinetics of CP 3-O- and 1-O-glucuronidation in pooled HLMs and by expressed human UGTs, and 3) confirming the contribution of the UGTs for CP O-glucuronidation from inhibition studies with UGT selective substrates as inhibitors and correlation studies in donor human HLMs. We now report our results from phenotyping with expressed UGTs and from inhibition and correlation studies which indicate that UGT2B7 is the primary UGT isoform catalyzing CP 3-O- and 1-O-glucuronidation.
Materials and Methods

**Chemicals and Reagents.** Unless otherwise indicated, reagents including CP, TRIS HCl, UDPGA, magnesium chloride, alamethicin, dimethyl sulfoxide, AZT, serotonin, propofol, hyodeoxycholic acid were purchased from Sigma-Aldrich (St. Louis, MO). AZT glucuronide sodium salt was purchased from Toronto Research Chemicals, Inc. (Ontario, Canada). HPLC grade acetonitrile and methanol were obtained from J.T. Baker (Phillipsburg, USA). Pooled mixed gender HLM (n=20) and expressed UGTs are microsomes from baculovirus-infected insect cells expressing human UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A7, UGT1A8, UGT1A9, UGT1A10, UGT2B4, UGT2B7, UGT2B15, UGT2B17 (Supersomes®) and UGT control Supersomes® were purchased from BD Gentest (Woburn, MA). The expressed UGT2A enzymes, as well as UGT1A5, UGT2B10, UGT2B11 and UGT2B28 were not available from BD Gentest and thus were not tested. The pooled HLMs and expressed enzymes were kept frozen at -80°C and used shortly after receiving the enzymes. The protein content of all Supersomes® was 5 mg/mL. Human liver microsomes from 10 individual donors were purchased from BD Gentest (Woburn, MA). All other chemicals were of reagent grade or of the highest purity available commercially.

**Chloramphenicol O-Glucuronidation by Pooled HLMs.** CP (300 μM) was incubated with alamethicin-activated pooled HLMs fortified with 20 mM UDPGA as described (Chen et al., 2007). The CP 3-O- and CP 1-O-glucuronide concentrations in the incubation mixture were determined by LC/MS/MS using biosynthesized CP 3-O-glucuronide as the standard (Chen et
al., 2007). Very briefly, the quantitation of the CP glucuronides was achieved using a LC-MS/MS system, which was equipped with an Agilent model 1100 series HPLC system and a Sciex API 4000 tandem mass spectrometer (Applied Biosystems, Foster City CA). LC separation was performed on a Synergi Polar-RP column (Phenomenex, Torrance, CA) with a gradient consisting of mobile phase A water/formic acid (100:0.1, v/v) and mobile phase B acetonitrile/formic acid (100:0.1, v/v) at a flow rate 0.5 mL/min. Negative MRM mode was used to monitor CP, CP glucuronides and warfarin (internal standard) at mass transitions $m/z$ of 321.2/152.1 for CP; at $m/z$ of 497.2/193.1 for CP glucuronides; and warfarin at 307.1/250.1 (Chen et al., 2007). Incubations without UDPGA or with boiled pooled HLMs were performed simultaneously and served as negative controls. Zero-time, zero-protein, and zero-substrate incubations served as blanks.

**Chloramphenicol O-Glucuronidation by Expressed Human UGT Isoforms.** CP 3-O- and 1-O-glucuronidation was measured in alamethicin-activated Supersomes® expressing human UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A7, UGT1A8, UGT1A9, UGT1A10, UGT2B4, UGT2B7, UGT2B15 (all 5 mg protein/mL) or UGT2B17 (0.5 mg protein/mL) Supersomes® with 30 or 300 μM CP and 20 mM UDPGA. UGT Control Supersomes® (0.5 mg protein/mL) served as the negative control. Incubations were done in duplicate, and samples were directly analyzed by the described LC-MS/MS method for pooled HLMs. All of the incubations were performed at 37°C for 50 min.

**Enzyme Kinetic Data Analysis.**
All data points represent the mean of triplicate estimations. The kinetic parameters for CP 3-O- and 1-O-glucuronidation were calculated by fitting the untransformed experimental data to the following enzyme kinetic equations using Prism 5.2 (GraphPad, SanDiego, CA), designed for nonlinear regression analysis. The selection of the “best-fit” kinetic model was based on comparison of the sum-of-squared residuals, SD of fit, coefficient of determination ($r^2$), and F test (Prism 5.2, GraphPad, San Diego, CA).

