

## SHORT COMMUNICATION

### **Brain Uptake of [<sup>13</sup>C] and [<sup>14</sup>C]Sucrose Quantified by Microdialysis and Whole Tissue Analysis in Mice**

Faleh Alqahtani, Ekram Ahmed Chowdhury, Raktima Bhattacharya, Behnam Noorani, Reza Mehvar, and Ulrich Bickel

*Department of Pharmaceutical Sciences (F.A., E.A.C., R.B., B.N., U.B.) and Center for Blood-Brain Barrier Research, School of Pharmacy, Texas Tech University Health Sciences Center, Amarillo, Texas (F.A., E.A.C., R.B., R.M., U.B.); and Department of Biomedical and Pharmaceutical Sciences, Chapman University, School of Pharmacy, Irvine, California (R.M.)*

Primary Laboratory of Origin: Department of Pharmaceutical Sciences, School of Pharmacy, Texas Tech University Health Sciences Center, Amarillo, TX, USA

**RUNNING TITLE: Microdialysis of Sucrose in Brain**

*Corresponding author:* Ulrich Bickel,

Department of Pharmaceutical Sciences, School of Pharmacy, Texas Tech University Health Sciences Center, 1300 S Coulter Street, Amarillo, TX 79106, USA.

Telephone: (+1) 806-414-9236; Fax: (+1) 806-356-4034

E-mail: [Ulrich.Bickel@ttuhsc.edu](mailto:Ulrich.Bickel@ttuhsc.edu)

*Co-corresponding author:* Reza Mehvar,

Department of Biomedical and Pharmaceutical Sciences, Rinker Health Science Campus, Chapman University, 9401 Jeronimo Road, Irvine, CA 92168, USA.

Telephone: (+1) 714-516-5490; Fax: (+1) 714-516-5481

E-mail: [Mehvar@chapman.edu](mailto:Mehvar@chapman.edu)

Text pages: 14

Figures: 2

Tables: 0

References: 20

Words in Abstract: 183

Words in Introduction: 579

Words in Results & Discussion: 1305

**ABBREVIATIONS:** aCSF, artificial cerebrospinal fluid; AUC, area under the curve; BBB, blood-brain barrier; ECF, extracellular fluid; LC-MS/MS, liquid chromatography tandem mass spectrometry

## ABSTRACT

Among small, hydrophilic drug-like molecules, [ $^{14}\text{C}$ ]sucrose has long been considered the gold standard for determination of blood-brain barrier (BBB) permeability. However, we have recently shown in rats that, compared with LC-MS/MS analysis of stable isotope ( $^{13}\text{C}$ ) of sucrose, [ $^{14}\text{C}$ ]sucrose significantly overestimates the brain tissue concentration and uptake of sucrose by a factor of 6-7. This discrepancy is due to the presence of small quantities of lipophilic impurities in [ $^{14}\text{C}$ ]sucrose tracer solutions. Here, we utilized intracranial microdialysis to measure concentrations of both sucrose variants in brain extracellular fluid (ECF) after IV bolus administration to mice. Both markers displayed similar plasma profiles and ECF dialysate concentrations. However, total brain tissue concentrations and apparent brain uptake clearance of [ $^{14}\text{C}$ ]sucrose were, respectively, 4.1- and 3.6-fold higher than those of [ $^{13}\text{C}$ ]sucrose. Therefore, the contaminants of [ $^{14}\text{C}$ ]sucrose with higher permeability were likely sequestered by brain cells, which renders them non-dialyzable. It is concluded that although measurement of radioactivity overestimates the concentrations of intact sucrose in the brain tissue, the ECF radioactivity after microdialysis is a relatively accurate reflection of intact sucrose after the systemic administration of the [ $^{14}\text{C}$ ]sucrose marker.

## Introduction

The blood-brain barrier (BBB) represents a tight endothelial barrier protecting the central nervous system by controlled influx of substances from the circulation. The integrity of BBB is compromised in a wide range of diseases such as stroke, multiple sclerosis, Alzheimer's disease and brain cancer (Liebner et al., 2018). Therefore, BBB permeability is a key parameter that needs to be analyzed frequently in preclinical models. In small animals, functional damage to the BBB is typically assessed by brain tissue sampling after systemic injection of a marker that penetrates the healthy barrier poorly. Exact quantitative pharmacokinetic evaluation of passive tissue uptake is greatly facilitated when the analyte is metabolically stable in circulation, shows no significant binding to plasma or tissue proteins, is not a substrate of any of the multiple influx or efflux transport systems present at the BBB, and is nontoxic or at least does not affect the integrity of the BBB during the experiment. Experience shows that few small molecular weight chemicals match these criteria. For example, atenolol has long been considered a good model of hydrophilic drugs for measuring passive permeability across the intestinal epithelium and across the BBB (Hammarlund-Udenaes et al., 1997). However, recent work by the same group revealed that, in fact, the low brain uptake of atenolol is not primarily a consequence of poor membrane permeability, but is due to substantial active efflux transport (Chen et al., 2017).

