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## **Use of Cryopreserved Hepatocytes as Part of an Integrated Strategy to Characterize In Vivo Clearance for Peptide-Antibody Conjugate Inhibitors of Nav1.7 in Preclinical Species**

Robert S. Foti<sup>1\*</sup>, Kaustav Biswas<sup>2</sup>, Jennifer Aral<sup>2</sup>, Xuhai Be<sup>1</sup>, Loren Berry<sup>1</sup>, Yuan Cheng<sup>2</sup>, Kip Conner<sup>3</sup>, James R. Falsey<sup>2</sup>, Charles Glaus<sup>2</sup>, Brad Herberich<sup>2</sup>, Dean Hickman<sup>1</sup>, Tayo Ikotun<sup>2</sup>, Hongyan Li<sup>5</sup>, Jason Long<sup>2</sup>, Liyue Huang<sup>1</sup>, Les P. Miranda<sup>2</sup>, Justin Murray<sup>2</sup>, Bryan Moyer<sup>4</sup>, Chawita Netirojjanakul<sup>2</sup>, Thomas E. Nixey<sup>2</sup>, Kelvin Sham<sup>2</sup>, Marcus Soto<sup>5</sup>, Christopher M. Tegley<sup>2</sup>, Linh Tran<sup>5</sup>, Bin Wu<sup>2</sup>, Lin Yin<sup>2</sup>, and Dan A. Rock<sup>3\*</sup>

<sup>1</sup> Pharmacokinetics and Drug Metabolism, Amgen Research, 360 Binney St, Cambridge, MA 02142

<sup>2</sup> Therapeutic Discovery, Amgen Research, 1 Amgen Center Dr, Thousand Oaks, CA 91320

<sup>3</sup> Pharmacokinetics and Drug Metabolism, Amgen Research, 1120 Veterans Blvd, S. San Francisco, 94080

<sup>4</sup> Neuroscience, Amgen Research, 1 Amgen Center Dr, Thousand Oaks, CA 91320

<sup>5</sup> Pharmacokinetics and Drug Metabolism, Amgen Research, 1 Amgen Center Dr, Thousand Oaks, CA 91320

\*To Whom Correspondence Should Be Addressed

Robert S. Foti, Ph.D.  
Principal Scientist  
Pharmacokinetics and Drug Metabolism  
Amgen, Inc.  
360 Binney Street  
Cambridge, MA 02142  
[rfoti@amgen.com](mailto:rfoti@amgen.com)

Dan A. Rock, Ph.D.  
Executive Director  
Pharmacokinetics and Drug Metabolism  
Amgen, Inc.  
1120 Veterans Boulevard  
South San Francisco, CA 94080  
[drock@amgen.com](mailto:drock@amgen.com)

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### Corresponding Authors:

Robert S. Foti, Ph.D.  
Principal Scientist  
Pharmacokinetics and Drug Metabolism  
Amgen, Inc.  
360 Binney Street  
Cambridge, MA 02142  
[rfoti@amgen.com](mailto:rfoti@amgen.com)

Dan A. Rock, Ph.D.  
Executive Director  
Pharmacokinetics and Drug Metabolism  
Amgen, Inc.  
1120 Veterans Boulevard  
South San Francisco, CA 94080  
[drock@amgen.com](mailto:drock@amgen.com)

### Manuscript metrics:

Text pages: 20

Tables: 2

Figures: 7

References: 72

Words in the abstract: 225

Words in Introduction: 841

Words in Discussion: 2628

**Abbreviations:** FcRn, neonatal Fc receptor; LDL, low-density lipoprotein; NODA-GA, 1,4,7-triazacyclononane,1-glutaric acid-4,7-acetic acid; HIC, hydrophobic interaction chromatography; HC, heavy chain.

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## Abstract

The identification of non-opioid alternatives to treat chronic pain has received a great deal of interest in recent years. Recently, the engineering of a series of Nav1.7 inhibitory peptide-antibody conjugates has been reported and herein, the preclinical efforts to identify novel approaches to characterize the pharmacokinetic properties of the peptide conjugates are described. A cryopreserved plated mouse hepatocyte assay was designed to measure the depletion of the peptide-antibody conjugates from the media, with a correlation being observed between percent remaining in the media and in vivo clearance (Pearson  $r = -0.5525$ ). Physicochemical (charge and hydrophobicity), receptor-binding (FcRn) and in vivo pharmacokinetic data were generated and compared to the results from an in vitro our hepatocyte assay that was hypothesized to encompass all of the aforementioned properties. Correlations were observed between hydrophobicity, FcRn binding, depletion rates from the hepatocyte assay and ultimately, in vivo clearance. Subsequent studies identified potential roles for the low-density lipoprotein and mannose/galactose receptors in the association of the Nav1.7 peptide conjugates with mouse hepatocytes, though in vivo studies suggested that FcRn was still the primary receptor involved in determining the pharmacokinetics of the peptide conjugates. Ultimately, the use of the cryopreserved hepatocyte assay along with FcRn binding and hydrophobic interaction chromatography provided an efficient and integrated approach to rapidly triage molecules for advancement while reducing the number of in vivo pharmacokinetic studies.

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## Significance Statement

While multiple in vitro and in silico tools are available in small molecule drug discovery, pharmacokinetic characterization of protein therapeutics is still highly dependent upon the use of in vivo studies in preclinical species. The current work demonstrates the combined use of cryopreserved hepatocytes, hydrophobic interaction chromatography and FcRn binding to characterize a series of Nav1.7 peptide-antibody conjugates prior to conducting in vivo studies, thus providing a means to rapidly evaluate novel protein therapeutic platforms while concomitantly reducing the number of in vivo studies conducted in preclinical species.

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## Introduction

It has been estimated that over half a million people in the United States alone died from drug overdoses between 2001 and 2015, the majority of which were attributed to the growing opioid crisis (Humphreys, 2017). As such, the identification of non-opioid alternatives to treat chronic pain has become a rapidly emerging area of research in recent years. The Nav1.7 voltage-gated sodium ion channel (SCN9A) represents one key pharmacological target given the high degree of human genetic validation for its role in modulating pain states (Dib-Hajj et al., 2013; Emery et al., 2016; Vetter et al., 2017). Multiple examples of small molecule, peptide and antibody approaches to inhibit Nav1.7 activity have been reported in the literature. Recently, we have described the engineering of two series of inhibitory Nav1.7 peptide-antibody conjugates, linking peptides based on the GpTx-1 or JzTx-V toxins found in tarantula venom to non-targeting antibodies in order to increase the half-life and stability of the inhibitory peptides (Biswas et al., 2017; Moyer et al., 2018; Wu et al., 2018; Murray et al., 2019).

As with monoclonal antibodies, it is likely that the pharmacokinetics of peptide-antibody conjugates are also affected by both physicochemical properties, target-mediated drug disposition and receptor-mediated interactions (Datta-Mannan et al., 2015a). Key physicochemical properties that have been shown to contribute to pharmacokinetics of antibody therapeutics include molecular charge, isoelectric point, hydrodynamic radius, size, valency, and post-translational modifications (Boswell et al., 2010; Bumbaca et al., 2012; Liu, 2015; Schoch et al., 2015). Interactions with the neonatal Fc receptor (FcRn) are perhaps the most well-studied protein-receptor interaction for therapeutic proteins, as it facilitates intracellular recycling, resulting in the long serum half-life of IgG (Roopenian and Akilesh, 2007; Giragossian et al., 2013). Antibodies are pinocytosed into intracellular endosomes where the Fc region binds to FcRn under acidic conditions (pH ~ 5.5 – 6). The complex is protected from proteolytic degradation and recycled to the cell surface where the antibody is released at neutral pH (Antohe et al., 2001; Roopenian and Akilesh, 2007). Importantly, it has been shown that modification to the non-Fc portion of an antibody (such as drug conjugation) can alter FcRn binding (Wang et al., 2011). In addition

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to FcRn, there are other cell surface receptors, many expressed in the liver, that recognize post-translational modifications or aberrant proteins and thus may also function as determinants of recycling or elimination for protein therapeutics. Examples include the Stab/Scarb1 family of scavenger receptors, the mannose and galactose receptors, the LDL receptor and the asialoglycoprotein receptor 1 (Ashwell and Morell, 1974; Ashwell and Harford, 1982; Inamoto and Brown, 1991; Stahl, 1992; Terpstra et al., 2000; Allavena et al., 2004; Goetze et al., 2011).

While small molecule drug discovery has optimized many approaches over the years to predict pharmacokinetics prior to conducting in vivo studies, the characterization of pharmacokinetic properties for protein therapeutics is still highly dependent upon the use of preclinical species, with available in vitro and in silico tools only now beginning to emerge (Jaramillo et al., 2017; Avery et al., 2018). Further complicating the development of such tools is the fact that they are often based on the processes that underwrite the catabolism and disposition of monoclonal antibodies, though protein therapeutic developmental efforts now favor highly-engineered protein platforms where in vivo disposition may not be governed by the same mechanisms as monoclonal antibodies (Boehm et al., 1999; Vugmeyster et al., 2012; Ueda, 2014; Ramsland et al., 2015).

To that end, various in vitro and in silico tools are currently being developed to understand protein therapeutic disposition, including surface plasmon resonance to measure FcRn affinity, hydrophobic interaction chromatography to compare relative hydrophobicity and computational approaches that utilize either sequence-based or three-dimensional structural modeling to identify drug candidates (Dostalek et al., 2017). More recently, reports of cellular-based approaches to assess non-specific clearance pathways have been reported. Nonspecific binding to HEK293 or CHO cells has been correlated to high in vivo clearance (Datta-Mannan et al., 2012; Xu et al., 2013; Datta-Mannan et al., 2015b), and association with liver sinusoidal endothelial cells has been implicated in the rapid clearance of a series of IgG-ECD (extracellular domain) and IgG-scFv bispecific antibodies (Datta-Mannan et al., 2016).

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In this study, several of the aforementioned assays were retrospectively applied to a series of Nav1.7 peptide-antibody conjugates based on observations from biodistribution studies that suggested Nav1.7 peptide properties altered tissue distribution of the conjugate relative to the parent antibody. Experiments were designed to test the hypothesis that hepatocytes may play a role in the distribution and elimination of engineered proteins such as these Nav1.7 peptide-antibody conjugates. To this end, an in vitro approach utilizing cryopreserved plated mouse hepatocytes was implemented to measure the depletion of the peptide-antibody conjugates from the media, similar to the approach commonly used in small molecule stability assays. Physicochemical, receptor-binding and in vivo pharmacokinetic data were generated and compared to the results from the hepatocyte assay. Finally, receptor phenotyping and in vivo knockout studies were conducted to identify receptor-mediated pathways beyond FcRn which may contribute to antibody binding in vitro and subsequently to pharmacokinetic profiles in vivo.

## **Materials and Methods**

### **Materials.**

Biotinylated murine anti-human IgG Fc monoclonal antibody (Clone no. 1.35.1) and all antibodies or peptide-antibody conjugates were prepared and purified at Amgen, Inc. (Thousand Oaks, CA) as previously described (Murray et al., 2019). Stable labeled peptides used as internal standards were obtained from Mid-West Biotech, Inc. (Fishers, IN). Cryopreserved mouse hepatocytes and FcRn knockout mouse hepatocytes were purchased from Triangle Research Labs (Durham, NC). CM4 surface plasmon resonance chips were purchased from GE Healthcare Life Sciences (Pittsburgh, PA). Acetyl-low density lipoprotein, Dulbecco phosphate buffered saline, HALT Protease Inhibitor Cocktail (100x), Streptavidin Dynal M-280 magnetic beads and trypsin (TPCK treated) were purchased from Thermo Fisher Scientific (Waltham, MA). Oasis

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HLB 96-well  $\mu$ Elution Plates were obtained from Waters Corp. (Milford, MA). All other reagents were from Sigma-Aldrich (St. Louis, MO) and were of the highest grade possible.