The Michaelis–Menten equation,

$$v = \frac{(V_{\max} \times [S])}{(K_m + [S])}$$ \hspace{1cm} (1)

where $v$ is the rate of metabolite formation, $V_{\max}$ is the maximum velocity (as pmol product per min per mg microsomal or cell lysate protein), $K_m$ is the Michaelis constant (substrate concentration at 0.5 $V_{\max}$), and $[S]$ is the substrate concentration.

The two-enzyme Michaelis–Menten equation,

$$v = \left\{ \frac{(V_{\max1} \times [S])}{(K_{m1} + [S])} \right\} + \left\{ \frac{(V_{\max2} \times [S])}{(K_{m2} + [S])} \right\}$$ \hspace{1cm} (2)

where $K_{m1}$ and $K_{m2}$ are the apparent high and low affinity components and $V_{\max1}$ and $V_{\max2}$ are the apparent maximum velocities for the high and low affinity components, was used when the Eadie-Hofstee plot exhibited a biphasic shape (Houston and Kenworthy 2001; Court et al., 2001; Yamanaka et al., 2005).

The two-site model equation (Houston and Kenworthy, 2000; Uchaipichat et al., 2004),

$$\frac{v}{V_{\max}} = \frac{(S/K_s + \beta S^2/\alpha K_s^2)}{(1+2S/ K_s + S^2/ \alpha K_s^2)}$$ \hspace{1cm} (3)

where $K_s$ represents the binding affinity and $\alpha$ and $\beta$ are modifying factors that reflect
changes in $K_s$ and $K_p$ (the catalytic rate constant), respectively.

The intrinsic clearance ($\text{CL}_{\text{int}}$) was calculated as $\text{CL}_{\text{int}} = \frac{V_{\text{max}}}{K_m}$ and expressed as L per min per mg protein. Enzyme activities were calculated by dividing metabolite concentration by protein concentration and incubation time and expressed as picomoles per minute per mg of protein.

When the Eadie-Hofstee plot exhibited a sigmoidal “U” shaped curve, the Hill equation (eq. 4) was selected for estimating $S_{50}$ and $V_{\text{max}}$ (Weiss, 1997):

$$v = \frac{V_{\text{max}} \times S^n}{(S_{50}^n + S^n)}$$

(4)

where $S_{50}$ is the substrate concentration resulting in 50% $V_{\text{max}}$ (analogous to $K_m$ in the Michaelis-Menten equation) and $n$ is the Hill coefficient. The maximum clearance ($\text{CL}_{\text{max}}$) [$\text{CL}_{\text{max}} = \frac{V_{\text{max}} \times (n-1)/\{S_{50} \times n(n-1)^{1/n}\}}{S_{50}}$] provides an estimate of the highest clearance attained, i.e., when the enzyme is fully activated before saturation occurs (Houston and Kenworthy, 2000).

When the Eadie-Hofstee plot showed a hyperbolic shape with no clearly defined plateau at high substrate concentrations and a “hook” curve at high substrate concentrations, the uncompetitive substrate inhibition model (eq. 5) was used (Houston and Kenworthy, 2000; Venkatakrishnan et al. 2001).

$$v = \frac{V_{\text{max}} \times S}{(K_m + S) \times (1 + S/K_{si})}$$

(5)

where $K_{si}$ is the constant describing the substrate inhibition interaction.

**Inhibition of Chloramphenicol O-Glucuronidation by AZT, Serotonin, and Propofol.** The 3-O- and 1-O-glucuronidation of CP in pooled HLMs was studied using known
selective UGT isoform probe substrates as inhibitors (Miners et al. 2004; Miners et al. 2006). Of the expressed UGT isoforms glucuronidating CP at the 3-hydroxyl position, expressed UGT1A6, UGT1A9, and UGT2B7 had exhibited significant activity and were therefore used for the inhibition studies. Since only expressed UGT2B7 exhibited activity for glucuronidating CP at the 1-hydroxyl group, it was used for the inhibition studies. The selective inhibitor for UGT1A6 was serotonin (Krishnaswamy et al., 2003; Court, 2005), for UGT1A9, propofol (Court et al., 2001; Yamanaka et al., 2005), and for UGT2B7, hyodeoxycholic acid (Pillot et al. 1993, Barre et al. 2007), and AZT (Court et al., 2003). Serotonin and AZT were each dissolved in water, hyodeoxycholic acid was dissolved in 1:1 acetonitrile and water, and propofol in DMSO. The concentration range for hyodeoxycholic acid or propofol was 0.1 to 1000 µM, for serotonin 100-40000 µM and for AZT 0.1 to 5000 µM. All the incubations of inhibitors were carried out as described above for CP O-glucuronidation in pooled HLM. The 3-O-CP- and 1-O-CP-UGT activities at 100 µM CP in pooled HLMs in the absence of any probe substrate as inhibitor were used as controls. The uninhibited glucuronidation activities were calculated as a percentage of control activity.