The disaccharide sucrose comes close to be an ideal small molecule marker because, after systemic administration, it is not metabolized or bound to proteins. Additionally, to date, there is no evidence that sucrose is the substrate of facilitative or active transport at the mammalian BBB. While members of the SLC45 family of proton-coupled transporters have recently been implicated as putative sucrose transporters under special conditions in mammalian tissues, including kidney and testis (Bartolke et al., 2014; Vitavska et al., 2016; Vitavska and Wieczorek, 2017), these

transport proteins have higher affinity to monosaccharides, in particular for glucose and fructose (Bartolke et al., 2014). However, relevant expression levels or physiological functions at the BBB have not been found for any of these transporters. Moreover, transport of sucrose would likely be blunted by physiological blood glucose levels. Therefore, sucrose has been used in the BBB field for decades in form of internally labeled radioactive tracers, most frequently as [ $^{14}\text{C}$ ]sucrose. However, we have recently demonstrated that measurement of brain uptake of [ $^{14}\text{C}$ ]sucrose, based on its plasma and total brain concentrations, is compromised by the presence of low quantities of lipophilic, BBB permeable impurities, which make up 80% or more of the total radioactivity in brain after IV administration (Miah et al., 2017b). Based on direct comparison, we suggested the use of nonradioactive [ $^{13}\text{C}$ ]sucrose and UPLC-MS/MS analysis as a better alternative (Miah et al., 2016; Miah et al., 2017a; Miah et al., 2017b).

Intracranial microdialysis is an excellent experimental approach to measure in vivo concentrations of analytes in brain interstitial fluid, and [ $^{14}\text{C}$ ]sucrose has been used repeatedly as a test or reference substance in microdialysis (Terasaki et al., 1992; Groothuis et al., 1998; Diserbo et al., 2002). We wondered if the data obtained in such experiments could also be biased by the radioactive impurities. The present study was designed to address that question by comparing brain uptake of [ $^{14}\text{C}$ ] and [ $^{13}\text{C}$ ]sucrose in two groups of mice after IV bolus administration of the markers. The comparison was based on measurement of [ $^{14}\text{C}$ ] and [ $^{13}\text{C}$ ]sucrose in plasma, whole brain tissue, and brain dialysate samples.

## Materials and Methods

**Chemicals and Reagents.** [ $^{13}\text{C}_{12}$ ]sucrose (all 12 carbons labeled with  $^{13}\text{C}$ ) and [ $^{13}\text{C}_6$ ]sucrose (only the 6 carbons in the fructose moiety labeled) were purchased from Omicron Biochemicals (South Bend, IN, USA). [ $^{14}\text{C}(\text{U})$ ]Sucrose (uniformly labeled, specific activity 435

mCi/mmol) was supplied by PerkinElmer (Boston, Massachusetts, USA). LC-MS/MS-grade solvents and analytical grade chemicals were obtained from reliable commercial sources.

**Animals.** Male C57BL/6J mice, aged 8-12 weeks, weight 25-30 g, were purchased from Jackson Laboratory (Bar Harbor, ME, USA) and housed in ventilated cages in a temperature- and humidity-controlled room with 12-h dark-light cycles and free access to food and water. All animal procedures were approved by the Institutional Animal Care and Use Committee at Texas Tech University Health Sciences Center and complied with NIH guidelines for care and use of animals.

**LC-MS/MS Analysis of [<sup>13</sup>C<sub>12</sub>]Sucrose in Plasma, Dialysate, and Brain Samples.** The permeability marker [<sup>13</sup>C<sub>12</sub>]sucrose was quantified by LC-MS/MS according to our previously published protocol, using [<sup>13</sup>C<sub>6</sub>]sucrose as the internal standard (IS) (Miah et al., 2016). The sample preparation methods for plasma and brain homogenate were according to the published method. Before analysis, microdialysate samples were diluted 1:9 in acetonitrile: water (80:20) containing 2 ng/mL IS. Standard curves were prepared in blank plasma (10-1,000 ng/mL), blank brain homogenate (1-200 ng/mL), and artificial cerebrospinal fluid (aCSF) (1-300 ng/mL). We validated [<sup>13</sup>C<sub>12</sub>]sucrose analysis in aCSF/brain dialysate matrix. The percent recovery ( $\pm$  SD) from aCSF samples at 10 ng/mL was  $93.5 \pm 10.4$  and  $100 \pm 3$  for [<sup>13</sup>C]sucrose and [<sup>13</sup>C<sub>6</sub>]sucrose (IS), respectively. Inter- and intra-run accuracy was within 94%-103% for all tested concentrations in the aCSF matrix, and sample precision values (R.S.D) were  $\leq 9\%$  for all concentrations within the range of the standard curve (1-300 ng/mL). The  $r^2$  of the linear regression analysis of standards was  $\geq 0.99$ .