### **Mouse Pharmacokinetics and Biodistribution Studies.**

Animals were cared for in accordance to the *Guide for the Care and Use of Laboratory Animals*, 8<sup>th</sup> Edition (National Research Council, 2011). All in vivo pharmacokinetic and biodistribution studies were conducted at an AAALAC International accredited facility and were approved by the Amgen Institutional Animal Care and Use Committee (Thousand Oaks, CA). Mice were maintained on a 12:12 h light:dark cycle, and ambient temperature and humidity were kept between 68 – 79 °F and 30 – 70%, respectively. Animals were allowed access to pelleted feed (Envigo Teklad, 2920X irradiated, Indianapolis, IN) and reverse osmosis-purified water *ad libitum*.

**Pharmacokinetic Studies.** Studies to characterize the intravenous clearance of a set of peptide-antibody conjugates were performed in male CD-1 mice (8 – 12 weeks old; Charles River Laboratories, Hollister, CA). Peptide-antibody conjugates were formulated in 10 mM sodium acetate with 9% sucrose (pH 5.2) at 1 mg/mL. Dosing (5 mg/kg) was via intravenous bolus tail vein injection. Blood samples were collected at pre-determined time points up to 168 hours post-dose by submandibular venipuncture. Whole blood was collected and placed into Sarstedt Microvette® 500 plasma tubes containing K2-EDTA and inverted several times prior to centrifugation at 14,000 x *g* for 5 minutes. The resulting plasma was stored at -70 °C until analysis. The pharmacokinetics of **aDNP** and Conjugate **4** were also evaluated in cynomolgus monkeys following a 3 mg/kg intravenous dose (10 mM sodium acetate with 9% sucrose (pH 5.2)). Blood samples were collected through 672 hours post-dose and processed as noted above.

**In Vivo Biodistribution Studies.** Representative anti-DNP antibodies were radiolabeled with <sup>64</sup>Cu as previously described (Murray et al., 2019). In brief, antibodies were modified via conjugation to NODA-

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GA and incubated with  $^{64}\text{CuCl}_2$  (1-2 mCi in 5-10  $\mu\text{L}$ , 0.1 M HCl) for 1 hour at 37 °C. The reaction was quenched with 10 mM EDTA for 5 min to remove any unchelated  $^{64}\text{Cu}$ . Thin layer chromatography with radiochemical detection was used to determine labeling efficiency as previously noted, with all conjugates exhibiting greater than 95% purity.

Mice (male, CD-1) were administered intravenous bolus tail vein injections with 100  $\mu\text{Ci}$  of radiolabeled conjugate in 100-125  $\mu\text{L}$  of saline for a total dose of 30 – 40 mg/kg. Mice were anesthetized using 1–2 % isoflurane delivered in 100% oxygen gas, provided with supplemental heating, and were monitored for physical well-being over the course of the PET scan acquisition. All images were acquired using a Siemens Inveon scanner (Siemens, Erlangen, Germany) at 3 hours post dose. Data analysis incorporated Siemens IAW and IRW software applications.

**Hepatocyte Depletion Assay.** An in vitro cellular assay using cryopreserved, plated CD-1 mouse or cynomolgus monkey hepatocytes was designed to monitor the depletion of the antibody-peptide conjugates from the hepatocyte media. In brief, hepatocytes (Triangle Research Labs, Durham, NC) were plated at 1 million cells per well, allowed to attach for 2 hours and subsequently washed with DMEM buffer (Sigma Aldrich, St. Louis, MO) containing 5% fetal bovine serum. The peptide-antibody conjugates (diluted in DMEM buffer containing 5% fetal bovine serum) were added to the hepatocytes (20  $\mu\text{g/mL}$ , final concentration) and incubated for up to 3 hours at 37 °C under 5%  $\text{CO}_2$ . Aliquots were removed at 0, 30, 60, 90, 120 and 150 minutes and immediately frozen for subsequent analysis. Following the 150 minute timepoint, the hepatocytes were washed three times with DMEM buffer containing 5% fetal bovine serum and lysed by addition of 0.5 mL lysis buffer. Lysed contents were collected and frozen for subsequent ELISA analysis. In general, cellular viability was > 90% over the course of the incubation.

**FcRn Binding.** The affinity of the peptide-antibody conjugates towards FcRn was measured by surface plasmon resonance using FcRn conjugated to a CM4 chip via an amine tether (1000 – 2000 RU level of

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immobilization) on a Biacore T200 (Biacore, GE Healthcare, Pittsburgh, PA). Affinity measurements were carried out at 25 °C using a flow rate of 50  $\mu$ L/min, a peptide-antibody concentration of between 0.1 – 1  $\mu$ M and association and dissociation phases of 1.5 minutes and 10 minutes, respectively. Responses (RU) were monitored at pH 5.5 and pH 7.4 using phosphate buffered saline (0.005% P20) and 10 mM sodium acetate / 150 mM NaCl / 0.005% P20 (pH 5.5 or pH 7.4) as running and sample buffers. Binding affinity is expressed as RU values as determined in the Biacore evaluation software.

**HIC Chromatography.** Peptide-antibody conjugates were analyzed by hydrophobic interaction chromatography (HIC) using a Dionex ProPac HIC-10, 5  $\mu$ m, 300A, 2.1 x 100 mm column attached to an Agilent analytical HPLC by eluting with buffer A (1.0 M  $(\text{NH}_4)_2\text{SO}_4$ , 20 mM NaOAc, pH 5) and buffer B (20 mM NaOAc, 10%  $\text{CH}_3\text{CN}$ , pH 5, at a 0.4 mL/min flow rate). The gradient was held at 0% B from 0-2 min, ramped 0-100% B from 2-12 min, held at 100% B from 12-13 min, ramped 100-0% B from 13-14 min, and re-equilibrated at 0% B from 14-18 min. Samples were diluted 10-fold in buffer A prior to injection, and 5-30  $\mu$ g of material were injected.

**Receptor Phenotyping Experiments.** To evaluate the role of specific cell-surface receptors in the depletion of the peptide-antibody conjugates, the unconjugated control antibody (**aDNP**) or Conjugate **4** were incubated in the above mentioned hepatocyte assay in the presence or absence of selective inhibitors. Acetyl-low density lipoprotein (Scarb/Stab1; 20  $\mu$ g/mL), ovalbumin (mannose and galactose receptors; 1 mg/mL), chondroitin (class A scavengers; 1 mg/mL) or IgG (FcRn; 500  $\mu$ g/mL) were added to the plated hepatocytes and allowed to incubate for 5 minutes prior to adding **aDNP** or Conjugate **4** (20  $\mu$ g/mL; final concentration). Incubations were carried out for 2.5 hours at 37 °C under 5%  $\text{CO}_2$ , at which point an aliquot of media was sampled and frozen for subsequent ELISA. Based on preliminary data, acetylated LDL and ovalbumin were selected for follow up studies. A range of inhibitor concentrations (acetylated LDL, 0.02

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– 0.5 mg/mL; ovalbumin, 0.3 – 10 mg/mL) was used to more definitively assess the contribution of these two scavenger pathways to the disposition of Conjugate **4**.

**In vivo KO Studies.** Pharmacokinetic studies were also carried out in knockout mouse models to further evaluate the potential mechanisms responsible for in vivo clearance. FcRn (B6.129P2-*B2m*<sup>tm1Unc</sup>/J, stock# 002087), low-density lipoprotein receptor (Ldlr<sup>tm1Her</sup>/J, stock# 002207) and mannose receptor (B6.129P2-*Mrc1*<sup>tm1Mnz</sup>/J, stock# 007620) knockout models were obtained from the Jackson Laboratory (Bar Harbor, ME) while asialoglycoprotein receptor 1 (ASGR1) knockout mice (B6.129S4-*Asgr1*<sup>tm1Sau</sup>/SaubJxmJ, stock# 009105) were obtained from Charles River Laboratories (Wilmington, MA). Knockout or wild-type mice received a 2 mg/kg intravenous bolus dose of either **aDNP** or Conjugate **4** (0.4 mg/mL in 10 mM sodium acetate with 9% sucrose (pH 5.2)) via tail vein injection. Blood sampling and processing details were similar to those described above.

**Measurement of Test Articles from In Vitro Studies.** Peptide-antibody conjugate levels from in vitro studies were detected and quantitated by ELISA. A capture plate was prepared by coating a high binding Costar ELISA plate (Sigma Aldrich, St. Louis, MO) with an anti-DNP 3A4 monoclonal antibody (2 µg/mL, final concentration) in phosphate buffered saline. Capture plates were washed (3x) and sample aliquots from in vitro studies (100 µL) were added and incubated for 1 hour at room temperature followed by a second wash step (5x) prior to the addition of a detection antibody. Analytes were detected by addition of a horse radish peroxidase conjugated mouse anti-human Fc monoclonal antibody (7.4 µg/mL, final concentration; 1 hour incubation at room temperature). Detection plates were washed (5x), followed by addition of TMB peroxidase substrate and TMB peroxidase substrate solution B (KPL SeraCare, Milford MA). Plates were incubated for 20 minutes and quenched with 2N sulfuric acid. Luminescence was read at 450 nm (reference wavelength, 650 nm) on a SpectraMax M2 plate reader (Molecular Devices,

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Sunnyvale, CA). Standard curves were prepared from 2.4 – 2500 ng/mL and test article concentrations extrapolated from the standard curve response.

**Measurement of Test Articles in Serum from Pharmacokinetic Studies.** Total and intact peptide-antibody conjugates from pharmacokinetic studies were evaluated using immunocapture followed by tandem mass spectrometry as previously described (Murray et al., 2019). In brief, an aliquot of each plasma or serum sample was diluted 1:1 (v/v) with Dulbecco phosphate buffered saline, (1x DPBS) with 2x HALT protease inhibitor cocktail. Biotinylated murine anti-human IgG Fc monoclonal antibody (Clone no. 1.35.1) immobilized on Streptavidin Dynal M-280 magnetic beads was added as immunoaffinity capture reagent to a 96-well plate format. Samples were vortex-mixed for 1 hour at room temperature and the beads subsequently washed with Tris-HCl buffer (pH 7.5). Reduction of the samples was accomplished by addition of TCEP solution (7.5 mM in 250 mM Tris-HCl, 8 M urea, pH 7.5) containing the relevant internal standard as noted below. Samples were alkylated with iodoacetamide for 45 minutes at 60 °C and subsequently digested with trypsin (500 µg/mL) in a microwave set to 30 °C / 400 W for 10 minutes. Samples were quenched with 20% formic acid and centrifuged at 4500 rpm for 10 minutes prior to analysis by liquid chromatography-tandem mass spectrometry (LC-MS/MS).

LC-MS/MS analysis of the peptide analytes was achieved using an Acquity UHPLC (Waters Corporation, Milford, MA) coupled to a Sciex QTRAP® 5500 mass spectrometer (AB Sciex, Foster City, CA) that was operated in the electrospray positive ion MRM mode. A Phenomenex Aeris C18 column (2.1x 100mm, 1.7 µm; Phenomenex, Carlsbad, CA) held at 70 °C was used for chromatographic separation of the peptide analytes. Total test article (antibody or peptide conjugate) was quantitated using a universal surrogate peptide (peptide sequence: VVSVLTVLHQDWLNGK) corresponding to the Fc region of human IgG1 monoclonal antibodies together with the corresponding stable label isotopic standard ([<sup>13</sup>C<sub>6</sub>, <sup>15</sup>N]-leucine; peptide sequence: VVSV\*LTVLHQDW\*LNGK). Specific peptide sequences (Supplemental Table 1) were utilized to quantitate intact peptide-antibody conjugates.

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Total and intact analyte concentrations were calculated by comparing peak area ratios to calibration curves prepared in a similar fashion using a  $1/x^2$  weighted linear least-squares regression. Pharmacokinetic parameters were estimated by noncompartmental analysis using Watson LIMST<sup>TM</sup>.