**Correlation Analysis.** The 3-O- and 1-O-glucuronides produced from 30 or 3000 µM CP concentrations were measured in a bank of HLMs isolated from 10 individual donors, using the same incubation conditions as previously described for CP O-glucuronidation by pooled HLM. Other UGT activities measured included propofol glucuronidation (50 µM propofol; UGT1A9 substrate), AZT glucuronidation (2000 µM AZT; UGT2B7 substrate), and serotonin
glucuronidation (4000 µM serotonin; UGT1A6 substrate). The concentrations of these probe substrates approximated their $K_m$ values in pooled HLMs, which were 1331 µM for AZT, 90 µM for propofol and 4854 µM for serotonin (data not shown). AZT, propofol, serotonin, and their glucuronides were analyzed by the previously described LC-MS/MS method for the glucuronidation of CP. The AZT glucuronide was analyzed with a standard curve of AZT glucuronide (Toronto Research Chemicals, Inc., Ontario, Canada) spiked in the incubation matrix by LC-MS/MS in positive ionization MRM mode at mass transitions $m/z$ 444/268. The concentrations of serotonin, propofol and their glucuronides in the incubation mixture were determined using the method as described by Yamanaka (Yamanaka et al., 2005) and analyzed by the described LC-MS/MS method with serotonin and propofol standard curves. Serotonin and propofol glucuronides were monitored in negative MRM mode at mass transitions $m/z$ 351/175 and $m/z$ 353/177, respectively.

Correlation analyses between these CP-UGT activities and the other UGT activities in 10 individual donor HLMs were determined by Spearman’s rank method. When the $r_s$ value was greater than or equal to 0.5 and the $P$ value was less than 0.05, the correlations were considered statistically significant.

**Results**

**Chloramphenicol O-Glucuronidation by Expressed Human UGTs.** UGT1A1, UGT1A3, UGT1A4, UGT1A6, UGT1A7, UGT1A8, UGT1A9, UGT1A10, UGT2B4, UGT2B7, UGT2B15 and UGT2B17 were screened for CP 3-O- and 1-O-glucuronidation activity at 30 µM CP
(Ogilvie et al., 2008), the approximate therapeutic plasma concentration (National Toxicology Program Report 2002) and at 300 μM CP, the approximate toxic plasma concentration (National Toxicology Program Report 2002) (Fig 2). CP 3-O-glucuronidation was observed for UGT 1A6, 1A9, 2B4 and 2B7 with activities >1 pmol/min/mg at 30 μM CP concentration. At 300 μM CP concentration, the additional isoforms UGT 1A3 and 2B17 were observed to catalyze the 3-O-glucuronidation of CP (Fig 2A). The isoforms UGT 1A1, 1A4, 1A7, 1A8, 1A10 and 2B15 did not form the CP 3-O-glucuronide. Kinetic studies were not performed with UGT2B4 and UGT2B17 because these enzymes have exhibited low or negligible activity towards drugs and other xenobiotics (Miners et al., 2006). Thus, CP 3-O-glucuronidation kinetics were characterized for UGT 1A6, 1A9 and 2B7. As shown in Fig. 2A and 2B, UGT2B7 showed highest CP 3-O- and 1-O-glucuronosyltransferase activities at 30 and 300 μM concentrations of CP. Negligible activities for the formation of CP 1-O-glucuronide were observed for all of the other expressed UGT isoforms.