**Liquid Scintillation Counting.** All radioactive samples were measured in a Beckman LS6500 liquid scintillation counter with appropriate window settings for <sup>14</sup>C. Quench monitoring (H-number) and automatic quench correction was applied to convert counts per minute into

disintegrations per minute (dpm/min). While aCSF and dialysate samples were measured in 5 mL of ScintiSafe 30%, plasma and brain samples were first solubilized in one mL of Soluene®350 at 65°C overnight before adding 15 mL Hionic-Fluor scintillation fluid.

**Microdialysis Probe Construction and In Vitro Recovery of [<sup>13</sup>C] and [<sup>14</sup>C]Sucrose.** Y-shaped self-built dialysis probes with a 10 kDa molecular weight cutoff, 280 μm o.d. and 2 mm exchange length were constructed as previously described (Sumbria et al., 2011). Random samples of probes ( $n = 4$ ) from each batch were selected to test the in vitro recovery rate of the probes. The probes were immersed in a 10-mL beaker containing 1 μg/mL [<sup>13</sup>C<sub>12</sub>]sucrose or 0.5 μCi/mL [<sup>14</sup>C]sucrose dissolved in aCSF that was well stirred and maintained at 37°C. Before dialysis, two 30-μL ([<sup>13</sup>C<sub>12</sub>]sucrose) or 50-μL ([<sup>14</sup>C]sucrose) aliquots of the solution in the beaker were taken and 20-μL aliquots were used for analysis of the reference concentration ( $C_e$ ). The probes were then perfused with aCSF at a flow rate of 2 μL/min. 20-μL dialysate samples were collected at intervals of 10 min over a period of 60 min for measurement of the outflow concentration ( $C_{out}$ ). In vitro recovery in the dialysate,  $R_{(in\ vitro)}$ , was calculated from the  $\frac{C_{out}}{C_e}$  ratio, where  $C_e$  is the average of duplicate measurements.

**Microdialysis Probe Implantation and In Vivo Recovery of [<sup>13</sup>C]Sucrose.** Mice ( $n = 4$ ) were anesthetized with isoflurane (4.0% for induction, then 1-1.5% for maintenance) in 70% N<sub>2</sub>O/ 30%O<sub>2</sub> using a facemask and a Surgivet vaporizer. The head was fixed in a stereotaxic frame (David Kopf Instruments, Tujunga, CA, USA), and the probe implanted into the striatum exactly as described before (Sumbria et al., 2011). After a 24 h recovery period, the mice were re-anesthetized with isoflurane, and the implanted probe was equilibrated with aCSF for 60 min under a flow rate of 2 μL/min. To determine the in vivo recovery of [<sup>13</sup>C]sucrose, a retrodialysis experiment in four animals was conducted by perfusion of the randomly selected probes ( $n = 4$ )

with aCSF containing 100 ng/mL of [<sup>13</sup>C]<sub>12</sub>]sucrose. An aliquot of the perfusate was taken for analysis of the inlet concentration,  $C_{in}$ . Six dialysis samples were collected at 15 min intervals over 90 min to measure  $C_{out}$ . The in vivo recovery,  $R_{(in vivo)}$ , which is equivalent to loss upon retrodialysis, was calculated as  $R_{(in vivo)} = \frac{C_{in} - C_{out}}{C_{in}}$ .