## Results

Pharmacokinetic studies in wild-type mice were carried out in support of lead candidate identification efforts for a series of Nav1.7 inhibitory peptide-antibody conjugates with varying peptide identity and formal charge, linker length and architecture, or antibody attachment site. Following a 5 mg/kg intravenous dose, the in vivo clearance of the unconjugated **aDNP** antibody was 0.0006 L/hr/kg. Clearance values for the conjugated antibodies ranged from 0.0013 L/hr/kg for conjugates **12** and **13** to 0.079 L/hr/kg for conjugate **2** (Table 1). The ratio of intact AUC to total AUC for the peptide conjugates ranged from < 0.1 for conjugate **2** to 0.76 for conjugate **5**. The highest clearance values and lowest  $AUC_{\text{intact}} / AUC_{\text{total}}$  ratios were observed for conjugates **1** and **2**, the two peptide conjugates with a peptide charge of +6. Lower clearance values and higher  $AUC_{\text{intact}} / AUC_{\text{total}}$  ratios were observed for peptide conjugates with a peptide charge of +1 or +2 as compared to those with a +6 charge, though no statistically significant difference was observed in clearance or  $AUC_{\text{intact}} / AUC_{\text{total}}$  between peptide conjugates with a peptide charge of +1 versus a peptide charge of +2 (Figure 1a). The stability and pharmacokinetic properties of antibodies is also known to be affected by the global physicochemical properties of the molecules. To evaluate the correlation between the hydrophobicity of the Nav1.7 peptide-conjugates and in vivo clearance or depletion from the in vitro hepatocyte assay, hydrophobic-interaction chromatography analyses were performed with the peptide-conjugates. A significant correlation (Pearson  $r = 0.8005$ ,  $N = 12$ ,  $P = 0.0018$ ) was observed between the HIC retention time and the clearance measured from in vivo pharmacokinetic studies (Figure 1b).

To characterize the large differences observed for in vivo clearance between the unconjugated **aDNP** antibody and the corresponding peptide-antibody conjugates, image-based biodistribution studies were carried out using <sup>64</sup>Cu-NODA labeled antibodies. <sup>64</sup>Cu-NODA imaging data suggested that at 3 hours

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post dose, both the unconjugated aDNP antibody (**aDNP**) and a representative symmetrically conjugated divalent peptide-antibody conjugate (**P<sub>2</sub>S-aDNP**; peptide charge: +6) molecule were constrained primarily to the vasculature, with the bladder being the main secondary site of accumulation (Figure 2). Conversely, a representative asymmetrically conjugated divalent peptide-antibody conjugate containing two peptides on a single mAb HC (**P<sub>2</sub>A-aDNP**; peptide charge: +6), or symmetrically conjugated tetravalent peptide-antibody conjugate with containing two peptides on each HC (**P<sub>4</sub>S-aDNP**; peptide charge: +6) demonstrated rapid distribution out of the central compartment and accumulation predominantly into the liver at 1 hour post injection (Figure 2).

Based on the results from the biodistribution studies, it was hypothesized that binding and/or uptake into hepatocytes may contribute to the elimination of the peptide-antibody conjugates with high clearance values in vivo. To test this hypothesis, a diverse chemical series of peptide-antibody conjugates were initially incubated with suspended mouse hepatocytes and the percent remaining in the media monitored over time. Exploratory experiments (data not shown) suggested a limited dynamic range for the assay and as such, subsequent experiments were carried out with cryopreserved plated mouse hepatocytes and the percent remaining antibody vs time in the media quantitated by ELISA. After incubation of the unconjugated or conjugated antibodies for up to 3 hours in the plated hepatocyte assay, 38 – 93% of the starting antibody concentration remained in the media (Table 1). The percent remaining in the media exhibited a significant correlation with the observed in vivo clearance (Pearson  $r = -0.5525$ ,  $N = 15$ ,  $P = 0.0327$ ), with more rapid depletion from the media correlating with higher in vivo clearance (Figure 3a). A significant correlation was also observed between HIC retention time and percent remaining in the media from the hepatocyte assay (Pearson  $r = -0.7185$ ,  $N = 15$ ,  $P = 0.0085$ ; Figure 3b).

The role of FcRn as a recycling pathway for IgG antibodies is well documented (Roopenian and Akilesh, 2007). Surface plasmon resonance studies were carried out at pH 5.5 and pH 7.4 in order to assess FcRn binding of our conjugate panel in vitro. Positive correlations were noted between FcRn binding at pH 7.4 (Pearson  $r = 0.9107$ ,  $N = 7$ ,  $P = 0.0044$ ) or pH 5.5 (Pearson  $r = 0.8088$ ,  $N = 7$ ,  $P = 0.0276$ ) and the

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in vivo clearance for the peptide-antibody conjugates (Figures 4a and 4b). A positive correlation was observed also observed between FcRn binding at pH 7.4 (Pearson  $r = 0.7114$ ,  $N = 6$ ,  $P = 0.1129$ ) or pH 5.5 (Pearson  $r = 0.8449$ ,  $N = 6$ ,  $P = 0.0342$ ) and the HIC retention time (Figures 4c and 4d), though the correlation at pH 7.4 was not statistically significant. Finally, a strong negative correlation was observed between FcRn binding at pH 7.4 and the percent remaining in the media from the aforementioned hepatocyte assay (Pearson  $r = -0.9455$ ,  $N = 7$ ,  $P = 0.0013$ ; Figure 4e). Conversely, no correlation was observed between FcRn binding at pH 5.5 the percent remaining in the hepatocyte assay media (Pearson  $r = -0.3902$ ,  $N = 7$ ,  $P = 0.3868$ ; Figure 4f). A summary of the statistical analysis for the relationships between in vivo clearance, percent remaining in hepatocyte media, HIC retention time, and FcRn binding at pH 7.4 or 5.5 is shown in Table 3. Initial attempts to assess the translatability of the aforementioned correlations to cynomolgus monkeys are shown in open circles in Figures 1, 3 and 4, though due to the limited nature of the data set, no statistical analyses were performed.

To further explore the role of cell surface receptors in the non-target-mediated binding and/or uptake of the Nav1.7 peptide-antibody conjugates to plated hepatocytes, receptor phenotyping experiments were designed to evaluate the individual contributions of surface receptors known to play a role in binding antibodies and proteins (Ashwell and Harford, 1982; Terpstra et al., 2000). Similar to earlier experiments, **aDNP** was not depleted from the media of plated hepatocytes while only 33% of Conjugate **4** remained in the media after 2.5 hrs (Figure 5). Incubation of Conjugate **4** in FcRn knockout hepatocytes resulted in a lack of depletion from the media, similar to what was observed for **aDNP** in control hepatocytes. Inclusion of acetylated LDL or ovalbumin in the incubation increased the percent remaining Conjugate **4** to 88% or 69%, respectively, suggesting a role for the Stab/Scarb1 receptors or mannose/galactose receptors in depleting the peptide-antibody conjugates from the media. Follow on experiments with increasing amounts of acetylated LDL resulted in complete inhibition of Conjugate **4** uptake ( $IC_{50} = 0.024$  mg/mL; Figure 6) while addition of ovalbumin reached a maximum inhibition of 64% of control uptake (estimated  $IC_{50} = 1.91$

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mg/mL; Figure 6). Inclusion of chondroitin (class A scavengers) or IgG (multiple receptors) had no measurable effect on the depletion of Conjugate **4** from the media.

Finally, pharmacokinetic studies were carried out in knockout mouse models to evaluate the in vitro to in vivo translation of the results from hepatocyte experiments. Experiments utilized FcRn, LDL receptor and mannose receptor knockout models, as well as a knockout mouse model for ASGR1, a receptor expressed on the surface of hepatocytes that has been shown to be involved in protein uptake (Inamoto and Brown, 1991). The pharmacokinetics of **aDNP** and Conjugate **4** (both total and intact conjugate) were evaluated in each strain. The clearance of **aDNP**, Conjugate **4**<sub>total</sub> and Conjugate **4**<sub>intact</sub> increased 15.3, 49.3 and 6.81-fold, respectively, in FcRn knockout mice (Table 2; Figure 7). A modest increase (2.18-fold) in the clearance of Conjugate **4**<sub>total</sub> was observed in mannose receptor knockout mice. Clearance and AUC values for all three analytes were within 2-fold of wild-type mice for the LDL receptor and ASGR1 knockout models. A model to evaluate the contribution of the Stab/Scarb1 pathway was not available at the time this manuscript was prepared.

## Discussion

The identification of non-opioid alternatives to treat chronic pain is an area of intense scientific research owing in no small part to the addiction risks and societal burdens associated with the current opioid-based standard of care (Dunn et al., 2016; Humphreys, 2017; Kaye et al., 2017). The Nav1.7 voltage-gated sodium ion channel (SCN9A) represents one potential pharmacological target with convincing human genetic validation for the reduction of pain without the use of opioids (Dib-Hajj et al., 2013; Emery et al., 2016; Vetter et al., 2017). Humans expressing loss-of-function mutations in the SCN9A gene have a congenital insensitivity to pain while those with gain-of-function mutations are often diagnosed with primary erythromelalgia or paroxysmal extreme pain disorder (Yang et al., 2004; Cox et al., 2006;

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Fertleman et al., 2006; Goldberg et al., 2007; Faber et al., 2012). As such, significant efforts have been placed on identifying Nav1.7 inhibitors across a range of therapeutic modalities, including small molecules, peptides and antibodies.

Previous reports have described internal efforts to identify potent inhibitory peptides of Nav1.7 based on GpTx-1 and JzTx-V peptide scaffolds that have been identified and isolated from tarantula spider venom (Biswas et al., 2017; Moyer et al., 2018; Wu et al., 2018). As many therapeutic peptides suffer from poor pharmacokinetic profiles due to proteolysis and/or renal filtration (Diao and Meibohm, 2013; Di, 2015), a half-life extension approach was adopted whereby the inhibitory peptides were conjugated via various linkers to non-targeting antibodies to increase their molecular weight and hydrodynamic radius while concurrently taking advantage of antibody recycling mechanisms (Biswas et al., 2017; Murray et al., 2019). An inherent challenge to this approach is the sub-optimal pharmacokinetics often observed with modified antibodies and the subsequent need to conduct numerous in vivo pharmacokinetic studies, given the nascency of in vitro approaches to prioritize protein therapeutics for in vivo studies (Vugmeyster et al., 2012; Jaramillo et al., 2017). To that end, we set out to develop an integrated in vitro strategy to optimize the pharmacokinetic properties of Nav1.7 peptide-antibody conjugates prior to conducting in vivo pharmacokinetic studies, with an eye towards more rapidly identifying a lead candidate to progress into late stage preclinical studies while concurrently fostering the “3Rs principle” (refine, reduce, replace) with regard to animal studies (Russell, 1995).

In conjunction with lead candidate identification efforts, intravenous pharmacokinetic studies were conducted across a panel of Nav1.7 peptide-antibody conjugates to set a baseline for which to characterize in vitro tools. The in vivo clearance of the unconjugated, non-targeting aDNP antibody (**aDNP**) was 0.0006 L/hr/kg, within the range of what would be expected for a typical antibody in the absence of target mediated clearance (Table 1). Conjugation of the aDNP antibody with Nav1.7 inhibitory peptides resulted in an approximate 22- to 132-fold increase in its clearance, suggesting that the nature of the peptide and linker had a significant effect on the clearance of the antibody and provided a sufficiently wide range of clearance

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values to compare to in vitro predictors. In addition to total peptide-antibody conjugate clearance, the ratio of intact to total conjugate ( $AUC_{\text{intact}} / AUC_{\text{total}}$ ) was also measured as it has been previously shown that proteolytic cleavage of peptide conjugates can result in a loss of pharmacological activity (Hecht et al., 2012; Hager et al., 2013). The observed  $AUC_{\text{intact}} / AUC_{\text{total}}$  ratios ranged from 0.1 to 0.76, with the lowest ratios being attributed to the two peptide-antibody conjugates with a peptide charge of +6 (conjugates **1** and **2**). As these two conjugates also exhibited the highest clearance values in vivo, it was hypothesized that the charge of the peptide was a key determinant of the in vivo pharmacokinetic characteristics of the corresponding peptide conjugate.