The Michaelis-Menten model for the formation of the CP 3-O-glucuronide by expressed UGT2B7 (Fig 3A) was best fitted to the experimental data (Fig 3A, Eadie-Hofstee plot insert), and the derived kinetic parameters were $K_m = 109.1 \pm 5.2 \mu M$ and $V_{max} = 46.9 \pm 0.4$ pmole/min/mg of protein and are summarized in Table 1. The apparent $CL_{int}$ for the formation of 3-O-CP glucuronide by UGT2B7 is 0.43 L/min/mg of protein. The sigmoidal model (Hill equation, Weiss, 1997) for the UGT1A9 catalyzed CP 3-O-glucuronidation was best fitted to the experimental data (Fig 4A, Eadie-Hofstee plot inset) and the derived kinetic parameters for $S_{50}$
was 714.4± 30.2 µM and 41.9 ± 0.6 pmole/min/mg of expressed protein, with a positive Hill coefficient of 1.4 suggestive of positive cooperativity kinetics (Houston and Kenworthy, 2000) and are summarized in Table 1. The apparent CL_{max} for the formation of 3-O-CP glucuronide by UGT1A9 is 0.059 L/min/mg of protein. Similarly for UGT1A6 catalyzed CP 3-O-glucuronidation, the Hill equation was best fitted to the experimental data (Fig 4B, Eadie-Hofstee plot inset) and the derived kinetic parameters for S_{50} was 1557.0 ± 122.0 µM, V_{max} 12.9 ± 0.4 pmole/min/mg of protein and Hill coefficient of 1.2, with an apparent CL_{max} of 0.0053 L/min/mg of protein and summarized in Table 1.

On the other hand, 1-O-CP glucuronidation by UGT2B7 exhibited atypical kinetics characteristic of substrate inhibition when initially fitted to the Michaelis-Menten equation (Fig 3B, Eadie-Hofstee plot inset) (Houston and Kenworthy, 2000). Thus the substrate inhibition equation was best fitted to the experimental data and the derived kinetic parameters by UGT2B7 were for K_{m} 115.0± 14.2 µM and V_{max} 5.4 ± 0.2 pmole/min/mg of expressed protein and are summarized in Table 1. The apparent CL_{int} value was 0.047 L/min/mg of protein.

Kinetics of Chloramphenicol O-Glucuronidation by Pooled Human Liver Microsomes.

CP 3-O-glucuronidation by pooled HLM exhibited atypical kinetics was characteristic of biphasic kinetics (Fig 5A, Eadie-Hofstee plot inset) (Houston and Kenworthy, 2001). The glucuronidation data were therefore analyzed using the two-enzyme Michaelis-Menten equation (eq. 2), the two-site model equation (eq. 3) or the Hill equation (eq. 4). The two-enzyme
Michaelis–Menten equation was best fitted to the experimental data and the derived kinetic parameters for the high-affinity components (\(K_{m1}\) and \(V_{max1}\)) were \(46.0 \pm 27.0\) µM and \(37.9 \pm 13.0\) pmol/min/mg of protein, respectively, whereas those for the low-affinity components (\(K_{m2}\) and \(V_{max2}\)) were \(1027.0 \pm 129.8\) µM and \(232.0 \pm 11.2\) pmol/min/mg of protein, respectively, and are summarized in Table 1. The apparent intrinsic clearance \(CL_{int1}\) for the high affinity component of CP 3-O-glucuronidation in pooled HLM was \(0.82\) L/min/mg of protein.

CP 1-O-glucuronidation by pooled HLM also exhibited substrate inhibition kinetics (Fig 5B, Eadie-Hofstee plot inset) (Hutzler and Tracy, 2002). The substrate inhibition equation was fitted to the experimental data and the derived kinetic constants for \(K_m\) were \(408.2 \pm 41.4\) µM and \(V_{max}\) \(16.7 \pm 0.6\) pmole/min/mg of expressed protein (Table 1). The apparent \(CL_{int}\) value was \(0.041\) L/min/mg of protein.

**Inhibition of Chloramphenicol O-Glucuronidation by AZT, Hyodeoxycholic Acid, Serotonin, and Propofol.**

At 5000 µM, AZT inhibited the formation of 3-O-CP-glucuronide by approximately 30% and 76% of 1-O-CP glucuronide production by pooled HLM at 100 µM CP (Fig 6A). When this inhibition study was repeated with expressed UGT2B7, >75% inhibition was observed for CP 3-O- and 1-O-glucuronidation (Fig. 6A). Hyodeoxycholic acid at 1000 µM exhibited >90% inhibition for both CP 3-O-glucuronidate and CP 1-O-glucuronide formation in pooled HLMs (Fig. 6B). As for the other inhibitors, propofol inhibited CP 3-O-glucuronide formation ~60%, but <20% inhibition for CP 1-O-glucuronide formation, whereas serotonin exhibited
insignificant inhibition (<5%) for both glucuronides by pooled HLMs (data not shown). The control activities of the UGTs for the formation of 3-O-CP and 1-O-CP glucuronides at 100 µM CP in pooled HLMs were determined to be 25.2 pmole/min/mg and 4.0 pmole/min/mg of protein.