**Analysis of Brain Uptake of [<sup>13</sup>C] and [<sup>14</sup>C]Sucrose by Measurement of Concentrations in Plasma, Whole Brain and Microdialysate.** To measure brain uptake of [<sup>14</sup>C]sucrose or [<sup>13</sup>C]<sub>12</sub>]sucrose after IV administration, microdialysis probes were implanted and equilibrated with aCSF for 60 min as described above. Subsequently, both jugular veins of mice were exposed. A bolus dose of 40 μCi/animal of [<sup>14</sup>C]sucrose ( $n = 5$ ) or 10 mg/kg of [<sup>13</sup>C]<sub>12</sub>]sucrose ( $n = 5$ ) in 50 μL physiological saline was injected into one jugular vein. Sequential blood samples of 30 μL each were drawn from the contralateral vein at 1, 5, 10, 20, 30, 60, and 120 min after injection. Simultaneously, the brain dialysates were collected every 10 min or 15 min for [<sup>14</sup>C]sucrose or [<sup>13</sup>C]<sub>12</sub>]sucrose, respectively. After the 120 min blood sample was collected, transcardiac perfusion with phosphate buffered saline (pH 7.4) was performed through the left ventricle of the heart at a flow rate of 2 mL/min for 10 min. At the beginning of vascular washout, both jugular veins were cut open to ensure free outflow of blood from the brain and to visually check that the outflowing perfusion fluid was clear of blood by the end of perfusion. Next, the forebrains were collected and cleaned of meninges, separated into hemispheres and weighed. The blood samples were centrifuged at 5,000 g for 10 min at 4°C, and plasma was separated. For the [<sup>13</sup>C]<sub>12</sub>]sucrose group, plasma, brain, and dialysate samples were processed as described above and stored at -80°C until the day of analysis by the UPLC-MS/MS method. Samples in the [<sup>14</sup>C]sucrose series were processed as described above. All samples were analyzed within one month of



collection. Our recent study (Chowdhury et al., 2018) confirmed stability of sucrose in plasma and brain samples when stored at -80°C for at least five months and one month, respectively.

**Pharmacokinetic and Statistical Analysis.** To compare the data of [<sup>14</sup>C]sucrose and [<sup>13</sup>C]sucrose, the concentration of the markers in plasma, brain and aCSF were corrected for the injected dose and expressed as the percentage of injected dose (%ID) per mL or per gram. BBB permeability was calculated as the apparent brain uptake clearance,  $K_{in}$ , from the measured concentrations in total brain tissue after 120 min and the area under the plasma concentration time curve (AUC), using the equation  $K_{in} = \frac{C_{br}^{120}}{AUC_{plasma}^{0-120}}$ , where  $AUC_{plasma}^{0-120}$  is the AUC for [<sup>14</sup>C] or [<sup>13</sup>C]sucrose from time 0 to 120 min, calculated by the logarithmic trapezoidal rule.

The concentration of [<sup>14</sup>C] or [<sup>13</sup>C<sub>12</sub>]sucrose in brain extracellular fluid,  $C_{ECF}$ , was calculated from the measured dialysate concentration ( $C_{dialysate}$ ) and  $R_{in\ vivo}$ :  $C_{ECF} = \frac{C_{dialysate}}{R_{in\ vivo}}$ . The area under the ECF concentration-time course from time zero to 120 min,  $AUC_{ECF}^{0-120}$ , was then calculated by the linear trapezoidal method for both [<sup>14</sup>C]sucrose and [<sup>13</sup>C<sub>12</sub>]sucrose.

Data are presented as mean ± SD. The kinetic parameters of [<sup>14</sup>C] and [<sup>13</sup>C<sub>12</sub>]sucrose were compared by unpaired 2-sided t-tests. A  $P$  value < 0.05 was considered statistically significant.

## Results and Discussion

### In Vitro and In Vivo Recovery of [<sup>14</sup>C] and [<sup>13</sup>C]Sucrose with Microdialysis Probes.

Using a sample of the self-made probes, in vitro recovery rates for the isotopically labeled sucrose variants were not significantly different, as shown in Fig. 1, with values of 7.29% ± 0.56% and 7.03% ± 0.58% ( $n = 4$ ) for [<sup>14</sup>C] and [<sup>13</sup>C<sub>12</sub>]sucrose, respectively. Considering that the isotopic variants are chemically identical and the difference in molecular weights is minor (342.3 for [<sup>14</sup>C]sucrose vs. 354.2 for [<sup>13</sup>C<sub>12</sub>]sucrose), no significant difference in probe recovery would be

expected. The present values are similar to the in vitro extraction fraction of about 6% for [ $^{14}\text{C}$ ]sucrose, which we found previously using probes with the same design as applied here (Bungay et al., 2011).

Consistent with experimental evidence and the theory of microdialysis in tissues, in vivo recovery rates are substantially lower than those observed in vitro due to a number of factors, including the brain extracellular space fraction of about 0.2, the tortuosity of extracellular space, and potential analyte clearance from extracellular fluid. In the current study, we determined an in vivo recovery rate of  $3.88\% \pm 0.36\%$  ( $n = 4$ ) for [ $^{13}\text{C}_{12}$ ]Sucrose, which is very close to the value of 4.2% observed earlier (Sumbria, 2010) for [ $^{14}\text{C}$ ]sucrose 24 hr after implantation of similar probes in mice. However, direct comparison to published data from other laboratories is limited by variations in probe geometry, molecular weight cutoff, perfusion flow rate, and animal species. In the following section, we used the in vivo recovery rate of 3.88% to calculate the in vivo concentrations of sucrose variants in brain interstitial fluid from the dialysate concentrations.