The charge and resulting isoelectric point of protein therapeutics is a well-documented determinant of their in vivo pharmacokinetic attributes (Boswell et al., 2010; Bumbaca et al., 2012; Li et al., 2014; Datta-Mannan et al., 2015b). Proteins with net positive charges often tend to interact more readily with negatively charged cell surface components, resulting in a more rapid clearance and increased biodistribution compared to corresponding proteins with less positive charges (Bergmann et al., 1984; Pardridge et al., 1995; Lee and Pardridge, 2003; Herve et al., 2008). As the initial JzTx-V peptides were highly basic with a positive charge of +6 and isoelectric points between 10.5 and 11, it can be hypothesized that conjugating multiple peptides to the aDNP antibody would result in a substantial increase to the overall pI of the peptide-antibody conjugate and negatively affect the resulting pharmacokinetic properties of the conjugates. Indeed, lower clearance values were observed when the net charge on the peptide was reduced to +1 or +2, though we were unable to further differentiate clearance values when comparing peptide-antibody conjugates with +1 vs +2 charged peptides (Figure 1a), suggesting that additional assays were needed to efficiently differentiate the conjugates prior to advancement to in vivo studies. Similar results were reported by Murray et al., where reductions in the positive nature of the conjugated peptide resulted in decreased clearance and increased exposure for Nav1.7 peptide-antibody conjugates (Murray et al., 2019). In addition to charge and isoelectric point, the structural stability and pharmacokinetic properties of antibodies is also known to be related to the overall hydrophobicity of the molecule (Deng et al., 2012).

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When the series of Nav1.7 peptide-conjugates were analyzed by hydrophobic interaction chromatography, a trend was observed between longer HIC retention times and increased in vivo clearance (Figure 1b). In a similar vein, decreases in clearance for a series of anti-CD70 antibody-drug conjugates were observed as modifications were made to the linker region to minimize the overall hydrophobicity of the antibody-drug conjugate (Lyon et al., 2015). As the analytical approach is relatively straightforward and requires a minimal amount of material, it can be readily incorporated into structural optimization efforts prior to progressing molecules to in vivo pharmacokinetic studies.

As mentioned above, cationic proteins with higher isoelectric points tend to more readily distribute into tissues as compared to their uncharged or negatively charged counterparts (Bickel et al., 1994; Pardridge et al., 1995; Khawli et al., 1996; Pardridge et al., 1998; Hong et al., 1999; Kobayashi et al., 1999; Lee and Pardridge, 2003; Boswell et al., 2010). As binding to target was not a biodistribution mechanism for the non-targeting aDNP antibodies, biodistribution studies were carried out using whole-body positron emission tomography with representative  $^{64}\text{Cu}$ -NOTA labeled Nav1.7 peptide antibody conjugates to determine if modifications to the peptide affected the broader biodistribution pattern of the peptide-antibody conjugate. The distribution of the unconjugated antibody or a representative peptide-antibody conjugate with two symmetrically conjugated peptides (**P<sub>2</sub>S-aDNP**; peptide charge: +6) following intravenous administration was generally constrained to the vasculature with a small amount found in the bladder (Figure 2). However, when the aDNP antibody was conjugated with two peptides in an asymmetrical fashion or with four peptides (**P<sub>2</sub>A-aDNP** or **P<sub>4</sub>S-aDNP**; peptide charge: +6), a rapid distribution to the liver was observed after 3 hours (Figure 2), suggesting that the molecular asymmetry may shunt the antibody conjugate towards specific pattern-recognition receptors on a given tissue such as the liver, a finding which may prove useful in general antibody-based protein engineering efforts (Szabo et al., 2006; Mullen et al., 2015). Unfortunately, while hepatic elimination is often considered when evaluating the elimination of small molecule therapeutics, it is generally overlooked in the pharmacokinetic assessment of protein therapeutics. This approach may be perfectly acceptable when the protein of interest is an

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unmodified monoclonal antibody, however as mentioned, engineered platforms such as antibody-conjugates, Fc-fusion proteins and bispecific antibodies may be more readily recognized as modified proteins and thus directed to the liver for elimination through multiple scavenging pathways (Liu, 2018). To this point, Datta-Mannan and colleagues have recently demonstrated the role of liver sinusoidal endothelial cells in the clearance of modified bispecific antibodies in cynomolgus monkeys (Datta-Mannan et al., 2016). Based on our biodistribution data and the findings from Datta-Mannan and colleagues, we hypothesized that hepatocytes may also play a role in the elimination of the Nav1.7 peptide-antibody conjugates and could serve as a readily available in vitro system with which to rank-order prospective peptide conjugates for in vivo evaluation.

The use of cryopreserved hepatocytes to evaluate the metabolic stability of small molecule therapeutics is commonplace in most ADME groups as hepatocytes contain the full complement of drug metabolizing enzymes and cofactors involved in the clearance of small molecules (Li, 2001). In addition to small molecule metabolism, hepatocytes also express numerous intracellular and cell surface receptors such as FcRn, ASGR1, mannose, galactose and LDL receptors, in addition to other scavenger receptors, that are known to play a role in protein recycling and elimination (Matsuura et al., 1982; Kindberg et al., 1990; Lagace et al., 2006; Pyzik et al., 2017). It is thus plausible that hepatocytes may represent a holistic system with which to predict the contribution of nonspecific elimination pathways for modified protein therapeutics such as the Nav1.7 peptide-antibody conjugates. Upon incubating a series of Nav1.7 peptide conjugates with cryopreserved, plated mouse hepatocytes and monitoring depletion from the media, a correlation was observed between the percent remaining peptide-antibody conjugate in the media at the end of the incubation and the in vivo clearance, with more rapid depletion from the media correlating with higher in vivo clearance (Figure 3a). Correspondingly, after washing, lysing and collecting the remaining cellular content, conjugates with higher in vivo clearance were observed to have more drug related material bound to or taken up into the hepatocytes as opposed to those with lower in vivo clearance (data not shown). A negative correlation was also observed between the HIC retention time and percent remaining peptide-

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antibody conjugate in the media from the hepatocyte assay, suggesting that hydrophobic interactions may be a key determinant of the depletion of the conjugates from the media (Figure 3b). Owing to their general ease of use, an similar approach using cryopreserved hepatocytes in suspension was evaluated, but the dynamic range was found to be limited (data not shown). Gene and protein expression patterns in hepatocytes have been shown to be highly dependent on thawing protocols and cellular attachment, and as such, it is possible that plated hepatocytes present a more amenable environment in which to study protein-cellular interactions (Waring et al., 2003).

While the ability of the plated hepatocyte approach to evaluate the in vivo clearance of this series of peptide-antibody conjugates in mice appears robust, the true value (similar to in vitro to in vivo extrapolation approaches with small molecules) will lie in the utility of the system to work with higher-order preclinical species and ultimately, to predict human pharmacokinetics. To this end we have extended the current analysis to include pharmacokinetics of **aDNP** and Conjugate **4** in cynomolgus monkeys. The initial results, shown in Figures 1, 3 and 4 suggest that similar trends may exist in cynomolgus monkey hepatocytes as was observed with mouse, though obviously the data set will need to be significantly expanded before any solid conclusions can be made.

The contribution of the neonatal Fc receptor (FcRn) to the pharmacokinetics of antibody therapeutics is well documented and often a key focus of antibody engineering efforts to optimize pharmacokinetic properties (Roopenian and Akilesh, 2007; Giragossian et al., 2013). In order to fully take advantage of the FcRn recycling pathway, the binding of the Fc region to FcRn must be balanced in such a way that the antibody rapidly binds to FcRn at pH 5.5 in the endosome but readily dissociates from the receptor at pH 7.4 when the complex is returned to the cell surface. When surface plasmon resonance was used to measure the binding of the Nav1.7 peptide conjugates to mouse FcRn at pH 5.5 or 7.4, a positive correlation was observed between FcRn binding at both pH 5.5 or pH 7.4 and in vivo mouse clearance (Figures 4a and 4b). Of particular interest for the Nav1.7 peptide-antibody conjugates was the similarity of the antibody backbone across the series, suggesting that peptide conjugation to non-Fc regions of the

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antibody were affecting FcRn binding, similar to what has been previously reported for a set of monoclonal antibodies with identical Fc sequences but varied Fab domains (Wang et al., 2011). A correlation was also observed between the percent remaining peptide-antibody conjugate in the media and FcRn binding affinity at pH 7.4 (Figure 4e) but not pH 5.5, suggesting that the elimination of the peptide conjugates from the hepatocyte media may be a combined function of physicochemical properties and release of the protein from FcRn at the cell surface.

Modifying monoclonal antibodies by conjugating charged peptides to the antibody also opens the possibility that the peptide-antibody conjugate will be recognized as an aberrant protein by other cell surface receptors, as is often the case with scavenger receptors expressed on hepatocytes and liver sinusoidal endothelial cells (Datta-Mannan et al., 2016). To this end, we sought to further characterize other potential receptor-mediated pathways that may be contributing to the observed media depletion in hepatocytes and subsequent *in vivo* clearance. Similar to phenotyping approaches used for small molecule drug metabolizing enzymes, inhibitors of various cell surface receptors were added to plated hepatocyte incubations prior to addition of the Nav1.7 peptide-antibody conjugates (Figure 5). After addition of acetylated LDL or ovalbumin, a decrease in the rate of media depletion for Conjugate **4** was observed, suggesting a role for either the Stab/Scarb1 receptor family (acetylated LDL) or the mannose and galactose receptors (ovalbumin) (Smedsrod et al., 1990; Krieger, 2001). As a confirmatory experiment, IC<sub>50</sub> curves were generated for the depletion of Nav1.7 peptide-antibody conjugates in the presence of increasing amounts of acetylated LDL or ovalbumin, with acetylated LDL potently inhibiting the depletion of the Nav1.7 peptide-antibody conjugates from the hepatocyte media (Figure 6). The primary role of the LDL receptor is the endocytosis of cholesterol-bound low density lipoprotein in the liver while the mannose and galactose receptors are generally involved in the elimination of proteins with complex glycosylation patterns and are expressed on multiple cell types including those in the liver (Ashwell and Harford, 1982; Goetze et al., 2011; Yang et al., 2015). Conjugate **4** depletion was completely abolished when FcRn

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knockout hepatocytes were used, further supporting the importance of the FcRn pathway in the disposition of the Nav1.7 peptide-conjugates.

Finally, to determine the translatability of the in vitro receptor phenotyping efforts to in vivo outcomes, the pharmacokinetics of **aDNP** or conjugate **4** were determined in FcRn, LDL receptor, mannose receptor, or ASGR1 knockout mouse models and compared to the pharmacokinetics measured in wild-type mice. As expected, knocking out FcRn had the greatest effect on the pharmacokinetics of **aDNP** or conjugate **4**, with clearance values increasing 15.3-fold or 49.3-fold, respectively, in FcRn knockout mice relative to wild-type mice. Interestingly, a small increase (2.18-fold) in the clearance of Conjugate **4**<sub>total</sub> was also observed in mannose receptor knockout mice (Figure 7). The data suggests that while multiple receptor-mediated pathways may play a role in the pharmacokinetics and distribution of the Nav1.7 peptide-antibody conjugates, interactions with FcRn are still the primary determinant of the observed in vivo pharmacokinetics.

The range of pharmacokinetics of the Nav1.7 peptide-antibody conjugates described in this paper implicates an important role for the inhibitory peptide in determining the in vivo properties of the peptide-antibody conjugate. Taken in its entirety, the data suggests that an integrated strategy combining hydrophobicity, FcRn affinity and hepatocyte association rates is an efficient way to prioritize Nav1.7 peptide-antibody conjugates for further evaluation and supports the hypothesis that hepatocytes may play a role in the disposition of protein therapeutics. While caution should be exercised in broadly applying such an approach to other engineered proteins especially when target-mediated disposition may be involved, the data presented herein represents an additional resource to more rapidly triage novel protein therapeutic platforms while concomitantly reducing the number of in vivo studies conducted in preclinical species.

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## **Acknowledgements**

The authors would like to thank Drs. Larry C. Wienkers and Josh T. Pearson for valuable discussions and insights throughout the preparation of this manuscript as well as Dr. Bill McCarty for assistance with statistical analyses.