**Correlation Study in Human Liver Microsomes**

Correlation experiments were conducted with pooled alamethicin-activated HLM from 10 human donor livers at 30 and 3000 µM CP concentrations. As shown in Fig. 7, CP 3-O- and 1-O-glucuronidation in the individual donor HLMs were significantly correlated with AZT glucuronidation activity at 30 and 3000 µM (Fig 7A-D) \( (r_s = 0.85, 0.83 \text{ and } r_s 0.65, 0.79, \) respectively) and summarized in Table 2, but were not correlated with propofol or serotonin glucuronidation activity at substrate concentration of 30 µM \( (r_s < 0.5), \) except for propofol glucuronidation activity at 3000 µM CP \( (r_s = 0.71) \) (Table 2). These results indicate UGT2B7 is the primary source of 3-O- and 1-O-glucuronides with a lesser contribution by UGT1A9 at CP concentrations > 30 µM.
Discussion

We have previously identified CP 3-O- and 1-O-glucuronides generated by pooled HLMs as the only glucuronide metabolites in humans (Chen et al., 2007). In this study, we have identified the main UGT isoforms responsible for the CP 3-O- and 1-O-glucuronidations by HLM. Phenotyping with expressed UGT isozymes showed highest activity for UGT2B7 as the primary UGT isoform catalyzing CP 3-O-glucuronidation with lesser contributions from UGT1A6, UGT1A9 and other UGT isoforms at 30 and 300 µM CP. Of the isoforms capable of metabolizing CP at physiological plasma concentrations, UGT 1A6, 1A9 and 2B7 are expressed in the liver and may potentially contribute to its hepatic clearance. However, the low activities for UGT 1A6, 1A9 and 2B4 suggest that involvement of these isoforms is likely to be minor at best. UGT2B7 was the only isoform catalyzing CP 1-O-glucuronidation at either CP concentration.

The kinetic profiles for CP 3-O-glucuronidation by expressed UGT 2B7 are typical of the single-enzyme Michaelis-Menten model, whereas expressed UGT1A6 and UGT1A9 are characteristic of the sigmoidal (Hill) model. CP 1-O-glucuronidation by expressed UGT2B7 displayed substrate inhibition and the kinetics were derived using the substrate inhibition equation. The CL_{int} for 1-O-CP glucuronide in pooled HLMs was similar to that for UGT2B7 suggesting this isoform was the only contributor. The 10-20-fold greater CL_{int} value for the CP 3-O-glucuronide than for the 1-O-CP glucuronide by pooled HLM and expressed UGT2B7 implies that the 3-OH site of CP is glucuronidated more readily than the more sterically hindered benzylic OH (Fig. 1). The K_{m} values for CP glucuronidation by expressed UGT1A6 and
UGT1A9 were helpful for confirming the minor involvement of these isoforms in CP O-glucuronidation.

The kinetics for the formation of CP 3-O-glucuronide by pooled HLM were characteristic for the two-enzyme Michaelis-Menten model with high and low affinity components. The mean $K_m$ value for the high-affinity component was 20-fold lower than the $K_m$ for the low-affinity reaction, and apparent $Cl_{int}$ differed four-fold. Comparisons of the enzyme kinetic parameters between pooled HLMs and expressed UGT2B7 were made to further determine the relative contribution of UGT2B7 to CP 3-O-glucuronidation. A substrate that is glucuronidated solely by a single expressed UGT isoform should have an apparent $K_m$ similar to the apparent $K_m$ for pooled HLM (Court et al., 2003). Based on the results of the present study, the apparent high affinity $K_m$ value (~46 µM) was similar to the apparent $K_m$ for UGT2B7 (~109 µM), which suggests that in human liver, UGT2B7 may be the predominant UGT isoform catalyzing CP 3-O-glucuronidation. When the mean $K_m$ and $V_{max}$ values for the high- and low-affinity components of CP 3-O-glucuronidation by HLM were substituted into the two-enzyme Michaelis–Menten equation, the high- affinity component (UGT2B7) is responsible for ~90% of CP 3-O-glucuronidation activity at 30 µM and ~18% of activity at 3000 µM CP, and as a result contributes significantly to the higher $Cl_{int}$ value by pooled HLM at low CP concentrations. Hence, CP was 3-O-glucuronidated in pooled HLM much more rapidly by UGT2B7 than by either UGT1A6, or UGT1A9, whose $Cl_{int}$ were 10-100-fold less, consistent with their minor role in the clearance of CP at therapeutic plasma concentrations. It is likely that the low-affinity
component of hepatic CP 3-O-glucuronidation, which is comprised of the minor UGT isoform activities are also collectively contributing to CP 3-O-glucuronidation at CP concentrations >30 µM and would therefore be expected to make minor contribution (<10%) to CP clearance in patients receiving this drug. Furthermore, UGT 1A6 and 1A9 have \( S_{50} \) values of similar order to the low-affinity component of HLM CP 3-O-glucuronidation. It should be noted that CP exhibits minor non-specific binding to pooled HLMs (unpublished data), precluding nonspecific microsomal binding as a cause of atypical kinetics. CP 1-O-glucuronidation in both pooled HLM and UGT2B7 exhibited substrate inhibition kinetics (Hutzler and Tracy, 2002), which implies binding of more than one substrate molecule in the active site.