**Brain Uptake of [ $^{14}\text{C}$ ]Sucrose versus [ $^{13}\text{C}_{12}$ ]Sucrose.** The plasma and ECF concentration-time profiles of [ $^{14}\text{C}$ ] and [ $^{13}\text{C}_{12}$ ]sucrose are presented in Fig. 2.A. Individual concentration-time courses of both variants were overlapping for both plasma and ECF samples. The AUC values ( $\text{AUC}_{\text{plasma}}$  and  $\text{AUC}_{\text{ECF}}$ ) and the ratio of  $\text{AUC}_{\text{ECF}}$  to  $\text{AUC}_{\text{plasma}}$ , calculated for each animal, are presented in Fig. 2.B. Statistical analysis revealed no significant differences between the [ $^{14}\text{C}$ ]sucrose and [ $^{13}\text{C}_{12}$ ]sucrose groups ( $n = 5/\text{group}$ ) for either  $\text{AUC}_{\text{plasma}}$  ( $604 \pm 169$  vs.  $521 \pm 121$  %ID  $\text{mL}^{-1}$  min),  $\text{AUC}_{\text{ECF}}$  ( $55.6 \pm 18.7$  vs.  $37.7 \pm 8.7$  %ID  $\text{mL}^{-1}$  min) or the AUC ratios ( $9.37 \pm 2.88$  vs.  $7.76 \pm 3.10$ ). Regarding the plasma concentrations, this finding is in agreement with our previous study in rats (Miah et al., 2017b), which showed no significant difference between [ $^{14}\text{C}$ ]sucrose and [ $^{13}\text{C}_{12}$ ]sucrose in terms of plasma profiles and  $\text{AUC}_{\text{plasma}}$

values. The terminal total brain tissue concentration ( $C_{br}^{120}$ ), and the apparent brain uptake clearance ( $K_{in}$ ) values of [ $^{14}\text{C}$ ]sucrose and [ $^{13}\text{C}_{12}$ ]sucrose are depicted in Fig. 2.C and 2.D, respectively. The average concentration in total brain tissue of the [ $^{14}\text{C}$ ]sucrose group ( $0.195 \pm 0.062$  %ID/g) was 4.1-fold higher ( $P = 0.0010$ ) than that in the [ $^{13}\text{C}_{12}$ ]sucrose group ( $0.0474 \pm 0.0208$  %ID/g), and the corresponding  $K_{in}$  value ( $\mu\text{L}\cdot\text{min}^{-1}\cdot\text{g}^{-1}$ ) was 3.6-fold higher for [ $^{14}\text{C}$ ]sucrose ( $0.325 \pm 0.052$  vs.  $0.0913 \pm 0.0587$ ;  $P < 0.0001$ ). These relations between the isotopic variants are similar to the data we reported previously in total brain tissue in the rat (Miah et al., 2017b), and they confirm the statement that [ $^{14}\text{C}$ ]sucrose substantially overestimates brain uptake of sucrose when brain tissue is used as the source for analysis. However, the comparable concentration-time courses and AUC values in brain ECF for both isotopes suggest that the fraction of  $^{14}\text{C}$  radioactivity, which is present in ECF, behaves similar to [ $^{13}\text{C}_{12}$ ]sucrose and can be sampled by the microdialysis technique. In contrast, the larger fraction of  $^{14}\text{C}$  radioactivity, which accounts for the higher tissue concentrations of that tracer, must consist of non-dialyzable material.

Our recent work (Miah et al., 2017b) and previous studies by others (Preston et al., 1998) suggest that the overestimation of brain concentrations and  $K_{in}$  values of [ $^{14}\text{C}$ ]sucrose are due to presence of minor lipophilic impurities in the stock solution of the marker, which easily penetrate the brain. Once beyond the brain endothelial cells and in brain interstitial fluid, the lipophilic substances will rapidly enter the intracellular compartment of brain cells (neurons, astroglia, microglia, oligodendroglia) and are no longer available for microdialysis sampling. However, the intact sucrose is expected to be confined to the brain interstitial fluid. Indeed, the observed total brain tissue concentration of [ $^{13}\text{C}_{12}$ ]sucrose ( $0.0454 \pm 0.0295$  %ID/g) was equal to 18% of that in the ECF ( $0.250 \pm 0.123$  %ID/g) at 120 min, which is in agreement with a brain extracellular space of 20% (Nicholson and Sykova, 1998). This suggests no uptake of intact sucrose into brain cells

during the 2 h experimental period. However, the brain tissue radioactivity after [ $^{14}\text{C}$ ]sucrose administration ( $0.195 \pm 0.062$  %ID/g) was equal to 55% of its ECF concentration ( $0.354 \pm 0.198$  %ID/g) at 120 min, which is much higher than the expected 20%, suggesting intracellular distribution of the radioactivity.