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## **Authorship Contributions**

*Participated in Research Design:* Foti, Biswas, Glaus, Hickman, Murray, Miranda, Moyer, Rock

*Conducted Experiments:* Foti, Huang, Be, Berry, Conner, Ikotun, Soto

*Contributed New Reagents or Analytical Tools:* Murray, Aral, Cheng, Falsey, Herberich, Long, Netirojjanakul, Nixey, Sham, Tegley, Wu, Yin

*Performed Data Analysis:* Foti, Huang, Be, Berry, Conner, Li, Tran

*Wrote or Contributed to the Writing of the Manuscript:* Foti, Murray, Rock

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## References

- Allavena P, Chieppa M, Monti P, and Piemonti L (2004) From pattern recognition receptor to regulator of homeostasis: the double-faced macrophage mannose receptor. *Crit Rev Immunol* **24**:179-192.
- Antohe F, Radulescu L, Gafencu A, Ghetie V, and Simionescu M (2001) Expression of functionally active FcRn and the differentiated bidirectional transport of IgG in human placental endothelial cells. *Hum Immunol* **62**:93-105.
- Ashwell G and Harford J (1982) Carbohydrate-specific receptors of the liver. *Annu Rev Biochem* **51**:531-554.
- Ashwell G and Morell AG (1974) The role of surface carbohydrates in the hepatic recognition and transport of circulating glycoproteins. *Adv Enzymol Relat Areas Mol Biol* **41**:99-128.
- Avery LB, Wade J, Wang M, Tam A, King A, Piche-Nicholas N, Kavosi MS, Penn S, Cirelli D, Kurz JC, Zhang M, Cunningham O, Jones R, Fennell BJ, McDonnell B, Sakorafas P, Apgar J, Finlay WJ, Lin L, Bloom L, and O'Hara DM (2018) Establishing in vitro in vivo correlations to screen monoclonal antibodies for physicochemical properties related to favorable human pharmacokinetics. *MAbs* **10**:244-255.
- Bergmann P, Kacenelenbogen R, and Vizet A (1984) Plasma clearance, tissue distribution and catabolism of cationized albumins with increasing isoelectric points in the rat. *Clin Sci (Lond)* **67**:35-43.
- Bickel U, Lee VM, Trojanowski JQ, and Pardridge WM (1994) Development and in vitro characterization of a cationized monoclonal antibody against beta A4 protein: a potential probe for Alzheimer's disease. *Bioconjug Chem* **5**:119-125.
- Biswas K, Nixey TE, Murray JK, Falsey JR, Yin L, Liu H, Gingras J, Hall BE, Herberich B, Holder JR, Li H, Ligutti J, Lin MJ, Liu D, Soriano BD, Soto M, Tran L, Tegley CM, Zou A, Gunasekaran K, Moyer BD, Doherty L, and Miranda LP (2017) Engineering Antibody Reactivity for Efficient Derivatization to Generate NaV1.7 Inhibitory GpTx-1 Peptide-Antibody Conjugates. *ACS Chem Biol* **12**:2427-2435.

DMD# 087742

- Boehm MK, Woof JM, Kerr MA, and Perkins SJ (1999) The Fab and Fc fragments of IgA1 exhibit a different arrangement from that in IgG: a study by X-ray and neutron solution scattering and homology modelling. *J Mol Biol* **286**:1421-1447.
- Boswell CA, Tesar DB, Mukhyala K, Theil FP, Fielder PJ, and Khawli LA (2010) Effects of charge on antibody tissue distribution and pharmacokinetics. *Bioconjug Chem* **21**:2153-2163.
- Bumbaca D, Boswell CA, Fielder PJ, and Khawli LA (2012) Physiochemical and biochemical factors influencing the pharmacokinetics of antibody therapeutics. *AAPS J* **14**:554-558.
- Cox JJ, Reimann F, Nicholas AK, Thornton G, Roberts E, Springell K, Karbani G, Jafri H, Mannan J, Raashid Y, Al-Gazali L, Hamamy H, Valente EM, Gorman S, Williams R, McHale DP, Wood JN, Gribble FM, and Woods CG (2006) An SCN9A channelopathy causes congenital inability to experience pain. *Nature* **444**:894-898.
- Datta-Mannan A, Chow CK, Dickinson C, Driver D, Lu J, Witcher DR, and Wroblewski VJ (2012) FcRn affinity-pharmacokinetic relationship of five human IgG4 antibodies engineered for improved in vitro FcRn binding properties in cynomolgus monkeys. *Drug Metab Dispos* **40**:1545-1555.
- Datta-Mannan A, Croy JE, Schirtzinger L, Torgerson S, Breyer M, and Wroblewski VJ (2016) Aberrant bispecific antibody pharmacokinetics linked to liver sinusoidal endothelium clearance mechanism in cynomolgus monkeys. *MAbs* **8**:969-982.
- Datta-Mannan A, Lu J, Witcher DR, Leung D, Tang Y, and Wroblewski VJ (2015a) The interplay of non-specific binding, target-mediated clearance and FcRn interactions on the pharmacokinetics of humanized antibodies. *MAbs* **7**:1084-1093.
- Datta-Mannan A, Thangaraju A, Leung D, Tang Y, Witcher DR, Lu J, and Wroblewski VJ (2015b) Balancing charge in the complementarity-determining regions of humanized mAbs without affecting pI reduces non-specific binding and improves the pharmacokinetics. *MAbs* **7**:483-493.
- Deng R, Jin F, Prabhu S, and Iyer S (2012) Monoclonal antibodies: what are the pharmacokinetic and pharmacodynamic considerations for drug development? *Expert Opin Drug Metab Toxicol* **8**:141-160.

DMD# 087742

- Di L (2015) Strategic approaches to optimizing peptide ADME properties. *AAPS J* **17**:134-143.
- Diao L and Meibohm B (2013) Pharmacokinetics and pharmacokinetic-pharmacodynamic correlations of therapeutic peptides. *Clin Pharmacokinet* **52**:855-868.
- Dib-Hajj SD, Yang Y, Black JA, and Waxman SG (2013) The Na(V)1.7 sodium channel: from molecule to man. *Nat Rev Neurosci* **14**:49-62.
- Dostalek M, Prueksaritanont T, and Kelley RF (2017) Pharmacokinetic de-risking tools for selection of monoclonal antibody lead candidates. *MAbs* **9**:756-766.
- Dunn LK, Durieux ME, and Nemergut EC (2016) Non-opioid analgesics: Novel approaches to perioperative analgesia for major spine surgery. *Best Pract Res Clin Anaesthesiol* **30**:79-89.
- Emery EC, Luiz AP, and Wood JN (2016) Nav1.7 and other voltage-gated sodium channels as drug targets for pain relief. *Expert Opin Ther Targets* **20**:975-983.
- Faber CG, Hoeijmakers JG, Ahn HS, Cheng X, Han C, Choi JS, Estacion M, Lauria G, Vanhoutte EK, Gerrits MM, Dib-Hajj S, Drenth JP, Waxman SG, and Merkies IS (2012) Gain of function Nav1.7 mutations in idiopathic small fiber neuropathy. *Ann Neurol* **71**:26-39.
- Fertleman CR, Baker MD, Parker KA, Moffatt S, Elmslie FV, Abrahamsen B, Ostman J, Klugbauer N, Wood JN, Gardiner RM, and Rees M (2006) SCN9A mutations in paroxysmal extreme pain disorder: allelic variants underlie distinct channel defects and phenotypes. *Neuron* **52**:767-774.
- Giragossian C, Clark T, Piche-Nicholas N, and Bowman CJ (2013) Neonatal Fc receptor and its role in the absorption, distribution, metabolism and excretion of immunoglobulin G-based biotherapeutics. *Curr Drug Metab* **14**:764-790.
- Goetze AM, Liu YD, Zhang Z, Shah B, Lee E, Bondarenko PV, and Flynn GC (2011) High-mannose glycans on the Fc region of therapeutic IgG antibodies increase serum clearance in humans. *Glycobiology* **21**:949-959.
- Goldberg YP, MacFarlane J, MacDonald ML, Thompson J, Dube MP, Mattice M, Fraser R, Young C, Hossain S, Pape T, Payne B, Radomski C, Donaldson G, Ives E, Cox J, Younghusband HB, Green R, Duff A, Boltshauser E, Grinspan GA, Dimon JH, Sibley BG, Andria G, Toscano E, Kerdraon J,

DMD# 087742

- Bowsher D, Pimstone SN, Samuels ME, Sherrington R, and Hayden MR (2007) Loss-of-function mutations in the Nav1.7 gene underlie congenital indifference to pain in multiple human populations. *Clin Genet* **71**:311-319.
- Hager T, Spahr C, Xu J, Salimi-Moosavi H, and Hall M (2013) Differential enzyme-linked immunosorbent assay and ligand-binding mass spectrometry for analysis of biotransformation of protein therapeutics: application to various FGF21 modalities. *Anal Chem* **85**:2731-2738.
- Hecht R, Li YS, Sun J, Belouski E, Hall M, Hager T, Yie J, Wang W, Winters D, Smith S, Spahr C, Tam LT, Shen Z, Stanislaus S, Chinookoswong N, Lau Y, Sickmier A, Michaels ML, Boone T, Veniant MM, and Xu J (2012) Rationale-Based Engineering of a Potent Long-Acting FGF21 Analog for the Treatment of Type 2 Diabetes. *PLoS One* **7**:e49345.
- Herve F, Ghinea N, and Scherrmann JM (2008) CNS delivery via adsorptive transcytosis. *AAPS J* **10**:455-472.
- Hong G, Bazin-Redureau MI, and Scherrmann JM (1999) Pharmacokinetics and organ distribution of cationized colchicine-specific IgG and Fab fragments in rat. *J Pharm Sci* **88**:147-153.
- Humphreys K (2017) Avoiding globalisation of the prescription opioid epidemic. *Lancet* **390**:437-439.
- Inamoto T and Brown WR (1991) IgG is associated with the asialoglycoprotein receptor in the human liver. *Hepatology* **14**:1070-1075.
- Jaramillo CAC, Belli S, Cascais AC, Dudal S, Edelmann MR, Haak M, Brun ME, Otteneder MB, Ullah M, Funk C, Schuler F, and Simon S (2017) Toward in vitro-to-in vivo translation of monoclonal antibody pharmacokinetics: Application of a neonatal Fc receptor-mediated transcytosis assay to understand the interplaying clearance mechanisms. *MAbs* **9**:781-791.
- Kaye AD, Jones MR, Kaye AM, Ripoll JG, Galan V, Beakley BD, Calixto F, Bolden JL, Urman RD, and Manchikanti L (2017) Prescription Opioid Abuse in Chronic Pain: An Updated Review of Opioid Abuse Predictors and Strategies to Curb Opioid Abuse: Part 1. *Pain Physician* **20**:S93-S109.

DMD# 087742

- Khawli LA, Glasky MS, Alauddin MM, and Epstein AL (1996) Improved tumor localization and radioimaging with chemically modified monoclonal antibodies. *Cancer Biother Radiopharm* **11**:203-215.
- Kindberg GM, Magnusson S, Berg T, and Smedsrod B (1990) Receptor-mediated endocytosis of ovalbumin by two carbohydrate-specific receptors in rat liver cells. The intracellular transport of ovalbumin to lysosomes is faster in liver endothelial cells than in parenchymal cells. *Biochem J* **270**:197-203.
- Kobayashi H, Le N, Kim IS, Kim MK, Pie JE, Drumm D, Paik DS, Waldmann TA, Paik CH, and Carrasquillo JA (1999) The pharmacokinetic characteristics of glycolated humanized anti-Tac Fabs are determined by their isoelectric points. *Cancer Res* **59**:422-430.
- Krieger M (2001) Scavenger receptor class B type I is a multiligand HDL receptor that influences diverse physiologic systems. *J Clin Invest* **108**:793-797.
- Lagace TA, Curtis DE, Garuti R, McNutt MC, Park SW, Prather HB, Anderson NN, Ho YK, Hammer RE, and Horton JD (2006) Secreted PCSK9 decreases the number of LDL receptors in hepatocytes and in livers of parabiotic mice. *J Clin Invest* **116**:2995-3005.
- Lee HJ and Pardridge WM (2003) Monoclonal antibody radiopharmaceuticals: cationization, pegylation, radiometal chelation, pharmacokinetics, and tumor imaging. *Bioconjug Chem* **14**:546-553.
- Li AP (2001) Screening for human ADME/Tox drug properties in drug discovery. *Drug Discov Today* **6**:357-366.
- Li B, Tesar D, Boswell CA, Cahaya HS, Wong A, Zhang J, Meng YG, Eigenbrot C, Pantua H, Diao J, Kapadia SB, Deng R, and Kelley RF (2014) Framework selection can influence pharmacokinetics of a humanized therapeutic antibody through differences in molecule charge. *MAbs* **6**:1255-1264.
- Liu L (2015) Antibody glycosylation and its impact on the pharmacokinetics and pharmacodynamics of monoclonal antibodies and Fc-fusion proteins. *J Pharm Sci* **104**:1866-1884.
- Liu L (2018) Pharmacokinetics of monoclonal antibodies and Fc-fusion proteins. *Protein Cell* **9**:15-32.