The co-incubation of CP with 5000 µM AZT in pooled HLM was observed as 30% and 76% concentration-dependent inhibition for the 3-O- and 1-O-glucuronidation of CP, respectively. We attribute the partial inhibition observed for CP 3-O-glucuronidation to its inhibition by the long-chain unsaturated fatty acids (oleic acid, linoleic acid and arachidonic acid) present (as phospholipids) in pooled HLMs which when released from the microsomal membrane during the course of an incubation, act as potent inhibitors of hepatic UGT2B7 (\( K_i \) 0.2-2 µM), competing with AZT (\( K_m \) 1331 µM) for the inhibition of hepatic UGT2B7 catalyzed CP 3-O-glucuronidation (Rowland et al., 2007; Tsoutsikos et al., 2004). However, when CP was co-incubated with AZT and expressed UGT2B7 under the same incubation conditions as for HLMs, >75% inhibition was observed for both CP 3-O- and 1-O-glucuronidation, consistent with the lower content of inhibitory unsaturated long-chain fatty acids in the expressed UGT2B7
(Rowland et al., 2007; Tsoutsikos et al., 2004), resulting in a decreased inhibition of CP 3-O-glucuronidation by the unsaturated fatty acids. Because of the complexity of the 1-O-glucuronidation substrate inhibition kinetics of CP by UGT2B7, its inhibition kinetics were not studied further (Tsoutsikos et al., 2004). The 10-20-fold greater $\text{CL}_{\text{int}}$ for UGT2B7 catalyzed O-glucuronidation in pooled HLMs and expressed UGT2B7 of the primary 3-hydroxyl group than the more sterically hindered 1-hydroxyl group may also be contributing factors to the observed differences in the inhibition kinetics between CP 3-O- and 1-O-glucuronidation. When CP is coincubated with hyodeoxycholic acid in pooled HLM, >90% inhibition was shown at 1000 μM hyodeoxycholic acid for both the 3-O- and 1-O-glucuronidation of CP. Coincubation of CP with propofol demonstrated moderate inhibition (~60%) of CP 3-O-glucuronidation and < 20% for CP 1-O-glucuronidation in pooled HLM, and no inhibition (<5%) for serotonin. These results help to support UGT2B7 as the primary UGT isoform catalyzing both CP 3-O- and 1-O-glucuronide formations in pooled HLMs, and UGT1A9 as having a minor role in the O-glucuronidation of CP.

To assess the contribution of UGT2B7 to CP 3-O- and 1-O-glucuronidation, a correlation analysis was carried out between CP 3-O-glucuronidation vs AZT (UGT2B7), propofol (UGT1A9) or serotonin (UGT1A6) glucuronidation by 10 donor individual HLMs. A significant correlation occurred between CP 3-O- and 1-O-glucuronidation and AZT glucuronidation at 30 and 3000 μM CP, but only a significant correlation for propofol (UGT1A9) at 3000 μM CP concentration. No correlation was observed for serotonin (UGT1A6) at these CP concentrations.
These correlation results help to confirm that UGT2B7 is the likely isoform catalyzing the hepatic 3-O- and 1-O-glucuronidation of CP at therapeutic concentrations.

When CP was incubated with pooled human kidney microsomes, the apparent CL_{int} for CP 3-O-glucuronide was an order of magnitude less than pooled HLMs (data not reported), indicating the contribution of renal 3-O-glucuronidation to systemic CP clearance in the neonate is much less than hepatic 3-O-glucuronidation.