Although the manufacturer's data for [ $^{14}\text{C}$ ]sucrose state the stock solutions are >97% pure (based on HPLC), even minor lipophilic impurities that easily cross the BBB can significantly distort the brain concentrations of the intact marker because of the non-specificity of the total radioactivity measurements. Preston et al. (Preston et al., 1998) reported that these minor impurities, which amounted to ~2% of total activity in tracer stock solutions of [ $^{14}\text{C}$ ]sucrose were barely detectable by thin layer chromatography. Here we performed HPLC fractionation of the [ $^{14}\text{C}$ ]sucrose stock solution on a column suitable for carbohydrates. The chromatogram, shown in Supplemental Figure 1, confirmed the radiochemical purity provided by the manufacturer's certificate of analysis. The injected dose of radioactivity was quantitatively recovered in the collected fractions. However, while a plot of the chromatogram at full scale does not suggest the presence of impurities, plotting at a 100-fold reduced scale reveals multiple small peaks of radioactivity, which cumulatively amounted to 1.98% and 0.87% of total activity eluting in fractions before and after the peak representing intact [ $^{14}\text{C}$ ]sucrose, respectively (Supplemental Fig. 1). We did not attempt to identify the chemical nature of the peaks, but it is known that earlier retention times on this type of chromatographic system indicate more lipophilic compounds.

Although similar lipophilic impurities could also be present in the [ $^{13}\text{C}_{12}$ ]sucrose stock solutions, their potential presence does not impact estimation of the brain concentration or  $K_{in}$  of the marker because of the specificity of the analytical method (LC-MS/MS), which does not measure the impurities. On the other hand, applying HPLC fractionation before scintillation

counting, as described above for tracer stock, to each brain sample for measurement of intact [ $^{14}\text{C}$ ]sucrose is not feasible, because it would require the administration of at least 10-fold higher doses in the in vivo experiments, rendering it cost and time prohibitive.

The overestimation of brain levels of [ $^{14}\text{C}$ ]sucrose due to lipophilic impurities could lead to erroneous conclusions (Miah et al., 2017a). Using a hepatic ischemia-reperfusion injury model, we demonstrated a significant increase in the  $K_{in}$  of [ $^{14}\text{C}$ ]sucrose in injured rats when compared with sham-operated animals. Based on these data, it could have been concluded that hepatic ischemia-reperfusion causes damage to the BBB. However, no significant difference in  $K_{in}$  was detected in the same disease model when [ $^{13}\text{C}$ ]sucrose was used as the BBB permeability marker.

In conclusion, utilizing the brain microdialysis technique in parallel to total brain tissue sampling for the comparison of two sucrose isotopes, [ $^{14}\text{C}$ ]sucrose and [ $^{13}\text{C}_{12}$ ]sucrose, as BBB permeability markers, we have shown that both isotopic variants accumulate in brain extracellular space to a similar extent after IV administration. However, this contrasts with the total brain tissue concentrations, which are significantly higher after systemic [ $^{14}\text{C}$ ]sucrose injection. This discrepancy is likely due to presence of minor, but highly lipophilic, impurities in the radiotracer solution. It is concluded that although measurement of radioactivity overestimates the concentrations of intact sucrose in the brain tissue, the ECF radioactivity after microdialysis is a relatively accurate reflection of intact sucrose after the systemic administration of the [ $^{14}\text{C}$ ]sucrose marker.

### **Authorship Contributions**

*Participated in research design:* Alqahtani, Mehvar, and Bickel.

*Conducted experiments:* Alqahtani, Chowdhury, Noorani, and Bhattacharya.

*Performed data analysis:* Alqahtani, Chowdhury, Bhattacharya, Mehvar, and Bickel.

*Wrote or contributed to the writing of the manuscript:* Alqahtani, Mehvar, and Bickel.

