

Characterization of interactions among CYP1A2, CYP2B4, and NADPH-cytochrome P450 reductase: Identification of specific protein complexes

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Supplemental Table 1 – Information on the GFP, Rluc, and WT P450 System Protein Constructs

Supplemental Figure 1 – Dynafit 4 script used to predict both 7-ethoxyresorufin-O-dealkylation (EROD) and 7-pentoxoresorufin-O-dealkylation (PROD) when CYP1A2 and CYP2B4 exist as functional monomers that simply compete for limiting concentrations of NADPH-cytochrome P450 reductase (CPR).

Supplemental Table 1 – Information on the GFP, Rluc, and WT P450 System Protein Constructs

Construct	Parent vector	5' restriction site	3' restriction site	Sequence identifier	Peptide linker
CYP1A2-GFP	pGFP ² -N1	EcoRI	BamHI	NM_001171121.1:29-1576	WIPPVAT
CYP1A2-Rluc	pRluc-N2	EcoRI	BamHI	NM_001171121.1:29-1576	WIPTGAT
Unlabeled CYP1A2*	CYP1A2-GFP	EcoRI		NM_001171121.1:29-1579	
CYP2B4-GFP	pGFP ² -N1	NheI	EcoRI	NM_001170859.1:1-1473	EFSRVCRISLRYRGGPIPPVAT
CYP2B4-Rluc	pRluc-N2	NheI	EcoRI	NM_001170859.1:1-1473	EFSRVCRISLRYRGGPIPTGAT
Unlabeled CYP2B4*	CYP2B4-GFP	NheI		NM_001170859.1:1-1476	
POR-GFP	pGFP ² -N3	EcoRI	HindIII	NM_001160290.1:40-2076	KLAVPRARDPPVAT
POR-Rluc	pRluc-N1	EcoRI	HindIII	NM_001160290.1:40-2076	KLAVPRARDPTGAT
Unlabeled POR*	POR-GFP	EcoRI		NM_001160290.1:40-2079	

Vector information. Vectors for the expression of labeled proteins were generated by ligating full-length cDNA into the empty BRET vectors shown above. The primers used to amplify the DNA of interest included the indicated restriction sites immediately flanking the coding sequence except for CYP1A2-GFP and CYP1A2-Rluc, for which extra thymidine residues were included between the CYP1A2 CDS and the BamHI site to ensure the tag was in frame. The 3' primers did not include a stop codon.

Each tagged construct had DNA between the 3' restriction site and the GFP or Rluc start codon that resulted in the expression of short peptide linker between the protein of interest and its tag.

To generate vectors for the expression of wild type, unlabeled protein, site directed mutagenesis was used to introduce a stop codon to the immediate 3' end of the GFP-tagged protein's CDS, destroying the 3' restriction site. For CYP1A2 TGG was changed to TGA, for CYP2B4 GAA was mutated to TAA, and for POR AAG was replaced with TAG.

Supplemental Figure 1 – Dynafit 4 script used to predict both 7-ethoxyresorufin-O-dealkylation (EROD) and 7-pentoxyresorufin-O-dealkylation (PROD) when CYP1A2 and CYP2B4 exist as functional monomers that simply compete for limiting concentrations of NADPH-cytochrome P450 reductase (CPR). The terms used are defined as: CYP1A2 “A”; CYP2B4 “B”; CPR, “R”; substrate “S”. K_{ar} (the dissociation constant for $K_D^{CPR \cdot CYP1A2}$) was 0.08 μ M [11;28], and K_{br} (the $K_D^{CPR \cdot CYP2B4}$) was 0.01 μ M [11;23]. K_{as} , and K_{ars} are the dissociation constants for substrate binding to CYP1A2, and CPR•CYP1A2, respectively. K_{bs} , and K_{brs} are the dissociation constants for substrate binding to CYP2B4, and CPR•CYP2B4, respectively. The rate constants for product formation by the CPR•CYP1A2•S and CPR•CYP2B4•S are “ k_{ars} and k_{brs} , respectively. The values for k_{ars} and k_{brs} for PROD were 0.38/min and 1.25/min, and were determined from the single-transfection systems. The values for k_{ars} and k_{brs} for EROD were 8.67/min and 0.015/min, and were determined from the single-transfection systems. This script is based on the following assumptions: (a) Both CYP1A2 and CYP2B4 behave as functional monomers that compete for limiting CPR. (b) The binding of substrate does not significantly affect the affinity of CPR•CYP1A2 or CPR•CYP2B4. (c) The binding of CPR to either P450 does not affect the affinity of substrate for either P450. (d) The substrate concentration is saturating.

; Simple competition between two P450s for Limiting CPR. (Dynafit 4)
 ; Predicted activities for CYP1A2 and CYP2B4 when competing for limiting CPR.

[task]

data = velocities
 task = fit
 approximation = rapid-equilibrium
 model = michaelis menten

[mechanism]

A + R <====> A.R : Kar dissociation
 A + S <====> A.S : Kas dissociation
 B + R <====> B.R : Kbr dissociation
 B + S <====> B.S : Kbs dissociation
 A.R + S <====> A.R.S : Kars dissociation
 A.S + R <====> A.R.S : Kar dissociation
 B.R + S <====> B.R.S : Kbrs dissociation
 B.S + R <====> B.R.S : Kbr dissociation
 A.R.S --> P + A.R : kars
 B.R.S --> P + B.R : kbrs

[constants]

Kar = 0.08
 Kas = 0.2
 Kars = 0.2
 Kbr = 0.01
 Kbs = 0.2
 Kbrs = 0.2
 kars = 8.67
 kbrs = 0.015

[concentrations]

S = 20

[responses]

P = 1.0

[data]

directory ./interaction

sheet P45012.txt

variable R

file P45012.txt | concentration A = 0.051, B = 0.0454

[output]

directory ./interaction/output/models

[end]