DMD# 087742

- Lyon RP, Bovee TD, Doronina SO, Burke PJ, Hunter JH, Neff-LaFord HD, Jonas M, Anderson ME, Setter JR, and Senter PD (2015) Reducing hydrophobicity of homogeneous antibody-drug conjugates improves pharmacokinetics and therapeutic index. *Nat Biotechnol* **33**:733-735.
- Matsuura S, Nakada H, Sawamura T, and Tashiro Y (1982) Distribution of an asialoglycoprotein receptor on rat hepatocyte cell surface. *J Cell Biol* **95**:864-875.
- Moyer BD, Murray JK, Ligutti J, Andrews K, Favreau P, Jordan JB, Lee JH, Liu D, Long J, Sham K, Shi L, Stocklin R, Wu B, Yin R, Yu V, Zou A, Biswas K, and Miranda LP (2018) Pharmacological characterization of potent and selective NaV1.7 inhibitors engineered from *Chilobrachys jingzhao* tarantula venom peptide JzTx-V. *PLoS One* **13**:e0196791.
- Mullen LM, Chamberlain G, and Sacre S (2015) Pattern recognition receptors as potential therapeutic targets in inflammatory rheumatic disease. *Arthritis Res Ther* **17**:122.
- Murray JK, Wu B, Tegley CM, Nixey TE, Falsey JR, Herberich B, Yin L, Sham K, Long J, Aral J, Cheng Y, Netirojjanakul C, Doherty L, Glaus C, Ikotun T, Li H, Tran L, Soto M, Salimi-Moosavi H, Ligutti J, Amagasu S, Andrews KL, Be X, Lin MJ, Foti RS, Ilch CP, Youngblood B, Kornecook TJ, Karow M, Walker KW, Moyer BD, Biswas K, and Miranda LP (2019) Engineering NaV1.7 Inhibitory JzTx-V Peptides with a Potency and Basicity Profile Suitable for Antibody Conjugation To Enhance Pharmacokinetics. *ACS Chem Biol*.
- National Research Council (2011) *Guide for the Care and Use of Laboratory Animals: 8th Edition*. National Academies Press, Washington, D.C.
- Pardridge WM, Buciak J, Yang J, and Wu D (1998) Enhanced endocytosis in cultured human breast carcinoma cells and in vivo biodistribution in rats of a humanized monoclonal antibody after cationization of the protein. *J Pharmacol Exp Ther* **286**:548-554.
- Pardridge WM, Kang YS, Yang J, and Buciak JL (1995) Enhanced cellular uptake and in vivo biodistribution of a monoclonal antibody following cationization. *J Pharm Sci* **84**:943-948.
- Pyzik M, Rath T, Kuo TT, Win S, Baker K, Hubbard JJ, Grenha R, Gandhi A, Kramer TD, Mezo AR, Taylor ZS, McDonnell K, Nienaber V, Andersen JT, Mizoguchi A, Blumberg L, Purohit S, Jones

DMD# 087742

- SD, Christianson G, Lencer WI, Sandlie I, Kaplowitz N, Roopenian DC, and Blumberg RS (2017) Hepatic FcRn regulates albumin homeostasis and susceptibility to liver injury. *Proc Natl Acad Sci U S A* **114**:E2862-E2871.
- Ramsland PA, Hutchinson AT, and Carter PJ (2015) Therapeutic antibodies: Discovery, design and deployment. *Mol Immunol* **67**:1-3.
- Roopenian DC and Akilesh S (2007) FcRn: the neonatal Fc receptor comes of age. *Nat Rev Immunol* **7**:715-725.
- Russell WM (1995) The development of the three Rs concept. *Altern Lab Anim* **23**:298-304.
- Schoch A, Kettenberger H, Mundigl O, Winter G, Engert J, Heinrich J, and Emrich T (2015) Charge-mediated influence of the antibody variable domain on FcRn-dependent pharmacokinetics. *Proc Natl Acad Sci U S A* **112**:5997-6002.
- Smedsrod B, Melkko J, Risteli L, and Risteli J (1990) Circulating C-terminal propeptide of type I procollagen is cleared mainly via the mannose receptor in liver endothelial cells. *Biochem J* **271**:345-350.
- Stahl PD (1992) The mannose receptor and other macrophage lectins. *Curr Opin Immunol* **4**:49-52.
- Szabo G, Dolganiuc A, and Mandrekar P (2006) Pattern recognition receptors: a contemporary view on liver diseases. *Hepatology* **44**:287-298.
- Terpstra V, van Amersfoort ES, van Velzen AG, Kuiper J, and van Berkel TJ (2000) Hepatic and extrahepatic scavenger receptors: function in relation to disease. *Arterioscler Thromb Vasc Biol* **20**:1860-1872.
- Ueda T (2014) Next-generation optimized biotherapeutics - A review and preclinical study. *Biochim Biophys Acta* **1844**:2053-2057.
- Vetter I, Deuis JR, Mueller A, Israel MR, Starobova H, Zhang A, Rash LD, and Mobli M (2017) NaV1.7 as a pain target - From gene to pharmacology. *Pharmacol Ther* **172**:73-100.
- Vugmeyster Y, Xu X, Theil FP, Khawli LA, and Leach MW (2012) Pharmacokinetics and toxicology of therapeutic proteins: Advances and challenges. *World J Biol Chem* **3**:73-92.

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- Wang W, Lu P, Fang Y, Hamuro L, Pittman T, Carr B, Hochman J, and Prueksaritanont T (2011) Monoclonal antibodies with identical Fc sequences can bind to FcRn differentially with pharmacokinetic consequences. *Drug Metab Dispos* **39**:1469-1477.
- Waring JF, Ciurlionis R, Jolly RA, Heindel M, Gagne G, Fagerland JA, and Ulrich RG (2003) Isolated human hepatocytes in culture display markedly different gene expression patterns depending on attachment status. *Toxicol In Vitro* **17**:693-701.
- Wu B, Murray JK, Andrews KL, Sham K, Long J, Aral J, Ligutti J, Amagasu S, Liu D, Zou A, Min X, Wang Z, Ilch CP, Kornecook TJ, Lin MJ, Be X, Miranda LP, Moyer BD, and Biswas K (2018) Discovery of Tarantula Venom-Derived NaV1.7-Inhibitory JzTx-V Peptide 5-Br-Trp24 Analogue AM-6120 with Systemic Block of Histamine-Induced Pruritis. *J Med Chem* **61**:9500-9512.
- Xu Y, Roach W, Sun T, Jain T, Prinz B, Yu TY, Torrey J, Thomas J, Bobrowicz P, Vasquez M, Wittrup KD, and Krauland E (2013) Addressing polyspecificity of antibodies selected from an in vitro yeast presentation system: a FACS-based, high-throughput selection and analytical tool. *Protein Eng Des Sel* **26**:663-670.
- Yang J, Primack R, Frohn M, Wang W, Luan P, Retter MW, and Flynn GC (2015) Impact of glycation on antibody clearance. *AAPS J* **17**:237-244.
- Yang Y, Wang Y, Li S, Xu Z, Li H, Ma L, Fan J, Bu D, Liu B, Fan Z, Wu G, Jin J, Ding B, Zhu X, and Shen Y (2004) Mutations in SCN9A, encoding a sodium channel alpha subunit, in patients with primary erythralgia. *J Med Genet* **41**:171-174.

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## Figure Legends

**Figure 1. Effect of charge (A) and hydrophobicity (B) on Nav1.7 peptide-antibody conjugate pharmacokinetics.** A reduction in clearance was observed for peptide-antibody conjugates with a peptide charge of +1 or +2 as compared to a peptide charge of +6, while a Pearson correlation ( $r = 0.8005$ ) was observed between the HIC retention time and in vivo clearance. Mouse data is denoted with closed circles (●) while cynomolgus monkey data is denoted by open circles (○).

**Figure 2. Cu64-NOTA-labelled imaging studies of the unconjugated antibody (aDNP) and 3 peptide-antibody conjugates.** At 3 hours post-dose, aDNP and a representative symmetrical divalent conjugate (P<sub>2</sub>S-aDNP) were confined primarily to the central compartment while the asymmetrical divalent conjugate (P<sub>2</sub>A-aDNP) and a tetravalent peptide-antibody conjugate (P<sub>4</sub>S-aDNP) distributed rapidly to the liver.

**Figure 3. Percent of peptide-antibody conjugates remaining in the medium after 2.5 hours in plated mouse hepatocyte assays versus in vivo clearance from intravenous pharmacokinetic studies (A) or HIC retention time (B).** Peptide-conjugates exhibiting extensive association with hepatocytes (lower percent remaining in media) demonstrated higher in vivo clearance values than those with decreased association (higher percent remaining in media; Pearson  $r = -0.5525$ ). A correlation was also observed between HIC retention time and percent remaining in the hepatocyte media (Pearson  $r = -0.7185$ ). Mouse data is denoted with closed circles (●) while cynomolgus monkey data is denoted by open circles (○).

**Figure 4. Observed correlation between FcRn binding affinity of peptide conjugates at pH 7.4 or 5.5 (RUs) and in vivo clearance (A and B), HIC retention time (C and D) or percent remaining in hepatocyte media (E and F).** Peptide-conjugates with high affinity for FcRn tended to exhibit higher in vivo clearances, HIC retention times and depletion rates from hepatocyte media (Pearson  $r$ -values noted in Table 3. Mouse data is denoted with closed circles (●) while cynomolgus monkey data is denoted by open circles (○).

**Figure 5. Depletion of unconjugated antibody (aDNP) or Conjugate 4 from media of plated mouse hepatocyte assays.** aDNP demonstrated limited uptake into mouse hepatocytes while greater than 60% of Conjugate 4 was removed from the media over 2.5 hours. Uptake of Conjugate 4 into mouse hepatocytes was attenuated in FcRn knockout hepatocytes or by the addition of acetylated LDL (Stab/Scarb1) or ovalbumin (mannose/galactose receptors) to wild-type hepatocytes.

**Figure 6. IC<sub>50</sub> determination for the inhibition of Conjugate 4 association with mouse hepatocytes by acetylated LDL (Stab/Scarb1 receptor) or ovalbumin (mannose/galactose receptors).** Addition of acetylated LDL to wild \-type mouse hepatocytes resulted in complete inhibition of Conjugate 4 uptake (IC<sub>50</sub> = 0.024 mg/mL) while addition of ovalbumin reached a maximum inhibition of 64% of control uptake (IC<sub>50</sub> = 1.91 mg/mL).

**Figure 7. In vivo pharmacokinetics of aDNP and Conjugate 4 in wild-type, FcRn knockout, LDL-receptor knockout, mannose receptor knockout or ASGR1 knockout mouse models following 2 mg/kg intravenous administration.** Both aDNP and Conjugate 4 exhibited higher in vivo clearance values in FcRn knockout mice as opposed to wild-type controls. Conjugate 4 also exhibited higher in vivo clearance values in mannose receptor knockout mice. No difference was observed between wild-type and LDL-receptor or ASGR1 knockout mice.

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## Tables.