## References

- Bartolke R, Heinisch JJ, Wieczorek H, and Vitavska O (2014) Proton-associated sucrose transport of mammalian solute carrier family 45: an analysis in *Saccharomyces cerevisiae*. *Biochem J* **464**:193-201.
- Bungay PM, Sumbria RK, and Bickel U (2011) Unifying the mathematical modeling of in vivo and in vitro microdialysis. *J Pharm Biomed Anal* **55**:54-63.
- Chen X, Slättengren T, de Lange ECM, Smith DE, and Hammarlund-Udenaes M (2017) Revisiting atenolol as a low passive permeability marker. *Fluids and Barriers of the CNS* **14**.
- Chowdhury EA, Alqahtani F, Bhattacharya R, Mehvar R, and Bickel U (2018) Simultaneous UPLC-MS/MS analysis of two stable isotope labeled versions of sucrose in mouse plasma and brain samples as markers of blood-brain barrier permeability and brain vascular space. *J Chromatogr B Analyt Technol Biomed Life Sci* **1073**:19-26.
- Diserbo M, Agin A, Lamproglou I, Mauris J, Staali F, Multon E, and Amourette C (2002) Blood-brain barrier permeability after gamma whole-body irradiation: an in vivo microdialysis study. *Canadian journal of physiology and pharmacology* **80**:670-678.
- Dykstra KH, Hsiao JK, Morrison PF, Bungay PM, Mefford IN, Scully MM, and Dedrick RL (1992) Quantitative examination of tissue concentration profiles associated with microdialysis. *J Neurochem* **58**:931-940.
- Groothuis DR, Ward S, Schlageter KE, Itskovich AC, Schwerin SC, Allen CV, Dills C, and Levy RM (1998) Changes in blood-brain barrier permeability associated with insertion of brain cannulas and microdialysis probes. *Brain Res* **803**:218-230.

- Hammarlund-Udenaes M, Paalzow LK, and de Lange EC (1997) Drug equilibration across the blood-brain barrier--pharmacokinetic considerations based on the microdialysis method. *Pharm Res* **14**:128-134.
- Liebner S, Dijkhuizen RM, Reiss Y, Plate KH, Agalliu D, and Constantin G (2018) Functional morphology of the blood-brain barrier in health and disease. *Acta Neuropathol* **135**:311-336.
- Miah MK, Bickel U, and Mehvar R (2016) Development and validation of a sensitive UPLC-MS/MS method for the quantitation of [(13)C]sucrose in rat plasma, blood, and brain: Its application to the measurement of blood-brain barrier permeability. *J Chromatogr B Analyt Technol Biomed Life Sci* **1015-1016**:105-110.
- Miah MK, Bickel U, and Mehvar R (2017a) Effects of hepatic ischemia-reperfusion injury on the blood-brain barrier permeability to [(14)C] and [(13)C]sucrose. *Metab Brain Dis* **32**:1903-1912.
- Miah MK, Chowdhury EA, Bickel U, and Mehvar R (2017b) Evaluation of [(14)C] and [(13)C]Sucrose as Blood-Brain Barrier Permeability Markers. *J Pharm Sci* **106**:1659-1669.
- Morgan ME, Singhal D, and Anderson BD (1996) Quantitative assessment of blood-brain barrier damage during microdialysis. *J Pharmacol Exp Ther* **277**:1167-1176.
- Nicholson C and Sykova E (1998) Extracellular space structure revealed by diffusion analysis. *Trends Neurosci* **21**:207-215.
- Preston E, Foster DO, and Mills PA (1998) Effects of radiochemical impurities on measurements of transfer constants for [14C]sucrose permeation of normal and injured blood-brain barrier of rats. *Brain Res Bull* **45**:111-116.



Sumbria RK (2010) Intracerebral microdialysis studies of blood-brain barrier permeability and neurochemical parameters in a murine stroke model (doctoral dissertation), in: *Department of Pharmaceutical Sciences*, Texas Tech University Health Sciences Center, Lubbock, TX (<https://ttu-ir.tdl.org/ttu-ir/handle/2346/66921>).

Sumbria RK, Klein J, and Bickel U (2011) Acute depression of energy metabolism after microdialysis probe implantation is distinct from ischemia-induced changes in mouse brain. *Neurochem Res* **36**:109-116.

Terasaki T, Deguchi Y, Kasama Y, Pardridge WM, and Tsuji A (1992) Determination of in vivo steady-state unbound drug concentration in the brain interstitial fluid by microdialysis. *Int J Pharm* **81**:143-152.

Vitavska O, Edemir B, and Wiczorek H (2016) Putative role of the H(+)/sucrose symporter SLC45A3 as an osmolyte transporter in the kidney. *Pflugers Arch* **468**:1353-1362.

Vitavska O and Wiczorek H (2017) Putative role of an SLC45 H(+)/sugar cotransporter in mammalian spermatozoa. *Pflugers Arch* **469**:1433-1442.

## Footnotes

FA and EAC contributed equally to this work.

Current affiliation of RB is at Texas A&M Health Sciences Center, College of Pharmacy,  
College Station, TX

## Figure Legends

**Figure 1.** The in vitro recovery rate for [ $^{14}\text{C}$ ]sucrose and [ $^{13}\text{C}$ ]sucrose using self-made probes with 10 kDa molecular weight cut-off ( $n = 4$ ). Dialysate was sampled in 10 min intervals at a flow rate of 2  $\mu\text{L}/\text{min}$ . The same probes were used to determine recovery of both isotopes.