In summary, our data from incubation of CP with human expressed UGT isoforms, inhibition studies with UGT selective substrates, and correlation studies clearly indicate that CP was efficiently O-glucuronidated to its major CP 3-O-glucuronide by pooled HLMs and expressed UGT2B7, and to a lesser extent by UGT1A9, confirming that hepatic 3-O-glucuronidation by UGT2B7 plays a major role in the systemic clearance of CP at therapeutic plasma concentrations. As a result of low UGT isoform activity in infants < 6 months postnatal, clinical studies have shown that hepatic glucuronidation undergoes significant changes during neonatal development requiring age-related changes in drug therapy and dosages as a result of delayed systemic clearance of the drug. For example, the clearance of AZT by UGT2B7 is approximately 50% lower in neonates than found in children 14-99 days of age (Boucher, 1993), while the clearance of morphine by UGT2B7 in postoperative infants was significantly reduced during the neonatal period and in infants and children under the age of two years, which required an adjustment in the intravenous morphine dosage (Bouwmeester et al., 2003). The results from our study indicate that the clearance of CP primarily by UGT2B7 suggests that the systemic
clearance of CP is likely to be minimal in neonates and infants < 6 months postnatal which is consistent with the argument that reduced O-glucuronidation of CP is responsible for its delayed clearance and subsequent toxicity in newborns. Available evidence suggests that UGT2B7 polymorphism has little effect on drug glucuronidation (Coffman et al., 1998; Bhasker et al., 2000), but an influence on CP elimination cannot be discounted in this age group. This study highlights the fact that the identification of the liver UGT isoform(s) involved in drug clearance is crucial to understanding the age-dependent variations in plasma drug concentration that lead to altered drug efficacy and/or toxicity in infants, which might require an age-based dosage adjustment.
Reference


Acknowledgements. The author is grateful to Dr. Caroline Decker, Dr. Hong Gao and Dr. Hongying Gao from Vertex Pharmaceuticals for their assistance and helpful discussions.
Legends for Figures

Fig. 1. Chloramphenicol, and its major 3-O-glucuronide and minor 1-O-glucuronides

Fig. 2. CP-3-O-glucuronidation (A) and CP 1-O-glucuronidation (B) at 30 and 300 μM CP by a panel of 12 expressed human UGT isoforms in microsomes from baculovirus-infected insect cells. Each bar represents the mean of duplicate determinations.

Fig. 3. Kinetics of CP-3-O-glucuronidation (A) and CP-1-O-glucuronidation (B) by expressed human UGT2B7 in microsomes from baculovirus-infected insect cells. For CP-3-O-glucuronidation (A), the solid line represents fitting the data to the single-enzyme Michaelis-Menten equation and the Eadie-Hofstee plot is shown as an inset. For 1-O-CP glucuronidation (B), the solid line represents fitting the data to the substrate inhibition model and the Eadie-Hofstee plot is shown as an inset. The CP-3-O- and 1-O-glucuronidation activity was determined as described under Materials and Methods. The kinetic parameters are summarized in Table 1. Each data point represents the mean of triplicate determinations ± s.d.

Fig. 4. Kinetics of CP-3-O-glucuronidation activity by expressed human UGT1A9 (A) and UGT1A6 (B) in microsomes from baculovirus-infected insect cells represent fitting the data to the Hill equation and the Eadie-Hofstee plot is shown as an inset. The CP-3-O- glucuronidation activity was determined as described under Materials and Methods. The kinetic parameters are
summarized in Table 1. Each data point represents the mean of triplicate determinations ± s.d.

Fig. 5. Kinetics of CP-3-O-glucuronidation (A) and CP-1-O-glucuronidation (B) in pooled HLMs. For CP-3-O-glucuronidation (A), the solid line represents fitting the data to the two-enzyme Michaelis-Menten equation, and the Eadie-Hofstee plot is shown as an inset. For CP-1-O-glucuronidation (B), the solid line represents fitting the data to the substrate inhibition model and the Eadie-Hofstee plot is shown as an inset. The CP-3-O- and 1-O-glucuronidation activity was determined as described under Materials and Methods. The kinetic parameters are summarized in Table 1. Each data point represents the mean of triplicate determinations ± s.d.

Fig. 6. The inhibitory effects of AZT (A) and hyodeoxycholic acid (B) on the CP 3-O- and 1-O-glucuronidation activity were determined in pooled HLMs at 100 µM CP. Each data point represents the mean of duplicate determinations.

Fig. 7. The correlation between 3-O-CP (A,B) and 1-O-CP glucuronosyltransferase (C,D) activities in a bank of HLMs from 10 individual donors at 30 and 3000 µM CP and AZT glucuronosyltransferase activities at 2000 µM AZT. Each data point represents the mean of duplicate determinations.
Table 1. Best-fit Derived Kinetic Parameters for CP O-Glucuronidation by HLM and Baculovirus Expressed Human UGT Isoforms.