**Table 1. List of Peptide Conjugates.**

Conjugate	Ab Core	Linker*	Peptide	Peptide Charge	In Vivo Clearance (L/hr/kg)	AUCintact / AUCtotal	Percent Remaining in Hep Media	FcRn @ pH 7.4	FcRn @ pH 5.5	HIC Retention Time (min)
aDNP	aDNP D88C	N/A	N/A	N/A	0.0006	N/A	93	21.5	0.8	3.90
1	aDNP D88C	PEG11	AM-0422	6	0.0205	0.1	57.7			
2	aDNP E384C	bisPEG 23	AM-0422	6	0.079	ND	38			
3	aDNP D88CC	PEG11	AM-6122	2	0.00175	0.33	74	92.5	25.1	
4	aDNP D88C	GGs	AM-2752	2	0.0044	0.42	47.8	147	30.9	5.18
5	aDNP E384C	GGs	AM-2752	2	0.0032	0.76	88	47.7	239.2	5.69
6	aDNP D88C	GGs	AM-5454	1	0.0062	0.51	76			5.43
7	aDNP D88C	GGs	AM-5759	1	0.0024	0.27	74.9			5.45
8	aDNP D88C	GGs	AM-5458	1	0.0045	0.61	74.8			5.42
9	aDNP-3A4-F D88C	GGs	AM-6918	1	0.0037	0.36	72.6			5.61
10	aDNP-3A4-F D88C	GGs	AM-6262	1	0.0083	0.26	39	228.1	337	6.46
11	3B1HC-3A4LC D88C	GGs	AM-2752	2	0.0033	0.44	63.8	79.7	93.7	5.71
12	3B1HC-3A4LC D88C	GGs	AM-2752	2	0.0013	0.4	73.4	73.1	34.6	4.92
13	aDNP-3A4-F D88C	GGs	AM-8960	1	0.0013	0.24	75.6			5.43
14	aDNP-3A4-F D88C	PEG11	AM-8248	1	0.0118	0.28	37.6			6.65

\* PEG11: 11-unit polyethylene glycol; bisPEG23: bis-23-unit polyethylene glycol; GGs: glycine-glycine-serine.

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**Table 2. In Vivo Pharmacokinetic Parameters Following 2 mg/kg Intravenous Administration of Ab1 or Conjugate 4 to Wild-Type Mice and Receptor Knockout Mouse Models.**

Test Article	Mouse Strain	T1/2 (hr)	AUC <sub>0-168</sub> (μM *hr)	Cl (L/hr/kg)	V <sub>ss</sub> (L/kg)	AUC <sub>KO</sub> /AUC <sub>WT</sub>	CL <sub>KO</sub> /CL <sub>WT</sub>
aDNP	WT	191	11.4	0.0006	0.150	---	---
	FcRn <sub>KO</sub>	4.98	1.36	0.0092	0.063	0.12	15.3
	LDL <sub>KO</sub>	183	8.45	0.0007	0.199	0.74	1.16
	MR <sub>KO</sub>	240	12.3	0.0005	0.150	1.08	0.83
	ASGR1 <sub>KO</sub>	333	14.2	0.0009	0.142	1.24	1.50
Conjugate 4 (Total)	WT	164	2.79	0.0027	0.544	---	---
	FcRn <sub>KO</sub>	1.19	0.096	0.133	0.237	0.03	49.3
	LDL <sub>KO</sub>	118	4.67	0.0028	0.376	1.67	1.04
	MR <sub>KO</sub>	104	1.55	0.0059	0.759	0.56	2.18
	ASGR1 <sub>KO</sub>	197	3.57	0.0036	0.456	1.28	1.33
Conjugate 4 (Intact)	WT	26.3	0.943	0.0131	0.277	---	---
	FcRn <sub>KO</sub>	1.27	0.149	0.0892	0.164	0.16	6.81
	LDL <sub>KO</sub>	38.4	1.30	0.0103	0.286	1.38	0.79
	MR <sub>KO</sub>	24.7	1.12	0.0117	0.418	1.19	0.89
	ASGR1 <sub>KO</sub>	19.9	1.32	0.0092	0.174	1.39	0.70

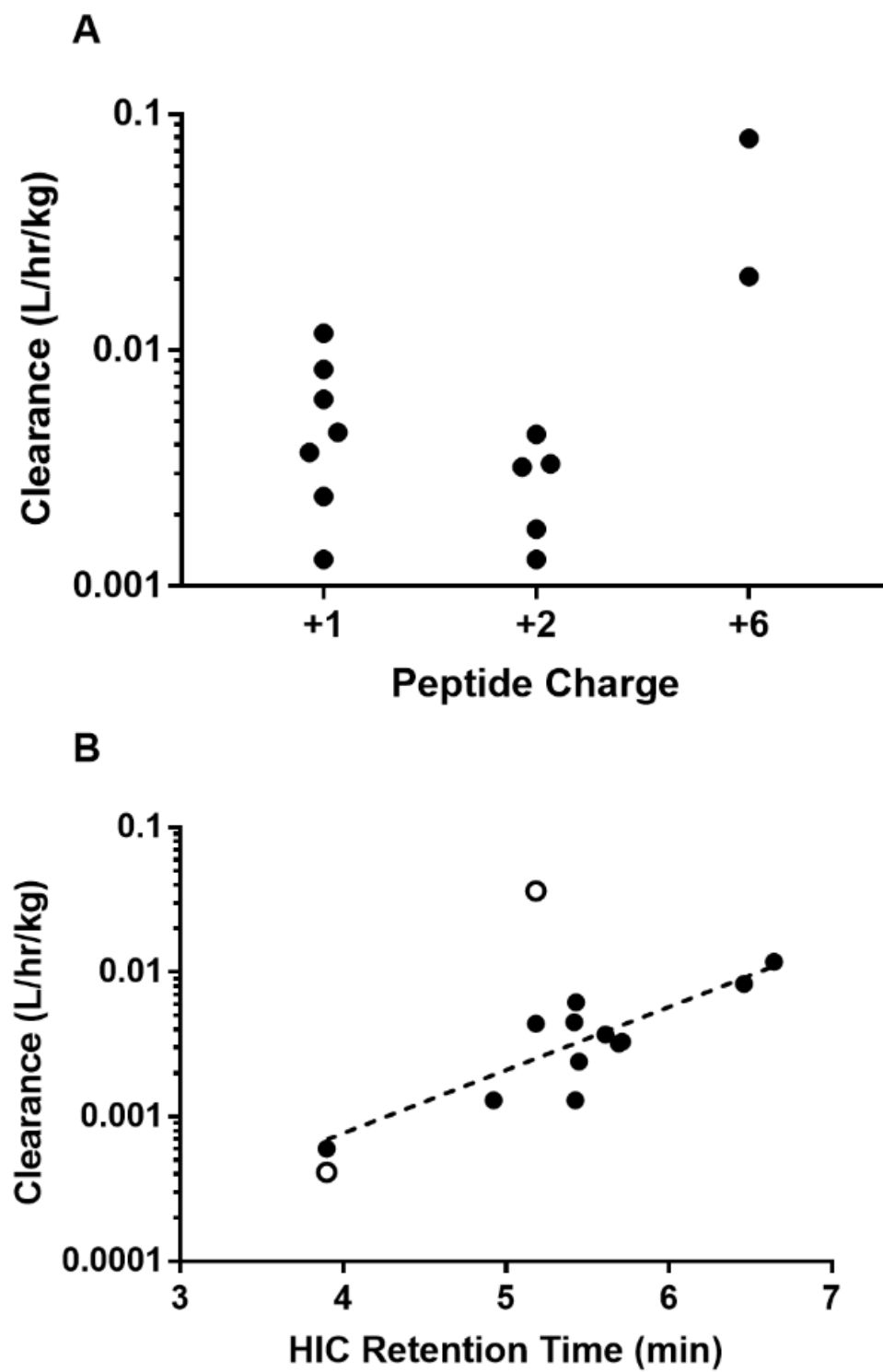
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**Table 3. Summary of Pearson Correlation Results For In Vivo Clearance, Percent Remaining in Hepatocyte Media, HIC Retention Time, and FcRn Binding at pH 7.4 or 5.5.**

<b>X-Axis Data</b>	<b>Y-Axis Data</b>	<b>Pearson r</b>	<b>N</b>	<b>P-value</b>	<b>Significant</b>
HIC RT	In Vivo Cl	0.8005	12	0.0018	Y
Percent Remaining in Media	In Vivo Cl	-0.5525	15	0.0327	Y
HIC RT	Percent Remaining in Media	-0.7185	15	0.0085	Y
FcRn (pH 7.4)	In Vivo Cl	0.9107	7	0.0044	Y
FcRn (pH 5.5)	In Vivo Cl	0.8088	7	0.0276	Y
FcRn (pH 7.4)	HIC RT	0.7144	6	0.1129	N
FcRn (pH 5.5)	HIC RT	0.8449	6	0.0342	Y
FcRn (pH 7.4)	Percent Remaining in Media	-0.9455	7	0.0013	Y
FcRn (pH 5.5)	Percent Remaining in Media	-0.3902	7	0.3868	N

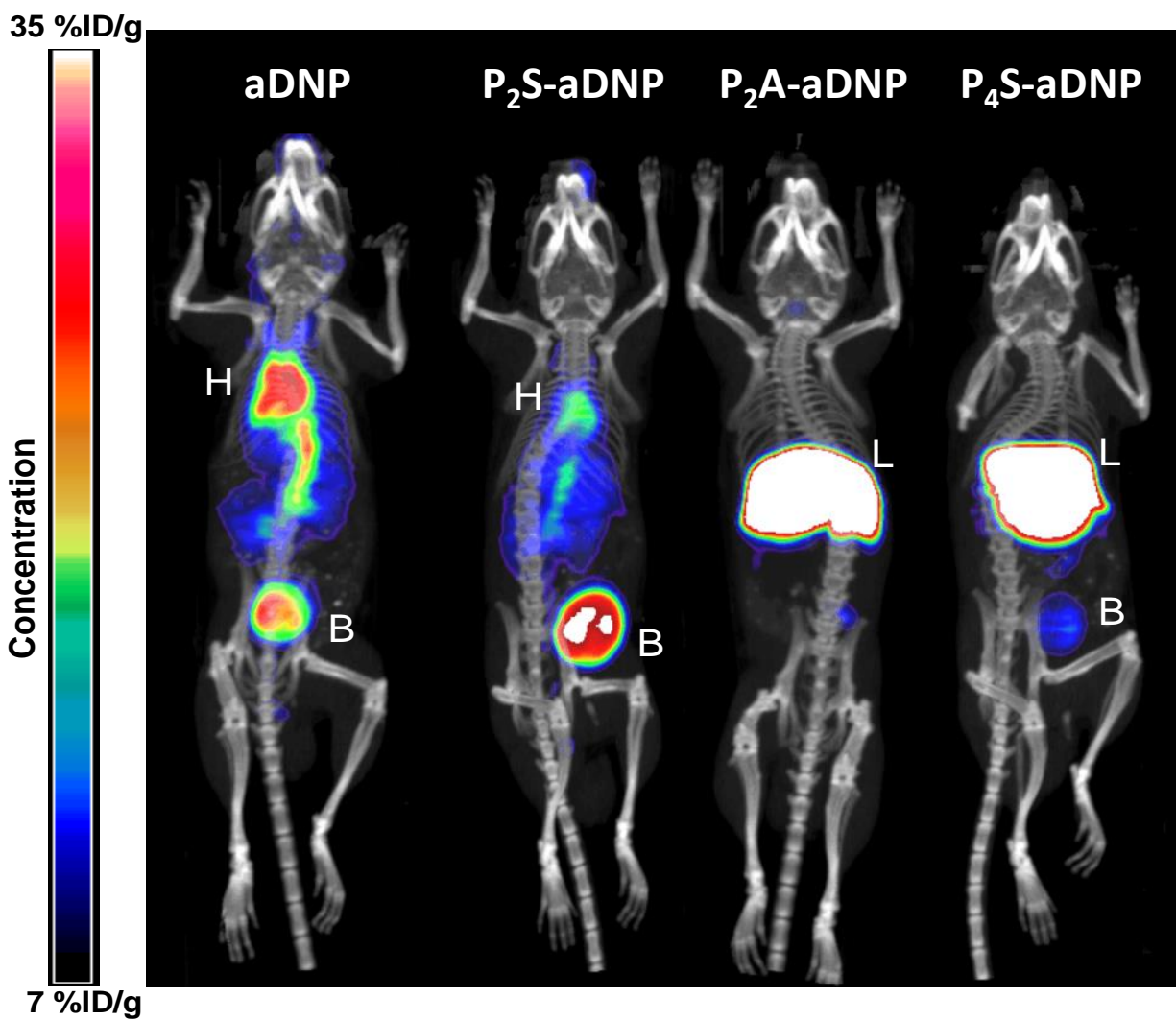
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**Figure 1.**