**Figure 2:** (A) Plasma ( $C_{\text{Plasma}}$ , solid lines) and ECF ( $C_{\text{ECF}}$ , dashed lines) concentration profiles for [ $^{13}\text{C}$ ] and [ $^{14}\text{C}$ ]sucrose from zero to 120 min after IV bolus injection; (B) AUC from zero to 120 min in extracellular fluid (ECF) and plasma and the ECF/plasma AUC ratio of [ $^{13}\text{C}$ ] and [ $^{14}\text{C}$ ]sucrose; (C) terminal total brain tissue concentration ( $C_{\text{br}}$ ) of [ $^{13}\text{C}$ ] and [ $^{14}\text{C}$ ]sucrose at 120 min; (D) apparent brain uptake clearance ( $K_{\text{in}}$ ) of [ $^{13}\text{C}$ ] and [ $^{14}\text{C}$ ]sucrose over 120 min. Data are presented for individual animals ( $n = 5$  for each marker). Additionally, mean values are presented as vertical bars for panels B-D.

\*\*\* $P < 0.001$  ; \*\*\*\* $P < 0.0001$ : Significantly different based on unpaired 2-tailed t-test.

FIGURE 1

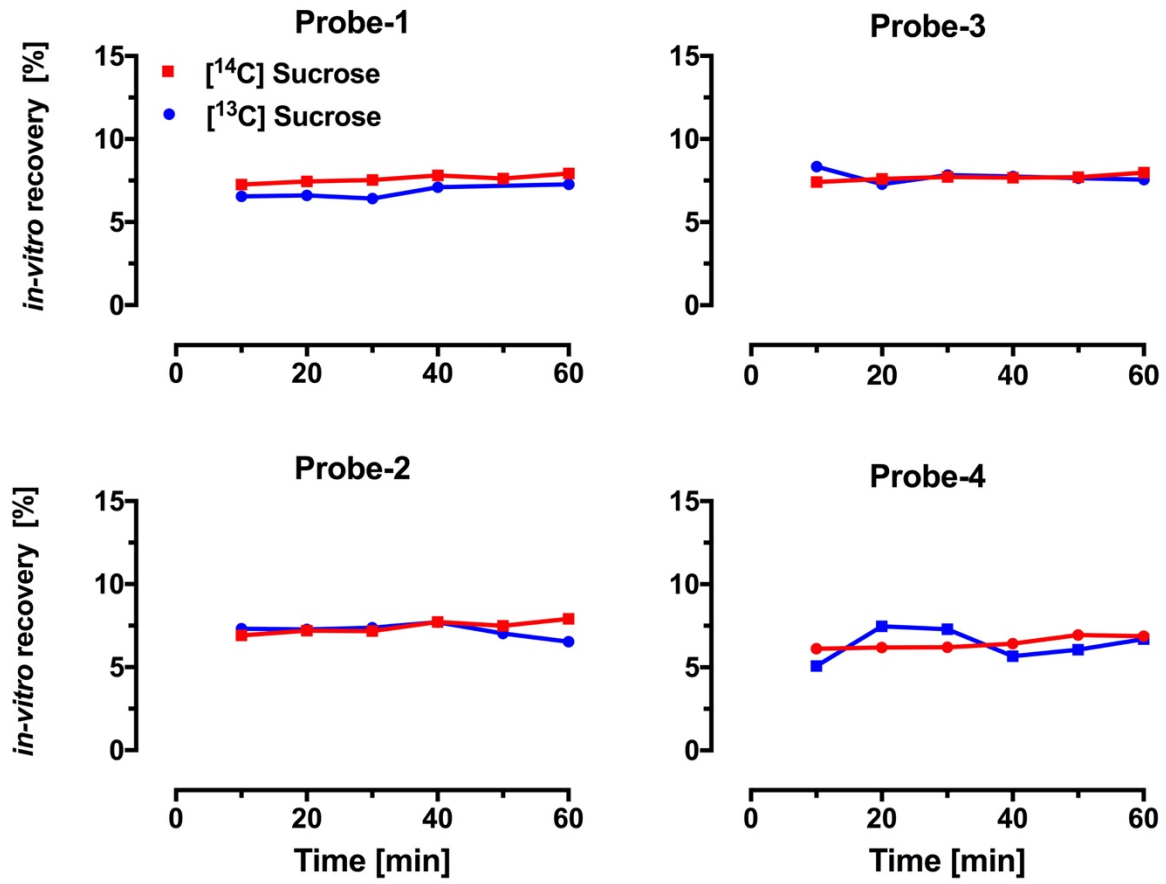


FIGURE 2