<table>
<thead>
<tr>
<th>UGT Isoform</th>
<th>1-O-CP Glucuronide Kinetics ± s.d. (n=3)</th>
<th>3-O-CP Glucuronide Kinetics ± s.d. (n=3)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>( K_m ) c (µM) ( V_{max} ) c (pmol/min/mg protein) ( CL_{int} ) c (L/min/mg protein)</td>
<td>( K_m ) or ( S_{50} ) c (µM) ( V_{max} ) (pmol/min/mg protein) ( CL_{int1} ) or ( CL_{max} ) b (L/min/mg protein)</td>
</tr>
<tr>
<td>Pooled HLM( ^d )</td>
<td>408.2±41.4</td>
<td>16.7±0.6</td>
</tr>
<tr>
<td>2B7</td>
<td>115.0±14.2</td>
<td>5.4±0.2</td>
</tr>
<tr>
<td>1A6</td>
<td>&lt;LOQ</td>
<td>-</td>
</tr>
<tr>
<td>1A9</td>
<td>&lt;LOQ</td>
<td>-</td>
</tr>
</tbody>
</table>

\( ^a \) \( CL_{int} \) calculated from Michaelis-Menten kinetics.

\( ^b \) \( CL_{max} \) calculated from Hill equation fitted kinetics.

\( ^c \) Best-fit kinetic parameters for 1-O-CP glucuronide calculated with substrate inhibition equation \( K_{si} \) (HLM) was 28500 ± 8200 µM and \( K_{si} \) (UGT2B7) was 8090 ± 1240 µM

\( ^d \) Best-fit \( K_m1 \) and \( K_m2 \) and \( V_{max1} \) and \( V_{max2} \) kinetic parameters calculated for 3-O-CP glucuronide with two-enzyme Michaelis-Menten equation.

\( ^e \) Best-fit kinetic parameters for 3-O-CP glucuronide calculated using Hill equation.

<LOQ: below Limit of Quantitation.

Each value represents best-fit values ± s.d.
Table 2. Correlation of Chloramphenicol Glucuronosyltransferase Activity with the Glucuronosyltransferase Activities of Marker Substrates

<table>
<thead>
<tr>
<th>Marker substrate Activity</th>
<th>3-O-CP UGT activity</th>
<th>I-O-CP UGT activity</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>CP = 30 µM</td>
<td>CP = 3000 µM</td>
</tr>
<tr>
<td></td>
<td>$r_s$</td>
<td>$P$</td>
</tr>
<tr>
<td>AZT glucuronidation</td>
<td>0.85</td>
<td>0.000$^a$</td>
</tr>
<tr>
<td>Propofol glucuronidation</td>
<td>0.44</td>
<td>0.180</td>
</tr>
<tr>
<td>Serotonin glucuronidation</td>
<td>-0.01</td>
<td>0.97</td>
</tr>
</tbody>
</table>

$r_s$ - Spearman rank order correlation coefficient

$^a$ The correlations were considered statistically significant when the $r_s$ value was greater than or equal to 0.50 and the $P$ value was less than 0.05.
Figure 1

Chloramphenicol

Chloramphenicol 1-O-Glucuronide

Chloramphenicol 3-O-Glucuronide
Figure 2

A: 3-O-CPGlu (major)

B: 1-O-CPGlu (minor)
Figure 3

A

3-O-CPGlu Formation (pmole/min/mg)

v/S

0 2 4 6 8 10 12
CP Concentrations (mM)

B

1-O-CPGlu Formation (pmole/min/mg)

v/S

0 2 4 6 8
CP Concentrations (mM)
Figure 4

A

3-O-CPGlu Formation (pmole/min/mg)

v/S

CP Concentrations (mM)

B

3-O-CPGlu Formation (pmole/min/mg)

v/S

CP Concentrations (mM)
Figure 5
Figure 6

**A**

Dose-response curves for CP-UGT activity (% of control) with different AZT concentrations (mM).

**B**

Dose-response curves for CP-UGT activity (% of control) with different hyodeoxycholic acid concentrations (μM).
Figure 7

(A) 3-O-CP-UGT Activity (pmol/min/mg protein) at 0.03 mM CP

(B) 3-O-CP-UGT Activity (pmol/min/mg protein) at 3 mM CP

(C) 1-O-CP-UGT Activity (pmol/min/mg protein) at 0.03 mM CP

(D) 1-O-CP-UGT Activity (pmol/min/mg protein) at 3 mM CP