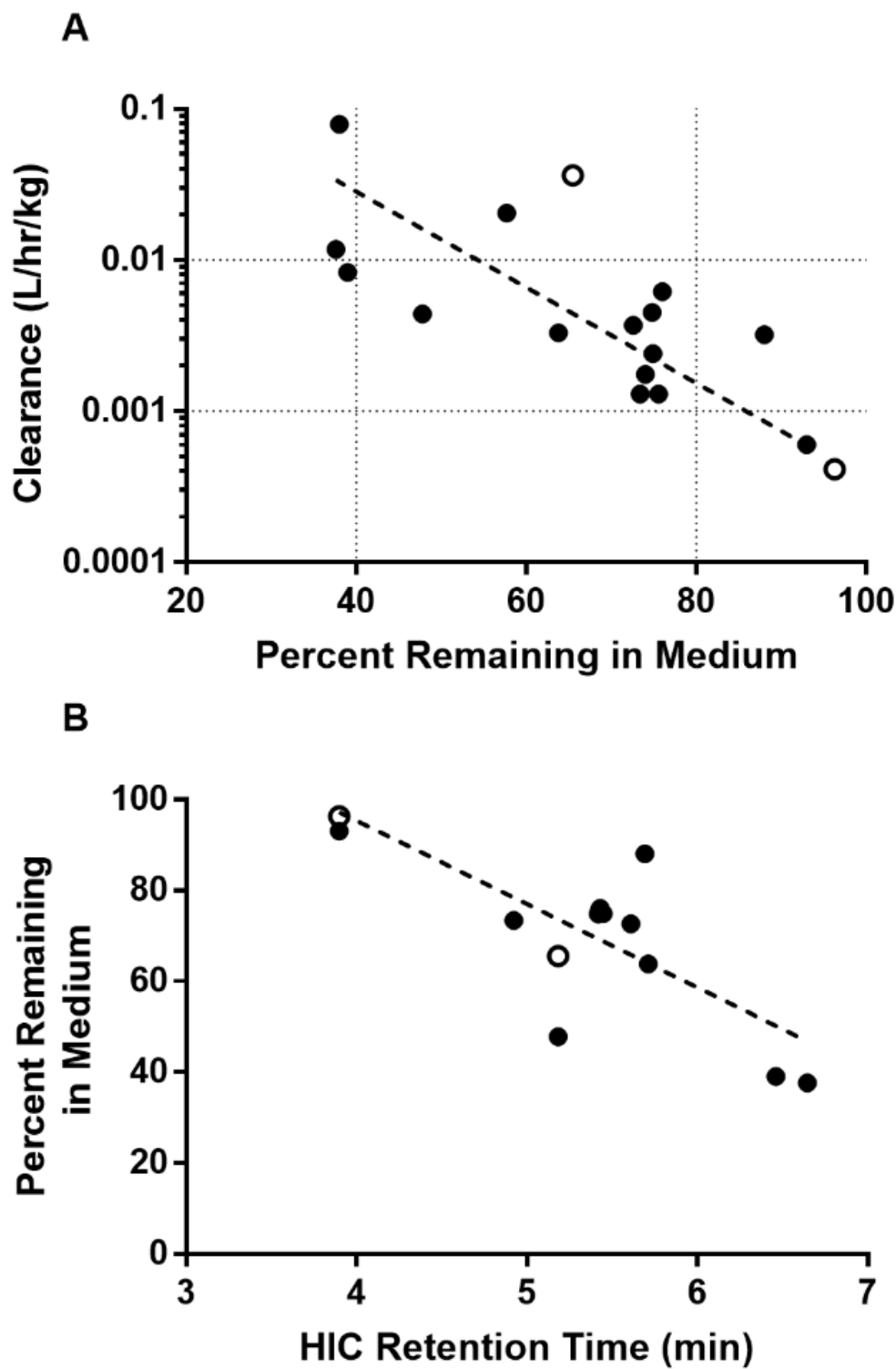
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**Figure 2.**



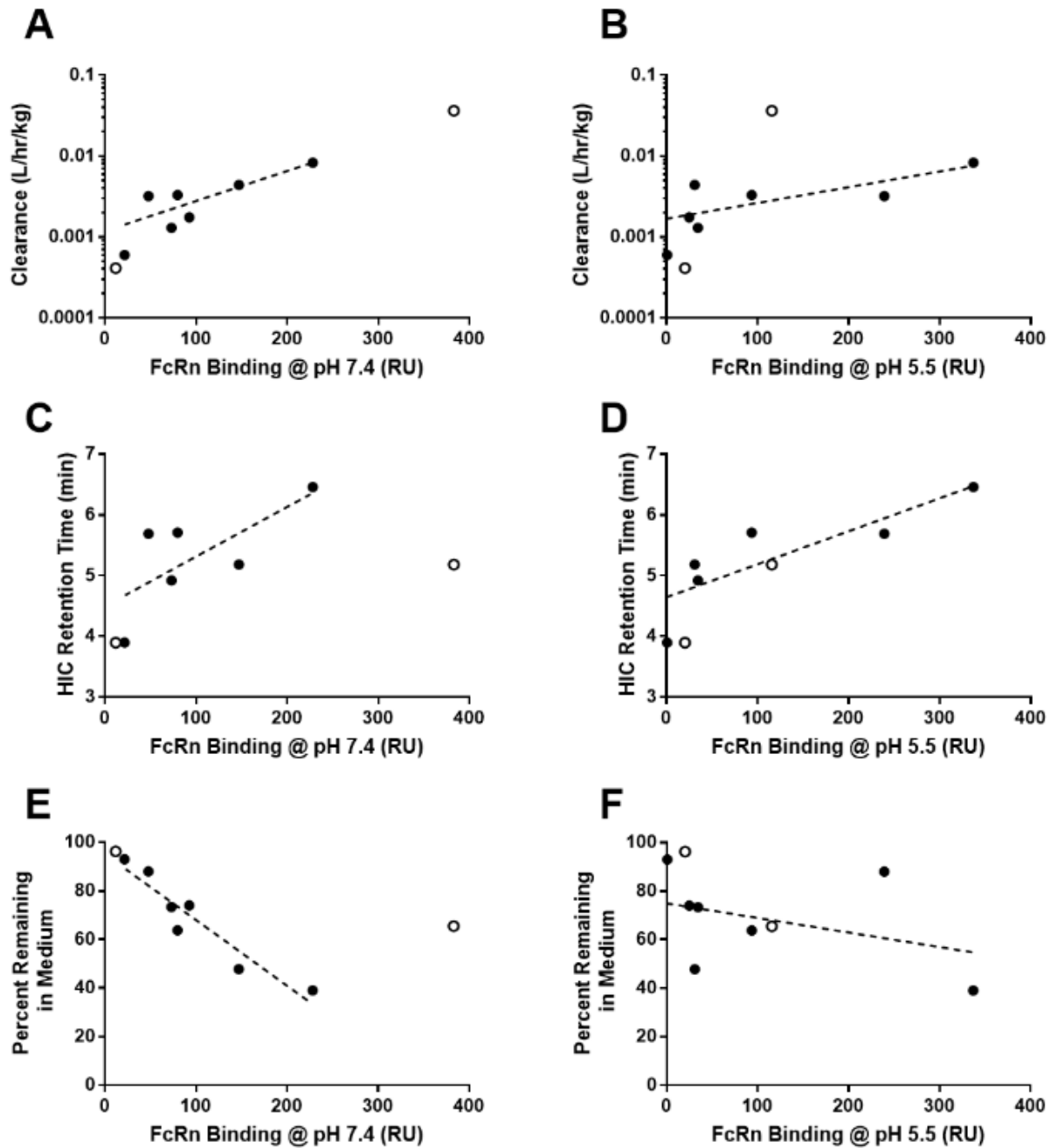
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**Figure 3.**



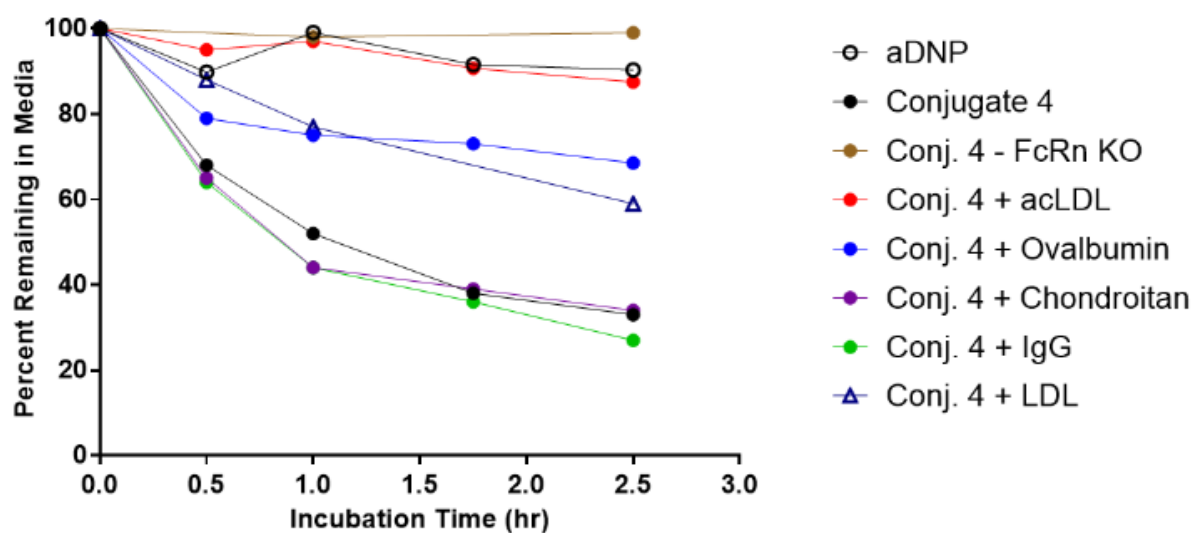
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**Figure 4.**



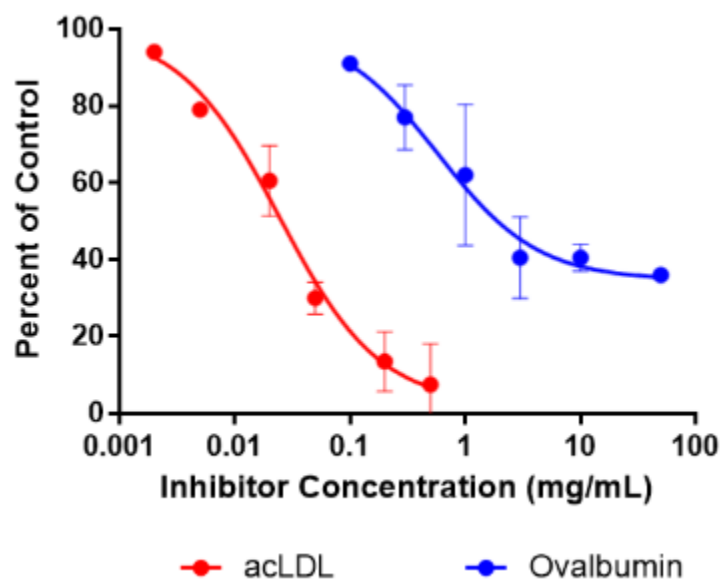
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**Figure 5.**



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**Figure 6.**



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**Figure 7.**

